

# World Journal of *Stem Cells*

*World J Stem Cells* 2019 December 26; 11(12): 1020-1141



**REVIEW**

- 1020** Induced pluripotent stem cells for therapy personalization in pediatric patients: Focus on drug-induced adverse events  
*Genova E, Cavion F, Lucafò M, Leo LD, Pelin M, Stocco G, Decorti G*
- 1045** Influence of olive oil and its components on mesenchymal stem cell biology  
*Casado-Díaz A, Dorado G, Quesada-Gómez JM*
- 1065** Three-dimensional cell culture systems as an *in vitro* platform for cancer and stem cell modeling  
*Chaicharoenaudomrung N, Kunhorm P, Noisa P*
- 1084** Small molecules for mesenchymal stem cell fate determination  
*Cheng YH, Dong JC, Bian Q*
- 1104** Mechanoreponse of stem cells for vascular repair  
*Tian GE, Zhou JT, Liu XJ, Huang YC*

**ORIGINAL ARTICLE****Basic Study**

- 1115** Anti-osteoarthritis effect of a combination treatment with human adipose tissue-derived mesenchymal stem cells and thrombospondin 2 in rabbits  
*Shin K, Cha Y, Ban YH, Seo DW, Choi EK, Park D, Kang SK, Ra JC, Kim YB*
- 1130** MiR-301a promotes embryonic stem cell differentiation to cardiomyocytes  
*Zhen LX, Gu YY, Zhao Q, Zhu HF, Lv JH, Li SJ, Xu Z, Li L, Yu ZR*

**ABOUT COVER**

Editorial Board Member of *World Journal of Stem Cells*, Manabu Akahane, MD, PhD, Associate Professor, Public Health, Health Management and Policy, Nara Medical University School of Medicine, Nara 634-8521, Japan

**AIMS AND SCOPE**

The primary aim of *World Journal of Stem Cells (WJSC, World J Stem Cells)* is to provide scholars and readers from various fields of stem cells with a platform to publish high-quality basic and clinical research articles and communicate their research findings online.

*WJSC* publishes articles reporting research results obtained in the field of stem cell biology and regenerative medicine, related to the wide range of stem cells including embryonic stem cells, germline stem cells, tissue-specific stem cells, adult stem cells, mesenchymal stromal cells, induced pluripotent stem cells, embryoid bodies, embryonal carcinoma stem cells, hemangioblasts, hematopoietic stem cells, lymphoid progenitor cells, myeloid progenitor cells, *etc.*

**INDEXING/ABSTRACTING**

The *WJSC* is now indexed in PubMed, PubMed Central, Science Citation Index Expanded (also known as SciSearch®), Journal Citation Reports/Science Edition, Biological Abstracts, and BIOSIS Previews. The 2019 Edition of Journal Citation Reports cites the 2018 impact factor for *WJSC* as 3.534 (5-year impact factor: N/A), ranking *WJSC* as 16 among 26 journals in Cell and Tissue Engineering (quartile in category Q3), and 94 among 193 journals in Cell Biology (quartile in category Q2).

**RESPONSIBLE EDITORS FOR THIS ISSUE**

Responsible Electronic Editor: *Yan-Xia Xing*

Proofing Production Department Director: *Yun-Xiaojuan Wu*

**NAME OF JOURNAL**

*World Journal of Stem Cells*

**ISSN**

ISSN 1948-0210 (online)

**LAUNCH DATE**

December 31, 2009

**FREQUENCY**

Monthly

**EDITORS-IN-CHIEF**

Tong Cao, Shengwen Calvin Li, Carlo Ventura

**EDITORIAL BOARD MEMBERS**

<https://www.wjgnet.com/1948-0210/editorialboard.htm>

**EDITORIAL OFFICE**

Jin-Lei Wang, Director

**PUBLICATION DATE**

December 26, 2019

**COPYRIGHT**

© 2019 Baishideng Publishing Group Inc

**INSTRUCTIONS TO AUTHORS**

<https://www.wjgnet.com/bpg/gerinfo/204>

**GUIDELINES FOR ETHICS DOCUMENTS**

<https://www.wjgnet.com/bpg/GerInfo/287>

**GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH**

<https://www.wjgnet.com/bpg/gerinfo/240>

**PUBLICATION MISCONDUCT**

<https://www.wjgnet.com/bpg/gerinfo/208>

**ARTICLE PROCESSING CHARGE**

<https://www.wjgnet.com/bpg/gerinfo/242>

**STEPS FOR SUBMITTING MANUSCRIPTS**

<https://www.wjgnet.com/bpg/GerInfo/239>

**ONLINE SUBMISSION**

<https://www.f6publishing.com>

## Basic Study

**Anti-osteoarthritis effect of a combination treatment with human adipose tissue-derived mesenchymal stem cells and thrombospondin 2 in rabbits**

Kyungha Shin, Yeseul Cha, Young-Hwan Ban, Da Woom Seo, Ehn-Kyoung Choi, Dongsun Park, Sung Keun Kang, Jeong Chan Ra, Yun-Bae Kim

**ORCID number:** Kyungha Shin (0000-0001-7843-4798); Yeseul Cha (0000-0002-5263-9290); Young-Hwan Ban (0000-0003-2219-3299); Da Woom Seo (0000-0002-1541-8800); Ehn-Kyoung Choi (0000-0002-6112-6622); Dongsun Park (0000-0001-6042-5949); Sung Keun Kang (0000-0003-4387-6363); Jeong Chan Ra (0000-0002-6982-5348); Yun-Bae Kim (0000-0003-1340-041X).

**Author contributions:** Shin K, Cha Y, and Ban YH performed animal experiments and assembled histological samples; Seo DW performed the cell differentiation study; Choi EK and Park D collected and analyzed the data; Kang SK prepared and characterized the stem cells; Ra CJ and Kim YB designed and interpreted the study, and wrote the manuscript.

**Supported by** Basic Science Research Program through the National Research Foundation of Korea funded by the Ministry of Science and ICT to Y.B.K., No. 2017R1A2A2A05069417

**Institutional review board**

**statement:** All human tissues were obtained with approval of the Institutional Review Board of K-Stem Cell (Seoul, Korea).

**Institutional animal care and use**

**committee statement:** All protocols and procedures of animal experiments complied with the

**Kyungha Shin, Yeseul Cha, Young-Hwan Ban, Da Woom Seo, Ehn-Kyoung Choi, Yun-Bae Kim,** College of Veterinary Medicine, Chungbuk National University, Cheongju 28644, Chungbuk, South Korea

**Dongsun Park,** Department of Biology Education, Korea National University of Education, Cheongju 28173, Chungbuk, South Korea

**Sung Keun Kang, Jeong Chan Ra,** Biostar Stem Cell Research Institute, R-BIO Co., Ltd., Seoul 07238, South Korea

**Corresponding author:** Yun-Bae Kim, DVM, PhD, Professor, College of Veterinary Medicine, Chungbuk National University, 1 Chungdaero (Gaesin-dong), Cheongju 28644, Chungbuk, South Korea. [solar93@cbu.ac.kr](mailto:solar93@cbu.ac.kr)

**Telephone:** +82-43-2613358

**Abstract****BACKGROUND**

Osteoarthritis (OA), a chronic age-related disease characterized by the slowly progressive destruction of articular cartilage, is one of the leading causes of disability. As a new strategy for treatment of OA, mesenchymal stem cells (MSCs) have the potential for articular cartilage regeneration. Meanwhile, thrombospondin 2 (TSP2) promotes the chondrogenic differentiation of MSCs.

**AIM**

To investigate whether TSP2 induces chondrogenic differentiation of human adipose-derived MSCs (hADMSCs) and potentiates the therapeutic effects of hADMSCs in OA rabbits.

**METHODS**

We investigated the chondrogenic potential of TSP2 in hADMSCs by analyzing the expression of chondrogenic markers as well as NOTCH signaling genes in normal and TSP2 small interfering RNA (siRNA)-treated stem cells. Anterior cruciate ligament transection surgery was performed in male New Zealand white rabbits, and 8 wk later, hADMSCs ( $1.7 \times 10^6$  or  $1.7 \times 10^7$  cells) were injected into the injured knees alone or in combination with intra-articular injection of TSP2 (100 ng/knee) at 2-d intervals. OA progression was monitored by gross, radiological, and histological examinations.

Institutional Animal Care and Use Committee of Laboratory Animal Research Center at Chungbuk National University (Chungbuk, Korea).

**Conflict-of-interest statement:** The author has no conflict of interest to declare.

**Open-Access:** This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

**Manuscript source:** Invited manuscript

**Received:** February 27, 2019

**Peer-review started:** February 27, 2019

**First decision:** August 1, 2019

**Revised:** October 14, 2019

**Accepted:** November 4, 2019

**Article in press:** November 4, 2019

**Published online:** December 26, 2019

**P-Reviewer:** Oltra E, Ventura C

**S-Editor:** Zhang L

**L-Editor:** Filipodia

**E-Editor:** Wu YXJ



## RESULTS

In hADMSC culture, treatment with TSP2 increased the expression of chondrogenic markers (SOX9 and collagen II) as well as NOTCH signaling genes (JAGGED1 and NOTCH3), which were inhibited by TSP2 siRNA treatment. *In vivo*, OA rabbits treated with hADMSCs or TSP2 alone exhibited lower degree of cartilage degeneration, osteophyte formation, and extracellular matrix loss 8 wk after cell transplantation. Notably, such cartilage damage was further alleviated by the combination of hADMSCs and TSP2. In addition, synovial inflammatory cytokines, especially tumor-necrosis factor- $\alpha$ , markedly decreased following the combination treatment.

## CONCLUSION

The results indicate that TSP2 enhances chondrogenic differentiation of hADMSCs *via* JAGGED1/NOTCH3 signaling, and that combination therapy with hADMSCs and TSP2 exerts synergistic effects in the cartilage regeneration of OA joints.

**Key words:** Osteoarthritis; Anterior cruciate ligament transection; Human adipose tissue-derived mesenchymal stem cell; Thrombospondin 2; Notch signaling; Cartilage regeneration

©The Author(s) 2019. Published by Baishideng Publishing Group Inc. All rights reserved.

**Core tip:** We demonstrated the role of thrombospondin 2 (referred to as TSP2) in the chondrogenic differentiation of human adipose-derived mesenchymal stem cells *in vitro* and in osteoarthritis therapy with the cells *in vivo*. TSP2 induced chondrogenic differentiation *via* JAGGED1/NOTCH signaling, and potentiated the cartilage-restoring efficacy of human adipose-derived mesenchymal stem cells.

**Citation:** Shin K, Cha Y, Ban YH, Seo DW, Choi EK, Park D, Kang SK, Ra JC, Kim YB. Anti-osteoarthritis effect of a combination treatment with human adipose tissue-derived mesenchymal stem cells and thrombospondin 2 in rabbits. *World J Stem Cells* 2019; 11(12): 1115-1129

**URL:** <https://www.wjgnet.com/1948-0210/full/v11/i12/1115.htm>

**DOI:** <https://dx.doi.org/10.4252/wjsc.v11.i12.1115>

## INTRODUCTION

Osteoarthritis (OA) of the knee is the most common form of arthritis, which causes pain, stiffness, and decreased function. OA is characterized by the degeneration of articular cartilage, mainly due to change in the activity of chondrocytes in favor of catabolic activity as well as reduced cartilage cellularity<sup>[1,2]</sup>. The capacity of adult articular chondrocytes to regenerate the normal cartilage matrix architecture declines with aging, due to cell death and abnormal responsiveness to anabolic stimuli. OA chondrocytes lose their capacity to secrete the specific components of the extracellular matrix (ECM), such as type II collagen (collagen II) or aggrecan. Currently, no treatment capable of markedly altering the progression of OA exists and therapeutic options are essentially pain management and surgical intervention<sup>[3]</sup>. Indeed, new innovative therapeutic strategies for cartilage protection/repair are currently being evaluated mainly based on stem cell-mediated approaches.

Mesenchymal stem cells (MSCs) isolated from various tissues such as bone marrow, adipose tissue, and umbilical cord blood are capable of self-renewal and can differentiate into chondrogenic lineage cells *in vitro* and *in vivo*<sup>[4,5]</sup>. Besides this property, MSCs produce diverse secretory factors such as growth factors (GFs) and neurotrophic factors, cytokines, and chemokines that mediate paracrine functions including tissue protection/regeneration and immunomodulation<sup>[6,7]</sup>.

Compared with bone marrow-derived MSCs (BMMSCs), adipose tissue-derived MSCs (ADMSCs) are more genetically stable in a long-term culture, and display a lower senescence ratio and higher proliferative capacity<sup>[8]</sup>. Recently, some studies have shown that autologous ADMSCs improve the function and pain of knee joints without causing adverse events and reduced cartilage defects by chondrocyte

regeneration in animal models of OA and in humans<sup>[9-11]</sup>.

Thrombospondin 2 (TSP2) is well known as a multi-functional protein with anti-angiogenic property that interacts with the ECM and various binding partners<sup>[12,13]</sup>. Recent studies have indicated that TSP2 is also involved in cartilage biology. TSP2 is expressed in chondrocytes<sup>[14]</sup>, and its deficiency results in connective tissue abnormalities in mice<sup>[15]</sup>. In embryonic and adult mice, TSP2 is distributed in the areas of chondrogenesis. Moreover, it was previously demonstrated that TSP2 from human umbilical cord blood-mesenchymal stem cells (hUCBMSCs) promote chondrogenic differentiation of chondroprogenitor cells through paracrine action<sup>[16]</sup>.

Meanwhile, various studies have shown that NOTCH signaling plays a role in chondrogenic differentiation<sup>[17-21]</sup>, and that NOTCH receptors are expressed in articular cartilage and are involved in chondrogenesis<sup>[22,23]</sup>. Recently, several studies have characterized signaling pathways involved in chondrogenic differentiation of MSCs<sup>[24,25]</sup>. Indeed, chondroprogenitor cells and MSCs express NOTCH signaling components. Systemic inhibition of NOTCH leads to reduced cartilage formation *in vivo*<sup>[26]</sup>, and NOTCH inhibition by N-[N-(3,5-difluorophenacetyl)-L-alanyl]-S-phenylglycine t-butyl ester (DAPT) results in blockage of chondrogenesis *in vitro*<sup>[27,28]</sup>. Similarly, it has been observed that TSP2 enhances chondrogenic differentiation of chondroprogenitor cells *via* NOTCH signaling, which is inhibited by DAPT treatment<sup>[16]</sup>.

In the present study, the effect of TSP2 on chondrogenic differentiation of human ADMSCs (hADMSCs) was confirmed using TSP2 small interfering RNA (siRNA)-treated hADMSCs *in vitro*. Furthermore, based on the effectiveness of hADMSCs and TSP2 in cartilage regeneration, their synergistic effects on clinical improvement in a rabbit model of experimental OA were investigated.

## MATERIALS AND METHODS

### Preparation of hADMSCs

All human tissues were obtained with approval of the Institutional Review Board of K-Stem Cell (Seoul, Korea) (Protocol No. KS-2015-01-001). hADMSCs were prepared under Good Manufacturing Practice conditions (Biostar Stem Cell Research Institute of R-BIO Co., Ltd., Seoul, Korea). In brief, human abdominal subcutaneous fat tissues were obtained by simple liposuction from a 53-year-old female donor after obtaining informed consent<sup>[29]</sup>. The adipose tissues, after digestion with collagenase I, were suspended in Dulbecco's modified Eagle's medium (Invitrogen, Grand Island, NY, United States) containing 0.2 mM ascorbic acid and 10% fetal bovine serum (FBS). After overnight culture, the cell medium was changed to Keratinocyte-SFM (serum-free keratinocyte medium; Invitrogen) containing 0.2 mM ascorbic acid, 0.09 mM calcium, 5 ng/mL recombinant epidermal growth factor, and 5% FBS. When the cells reached 90% confluency, they were subculture-expanded in Keratinocyte-SFM medium until passage 3. FBS from cultured stem cells was completely removed by several washes with phosphate-buffered saline (PBS), which was verified by testing the albumin level below the measurement limit using a bovine albumin enzyme-linked immunosorbent assay (ELISA) quantification kit (Bethyl Laboratories, Montgomery, TX, United States).

### siRNA treatment

TSP2 siRNAs were used to knock down TSP2 expression in hADMSCs during chondrogenic differentiation. TSP2 siRNAs (Thermo Fisher Scientific, Lafayette, CO, United States) at a final concentration of 25 nM were transfected into hADMSCs using DharmaFECT reagent 1 (Thermo Fisher Scientific) according to the manufacturer's instructions. Transfected hADMSCs were maintained for 24 h in 10% FBS-containing medium.

### In vitro chondrogenic differentiation

Chondrogenesis was performed in micromass pellet cultures. hADMSCs ( $2 \times 10^5$  cells) were harvested, resuspended in STEMPRO<sup>®</sup> Chondrogenesis Differentiation Kit medium (Life Science, Carlsbad, CA, United States) in a 15 mL polypropylene tube, and centrifuged at 500 g for 5 min at room temperature. TSP2 siRNA-transfected hADMSCs were mixed with various concentrations (0, 10 or 100 ng/mL) of recombinant human TSP2 (R&D Systems, Minneapolis, MN, United States). The cell pellets were cultured in chondrogenic medium and maintained (37 °C, 5% CO<sub>2</sub>) for 14 d, in which the medium was replaced with fresh medium every 3 d.

### Real-time reverse transcription-PCR

Total RNA was isolated from hADMSCs culture pellets using TRIzol<sup>®</sup> reagent (Life

Technologies, Carlsbad, CA, United States). RNA (1 µg) was reverse-transcribed into complementary DNA using SuperScript® III First-Strand Synthesis System (Life Technologies). Glyceraldehyde-3-phosphate dehydrogenase was used as a reference gene. Real-time PCR was performed using Power SYBR® Green PCR Master Mix (Life Technologies). The primers used for real-time PCR are described in [Supplementary Table 1](#) (Genotech, Daejeon, Korea).

### **Immunohistochemistry**

Chondrogenic pellets of hADMSCs and TSP2 siRNA-treated hADMSCs were fixed for 24 h in 4% paraformaldehyde, frozen in OCT compound, and 5-µm cryosections were prepared. For immunohistochemistry, non-specific antibody-binding sites were blocked by incubating the slides in PBS with 5% bovine serum albumin (BSA) for 1 h. The slides were incubated overnight at 4 °C with primary antibody against SOX9 (1:200, rabbit monoclonal; EMD Millipore, Temecula, CA, United States), JAGGED1 (1:200, mouse monoclonal; Santa Cruz, Dallas, TX, United States), NOTCH3 (1:200, mouse monoclonal; Abnova, Taipei, Taiwan) or collagen II (1:200, mouse monoclonal; EMD Millipore) diluted in 0.1% Tris-buffered saline with Tween-20 with 1% BSA. Secondary antibodies conjugated with Alexa Fluor-488 or -594 (1:200, mouse polyclonal; Life Technologies) for 2 h at room temperature. All pellets were examined immediately after staining and photographed with a laser-scanning confocal microscope (LSM710; Zeiss, Oberkochen, Germany).

### **In vivo experimental design**

Adult male New Zealand white rabbits weighing  $3.2 \pm 0.4$  kg were obtained from Daehan-Biolink (Eumseong, Korea). The animals housed in a room with a constant temperature ( $22 \pm 2$  °C), relative humidity of  $55 \pm 10\%$ , and a 12-h light/dark cycle. The rabbits were fed standard chow and purified water *ad libitum*.

OA was induced surgically *via* anterior cruciate ligament transection (ACLT) of right hind knee, except sham control rabbits, under inhalation anesthesia with isoflurane (Sigma-Aldrich, St. Louis, MO, United States). For the ACLT procedures, a 4-cm skin and capsular incision was carried out and right ACLs were exposed through a medial para-patellar cut. To achieve optimal visualization of the ACLs, the patellar bone was displayed laterally and the knee was placed in full flexion. ACL removal was performed by cutting its attachment on the medial aspect of the lateral femoral condyle. The stifle was moved in a drawer test to ensure that the entire cruciate ligament had been excised. The incision was sutured in a routine fashion. After each operation, antibiotic (Foxolin®; Samjin Pharm, Seoul, Korea) and analgesic (Maritrol®; Jeil Pharm, Daegu, Korea) treatments were given immediately after the surgery and for 3 d thereafter. All surgical procedures were performed under general anesthesia and sterile conditions.

After ACLT surgery, the rabbits ( $n = 6$ /group) were subjected to a forced-exercise (walking) for 15 min every day 5 d a week for 8 wk to aggravate OA. The OA rabbits were randomly divided into five groups, and treated with hADMSCs ( $1.7 \times 10^6$  or  $1.7 \times 10^7$  cells/0.5 mL/knee) and/or TSP2 (100 ng/0.1 mL/knee). hADMSCs were transplanted once, and TSP2 was injected intra-articularly at 2-d intervals into the hind limb joints underwent ACLT for 8 wk, during which the joints were X-ray-photographed and synovial fluid was collected. Animals were sacrificed 8 wk after hADMSCs administration to investigate the effects of stem cells and TSP2 on the different compartments of the knee joint. Femoral condyles and tibial plateau were isolated for gross and microscopic examinations. All protocols and procedures of animal experiments complied with the Institutional Animal Care and Use Committee of Laboratory Animal Research Center at Chungbuk National University, Korea (Approval No. CBNUA-743-14-01).

### **Analysis of proinflammatory cytokines in synovial fluid**

At 1, 2, 4, and 8 wk after cell transplantation, synovial fluid was collected from ACLT knees of rabbits using sterile techniques. After centrifugation to remove cellular debris, the samples were analyzed for tumor-necrosis factor- $\alpha$  (TNF- $\alpha$ ), interleukin-1 $\beta$  (IL-1 $\beta$ ), and IL-6 by commercially available ELISA kits (TNF- $\alpha$ : E-EL-RB0011, Elabscience, St. Charles, MO, United States; IL-1 $\beta$ : E-EL-RB0013, Elabscience; IL-6: E-EL-RB0014, Elabscience), according to the manufacturer's instructions.

### **Radiological evaluation**

Knee joint radiographs were taken with a portable X-ray machine (Rextar EXO1414; Posdion, Seoul, Korea) before and 2, 4, and 8 wk after cell transplantation. For antero-posterior view, rabbits were placed in dorsal recumbency with legs extended caudally. Radiological OA was assessed by means of the Kellgren and Lawrence (KL) grading system<sup>[30]</sup>. This system is based on radiological features including the

presence of osteophyte (0-5), deformity of subchondral bone (0-5), and sharpening of tibial plateau (0-5).

### Gross examination

After sacrifice, macroscopic assessment of the knee joint surfaces of the rabbits was performed to score cartilage lesions. The scored was based on the protocol of the International Cartilage Research Society (ICRS) for cartilage repair<sup>[30,31]</sup> using criteria of roughness of surface (0-8), adhesion with meniscus (0-8), hypertrophy of joint (0-8), and osteophyte (0-8).

### Microscopic examination

After gross examination on the joint surfaces, distal femurs were excised with the knee joints intact. All samples were fixed in 10% neutral buffered formalin and decalcified in 10% EDTA-buffered saline (pH 8.0). Five- $\mu$ m paraffin-embedded sections were stained with hematoxylin and eosin (H&E) or Safranin O/Fast green to assess general morphology and proteoglycan contents, respectively. Separately, collagen II, accounting for about a half of cartilage ECM, was immuno-stained with a specific antibody. The evaluation was scored based on the criteria of the Osteoarthritis Research Society International (OARSI) that include intensity of ECM staining (0-6), structure (0-11), cellularity (0-4), and cluster formation (0-3)<sup>[32]</sup>.

### Data and statistical analysis

Data are presented as mean  $\pm$  standard error of the mean. Statistical significance between groups was determined by one-way analysis of variance followed by post-hoc Tukey's multiple-comparison tests using SAS program (version 6.12).  $P < 0.05$  was considered statistically significant.

## RESULTS

### TSP2-mediated chondrogenesis via NOTCH signaling

The correlation between TSP2 expression and chondrogenic potential of hADMSCs was analyzed by inhibiting the expression of TSP2 with siRNA. Treatment with TSP2 siRNA greatly decreased the expressions of JAGGED1 and NOTCH3 to 12% and 18% of the control levels, respectively (Figures 1-3). Furthermore, the expressions of SOX9 (a chondrogenic transcription factor) and collagen II (a cartilage-specific marker) also markedly decreased in TSP2 siRNA-transfected hADMSCs.

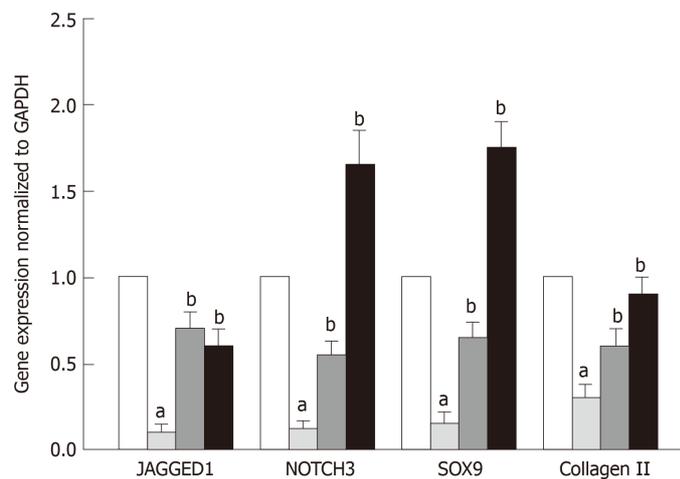
Next, the chondrogenic potential of TSP2 siRNA-transfected hADMSCs pellet cultures was analyzed in the presence of exogenous recombinant human TSP2 during differentiation. Treatment with 10 ng/mL TSP2 significantly recovered the expression of GAGGED1, NOTCH3, SOX9, and collagen II to 53%-71% compared to their control levels. A higher dose (100 ng/mL) of TSP2 fully recovered the expression of NOTCH3, SOX9, and collagen II. The change in mRNA expression was confirmed by immunohistochemical analysis of proteins, *i.e.*, the production of chondrogenic proteins blocked by siRNA was reconstituted by exogenous TSP2 treatment (Figures 2 and 3). From these results, it was suggested that chondrogenic differentiation of hADMSCs should be mediated by TSP2 through NOTCH signaling.

### Change in proinflammatory cytokines

In order to evaluate the inflammatory reaction caused by OA progression, the levels of pro-inflammatory cytokines such as TNF- $\alpha$ , IL-1 $\beta$ , and IL-6 in synovial fluid were quantified using ELISA. In ACLT knees, the cytokines markedly increased during 8-wk OA-induction and lasted for an additional 8 wk (Figure 4). Notably, the increases in TNF- $\alpha$  and IL-1 $\beta$  levels in ACLT rabbits were markedly attenuated as early as 1 wk after hADMSC treatment, reaching normal levels with a high dose ( $1.7 \times 10^7$ ) of hADMSCs. TSP2 also attenuated the increase in TNF- $\alpha$  level. Interestingly, IL-6 was fully suppressed in a short time (1 wk) by hADMSCs or TSP2 treatment. In the combination treatment, TSP2 further enhanced the effect of  $1.7 \times 10^6$  hADMSCs on TNF- $\alpha$ , but did not affect the effects on IL-1 $\beta$  and IL-6 levels. Taken together, these data indicate that TSP2 lower the levels of TNF- $\alpha$  and IL-6, and hADMSCs have inhibitory capacity on pro-inflammatory cytokines including TNF- $\alpha$ , IL-1 $\beta$ , and IL-6.

### Radiological findings

At 8 wk after surgery before treatment with hADMSCs and TSP2, all knees subjected to ACLT showed radiological signs of OA including osteophytes in medial femoral condyle, deformity of medial tibia, and sharpening of the tibial spine. Based on the modified KL grading system, the total lesion scores ranged from 4.8 to 6.5 (maximum score = 15), and further increased in ACLT rabbits without treatment up to a score of



**Figure 1 Expression of mRNA of NOTCH signaling and chondrogenic genes in pellet cultures.** White: Normal hADMSCs; Light grey: TSP2 siRNA-transfected hADMSCs; Dark gray: TSP2 siRNA-transfected hADMSCs + 10 ng/mL TSP2; Black: TSP2 siRNA-transfected hADMSCs + 100 ng/mL TSP2. <sup>a</sup>Significantly different from normal hADMSCs,  $P < 0.05$ . <sup>b</sup>Significantly different from TSP2 siRNA-transfected hADMSCs,  $P < 0.05$ . hADMSCs: Human adipose-derived mesenchymal stem cells; TSP2: Thrombospondin 2.

$9.3 \pm 0.6$  during the additional 8 wk (Figure 5). Representative images of articular osteophytes and deformities and each score of lesions 8 wk after cell treatment are shown in Figure 6 and Table 1, respectively. However, treatment with hADMSCs remarkably suppressed the progression of OA signs in a dose-dependent manner to a score of  $7.2 \pm 0.3$  at a high dose ( $1 \times 10^7$  cells). Notably, TSP2 alone significantly attenuated the OA lesions (score  $7.2 \pm 0.8$ ), and further enhanced the effects of hADMSCs following combination treatment (score  $6.0 \pm 0.4$ ).

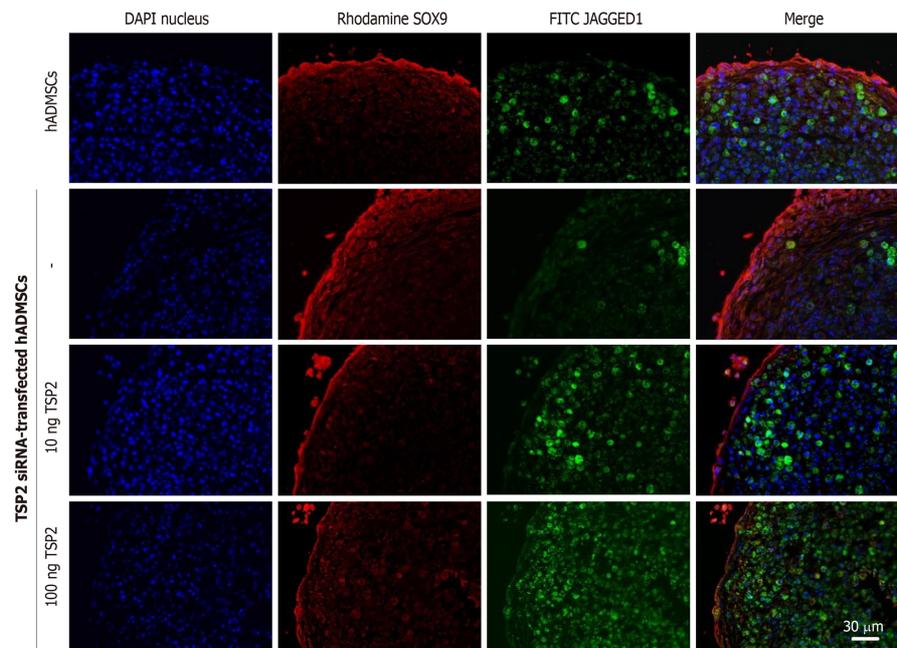
### Gross findings

At 8 wk post-treatment, exposed subchondral bone exhibited large fissures and erosions on lateral femoral condyle and tibial plateau in ACLT rabbits (Figure 7). In addition, severe deformities including osteophytes as well as adhesion of meniscus to cartilage surface were observed, leading to a total ICRS score of  $20.0 \pm 3.1$  (Table 2). In contrast, treatment with a high dose ( $1.7 \times 10^7$ ) of hADMSCs greatly attenuated the ACLT-induced osteoarthritic lesions to a mean score of  $8.5 \pm 2.5$ , although a low dose ( $1 \times 10^6$ ) of cells did not exert a significant effect. It is of interest to note that TSP2 also significantly protected against OA gross lesions (score,  $11.5 \pm 4.5$ ), and further improved when co-administered with hADMSCs (score,  $7.5 \pm 0.5$ ).

### Microscopic findings

Histological observations were quantified according to the modified OARSI scale. In sham control cartilage, H&E and Safranin O/Fast green staining as well as collagen II immunostaining demonstrated normal features, *i.e.*, the surfaces of articular cartilage were not fissured nor cracked, and the ECM and cartilage remained intact and dense from superficial to deep zones (Figure 8). In contrast, the articular cartilages in ACLT knees displayed severely rough surface, decreased chondrocytic density, clusters of chondrocytes, and loss of ECM such as proteoglycan (stained red with Safranin O) and collagen II (stained brown with collagen II antibody). The total modified OARSI score of ACLT knees without treatment reached  $23.0 \pm 5.0$  (maximum score = 30) (Table 3). In parallel with the gross lesions, treatment with a high dose ( $1.7 \times 10^7$ ) of hADMSCs drastically attenuated the ACLT-induced microscopic lesions to a score of  $8.5 \pm 3.5$ , in comparison with a minimal effect of a low dose ( $1 \times 10^6$ ) of the cells. Noteworthy, TSP2 also exhibited a potent activity (score  $10.5 \pm 2.5$ ), and synergistically enhanced the efficacy of hADMSCs (score  $7.5 \pm 0.6$ ). Indeed, following treatment with a high dose of hADMSCs or co-administration of a low dose of the cells and TSP2, the improved tissues were hyaline-like and showed thicker and smoother surface, in addition to intact integrity comparable to sham-operated cartilages.

## DISCUSSION



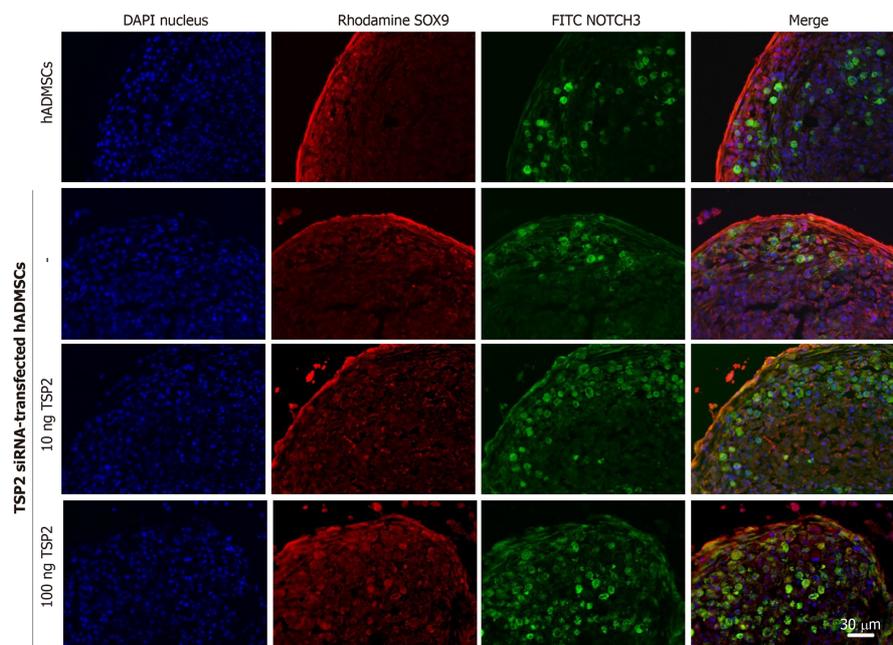
**Figure 2** Production of SOX9 (chondrogenic transcription factor) and JAGGED1 (NOTCH ligand) proteins in normal hADMSCs and TSP2 siRNA-transfected hADMSCs treated with TSP2 (10 or 100 ng/mL). hADMSCs: Human adipose-derived mesenchymal stem cells (200 ×); TSP2: Thrombospondin 2.

OA is one of the major chronic diseases leading to musculoskeletal morbidity and functional loss<sup>[33]</sup>. The etiology of OA is not completely understood. However, trauma, age, and genetic factors have been considered risk factors<sup>[34]</sup>. The ACLT model is widely validated in investigating OA disease, because it determines similar biomechanical and pathological changes to those seen in humans<sup>[35]</sup>. Previous studies have proven that 8-20 wk after ACLT in rabbits, the knee joints show diverse signs of OA with cartilaginous damage and osteophytes<sup>[36-38]</sup>. In the present study, however, we applied the ACLT to adult rabbits followed by daily 15-min forced exercise to induce mechanical inflammation in a relatively short-term period. At 8 wk after ACLT surgery, the rabbit knees displayed moderate osteoarthritic lesions according to the KL grading system in radiological analysis, compared to mild injury without exercise (data not shown).

Many cytokines have been implicated in the pathogenesis of OA. IL-1 $\beta$  inhibits the synthesis of ECM of hyaline cartilage and promotes resorption of the cartilage matrix by stimulating the synthesis of matrix metalloproteases (MMPs), which digest ECM components<sup>[39,40]</sup>. IL-1 $\beta$  also stimulates chondrocytes and synoviocytes to produce IL-6, which contributes to various manifestations of inflammatory arthropathies<sup>[41]</sup>. In addition, IL-1 $\beta$ -induced IL-6 secretion may be a required cofactor for the inhibition of proteoglycan synthesis by IL-1 $\beta$ <sup>[42]</sup>. TNF- $\alpha$  is a macrophage-derived cytokine that, like IL-1 $\beta$ , promotes the degradation of cartilage and suppresses the synthesis of the primary building blocks of cartilaginous matrix, proteoglycans<sup>[43]</sup>. TNF- $\alpha$  increases IL-1 $\beta$  and MMP synthesis by synovial cells<sup>[43]</sup>. TNF- $\alpha$ , therefore, promotes cartilage catabolism and decreases matrix synthesis directly or by inducing other cytokines.

In OA knees, all the cytokines markedly increased after ACLT surgery and 8-wk exercise, and were maintained at high levels for an additional 8 wk, which probably facilitated degradation of cartilaginous matrix as shown in the decreased safranin O-stained proteoglycans and collagen II immunoreactivity. Notably, the increases in TNF- $\alpha$ , IL-1 $\beta$ , and IL-6 levels in OA rabbits were drastically attenuated by hADMSCs to near normal levels as early as 1 wk after cell transplantation. Such an early suppression of inflammatory cytokines might be related to the effective protection against the progress of bone and cartilage lesions and deformities as observed by radiological observations. Although we injected the cells when the OA lesions were already formed (8 wk after ACLT surgery) to investigate their therapeutic potential rather than preventive-mode activity, it was demonstrated that earlier injection of MSCs was more beneficial in a donkey model<sup>[44]</sup>.

Interestingly, TSP2 also attenuated the increase in TNF- $\alpha$  level, in addition to full blocking of IL-6 secretion, indicating that TSP2 itself has OA-protective potential *via* anti-inflammatory activity. In the combination treatment, TSP2 further enhanced the effect of low dose ( $1.7 \times 10^6$ ) hADMSCs on TNF- $\alpha$  without affecting the effects on IL-



**Figure 3** Production of SOX9 (chondrogenic transcription factor) and NOTCH3 (NOTCH receptor) proteins in normal hADMSCs and TSP2 siRNA-transfected hADMSCs treated with TSP2 (10 or 100 ng/mL). hADMSCs: Human adipose-derived mesenchymal stem cells (200 ×); TSP2: Thrombospondin 2.

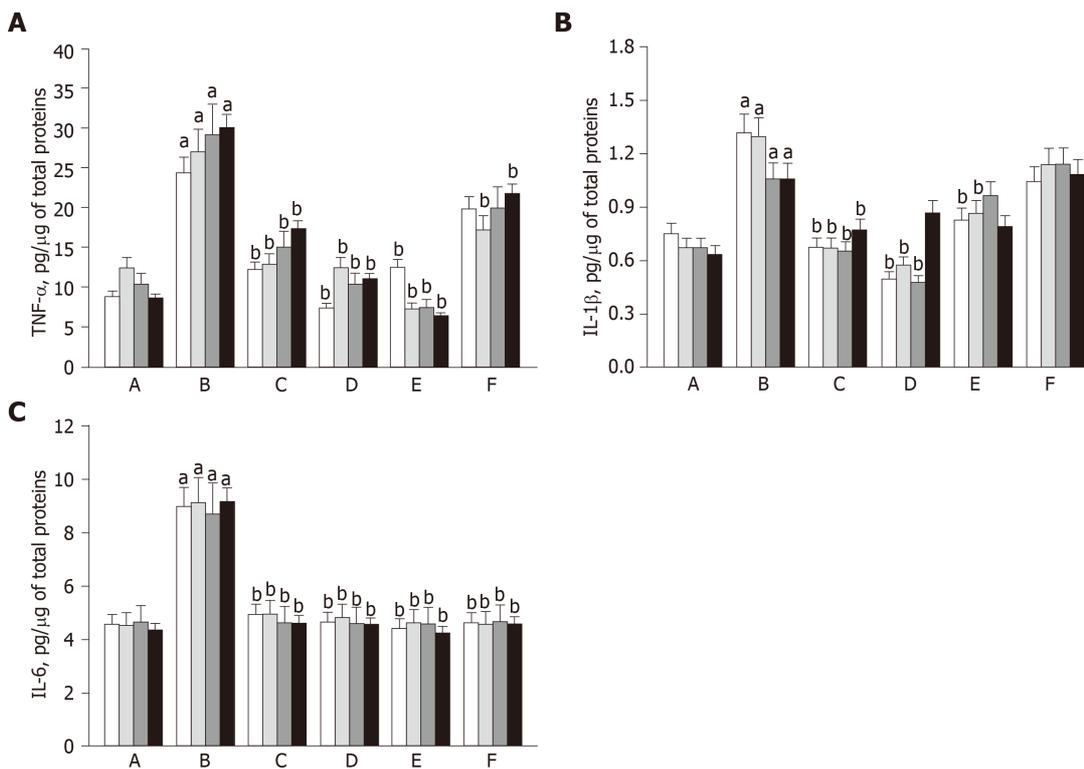
$1\beta$  and IL-6 levels. These data suggest that the combination therapy of hADMSCs and TSP2 may be effective in inhibiting the digestion of cartilage ECM by reducing inflammatory cytokines, especially TNF- $\alpha$ .

ADMSCs can be isolated from various organs that have a limited amount of adipose tissue, and are easy to be culture-expanded. Therefore, obtaining autologous ADMSCs, compared to those derived from bone marrow or other tissues, is believed to be less aggressive and quantitatively advantageous<sup>[45,46]</sup>. In the meantime, the number of ADMSC-based Investigational New Drug applications submitted to the US Food and Drug Administration as well as clinical trials registered worldwide significantly increased from 2011, indicating that adipose tissue is becoming an attractive cell source for MSC-based therapeutic application<sup>[47]</sup>.

It is believed that proliferation of remaining host synovial chondrocytes as well as chondrocytic differentiation of transplanted stem cells are key factors for the restoration of OA cartilages. It is well known that among these GFs, transforming growth factor- $\beta$ , insulin-like growth factor, and fibroblast growth factor are potent regulators of proliferation and differentiation of chondrocytes<sup>[44,45]</sup>. Notably, it was recently revealed that TSP2 contributes to the chondrogenic potential of hUCBMSCs for the regeneration of cartilage defects<sup>[48]</sup>. Moreover, TSP2 is released from hBMMSCs and hUCBMSCs themselves<sup>[16]</sup>, and potentiates NOTCH signaling for self-chondrogenic differentiation<sup>[18,20,49,50]</sup>. Therefore, it has been hypothesized that the chondrogenic potential of hADMSCs may be enhanced by TSP2 *via* NOTCH signaling.

Indeed, we showed that TSP2 regulated the chondrogenic differentiation of hADMSCs, which was mediated by the NOTCH signaling pathway. This hypothesis was elucidated in TSP2 siRNA-transfected cells, in which the expression of JAGGED1 and NOTCH3 was blocked, and eventually, the genes and proteins of SOX9 and collagen II were downregulated. However, the blocking of chondrogenic differentiation of hADMSCs by TSP2 siRNA was fully recovered by exogenous TSP2, confirming the involvement of TSP2 in MSC chondrogenic differentiation as previously reported<sup>[18,20,49,50]</sup>.

According to *in vitro* studies, it was expected that combination treatment with hADMSCs and TSP2 may lead to enhanced efficacy in the recovery of OA. hADMSCs alone improved the articular lesions in radiological, gross, and microscopic observations in a dose-dependent manner, *i.e.*,  $1.7 \times 10^7$  cells were enough for full recovery in 8 wk, but  $1.7 \times 10^6$  cells displayed a partial effect. More specifically, the high-dose ( $1.7 \times 10^7$ ) hADMSCs greatly restored the cartilage structure as well as the integrity of ECM including proteoglycan and collagen II. It is of interest to note that co-administration of TSP2 and hADMSCs exhibited synergistic effects, *i.e.* repeated injections of TSP2 (100 ng/knee at 2-d intervals) increased the efficacy of low-dose



**Figure 4** Concentration of pro-inflammatory cytokines in the synovial fluid 1 wk (white), 2 wk (light grey), 4 wk (dark grey), and 8 wk (black) after cell transplantation. A: Sham control; B: ACLT alone; C: Low-dose ( $1.7 \times 10^6$ ) hADMSCs; D: High-dose ( $1.7 \times 10^7$ ) hADMSCs; E: Low-dose ( $1.7 \times 10^6$ ) hADMSCs + TSP2 (100 ng); F: TSP2. <sup>a</sup>Significantly different from sham control,  $P < 0.05$ . <sup>b</sup>Significantly different from ACLT alone,  $P < 0.05$ . hADMSCs: Human adipose-derived mesenchymal stem cells; TSP2: Thrombospondin 2.

( $1.7 \times 10^6$ ) cells similar to or higher than that of 10-fold higher-dose ( $1.7 \times 10^7$ ) cells alone. In fact, however, TSP2 alone exerted beneficial effects to some extent, which may come from its paracrine action for the differentiation and proliferation of remaining host chondroprogenitor cells<sup>[16]</sup>, in addition to anti-inflammatory potential *via* cytokine regulation.

Actually, there are controversies on the long-term survival, attachment to the injured articular surface, and differentiation into chondrocytes of transplanted autologous or allogeneic human stem cells due to limited clinical studies on those issues. However, it has been demonstrated that autologous equine BMMSCs survive longer than 6 mo and integrates within the existing articular cartilage<sup>[44]</sup>. Therefore, it is anticipated that similar long-term survival and participation of human stem cells in the reparative process of human OA joints would be practical. In our recent study, heterologous human ADMSCs transplanted to rabbits' synovial cavity were also found to attach on the injured articular surface, survive some periods, and recover OA cartilages by secreting GFs, as inferred from the fact that direct injection of conditioned medium of hADMSCs exerted OA-improving activity to some extent (unpublished results). Despite the high activities of hADMSCs shown in this study, the potential, especially in proliferation and chondrogenic efficiency, of MSCs from individuals are somewhat different according to the age and health conditions. Thus, we selected healthy stem cells from a single individual to obtain consistent results.

Taken together, in this study, ACLT plus exercise were carried out to induce moderate to severe OA conditions to explore the efficacy of hADMSCs and TSP2. Our investigations demonstrated enhanced beneficial effects of the combination treatment of hADMSCs plus TSP2, showing well-organized tissues with low cytokine contents, smooth articular surfaces, and abundant ECM contents in diverse radiological, gross, and microscopic evaluations. As an underlying mechanism on the role of TSP2, we propose its potential to stimulate the chondrogenic differentiation of hADMSCs as well as anti-inflammatory activity.

In conclusion, TSP2 enhanced chondrogenic differentiation of hADMSCs *via* JAGGED1/NOTCH3 signaling, and the combination therapy with hADMSCs and TSP2 exerted synergistic efficacy in the cartilage regeneration of OA joints of rabbits. It is suggested that the beneficial effects of human ADMSCs plus TSP2 combination treatment could be obtained in human patients.

**Table 1 Radiological evaluation scores 8 wk after cell transplantation**

Treatment (dose)	Osteophytes in medial femoral condyle (0-5)	Deformity of medial tibia (0-5)	Sharpening of tibial spine (0-5)	Total score (0-15)
ACLT alone	3.5 ± 0.2	3.2 ± 0.4	2.7 ± 0.3	9.3 ± 0.6
hADMSCs (low)	3.0 ± 0.4	3.0 ± 0.4	2.7 ± 0.2	8.7 ± 0.4
hADMSCs (high)	2.7 ± 0.2	2.3 ± 0.2	2.2 ± 0.2	7.2 ± 0.3 <sup>1</sup>
hADMSCs (low) + TSP2	2.3 ± 0.3	1.5 ± 0.3	2.5 ± 0.3	6.0 ± 0.4 <sup>1</sup>
TSP2 (100 ng)	2.5 ± 0.3	2.5 ± 0.3	2.3 ± 0.3	7.2 ± 0.8 <sup>1</sup>

<sup>1</sup>Significantly different from ACLT alone,  $P < 0.05$ . ACLT: Anterior cruciate ligament transection; hADMSCs: Human adipose-derived mesenchymal stem cells; TSP2: Thrombospondin 2.

**Table 2 Gross evaluation scores**

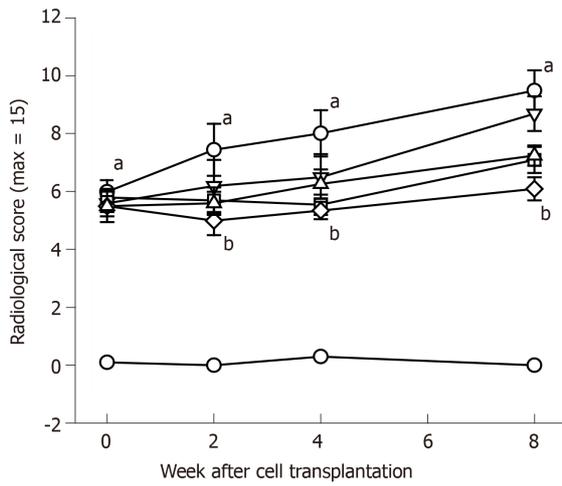
Treatment (dose)	Roughness of surface (0-8)	Adhesion with meniscus (0-8)	Hypertrophy of joint (0-8)	Osteophyte (0-8)	Total score (0-32)
ACLT alone	5.5 ± 0.5	4.5 ± 0.5	6.0 ± 0.4	4.0 ± 2.0	20.0 ± 3.1
hADMSCs (low)	4.0 ± 1.0	4.0 ± 1.0	5.0 ± 1.0	4.0 ± 1.2	17.0 ± 2.2
hADMSCs (high)	2.0 ± 1.1	2.5 ± 0.5	2.5 ± 1.5	1.5 ± 0.5	8.5 ± 2.5 <sup>1</sup>
hADMSCs (low) + TSP2	3.0 ± 0.2	1.5 ± 0.6	1.0 ± 0.4	2.0 ± 0.6	7.5 ± 0.5 <sup>1</sup>
TSP2 (100 ng)	1.5 ± 1.5	3.5 ± 0.5	3.5 ± 1.5	3.0 ± 1.1	11.5 ± 4.5 <sup>1</sup>

<sup>1</sup>Significantly different from ACLT alone,  $P < 0.05$ . ACLT: Anterior cruciate ligament transection; hADMSCs: Human adipose-derived mesenchymal stem cells; TSP2: Thrombospondin 2.

**Table 3 Microscopic evaluation scores**

Treatment (dose)	Structure (0-11)	Cellularity (0-4)	Cluster (0-3)	Proteoglycan (0-6)	Collagen II (0-6)	Total score (0-30)
ACLT alone	7.5 ± 1.5	3.0 ± 1.1	2.0 ± 1.0	6.0 ± 0.3	4.5 ± 1.5	23.0 ± 5.0
hADMSCs (low)	8.0 ± 0.5	1.0 ± 1.0	1.5 ± 1.5	4.5 ± 0.5	3.0 ± 0.4	18.0 ± 2.0
hADMSCs (high)	2.5 ± 1.5	2.0 ± 1.0	0.0 ± 0.0	2.5 ± 0.4	1.5 ± 0.5	8.5 ± 3.5 <sup>1</sup>
hADMSCs (low) + TSP2	1.0 ± 0.2	3.0 ± 0.5	0.0 ± 0.0	2.0 ± 0.3	1.5 ± 0.5	7.5 ± 0.6 <sup>1</sup>
TSP2 (100 ng)	1.5 ± 0.5	2.0 ± 0.2	1.5 ± 1.5	4.0 ± 0.7	1.5 ± 1.5	10.5 ± 2.5 <sup>1</sup>

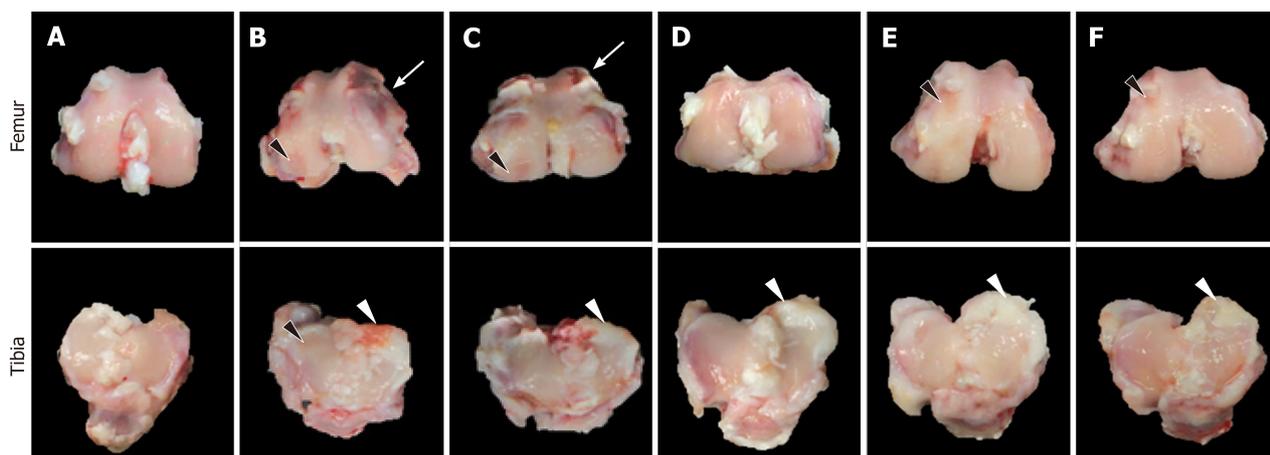
<sup>1</sup>Significantly different from ACLT alone,  $P < 0.05$ . ACLT: Anterior cruciate ligament transection; hADMSCs: Human adipose-derived mesenchymal stem cells; TSP2: Thrombospondin 2.



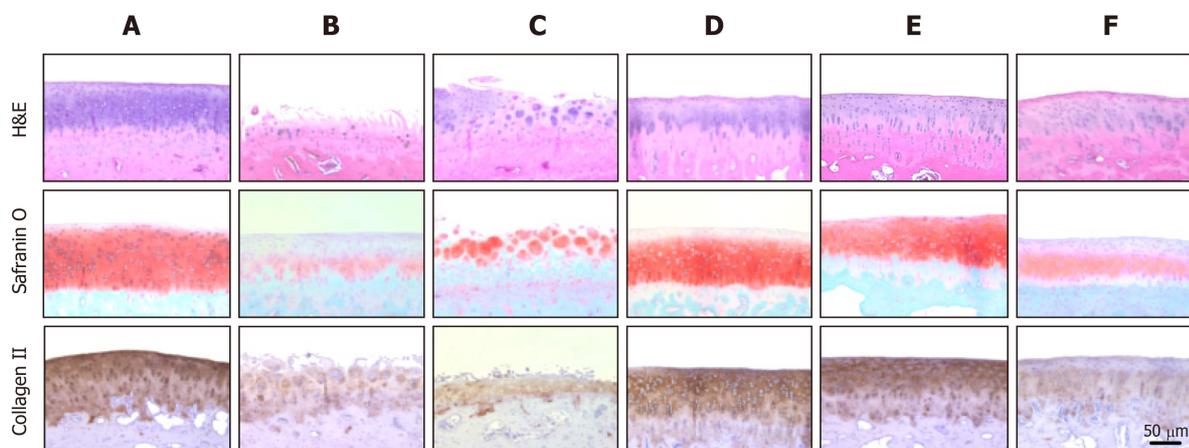
**Figure 5 Time-course radiological evaluation on knee joints.** Open circle: Sham control; Filled circle: Anterior cruciate ligament transection alone; Inverted triangle: Low-dose ( $1.7 \times 10^6$ ) hADMSCs; Square: High-dose ( $1.7 \times 10^7$ ) hADMSCs; Lozenge: Low-dose ( $1.7 \times 10^6$ ) hADMSCs + TSP2 (100 ng); Triangle: TSP2. <sup>a</sup>Significantly different from sham control,  $P < 0.05$ . <sup>b</sup>Significantly different from ACLT alone,  $P < 0.05$ . hADMSCs: Human adipose-derived mesenchymal stem cells; TSP2: Thrombospondin 2.



**Figure 6 Representative radiological images 8 wk after cell transplantation.** A: Sham control; B: Anterior cruciate ligament transection alone; C: Low-dose ( $1.7 \times 10^6$ ) hADMSCs; D: High-dose ( $1.7 \times 10^7$ ) hADMSCs; E: Low-dose ( $1.7 \times 10^6$ ) hADMSCs + TSP2 (100 ng); F: TSP2. Black arrowhead: Osteophytes in medial femoral condyle; White arrow: Deformity of medial tibia; White arrowhead: Sharpening of tibial spine. hADMSCs: Human adipose-derived mesenchymal stem cells; TSP2: Thrombospondin 2.



**Figure 7 Representative gross findings.** A: Sham control; B: Anterior cruciate ligament transection alone; C: Low-dose ( $1.7 \times 10^6$ ) hADMSCs; D: High-dose ( $1.7 \times 10^7$ ) hADMSCs; E: Low-dose ( $1.7 \times 10^6$ ) hADMSCs + TSP2 (100 ng); F: TSP2. Black arrowhead: Fissure and erosion; White arrowhead: Adhesion with meniscus; White arrow: Osteophyte. hADMSCs: Human adipose-derived mesenchymal stem cells; TSP2: Thrombospondin 2.



**Figure 8 Representative microscopic images stained with H&E (for structure, chondrocyte density, and cluster formation), safranin O (for proteoglycan contents) or collagen II antibody (for extracellular matrix).** A: Sham control; B: ACLT alone; C: Low-dose ( $1.7 \times 10^6$ ) hADMSCs; D: High-dose ( $1.7 \times 10^7$ ) hADMSCs; E: Low-dose ( $1.7 \times 10^6$ ) hADMSCs + TSP2 (100 ng); F: TSP2 (100×). hADMSCs: Human adipose-derived mesenchymal stem cells; TSP2: Thrombospondin 2.

## ARTICLE HIGHLIGHTS

### Research background

Mesenchymal stem cells (MSCs) reportedly differentiate into chondrocytes and have a potential for articular cartilage regeneration. Therefore, they may be a novel strategy for osteoarthritis (OA) treatment.

### Research motivation

Since MSCs can be differentiated into osteocytes, adipocytes, and other cells, in addition to chondrocytes, the efficiency for OA treatment is limited. We tried to increase the ratio of chondrocytic differentiation of human adipose-derived MSCs (hADMSCs) with thrombospondin 2 (TSP2), thereby enhancing OA therapeutic efficacy.

### Research objectives

The present study investigated whether TSP2 induces the chondrogenic differentiation of hADMSCs and potentiates the therapeutic effects of hADMSCs in OA rabbits.

### Research methods

*In vitro*, the chondrogenic potential of TSP2 in hADMSCs was investigated by analyzing the expression of chondrogenic markers as well as NOTCH signaling genes in normal and TSP2 siRNA-treated stem cells. *In vivo*, hADMSCs were injected into the injured knees of OA rabbits alone or in combination with TSP2, and OA progression was monitored *via* gross, radiological, and histological examinations.

### Research results

In hADMSC culture, TSP2 increased the expression of chondrogenic markers as well as NOTCH signaling genes, which were inhibited by TSP2 siRNA treatment. *In vivo*, combination treatment with hADMSCs and TSP2 not only attenuated cartilage degeneration, osteophyte formation, and extracellular matrix loss, but also decreased synovial inflammatory cytokines.

### Research conclusions

TSP2 enhances chondrogenic differentiation of hADMSCs *via* JAGGED1/NOTCH3 signaling, and combination therapy with hADMSCs and TSP2 exerts synergistic effects in the cartilage regeneration of OA joints.

### Research perspectives

We demonstrated the positive roles of TSP2 in the chondrogenic differentiation of hADMSCs and in OA therapy. TSP2 could be an adjunct therapeutic for enhancing the cartilage-restoring efficacy of hADMSCs.

## REFERENCES

- 1 **Bijlsma JW**, Berenbaum F, Lafeber FP. Osteoarthritis: an update with relevance for clinical practice. *Lancet* 2011; **377**: 2115-2126 [PMID: 21684382 DOI: 10.1016/S0140-6736(11)60243-2]
- 2 **Loeser RF**, Goldring SR, Scanzello CR, Goldring MB. Osteoarthritis: a disease of the joint as an organ. *Arthritis Rheum* 2012; **64**: 1697-1707 [PMID: 22392533 DOI: 10.1002/art.34453]
- 3 **Hunter DJ**. Pharmacologic therapy for osteoarthritis--the era of disease modification. *Nat Rev Rheumatol* 2011; **7**: 13-22 [PMID: 21079644 DOI: 10.1038/nrrheum.2010.178]
- 4 **Caplan AL**. The mesengenic process. *Clin Plast Surg* 1994; **21**: 429-435 [PMID: 7924141]
- 5 **Pittenger MF**, Mackay AM, Beck SC, Jaiswal RK, Douglas R, Mosca JD, Moorman MA, Simonetti DW, Craig S, Marshak DR. Multilineage potential of adult human mesenchymal stem cells. *Science* 1999; **284**: 143-147 [PMID: 10102814 DOI: 10.1126/science.284.5411.143]
- 6 **Meirelles Lda S**, Fontes AM, Covas DT, Caplan AL. Mechanisms involved in the therapeutic properties of mesenchymal stem cells. *Cytokine Growth Factor Rev* 2009; **20**: 419-427 [PMID: 19926330 DOI: 10.1016/j.cytogfr.2009.10.002]
- 7 **Salgado AJ**, Reis RL, Sousa NJ, Gimble JM. Adipose tissue derived stem cells secretome: soluble factors and their roles in regenerative medicine. *Curr Stem Cell Res Ther* 2010; **5**: 103-110 [PMID: 19941460 DOI: 10.2174/157488810791268564]
- 8 **Strioga M**, Viswanathan S, Darinskas A, Slaby O, Michalek J. Same or not the same? Comparison of adipose tissue-derived versus bone marrow-derived mesenchymal stem and stromal cells. *Stem Cells Dev* 2012; **21**: 2724-2752 [PMID: 22468918 DOI: 10.1089/scd.2011.0722]
- 9 **Black LL**, Gaynor J, Adams C, Dhupa S, Sams AE, Taylor R, Harman S, Gingerich DA, Harman R. Effect of intraarticular injection of autologous adipose-derived mesenchymal stem and regenerative cells on clinical signs of chronic osteoarthritis of the elbow joint in dogs. *Vet Ther* 2008; **9**: 192-200 [PMID: 19003780]
- 10 **Guercio A**, Di Marco P, Casella S, Cannella V, Russotto L, Purpari G, Di Bella S, Piccione G. Production of canine mesenchymal stem cells from adipose tissue and their application in dogs with chronic osteoarthritis of the humeroradial joints. *Cell Biol Int* 2012; **36**: 189-194 [PMID: 21936851 DOI: 10.1042/CBI20110304]
- 11 **Jo CH**, Lee YG, Shin WH, Kim H, Chai JW, Jeong EC, Kim JE, Shim H, Shin JS, Shin IS, Ra JC, Oh S, Yoon KS. Intra-articular injection of mesenchymal stem cells for the treatment of osteoarthritis of the knee: a proof-of-concept clinical trial. *Stem Cells* 2014; **32**: 1254-1266 [PMID: 24449146 DOI: 10.1002/stem.1634]
- 12 **Streit M**, Riccardi L, Velasco P, Brown LF, Hawighorst T, Bornstein P, Detmar M. Thrombospondin-2: a potent endogenous inhibitor of tumor growth and angiogenesis. *Proc Natl Acad Sci U S A* 1999; **96**: 14888-14893 [PMID: 10611308 DOI: 10.1073/pnas.96.26.14888]
- 13 **Bornstein P**, Armstrong LC, Hankenson KD, Kyriakides TR, Yang Z. Thrombospondin 2, a matricellular protein with diverse functions. *Matrix Biol* 2000; **19**: 557-568 [PMID: 11102746 DOI: 10.1016/S0945-053X(00)00104-9]
- 14 **Carron JA**, Bowler WB, Wagstaff SC, Gallagher JA. Expression of members of the thrombospondin family by human skeletal tissues and cultured cells. *Biochem Biophys Res Commun* 1999; **263**: 389-391 [PMID: 10491303 DOI: 10.1006/bbrc.1999.1380]
- 15 **Kyriakides TR**, Zhu YH, Smith LT, Bain SD, Yang Z, Lin MT, Danielson KG, Iozzo RV, LaMarca M, McKinney CE, Ginns EI, Bornstein P. Mice that lack thrombospondin 2 display connective tissue abnormalities that are associated with disordered collagen fibrillogenesis, an increased vascular density, and a bleeding diathesis. *J Cell Biol* 1998; **140**: 419-430 [PMID: 9442117 DOI: 10.1083/jcb.140.2.419]
- 16 **Jeong SY**, Kim DH, Ha J, Jin HJ, Kwon SJ, Chang JW, Choi SJ, Oh W, Yang YS, Kim G, Kim JS, Yoon JR, Cho DH, Jeon HB. Thrombospondin-2 secreted by human umbilical cord blood-derived mesenchymal stem cells promotes chondrogenic differentiation. *Stem Cells* 2013; **31**: 2136-2148 [PMID: 23843355 DOI: 10.1002/stem.1471]
- 17 **Watanabe N**, Tezuka Y, Matsuno K, Miyatani S, Morimura N, Yasuda M, Fujimaki R, Kuroda K, Hiraki Y, Hozumi N, Tezuka K. Suppression of differentiation and proliferation of early chondrogenic cells by Notch. *J Bone Miner Metab* 2003; **21**: 344-352 [PMID: 14586790 DOI: 10.1007/s00774-003-0428-4]
- 18 **Oldershaw RA**, Tew SR, Russell AM, Meade K, Hawkins R, McKay TR, Brennan KR, Hardingham TE. Notch signaling through Jagged-1 is necessary to initiate chondrogenesis in human bone marrow stromal cells but must be switched off to complete chondrogenesis. *Stem Cells* 2008; **26**: 666-674 [PMID: 18192230 DOI: 10.1634/stemcells.2007-0806]
- 19 **Mead TJ**, Yutzey KE. Notch pathway regulation of chondrocyte differentiation and proliferation during appendicular and axial skeleton development. *Proc Natl Acad Sci U S A* 2009; **106**: 14420-14425 [PMID: 19590010 DOI: 10.1073/pnas.0902306106]
- 20 **Oldershaw RA**, Hardingham TE. Notch signaling during chondrogenesis of human bone marrow stem

- cells. *Bone* 2010; **46**: 286-293 [PMID: 19406255 DOI: 10.1016/j.bone.2009.04.242]
- 21 **Haller R**, Schwanbeck R, Martini S, Bernoth K, Kramer J, Just U, Rohwedel J. Notch1 signaling regulates chondrogenic lineage determination through Sox9 activation. *Cell Death Differ* 2012; **19**: 461-469 [PMID: 21869831 DOI: 10.1038/cdd.2011.114]
- 22 **Hayes AJ**, Douthwaite GP, Webster SV, Archer CW. The distribution of Notch receptors and their ligands during articular cartilage development. *J Anat* 2003; **202**: 495-502 [PMID: 12846471 DOI: 10.1046/j.1469-7580.2003.00185.x]
- 23 **Douthwaite GP**, Bishop JC, Redman SN, Khan IM, Rooney P, Evans DJ, Houghton L, Bayram Z, Boyer S, Thomson B, Wolfe MS, Archer CW. The surface of articular cartilage contains a progenitor cell population. *J Cell Sci* 2004; **117**: 889-897 [PMID: 14762107 DOI: 10.1242/jcs.00912]
- 24 **Kolf CM**, Cho E, Tuan RS. Mesenchymal stromal cells. Biology of adult mesenchymal stem cells: regulation of niche, self-renewal and differentiation. *Arthritis Res Ther* 2007; **9**: 204 [PMID: 17316462 DOI: 10.1186/ar2116]
- 25 **Augello A**, De Bari C. The regulation of differentiation in mesenchymal stem cells. *Hum Gene Ther* 2010; **21**: 1226-1238 [PMID: 20804388 DOI: 10.1089/hum.2010.173]
- 26 **Dishowitz MI**, Mutyaba PL, Takacs JD, Barr AM, Engiles JB, Ahn J, Hankenson KD. Systemic inhibition of canonical Notch signaling results in sustained callus inflammation and alters multiple phases of fracture healing. *PLoS One* 2013; **8**: e68726 [PMID: 23844237 DOI: 10.1371/journal.pone.0068726]
- 27 **Karlsson C**, Jonsson M, Asp J, Brantsing C, Kageyama R, Lindahl A. Notch and HES5 are regulated during human cartilage differentiation. *Cell Tissue Res* 2007; **327**: 539-551 [PMID: 17093926 DOI: 10.1007/s00441-006-0307-0]
- 28 **Vujovic S**, Henderson SR, Flanagan AM, Clements MO. Inhibition of gamma-secretases alters both proliferation and differentiation of mesenchymal stem cells. *Cell Prolif* 2007; **40**: 185-195 [PMID: 17472726 DOI: 10.1111/j.1365-2184.2007.00426.x]
- 29 **Ra JC**, Shin IS, Kim SH, Kang SK, Kang BC, Lee HY, Kim YJ, Jo JY, Yoon EJ, Choi HJ, Kwon E. Safety of intravenous infusion of human adipose tissue-derived mesenchymal stem cells in animals and humans. *Stem Cells Dev* 2011; **20**: 1297-1308 [PMID: 21303266 DOI: 10.1089/scd.2010.0466]
- 30 **Kellgren JH**, Lawrence JS. Radiological assessment of rheumatoid arthritis. *Ann Rheum Dis* 1957; **16**: 485-493 [PMID: 13498603 DOI: 10.1136/ard.16.4.485]
- 31 **Pacifici M**. Introduction to the mini-review series "Articular cartilage: biology, pathology and repair". *Matrix Biol* 2014; **39**: 1 [PMID: 25172827 DOI: 10.1016/j.matbio.2014.08.004]
- 32 **Lavery S**, Girard CA, Williams JM, Hunziker EB, Pritzker KP. The OARSI histopathology initiative - recommendations for histological assessments of osteoarthritis in the rabbit. *Osteoarthritis Cartilage* 2010; **18** Suppl 3: S53-S65 [PMID: 20864023 DOI: 10.1016/j.joca.2010.05.029]
- 33 **Burnett BP**, Levy R, Cole BJ. Metabolic mechanisms in the pathogenesis of osteoarthritis. A review. *J Knee Surg* 2006; **19**: 191-197 [PMID: 16893158 DOI: 10.1055/s-0030-1248105]
- 34 **Chen FH**, Tuan RS. Mesenchymal stem cells in arthritic diseases. *Arthritis Res Ther* 2008; **10**: 223 [PMID: 18947375 DOI: 10.1186/ar2514]
- 35 **Vignon E**, Bejui J, Mathieu P, Hartmann JD, Ville G, Evreux JC, Descotes J. Histological cartilage changes in a rabbit model of osteoarthritis. *J Rheumatol* 1987; **14** Spec No: 104-106 [PMID: 3625662]
- 36 **Hefti FL**, Kress A, Fasel J, Morscher EW. Healing of the transected anterior cruciate ligament in the rabbit. *J Bone Joint Surg Am* 1991; **73**: 373-383 [PMID: 2002075 DOI: 10.2106/00004623-199173030-00008]
- 37 **Jean YH**, Wen ZH, Chang YC, Lee HS, Hsieh SP, Wu CT, Yeh CC, Wong CS. Hyaluronic acid attenuates osteoarthritis development in the anterior cruciate ligament-transected knee: Association with excitatory amino acid release in the joint dialysate. *J Orthop Res* 2006; **24**: 1052-1061 [PMID: 16583446 DOI: 10.1002/jor.20123]
- 38 **Grigolo B**, Lisignoli G, Desando G, Cavallo C, Marconi E, Tschon M, Giavaresi G, Fini M, Giardino R, Facchini A. Osteoarthritis treated with mesenchymal stem cells on hyaluronan-based scaffold in rabbit. *Tissue Eng Part C Methods* 2009; **15**: 647-658 [PMID: 19249964 DOI: 10.1089/ten.TEC.2008.0569]
- 39 **Evans CH**. The role of proteinases in cartilage destruction. *Agents Actions Suppl* 1991; **32**: 135-152 [PMID: 2069082 DOI: 10.1007/978-3-0348-7405-2\_19]
- 40 **Martel-Pelletier J**, Zafarullah M, Kodama S, Pelletier JP. In vitro effects of interleukin 1 on the synthesis of metalloproteinases, TIMP, plasminogen activators and inhibitors in human articular cartilage. *J Rheumatol Suppl* 1991; **27**: 80-84 [PMID: 1851231]
- 41 **Campbell IK**, Piccoli DS, Roberts MJ, Muirhead KD, Hamilton JA. Effects of tumor necrosis factor alpha and beta on resorption of human articular cartilage and production of plasminogen activator by human articular chondrocytes. *Arthritis Rheum* 1990; **33**: 542-552 [PMID: 2328032 DOI: 10.1002/art.1780330412]
- 42 **Seckinger P**, Yaron I, Meyer FA, Yaron M, Dayer JM. Modulation of the effects of interleukin-1 on glycosaminoglycan synthesis by the urine-derived interleukin-1 inhibitor, but not by interleukin-6. *Arthritis Rheum* 1990; **33**: 1807-1814 [PMID: 2175610 DOI: 10.1002/art.1780331208]
- 43 **Pelletier JP**, Roughley PJ, DiBattista JA, McCollum R, Martel-Pelletier J. Are cytokines involved in osteoarthritic pathophysiology? *Semin Arthritis Rheum* 1991; **20**: 12-25 [PMID: 1866626 DOI: 10.1016/0049-0172(91)90024-T]
- 44 **Mokbel AN**, El Tookhy OS, Shamaa AA, Rashed LA, Sabry D, El Sayed AM. Homing and reparative effect of intra-articular injection of autologous mesenchymal stem cells in osteoarthritic animal model. *BMC Musculoskelet Disord* 2011; **12**: 259 [PMID: 22085445 DOI: 10.1186/1471-2474-12-259]
- 45 **Banfi A**, Bianchi G, Galotto M, Cancedda R, Quarto R. Bone marrow stromal damage after chemo/radiotherapy: occurrence, consequences and possibilities of treatment. *Leuk Lymphoma* 2001; **42**: 863-870 [PMID: 11697641 DOI: 10.3109/10428190109097705]
- 46 **Muschler GF**, Nitto H, Boehm CA, Easley KA. Age- and gender-related changes in the cellularity of human bone marrow and the prevalence of osteoblastic progenitors. *J Orthop Res* 2001; **19**: 117-125 [PMID: 11332607 DOI: 10.1016/S0736-0266(00)00010-3]
- 47 **Mendicino M**, Bailey AM, Wonnacott K, Puri RK, Bauer SR. MSC-based product characterization for clinical trials: an FDA perspective. *Cell Stem Cell* 2014; **14**: 141-145 [PMID: 24506881 DOI: 10.1016/j.stem.2014.01.013]
- 48 **Jeong SY**, Ha J, Lee M, Jin HJ, Kim DH, Choi SJ, Oh W, Yang YS, Kim JS, Kim BG, Chang JH, Cho DH, Jeon HB. Autocrine Action of Thrombospondin-2 Determines the Chondrogenic Differentiation Potential and Suppresses Hypertrophic Maturation of Human Umbilical Cord Blood-Derived Mesenchymal Stem Cells. *Stem Cells* 2015; **33**: 3291-3303 [PMID: 26235673 DOI: 10.1002/stem.2120]

- 49 **Meng H**, Zhang X, Hankenson KD, Wang MM. Thrombospondin 2 potentiates notch3/jagged1 signaling. *J Biol Chem* 2009; **284**: 7866-7874 [PMID: [19147503](#) DOI: [10.1074/jbc.M803650200](#)]
- 50 **Meng H**, Zhang X, Lee SJ, Strickland DK, Lawrence DA, Wang MM. Low density lipoprotein receptor-related protein-1 (LRP1) regulates thrombospondin-2 (TSP2) enhancement of Notch3 signaling. *J Biol Chem* 2010; **285**: 23047-23055 [PMID: [20472562](#) DOI: [10.1074/jbc.M110.144634](#)]



Published By Baishideng Publishing Group Inc  
7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA  
Telephone: +1-925-2238242  
E-mail: [bpgoffice@wjgnet.com](mailto:bpgoffice@wjgnet.com)  
Help Desk: <https://www.f6publishing.com/helpdesk>  
<https://www.wjgnet.com>

