World Journal of *Stem Cells*

World J Stem Cells 2020 October 26; 12(10): 1050-1236





Published by Baishideng Publishing Group Inc

W J S C World Journal of Stem Cells

Contents

Monthly Volume 12 Number 10 October 26, 2020

REVIEW

1050	Inflammatory bowel disease: Therapeutic limitations and prospective of the stem cell therapy
	Mishra R, Dhawan P, Srivastava AS, Singh AB
1067	Mesenchymal stem cells as living anti-inflammatory therapy for COVID-19 related acute respiratory distress syndrome

Lin F, Ichim TE, Pingle S, Jones LD, Kesari S, Ashili S

- 1080 Urine-derived stem/progenitor cells: A focus on their characterization and potential Burdeyron P, Giraud S, Hauet T, Steichen C
- 1097 Stem cell-based therapies for fertility preservation in males: Current status and future prospects Liu HC, Xie Y, Deng CH, Liu GH

MINIREVIEWS

- 1113 Medical, ethical, and legal aspects of hematopoietic stem cell transplantation for Crohn's disease in Brazil Ruiz MA, Junior RLK, Piron-Ruiz L, Saran PS, Castiglioni L, Quadros LG, Pinho TS, Burt RK
- 1124 Still 'dwelling in the possibility' - critical update on stem cell therapy for acute on chronic liver failure Philips CA, Augustine P

ORIGINAL ARTICLE

Basic Study

1133 In vivo cardiac pacemaker function of differentiated human mesenchymal stem cells from adipose tissue transplanted into porcine hearts

Darche FF, Rivinius R, Rahm AK, Köllensperger E, Leimer U, Germann G, Reiss M, Koenen M, Katus HA, Thomas D, Schweizer PA

- 1152 Pericyte-like differentiation of human adipose-derived mesenchymal stem cells: An in vitro study Mannino G, Gennuso F, Giurdanella G, Conti F, Drago F, Salomone S, Lo Furno D, Bucolo C, Giuffrida R
- 1171 Enrichment of retinal ganglion and Müller glia progenitors from retinal organoids derived from human induced pluripotent stem cells - possibilities and current limitations

Freude KK, Saruhanian S, McCauley A, Paterson C, Odette M, Oostenink A, Hyttel P, Gillies M, Haukedal H, Kolko M

- 1184 Creating rat hepatocyte organoid as an in vitro model for drug testing He YT, Zhu XL, Li SF, Zhang BQ, Li Y, Wu Q, Zhang YL, Zhou YY, Li L, Qi YN, Bao J, Bu H
- 1196 Neurotrophic effects of dental pulp stem cells in repair of peripheral nerve after crush injury Wang DR, Wang YH, Pan J, Tian WD



Contents

Monthly Volume 12 Number 10 October 26, 2020

SYSTEMATIC REVIEWS

Proteomic profiling of various human dental stem cells - a systematic review 1214

Hosmani J, Assiri K, Almubarak HM, Mannakandath ML, Al-Hakami A, Patil S, Babji D, Sarode S, Devaraj A, Chandramoorthy HC



Contents

Monthly Volume 12 Number 10 October 26, 2020

ABOUT COVER

Editorial board member of World Journal of Stem Cells, Dr. Liu is a Distinguished Professor at The First Affiliated Hospital of Nanchang University in Jiangxi Province, China. Dr. Liu undertook his postgraduate training at the Medical College of Nanchang University, receiving his Master's degree in 1995 and his MD/PhD in 2003. He became Chief Physician in the Burn Institute of the First Affiliated Hospital of Nanchang University in 2004 and has held the position since. His ongoing research interests involve wound healing and regenerative medicine, particularly to study stem cell skin tissue engineering, wound healing and scarring, and the regulation of refractory diabetic wounds. (L-Editor: Filipodia)

AIMS AND SCOPE

The primary aim of World Journal of Stem Cells (WJSC, World J Stem Cells) is to provide scholars and readers from various fields of stem cells with a platform to publish high-quality basic and clinical research articles and communicate their research findings online. WJSC publishes articles reporting research results obtained in the field of stem cell biology and regenerative medicine, related to the wide range of stem cells including embryonic stem cells, germline stem cells, tissue-specific stem cells, adult stem cells, mesenchymal stromal cells, induced pluripotent stem cells, embryonal carcinoma stem cells, hemangioblasts, lymphoid progenitor cells, etc.

INDEXING/ABSTRACTING

The WJSC is now indexed in Science Citation Index Expanded (also known as SciSearch®), Journal Citation Reports/Science Edition, Biological Abstracts, BIOSIS Previews, PubMed, and PubMed Central. The 2020 Edition of Journal Citation Reports[®] cites the 2019 impact factor (IF) for WJSC as 3.231; IF without journal self cites: 3.128; Ranking: 18 among 29 journals in cell and tissue engineering; Quartile category: Q3; Ranking: 113 among 195 journals in cell biology; and Quartile category: Q3.

RESPONSIBLE EDITORS FOR THIS ISSUE

Production Editor: Yan-Xia Xing; Production Department Director: Yun-Xiaojian Wu; Editorial Office Director: Jin-Lei Wang.

NAME OF JOURNAL	INSTRUCTIONS TO AUTHORS		
World Journal of Stem Cells	https://www.wjgnet.com/bpg/gerinfo/204		
ISSN	GUIDELINES FOR ETHICS DOCUMENTS		
ISSN 1948-0210 (online)	https://www.wignet.com/bpg/GerInfo/287		
LAUNCH DATE	GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH		
December 31, 2009	https://www.wignet.com/bpg/gerinfo/240		
FREQUENCY	PUBLICATION ETHICS		
Monthly	https://www.wignet.com/bpg/GerInfo/288		
EDITORS-IN-CHIEF	PUBLICATION MISCONDUCT		
Carlo Ventura	https://www.wignet.com/bpg/gerinfo/208		
EDITORIAL BOARD MEMBERS	ARTICLE PROCESSING CHARGE		
https://www.wjgnet.com/1948-0210/editorialboard.htm	https://www.wjgnet.com/bpg/gerinfo/242		
PUBLICATION DATE	STEPS FOR SUBMITTING MANUSCRIPTS		
October 26, 2020	https://www.wjgnet.com/bpg/GerInfo/239		
COPYRIGHT	ONLINE SUBMISSION		
© 2020 Baishideng Publishing Group Inc	https://www.f6publishing.com		

© 2020 Baishideng Publishing Group Inc. All rights reserved. 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA E-mail: bpgoffice@wjgnet.com https://www.wjgnet.com



WJSC

World Journal of **Stem Cells**

Submit a Manuscript: https://www.f6publishing.com

World J Stem Cells 2020 October 26; 12(10): 1067-1079

DOI: 10.4252/wjsc.v12.i10.1067

ISSN 1948-0210 (online)

REVIEW

Mesenchymal stem cells as living anti-inflammatory therapy for COVID-19 related acute respiratory distress syndrome

Feng Lin, Thomas E Ichim, Sandeep Pingle, Lawrence D Jones, Santosh Kesari, Shashaanka Ashili

ORCID number: Feng Lin 0000-0001-7662-423X; Thomas E Ichim 0000-0001-5611-374X; Sandeep Pingle 0000-0003-0124-4093; Lawrence D Jones 0000-0002-8706-2014; Santosh Kesari 0000-0003-3772-6000; Shashaanka Ashili 0000-0001-6335-0954.

Author contributions: All authors contributed equally to this paper in conception, literature review and analysis, drafting, critical revision and editing, and approval of the final version.

Conflict-of-interest statement: No conflict of interest to report.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/licenses /by-nc/4.0/

Manuscript source: Unsolicited manuscript

Feng Lin, Thomas E Ichim, Sandeep Pingle, Lawrence D Jones, Shashaanka Ashili, Research and Development, CureScience, San Diego, CA 92121, United States

Santosh Kesari, Cancer Center, John Wayne Cancer Institute and Pacific Neuroscience Institute at Providence Saint John's Health Center, Santa Monica, CA 90404, United States

Corresponding author: Feng Lin, PhD, Research Scientist, Research and Development, CureScience, No. 10225 Barnes Canyon Road A207, San Diego, CA 92121, United States. flin@curescience.org

Abstract

Coronavirus disease 2019 (COVID-19), a pandemic disease caused by the severe acute respiratory syndrome coronavirus 2 (SARS-CoV2), is growing at an exponential rate worldwide. Manifestations of this disease are heterogeneous; however, advanced cases often exhibit various acute respiratory distress syndrome-like symptoms, systemic inflammatory reactions, coagulopathy, and organ involvements. A common theme in advanced COVID-19 is unrestrained immune activation, classically referred to as a "cytokine storm", as well as deficiencies in immune regulatory mechanisms such as T regulatory cells. While mesenchymal stem cells (MSCs) themselves are objects of cytokine regulation, they can secrete cytokines to modulate immune cells by inducing antiinflammatory regulatory Treg cells, macrophages and neutrophils; and by reducing the activation of T and B cells, dendritic and nature killer cells. Consequently, they have therapeutic potential for treating severe cases of COVID-19. Here we discuss the unique ability of MSCs, to act as a "living antiinflammatory", which can "rebalance" the cytokine/immune responses to restore equilibrium. We also discuss current MSC trials and present different concepts for optimization of MSC therapy in patients with COVID-19 acute respiratory distress syndrome.

Key Words: Mesenchymal stem cells; SARS-CoV-2; COVID-19; Cytokine storm; Acute respiratory distress syndrome; Immunomodulation

©The Author(s) 2020. Published by Baishideng Publishing Group Inc. All rights reserved.

Saisbideng® WJSC | https://www.wjgnet.com

Received: July 6, 2020 Peer-review started: July 6, 2020 First decision: July 30, 2020 Revised: August 13, 2020 Accepted: September 14, 2020 Article in press: September 14, 2020 Published online: October 26, 2020

P-Reviewer: Figueiredo CS, Li ZJ S-Editor: Zhang L L-Editor: A P-Editor: Xing YX



Core Tip: Coronavirus disease 2019 a disease caused by the severe acute respiratory syndrome coronavirus 2, is growing exponentially, with no treatments currently available. Preclinical and clinical studies have shown that mesenchymal stem cells (MSCs) work in reversing acute respiratory distress syndrome caused by other conditions such as influenza virus infection, or sepsis. In this review we discuss the unique ability of MSCs, to act as a "living anti-inflammatory", which can "rebalance" the cytokine/immune responses to restore equilibrium.

Citation: Lin F, Ichim TE, Pingle S, Jones LD, Kesari S, Ashili S. Mesenchymal stem cells as living anti-inflammatory therapy for COVID-19 related acute respiratory distress syndrome. *World J Stem Cells* 2020; 12(10): 1067-1079

URL: https://www.wjgnet.com/1948-0210/full/v12/i10/1067.htm **DOI:** https://dx.doi.org/10.4252/wjsc.v12.i10.1067

INTRODUCTION

The severe respiratory consequences of the coronavirus disease 2019 (COVID-19) are caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) with few effective treatments currently available. SARS-CoV-2 along with SARS-CoV and middle east respiratory syndrome coronavirus are coronaviruses that have caused significant human morbidity and mortality^[1]. We are currently in the middle of a SARS-CoV-2 global pandemic. As of August 7, 2020, based on a Johns Hopkins University Coronavirus Resource Center Report, there have been at least 19.4 million confirmed cases worldwide, with at least 722706 deaths, and a mortality rate reaching approximately 3.7%. In the United States alone, approximately 5 million people are infected resulting in 161810 deaths and a mortality rate of 3.3%. COVID-19 infection of the lungs leads to extraordinary intensive care unit resource utilization and mortality.

One of the ways COVID-19 produces morbidity and mortality is by severely impairing lung function, causing a condition called acute respiratory distress syndrome (ARDS). This is characterized by a rapid onset of cytokine storm widespread lung inflammation, and sepsis-like conditions. Currently there is no effective anti-viral treatment for COVID-19. However, a few therapeutic strategies have been tested in the clinic or in trials for the treatment of patients with COVID-19. The anti-viral remdesivir can reduce death risk of severe COVID-19 patients as much as 62% when compared with standard care alone. Other anti-viral drugs such as but not limited to lopinavir-ritonavir, favipiravir, chloroquine and hydroxychloroquine have been proposed to treat COVID-19. Many of these anti-viral agents are currently being tested in clinical trials. Anti-inflammatory drugs such as dexamethasone were found to have beneficial effects in critically ill COVID-19 patients. Additionally, tocilizumab and siltuximab are interleukin-6 inhibitors being studied as therapeutics in critically ill patients with severe respiratory failure and elevated serum IL-6. The immunomodulatory functions of mesenchymal stem cells (MSCs) have been well documented in recent years. While MSCs themselves are objects of cytokine regulation, they can secrete cytokines to modulate immune cells by inducing antiinflammatory regulatory Treg (T) cells, macrophages and neutrophils; and by reducing the activation of T and B cells, dendritic and nature killer (NK) cells. Consequently, they have therapeutic potential for treating severe cases of COVID-19.

Preclinical and clinical studies have shown that MSCs work in reversing ARDS caused by other conditions such as influenza virus infection, or sepsis. For example, MSCs derived from adipose^[2-5], bone marrow^[6-25], placental^[26], amniotic membrane^[27,28], umbilical cord^[29-35], menstrual blood^[36], and lung^[37,38] origin, as well as conditioned media^[39-46], have demonstrated reduction of pulmonary injury, and neutrophil accumulation. Early clinical trials^[47,48] have shown safety of systemic infusions and bronchial instillations of MSCs for treating ARDS and other pulmonary complications. Most recently small studies^[49,50] have also suggested that MSCs can be effective in COVID-19 ARDS; however, these approaches have not been well validated. In this review, we discuss the immunomodulatory effects of stem cells and the role of MSCs as potential therapeutic options for cytokine storm and/or ARDS in COVID-19.

Zaishidene® WJSC | https://www.wjgnet.com

COVID-19 AND ITS PATHOLOGY

SARS-CoV-2 enters host cells through the angiotensin converting enzyme 2 (ACE2) receptor. ACE2 expression was found to be high in the lung, heart, ileum, kidney and bladder^[51]. ACE2 is highly expressed on the apical side of lung epithelial cells in the alveolar space, which is how SARS-CoV-2 virus can likely enter and destroy these cells. This is probably why early lung injury was often seen in the distal airway. The issue now becomes in what way does the immune system respond to viral infection in the lung. Innate immunity in the airway mainly includes epithelial cells, alveolar macrophages and dendritic cells. They fight against the virus until adaptive immunity is initiated. T cell mediated responses are initiated by antigen presentation via dendritic cells and macrophages. CD4+ T cells activate B cells to promote the production of virus-specific antibody CD8+ T cells which can kill virus infected cells. Approximately 80% percent of patients with COVID-19 are asymptomatic or experience only mild symptoms such as fever, dry cough and shortness of breath. However, some patients deteriorate quickly and develop ARDS^[52]. Patients with severe diseases were reported to have increased plasma levels of proinflammatory cytokines, including IL-6, IL-8, IL-10, GM-CSF, macrophage inflammatory protein 1-alpha, and TNF-alpha^[53].

Information regarding the pathological findings in COVID-19 is still limited, although several reports on this topic have been published recently. Xu *et al*^[54] reported one case where the patient presented 15 d of symptoms. Biopsy samples from both lungs showed bilateral diffuse alveolar damage^[55] with cellular fibromyxoid exudates and hyaline membrane formation, indicating ARDS in both lungs. Noteworthy is the observation that the pathological features of COVID-19 greatly resemble those seen in SARS and middle east respiratory syndrome coronavirus infections^[56,57]. In addition, over-activation of T-cells was manifested by an increase of Th17 and high cytotoxicity of CD8 T cells, partially accounting for the severe immune injury in the patient.

Tian *et al*^[58] described the early histopathological features in two patients who underwent postmortem for lung cancer but were later discovered to have had COVID-19 upon resection. The results of the lung evaluation from the two patients exhibited only nonspecific histologic changes, including edema, proteinaceous exudate, hyperplastic pneumocytes, patchy inflammation, and multinucleated giant cells with no hyaline membrane. Given that the two patients were asymptomatic from COVID-19 at the time of postmortem, they were likely only in the early stages of acute lung injury from the infection^[58]. Tian *et al*^[59] also conducted another postmortem study of four COVID-19 patients with a minimum of 15 days of symptoms, demonstrated ARDS in all biopsies.

Two COVID-19 autopsies on the lungs of a 77-year-old man revealing diffuse alveolar damage, the most common histopathologic correlation with ARDS, and on the lungs of a 42-year-old man presenting no evidence of diffuse alveolar damage/ARDS^[60]. Magro et al^[61] demonstrated in a report of five cases that diffuse alveolar damage was not prominent with the presentation of hyaline membranes, inflammation, and type II pneumocyte hyperplasia, all hallmarks of classic ARDS. These pulmonary findings were accompanied by significant deposits of terminal complement components C5b-9, C4d, and mannose binding lectin-associated serine protease 2, in the microvasculature, consistent with sustained, systemic activation of the alternative and lectin-based complement pathways. There was colocalization of COVID-19 spike glycoproteins with C4d and C5b-9 in the interalveolar septa and the cutaneous microvasculature of two cases examined. This indicated the pathophysiologic importance of complement in COVID-19. The results suggest that at least a subset of severe COVID-19 infection involves a catastrophic, complementmedicated thrombotic microvascular injury syndrome with sustained activation of the alternative and lectin-based cascades, possible pathways apart from virus spike protein engagement.

In general, ARDS is a common manifestation of cytokine storms and as well could be the cause of death in many COVID-19 patients, although other mechanisms may also be involved. A better understanding of COVID-19 patients' underlying pathogenesis will pave the way for formulating a timely therapeutic strategy to reduce mortality.

Zaishidena® WJSC | https://www.wjgnet.com

MSCS IMMUNOMODULATORY EFFECTS

MSCs are fibroblast-like and multipotent stromal cells. Human MSCs are positive for a number of cell surface markers including CD73, CD44, CD90, and CD105 and negative for the hematopoietic markers of CD34, CD45 and HLA-DR^[62]. MSCs are traditionally isolated from bone marrow, and a variety of fetal, neonatal and adult tissues, including cord blood, peripheral blood, fetal liver and lung, adipose tissue, compact bone, dental pulp, dermis, endometrial, human islet, adult brain, skeletal muscle, amniotic fluid, synovium, and the circulatory system[63-65]. MSCs can differentiate into a variety of cell types of mesodermal origin, including osteoblasts, chondrocytes, cardiomyocytes, neural cells, smooth muscle cells and adipocytes^[62,66-68]. MSCs are likely the only stem cell type that possesses both regenerative and immunomodulatory capabilities. Consequently, they have been used widely in the treatment of many degenerative and inflammatory diseases.

One property that greatly increases the value of MSCs in therapeutic applications is their ability to modulate immune responses. MSCs can exert their immunomodulatory function by producing many molecules having immunomodulatory effects, these include prostaglandin E2 (PGE2)^[69], nitric oxide^[70], indolamine 2,3-dioxigenase (IDO)^[71], transforming growth factor beta^[72,73], IL-6^[74,75], hemoxygenase-1^[76], leukocyte inhibitory factor^[77], HLAG5 and chemokines^[78], PDL1/2^[79] and other surface markers-FasL^[80]. MSCs can escape the immune system because bone marrow derived MSCs (BM-MSCs) are not recognized by NK cells as they lack expression of HLA Class I surface markers. They also lack expression of HLA Class II antigens, which is desirable for transplantation applications.

The immunosuppressive activity of MSCs is well described, with recent reports providing some mechanistic insights into key soluble factors and receptors. Programmed death-ligand 1/CD274 also known as B7 Homolog 1 (B7-H1) has been shown to be expressed in cultured MSCs and is strongly upregulated following IFN-y stimulation. Combination therapy using rapamycin and MSCs induced immune tolerance to allografts, but monoclonal antibodies against B7-H1 were shown to abrogate this tolerance leading to allograft rejection^[81]. The immunomodulatory effects of MSCs were mediated in part through upregulation of regulatory immune cells including CD4+CD25+FoxP3+ T cells $^{[82,83]}$ and tolerogenic dendritic cells $^{[84]}$ and a decrease in alloantibody levels. MSCs that expressed B7H1 may also induce the apoptosis of activated T-cells as a co-culture of CD4+CD25- T cells with MSCs resulting in significant upregulation of programmed cell death-1 receptor (PD-1) on activated T cells^[85]. Similar results were reported by Chinnadurai et al^[86] who further examined the role of IFN- γ in the "licensing" of MSCs to inhibit the proliferation of activated T cells^[86]. Both MSCs and IFN-y licensed MSCs inhibited T-cell proliferation; however, only IFN-y licensed MSCs significantly inhibited Th1 cytokine (IFN-y, TNFa and IL-2) production as well as T-cell degranulation. This IFN-y licensed MSCs inhibitory effect on T-cells is thought to be dependent on IDO^[71]; however, Chinnadurai showed that MSC IDO catalytic function is dispensable with regard to MSC driven T-cell inhibition. Chinnadurai et al^[86] identified the B7-H1 PD1 pathways as essential effectors in blocking T-cell function. Further complexity was also suggested by a recent report that IFN-y treatment of MSCs upregulated HLA-DR /Class II MHC after 48 h, and MSCs ability to inhibit T cells through B7-H1 was dependent upon the presence of HLA-DR^[87].

MSCs express the adhesion molecules VCAM-1 and ICAM-1, which allow Tlymphocytes to adhere to their surface. Subsequently MSCs can affect them by molecules which have a short half-life. Their effect is in the immediate vicinity of the cell^[70]. Examples of such molecules include nitric oxide, PGE2, HGF^[88], and activation of receptor PD-1. MSCs reduce T cell proliferation between G0 and G1 cell cycle phases G^[89], and decrease the expression of IFNy of Th1 cells while increasing the expression of IL-4 of Th2 cells^[90]. MSCs also inhibit the proliferation of B-lymphocytes between G0 and G1 cell cycle phases.

A novel mechanism for MSC-induced immunosuppression was recently proposed by Obermajer and colleagues who showed that cells of the Th17 type, predominantly associated with the rejection of allogeneic solid organ grafts, can be directly converted into a regulatory T cell type^[91]. The induction of Tregs was preceded by development of a CD11b(hi)Gr1(int) myeloid-derived immunosuppressive cell-mediated Th17. They identified retinoic acid receptor-related orphan receptor γ as a common factor in the differentiation of T and Th17 cells. The identification of a specific subset of T cells IL-17A+Foxp3+ double-positive and ex-IL-17- producing IL-17A-Foxp3+ in this paper argues for direct conversion as the mechanism for MSC-mediated immuno-tolerance. This proposed mechanism where MSC-induced myeloid-derived immunosuppressive



cells act as mediator for immune tolerance without complete immunosuppression may have significant implications for therapeutic applications.

MSCs have an effect on macrophages, neutrophils, NK cells, mast cells and dendritic cells in innate immunity and effector T cells, regulatory T cells, and B cells in adaptive immunity illustrated in Figure 1. In severe COVID-19 patients, their immune responses to SARS-CoV-2 infection are usually over-activated. MSCs are able to exert their anti-inflammatory effect by regulating immune cells and balancing the immune responses. Furthermore, MSCs can migrate to the site of injury, where they polarize through GE2 macrophages into phenotype-2 which is characterized by an antiinflammatory effect^[92,93]. Further, PGE2 inhibits the ability of mast cells to degranulate and produce TNF-a. Proliferation and cytotoxic activity of NK cells are inhibited by PGE2 and IDO. MSCs also reduce the expression of NK cell receptors-NKG2D, NKp44 and NKp30^[94], MSCs inhibit respiratory flare and apoptosis of neutrophils by production of cytokines IL-6 and IL-8^[95]. Differentiation and expression of dendritic cell surface markers is inhibited by IL-6 and PGE2 of MSCs^[96]. The immunosuppressive effects of MSCs also depend on IL-10, but it is not certain whether they produce it alone, or only stimulate other cells to produce it^[97,98].

MSCS THERAPY FOR INHIBITION OF ACUTE INFLAMMATION AND CYTOKINE STORM

MSCs have been shown to possess a comprehensive and powerful immunomodulatory function to suppress excessive activation of the immune system, thus promoting endogenous repair by improving the microenvironment. There have been 13 MSCs therapies approved for treating a number of conditions (Table 1) outside of the United States, mainly in the EU, Japan, South Korea and India. Among the conditions, two adipose tissue derived MSC products, Alofisel® and Cupistem®, have been used for complex perianal fistulas in Crohn's disease. The underlying mechanism of action is the MSC immunomodulatory and anti-inflammatory effects at the inflammation sites. Specifically MSCs impair proliferation of activated lymphocytes and reduce the inflammatory cytokines. Two BM-MSC products, Prochymal® and Temcell® HS, have been used for treating GvHD, due to MSCs immunomodulatory effects.

Preclinical study has demonstrated that MSCs can inhibit the progress of acute inflammation in the lungs and alleviate symptoms of respiratory distress^[99]. The feasibility of utilizing MSCs for the treatment of ARDS has been demonstrated in animal models and extracorporeal lung models^[100]. MSCs of adipose, bone marrow, placental, amniotic membrane, umbilical cord, menstrual blood, and lung, origin, as well as conditioned media with secreted exosomes, have demonstrated a reduction of pulmonary injury and neutrophil accumulation. In a recent study using a sheep model of ARDS^[9], both endobronchial and intravenous administration of bone marrowderived multipotent adult progenitor cells were effective for the treatment of ARDS.

Additionally, an analysis of 342 systemic infusions and 57 bronchial instillations (204 recipients) of cells of various origins for ARDS and other pulmonary issues demonstrated safety in early human clinical trials^[47]. Recently, a study involving two patients with severe refractory ARDS, both showed improvement^[99]. Both patients received 2×10^6 cells per kilogram of body weight. Subsequently, each of the patients improved with resolution of respiratory, hemodynamic, and multiorgan failure. In parallel, a decrease was seen in multiple pulmonary and systemic markers of inflammation, including epithelial apoptosis, alveolar-capillary fluid leakage, proinflammatory cytokines, microRNAs, and chemokines. In vitro studies of the MSCs demonstrated a broad anti-inflammatory capacity, including suppression of T-cell responses and induction of regulatory phenotypes in T cells, monocytes, and neutrophils. Some of these in vitro potency assessments correlated with, and were relevant to, the observed in vivo actions.

RECENT SUCCESS OF MSCS FOR COVID-19 PATIENTS AND CLINICAL TRIALS

Currently, drugs alone or in combination with other therapeutic approaches have not afforded a cure; however a number of investigational drugs in clinical trials, including antivirals such as chloroquine/hydroxychloroquine, remdesivir, immune-based



Tab	Table 1 Approved mesenchymal stem cell therapies outside of United States in the past 20 years						
	Product name	Source	Autologous/Allogeneic	Indication	Company/Country		
1	Alofisel	Adipose tissue-derived stem cells	Allogeneic	Complex perianal fistuals in Crohn's disease	TiGenix NV/Takeda PharmaceuticalEU		
2	Chondrocytes-T- Ortho-ACI	Chondrocyte	Autologous	Cartilage damage, lesions and defects	Ortho Cell, Australia		
3	Spherox	Chondrocyte	Autologous	Symptomatic articular cartilage defects	CO.DON AG, Germany and EU		
4	Ossgrow	BM-MSCs	Autologous	Avascular necrosis	Regrow, India		
5	Stempeucel	BM-MSCs	Allogeneic	CLI	Stempeutics, India		
6	Porchymal	BM-MSCs	Allogeneic	GvHD in children	Osiris Therapeutics, Canada		
7	Temcell HS	BM-MSCs	Allogeneic	GvHD	JCR Pharmaceuticals, Japan		
8	NeuroNata-R	BM-MSCs	Autologous	Lou Gehrig's disease, or ALS	Corestem, Korea		
9	Cupistem	AT-MSCs	Autologous	Crohn's fistula	Anterogen, Korea		
10	Cartistem,	UC-blood-derived MSCs	Allogeneic	Damaged cartilage	Medipost Inc., Korea		
11	Cellgram-AMI	BM-MSCs	Autologous	Acute myocardial infarction	Pharmicell, Korea		
12	AstroStem	AT-MSCs	Autologous	Alzheimer's disease	Nature cell, Korea		
13	Stemilac	BM-MSCs	Autologous	Alzheimer's disease	Nipro and Sapporo Medical University, Japan		

MSCs: Mesenchymal stem cells; BM-MSCs: Bone marrow derived mesenchymal stem cells; EU: European Union; UC-MSCs: Umbilical cord mesenchymal stem cells; AT-MSCs: Adipose tissue-derived mesenchymal stem cells.

> therapies and adjunctive therapies have shown promise, particularly in mitigating certain systemic markers according to NIH COVID-19 Guidelines. Potential antiviral drugs: remdesivir, chloroquine or hydroxychloroquine, hydroxychloroquine plus azithromycin, lopinavir/ritonavir and other HIV protease inhibitors. Immune-based therapy under evaluation: Convalescent plasma, Immunoglobulins: SAR-CoV-2specific and non-SAR-CoV-2 specific, MSCs, Corticosteroids, Interferons alpha and beta, IL-1 and IL-6 inhibitors, Kinase inhibitors: Bruton's tyrosine kinase inhibitors and Janus kinase inhibitors; Adjunctive therapy: antithrombotic therapy, vitamin C and vitamin D, zinc supplementation.

> To date, there are over 50 clinical trials using MSCs to treat COVID-19 patients based on the registration in clinicaltrial.gov website, we listed the most relevant studies in Table 2. Umbilical cord MSCs (UC-MSCs), BM-MSCs, AT-MSCs and other MSCs, as well as exosomes from MSCs are used in clinical trials, among them UC-MSCs are the most desirable for treating severely compromised COVID-19 patients due to its rich and extensive source of stem cells, scalable expansion capability, and ability to be allogeneic as low MHC-I expression[49]. The dose and delivery times are also varied in different trials. Recent reviews have described the potential for and rationale of using different types of MSCs for treating severe COVID-19 patients to protect alveolar epithelial cells, to reclaim the pulmonary microenvironment, to induce anti-inflammatory macrophages, regulatory T and B cells, and regulatory dendritic cells. In addition, MSCs can inactivate T cells in order to prevent cytokine storm, prevent pulmonary fibrosis and cure lung dysfunction^[101-104]. Details of MSCs clinical trials for COVID-19 have also been discussed in other reviews^[101,104-106].

> MSCs have been used effectively to treat patients with COVID-19 in recent reports^[50,107,108]. The underlying processes involve preventing the cytokine storm from occurring as well as reversing the cytokine storm in compromised patients. A total of seven patients with COVID-19 were enrolled in the study^[50], the results have shown that MSCs significantly improved the functional outcome without observed adverse effects. The pulmonary function and symptoms of these patients were significantly improved in two days after MSC transplantation. Among them, two common and one severe patient recovered and were discharged within 10 d after the treatment. Compared to the placebo control group, the level of TNF-alpha was significantly decreased, and IL-10 increased in the MSCs treatment group. The gene profile showed MSCs were ACE2- and TMPRSS2- which indicate MSCs are free from COVID-19



Table 2 The most relevant clinical trials of mesenchymal stem cell treating COVID-19 patients							
NCT number	Cell type	Autologous	Phase	Sponsor			
NCT04490486	UC-MSCs	Allogeneic	Ι	Joshua M Hare, United States			
NCT04456361	UC-MSCs	Allogeneic	Ι	Instituto de Medicina Regenerativa, Mexico			
NCT04313322	UC -MSCs	Allogeneic	Ι	Stem Cells, Arabia			
NCT04288102	MSCs	NA	II	Beijing 302 Hospital			
NCT04346368	BM-MSCs	NA	I/II	Guangzhou Institute of Respiratory Disease			
NCT04366323	Adipose-derived MSCs	Allogeneic	I/II	Andalusian Network for Design and Translation of Advanced Therapies			
NCT04273646	UC-MSCs	Allogeneic	Ι	Wuhan Union Hospital, China			
NCT04349631	Adipose-derived MSCs	Autologous	II	Hope Biosciences			
NCT04339660	UC-MSCs	Allogeneic	I/II	Puren Hospital Affiliated to Wuhan University of Science and Technology			
NCT04366063	MSCs	NA	II/III	Royan Institute			
NCT04352803	Adipose-derived MSCs	Autologous	Ι	Regeneris Medical			
NCT04355728	UC-MSCs	Allogeneic	I/II	Camillo Ricordi			
NCT04366271	UC-MSCs	Allogeneic	II	Hospital Infantil Universitario Niño Jesús, Madrid, Spain			
NCT04348461	Adipose-derived MSCs	Allogeneic	Ι	Instituto de Investigación Sanitaria de la Fundación Jiménez Díaz			
NCT04345601	BM-MSCs	Allogeneic	Ι	Baylor College of Medicine			
NCT03042143	UC-MSCs (CD362 enriched)	Allogeneic	I/II	Belfast Health and Social Care Trust			
NCT04361942	MSCs	Allogeneic	II	Red de Terapia Celular			
NCT04269525	UC-MSCs	Allogeneic	II	Zhi-Yong Peng, Hospital			
NCT04333368	UC-MSCs	Allogeneic	Ι	Assistance Publique - Hôpitaux de Paris			
NCT04299152	Cord blood stem cells (CB-SC)	Allogeneic	II	Tianhe Stem Cell Biotechnologies Inc.			
NCT04341610	Adipose-derived MSCs	Allogeneic	I/II	Rigshospitalet, Denmark			
NCT04276987	Adipose MSC-derived exosomes (inhalation)	Allogeneic	Ι	Ruijin Hospital			
NCT03857841	BM-MSC derived extracellular vesicles (UNEX-42)	Allogeneic	Ι	United Therapeutics			

MSC: Mesenchymal stem cells; BM-MSC: Bone marrow derived mesenchymal stem cells; UC-MSCs: Umbilical cord mesenchymal stem cells.

infection. In another case report^[107], UC-MSCs were infused into a severely compromised COVID-19 patient. The pulmonary function and symptoms of the patient were significantly improved in 2 d after UC-MSCs transplantation. The patient recovered and was discharged in 7 d after treatment. The percentage and counts of lymphocyte subsets (CD3, CD4, and CD8 T cell) were increased, and the level of IL-6, TNF- α , and C-reactive protein was shown to have significantly decreased after UC-MSCs treatment. Guo et al^[108] reported a 31-patient trial with UC-MSCs infusion. After the first infusion of UC-MSCs, the SARS-CoV-2 PCR results of 30 patients (96.8%) became negative after a mean time of 10.7 d (SD, 4.2 d). Laboratory parameters tended to improve after UC-MSCs therapy compared to the status before treatment, including elevated lymphocyte count, decreased C-reactive protein and IL-6 levels. Thus far, the intravenous transplantation of MSCs has been shown to be effective for the treatment of patients with COVID-19 pneumonia, especially for the patients in critical condition.

Exosomes derived from MSCs have been studied in clinical trials for treating severely compromised COVID-19 patients^[104,109,110]. Exosomes (ExoFlo™) derived from allogeneic bone marrow MSCs in a single 15 mL dose were evaluated in a 24-patient trial^[110] for both safety and efficacy from days 1 to 14 post-treatment. No adverse events were observed; a survival rate of 83% was observed; 71 patents recovered, 13% remained critically ill though stable, and 16% patients expired for reasons unrelated to the treatment. Overall, after one treatment, patients' clinical status and oxygenation improved with an average pressure of arterial oxygen to fraction of inspired oxygen

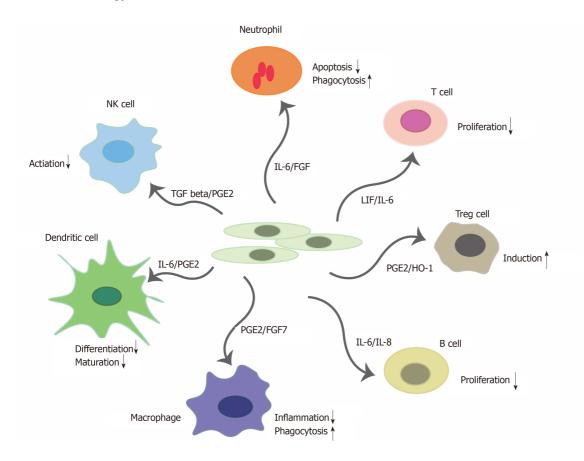


Figure 1 Immunomodulatory effects of mesenchymal stem cells on immune cells. Mesenchymal stem cells secrete cytokines to modulate immune cells by inducing anti-inflammatory regulatory Treg cells, macrophages and neutrophils, reducing the activation of Treg and B cells, dendritic and nature killer cells. T cell: Treg cell; PGE2: Prostaglandin E2; NK cell: Nature killer cell.

ratio (PaO_2/FiO_2) increase of 192%. Laboratory values revealed significant improvements in absolute neutrophil count and lymphopenia with average CD3⁺, CD4⁺, and CD8⁺ lymphocyte counts increasing by 46%, 45%, and 46%, respectively. Likewise, acute phase reactants declined, with mean C-reactive protein, ferritin, and D-dimer reduction of 77%, 43%, and 42%, respectively. The study demonstrated the excellent safety profile and capacity to restore oxygenation, downregulate cytokine storm, and reconstitute immunity. Exosome derived from MSCs is a promising therapeutic candidate for severely compromised COVID-19 patients.

CONCLUSION

The social burden of COVID-19 is growing with the global pandemic. However, there is no effective or curative therapy for COVID-19, and preventive vaccines, other than the vaccine recently approved in Russia, but for which there is limited information, are still under development and will not be available until next year. The most recent clinical trials with MSCs may fulfill the unmet medical need of COVID-19, to reduce the related ARDS and cytokine storm. There are several issues that need to be addressed in order to move forward: dose, delivery times, type of MSCs, efficacy and cost-effectiveness. An understanding of all facets of MSCs and pathomechanism of COVID-19 is necessary to fully translate the MSCs therapy into a meaningful treatment for COVID-19. The next therapeutic strategies may focus on a combination approach using two or more types of MSCs, certain type of MSCs, and immune-based therapies or antiviral therapies to achieve maximal therapeutic efficacy.

REFERENCES

 Guarner J. Three Emerging Coronaviruses in Two Decades. Am J Clin Pathol 2020; 153: 420-421 [PMID: 32053148 DOI: 10.1093/ajcp/aqaa029]



- 2 Jung YJ, Park YY, Huh JW, Hong SB. The effect of human adipose-derived stem cells on lipopolysaccharide-induced acute respiratory distress syndrome in mice. Ann Transl Med 2019; 7: 674 [PMID: 31930075 DOI: 10.21037/atm.2019.10.48]
- Chen CH, Chen YL, Sung PH, Sun CK, Chen KH, Chen YL, Huang TH, Lu HI, Lee FY, Sheu JJ, Chung 3 SY, Lee MS, Yip HK. Effective protection against acute respiratory distress syndrome/sepsis injury by combined adipose-derived mesenchymal stem cells and preactivated disaggregated platelets. Oncotarget 2017; 8: 82415-82429 [PMID: 29137274 DOI: 10.18632/oncotarget.19312]
- 4 Lu H, Cook T, Poirier C, Merfeld-Clauss S, Petrache I, March KL, Bogatcheva NV. Pulmonary Retention of Adipose Stromal Cells Following Intravenous Delivery Is Markedly Altered in the Presence of ARDS. Cell Transplant 2016; 25: 1635-1643 [PMID: 26609693 DOI: 10.3727/096368915X690189]
- 5 Zheng G, Huang L, Tong H, Shu Q, Hu Y, Ge M, Deng K, Zhang L, Zou B, Cheng B, Xu J. Treatment of acute respiratory distress syndrome with allogeneic adipose-derived mesenchymal stem cells: a randomized, placebo-controlled pilot study. Respir Res 2014; 15: 39 [PMID: 24708472 DOI: 10.1186/1465-9921-15-39]
- 6 Lu Z, Chang W, Meng S, Xu X, Xie J, Guo F, Yang Y, Qiu H, Liu L. Mesenchymal stem cells induce dendritic cell immune tolerance via paracrine hepatocyte growth factor to alleviate acute lung injury. Stem Cell Res Ther 2019; 10: 372 [PMID: 31801626 DOI: 10.1186/s13287-019-1488-2]
- Silva JD, de Castro LL, Braga CL, Oliveira GP, Trivelin SA, Barbosa-Junior CM, Morales MM, Dos Santos CC, Weiss DJ, Lopes-Pacheco M, Cruz FF, Rocco PRM. Mesenchymal Stromal Cells Are More Effective Than Their Extracellular Vesicles at Reducing Lung Injury Regardless of Acute Respiratory Distress Syndrome Etiology. Stem Cells Int 2019; 2019: 8262849 [PMID: 31531026 DOI: 10.1155/2019/8262849
- 8 Xu AL, Rodriguez LA 2nd, Walker KP 3rd, Mohammadipoor A, Kamucheka RM, Cancio LC, Batchinsky AI, Antebi B. Mesenchymal Stem Cells Reconditioned in Their Own Serum Exhibit Augmented Therapeutic Properties in the Setting of Acute Respiratory Distress Syndrome. Stem Cells Transl Med 2019; 8: 1092-1106 [PMID: 31219247 DOI: 10.1002/sctm.18-0236]
- Cardenes N, Aranda-Valderrama P, Carney JP, Sellares Torres J, Alvarez D, Kocyildirim E, Wolfram Smith JA, Ting AE, Lagazzi L, Yu Z, Mason S, Santos E, Lopresti BJ, Rojas M. Cell therapy for ARDS: efficacy of endobronchial versus intravenous administration and biodistribution of MAPCs in a large animal model. BMJ Open Respir Res 2019; 6: e000308 [PMID: 30713713 DOI: 10.1136/bmjresp-2018-000308]
- 10 Li L, Dong L, Zhang J, Gao F, Hui J, Yan J. Mesenchymal stem cells with downregulated Hippo signaling attenuate lung injury in mice with lipopolysaccharideinduced acute respiratory distress syndrome. Int J Mol Med 2019; 43: 1241-1252 [PMID: 30628652 DOI: 10.3892/ijmm.2018.4047]
- Mokhber Dezfouli MR, Jabbari Fakhr M, Sadeghian Chaleshtori S, Dehghan MM, Vajhi A, Mokhtari R. 11 Intrapulmonary autologous transplant of bone marrow-derived mesenchymal stromal cells improves lipopolysaccharide-induced acute respiratory distress syndrome in rabbit. Crit Care 2018; 22: 353 [PMID: 30572913 DOI: 10.1186/s13054-018-2272-x]
- 12 Schwede M, Wilfong EM, Zemans RL, Lee PJ, Dos Santos C, Fang X, Matthay MA. Effects of bone marrow-derived mesenchymal stromal cells on gene expression in human alveolar type II cells exposed to TNF-α, IL-1β, and IFN-γ. Physiol Rep 2018; 6: e13831 [PMID: 30136410 DOI: 10.14814/phy2.13831]
- 13 Masterson C, Devaney J, Horie S, O'Flynn L, Deedigan L, Elliman S, Barry F, O'Brien T, O'Toole D, Laffey JG. Syndecan-2-positive, Bone Marrow-derived Human Mesenchymal Stromal Cells Attenuate Bacterial-induced Acute Lung Injury and Enhance Resolution of Ventilator-induced Lung Injury in Rats. Anesthesiology 2018; 129: 502-516 [PMID: 29979191 DOI: 10.1097/ALN.00000000002327]
- 14 Park J, Jeong S, Park K, Yang K, Shin S. Expression profile of microRNAs following bone marrowderived mesenchymal stem cell treatment in lipopolysaccharide-induced acute lung injury. Exp Ther Med 2018; 15: 5495-5502 [PMID: 29904430 DOI: 10.3892/etm.2018.6118]
- 15 Pedrazza L, Cunha AA, Luft C, Nunes NK, Schimitz F, Gassen RB, Breda RV, Donadio MV, de Souza Wyse AT, Pitrez PMC, Rosa JL, de Oliveira JR. Mesenchymal stem cells improves survival in LPS-induced acute lung injury acting through inhibition of NETs formation. J Cell Physiol 2017; 232: 3552-3564 [PMID: 28112391 DOI: 10.1002/jcp.25816]
- 16 Yang Y, Hu S, Xu X, Li J, Liu A, Han J, Liu S, Liu L, Qiu H. The Vascular Endothelial Growth Factors-Expressing Character of Mesenchymal Stem Cells Plays a Positive Role in Treatment of Acute Lung Injury In Vivo. Mediators Inflamm 2016; 2016: 2347938 [PMID: 27313398 DOI: 10.1155/2016/2347938]
- Moodley Y, Sturm M, Shaw K, Shimbori C, Tan DB, Kolb M, Graham R. Human mesenchymal stem cells 17 attenuate early damage in a ventilated pig model of acute lung injury. Stem Cell Res 2016; 17: 25-31 [PMID: 27231985 DOI: 10.1016/j.scr.2016.05.005]
- Hayes M, Curley GF, Masterson C, Devaney J, O'Toole D, Laffey JG. Mesenchymal stromal cells are more 18 effective than the MSCs secretome in diminishing injury and enhancing recovery following ventilatorinduced lung injury. Intensive Care Med Exp 2015; 3: 29 [PMID: 26472334 DOI: 10.1186/s40635-015-0065-y]
- Monsel A, Zhu YG, Gennai S, Hao Q, Hu S, Rouby JJ, Rosenzwajg M, Matthay MA, Lee JW. Therapeutic 19 Effects of Human Mesenchymal Stem Cell-derived Microvesicles in Severe Pneumonia in Mice. Am J Respir Crit Care Med 2015; 192: 324-336 [PMID: 26067592 DOI: 10.1164/rccm.201410-1765OC]
- 20 Hao Q, Zhu YG, Monsel A, Gennai S, Lee T, Xu F, Lee JW. Study of Bone Marrow and Embryonic Stem Cell-Derived Human Mesenchymal Stem Cells for Treatment of Escherichia coli Endotoxin-Induced Acute Lung Injury in Mice. Stem Cells Transl Med 2015; 4: 832-840 [PMID: 25999518 DOI: 10.5966/sctm.2015-0006]
- 21 Devaney J, Horie S, Masterson C, Elliman S, Barry F, O'Brien T, Curley GF, O'Toole D, Laffey JG. Human mesenchymal stromal cells decrease the severity of acute lung injury induced by E. coli in the rat. Thorax 2015; 70: 625-635 [PMID: 25986435 DOI: 10.1136/thoraxjnl-2015-206813]
- Asmussen S, Ito H, Traber DL, Lee JW, Cox RA, Hawkins HK, McAuley DF, McKenna DH, Traber LD, 22 Zhuo H, Wilson J, Herndon DN, Prough DS, Liu KD, Matthay MA, Enkhbaatar P. Human mesenchymal stem cells reduce the severity of acute lung injury in a sheep model of bacterial pneumonia. Thorax 2014;



69: 819-825 [PMID: 24891325 DOI: 10.1136/thoraxinl-2013-204980]

- 23 Shalaby SM, El-Shal AS, Abd-Allah SH, Selim AO, Selim SA, Gouda ZA, Abd El Motteleb DM, Zanfaly HE, El-Assar HM, Abdelazim S. Mesenchymal stromal cell injection protects against oxidative stress in Escherichia coli-induced acute lung injury in mice. Cytotherapy 2014; 16: 764-775 [PMID: 24525173 DOI: 10.1016/j.jcyt.2013.12.006]
- Bustos ML, Huleihel L, Meyer EM, Donnenberg AD, Donnenberg VS, Sciurba JD, Mroz L, McVerry BJ, 24 Ellis BM, Kaminski N, Rojas M. Activation of human mesenchymal stem cells impacts their therapeutic abilities in lung injury by increasing interleukin (IL)-10 and IL-1RN levels. Stem Cells Transl Med 2013; 2: 884-895 [PMID: 24089414 DOI: 10.5966/sctm.2013-0033]
- 25 Rojas M, Parker RE, Thorn N, Corredor C, Iyer SS, Bueno M, Mroz L, Cardenes N, Mora AL, Stecenko AA, Brigham KL. Infusion of freshly isolated autologous bone marrow derived mononuclear cells prevents endotoxin-induced lung injury in an ex-vivo perfused swine model. Stem Cell Res Ther 2013; 4: 26 [PMID: 23497755 DOI: 10.1186/scrt174]
- 26 Yan X, Fu X, Jia Y, Ma X, Tao J, Yang T, Ma H, Liang X, Liu X, Yang J, Wei J. Nrf2/Keap1/ARE Signaling Mediated an Antioxidative Protection of Human Placental Mesenchymal Stem Cells of Fetal Origin in Alveolar Epithelial Cells. Oxid Med Cell Longev 2019; 2019: 2654910 [PMID: 31217836 DOI: 10.1155/2019/26549101
- 27 Cui P, Xin H, Yao Y, Xiao S, Zhu F, Gong Z, Tang Z, Zhan Q, Qin W, Lai Y, Li X, Tong Y, Xia Z. Human amnion-derived mesenchymal stem cells alleviate lung injury induced by white smoke inhalation in rats. Stem Cell Res Ther 2018; 9: 101 [PMID: 29650044 DOI: 10.1186/s13287-018-0856-7]
- Zhang S, Jiang W, Ma L, Liu Y, Zhang X, Wang S. Nrf2 transfection enhances the efficacy of human 28 amniotic mesenchymal stem cells to repair lung injury induced by lipopolysaccharide. J Cell Biochem 2018: 119: 1627-1636 [PMID: 28905450 DOI: 10.1002/jcb.26322]
- 29 Huang Z, Liu H, Zhang X, Wen G, Zhu C, Zhao Y, Niu W, Qin Y, Chen H, Bai C, Liu G. Transcriptomic analysis of lung tissues after hUC-MSCs and FTY720 treatment of lipopolysaccharide-induced acute lung injury in mouse models. Int Immunopharmacol 2018; 63: 26-34 [PMID: 30064040 DOI: 10.1016/j.intimp.2018.06.036]
- 30 Xuan YY, Wu YY, Xie YL, Chu JG, Li GX, Wang LP. Human Mesenchymal Stem/Stromal Cells From Human Umbilical Cord Ameliorate Acute Respiratory Distress Syndrome in Rats: Factors to Consider, Crit Care Med 2017; 45: e736-e737 [PMID: 28622233 DOI: 10.1097/CCM.00000000002401]
- 31 Lee FY, Chen KH, Wallace CG, Sung PH, Sheu JJ, Chung SY, Chen YL, Lu HI, Ko SF, Sun CK, Chiang HJ, Chang HW, Lee MS, Yip HK. Xenogeneic human umbilical cord-derived mesenchymal stem cells reduce mortality in rats with acute respiratory distress syndrome complicated by sepsis. Oncotarget 2017; 8: 45626-45642 [PMID: 28484089 DOI: 10.18632/oncotarget.17320]
- 32 Zhu H, Xiong Y, Xia Y, Zhang R, Tian D, Wang T, Dai J, Wang L, Yao H, Jiang H, Yang K, Liu E, Shi Y, Fu Z, Gao L, Zou L. Therapeutic Effects of Human Umbilical Cord-Derived Mesenchymal Stem Cells in Acute Lung Injury Mice. Sci Rep 2017; 7: 39889 [PMID: 28051154 DOI: 10.1038/srep39889]
- 33 Curley GF, Jerkic M, Dixon S, Hogan G, Masterson C, O'Toole D, Devaney J, Laffey JG. Cryopreserved, Xeno-Free Human Umbilical Cord Mesenchymal Stromal Cells Reduce Lung Injury Severity and Bacterial Burden in Rodent Escherichia coli-Induced Acute Respiratory Distress Syndrome. Crit Care Med 2017; 45: e202-e212 [PMID: 27861182 DOI: 10.1097/CCM.000000000002073]
- Chang Y, Park SH, Huh JW, Lim CM, Koh Y, Hong SB. Intratracheal administration of umbilical cord 34 blood-derived mesenchymal stem cells in a patient with acute respiratory distress syndrome. J Korean Med Sci 2014; 29: 438-440 [PMID: 24616596 DOI: 10.3346/jkms.2014.29.3.438]
- Moodley Y, Atienza D, Manuelpillai U, Samuel CS, Tchongue J, Ilancheran S, Boyd R, Trounson A. Human umbilical cord mesenchymal stem cells reduce fibrosis of bleomycin-induced lung injury. Am J Pathol 2009; 175: 303-313 [PMID: 19497992 DOI: 10.2353/ajpath.2009.080629]
- Xiang B, Chen L, Wang X, Zhao Y, Wang Y, Xiang C. Transplantation of Menstrual Blood-Derived Mesenchymal Stem Cells Promotes the Repair of LPS-Induced Acute Lung Injury. Int J Mol Sci 2017; 18 [PMID: 28346367 DOI: 10.3390/ijms18040689]
- 37 Wang L, Shi M, Tong L, Wang J, Ji S, Bi J, Chen C, Jiang J, Bai C, Zhou J, Song Y. Lung-Resident Mesenchymal Stem Cells Promote Repair of LPS-Induced Acute Lung Injury via Regulating the Balance of Regulatory T cells and Th17 cells. Inflammation 2019; 42: 199-210 [PMID: 30187337 DOI: 10.1007/s10753-018-0884-6]
- 38 Silva JD, Lopes-Pacheco M, Paz AHR, Cruz FF, Melo EB, de Oliveira MV, Xisto DG, Capelozzi VL, Morales MM, Pelosi P, Cirne-Lima E, Rocco PRM, Mesenchymal Stem Cells From Bone Marrow, Adipose Tissue, and Lung Tissue Differentially Mitigate Lung and Distal Organ Damage in Experimental Acute Respiratory Distress Syndrome. Crit Care Med 2018; 46: e132-e140 [PMID: 29116998 DOI: 10.1097/CCM.00000000002833
- 39 Su VY, Lin CS, Hung SC, Yang KY. Mesenchymal Stem Cell-Conditioned Medium Induces Neutrophil Apoptosis Associated with Inhibition of the NF-κB Pathway in Endotoxin-Induced Acute Lung Injury. Int J Mol Sci 2019: 20 [PMID: 31060326 DOI: 10.3390/ijms20092208]
- Mohammadipoor A, Antebi B, Batchinsky AI, Cancio LC. Therapeutic potential of products derived from mesenchymal stem/stromal cells in pulmonary disease. Respir Res 2018; 19: 218 [PMID: 30413158 DOI: 10.1186/s12931-018-0921-x
- Lee JH, Park J, Lee JW. Therapeutic use of mesenchymal stem cell-derived extracellular vesicles in acute 41 lung injury. Transfusion 2019; 59: 876-883 [PMID: 30383895 DOI: 10.1111/trf.14838]
- Abreu SC, Weiss DJ, Rocco PR. Extracellular vesicles derived from mesenchymal stromal cells: a 42 therapeutic option in respiratory diseases? Stem Cell Res Ther 2016; 7: 53 [PMID: 27075363 DOI: 10.1186/s13287-016-0317-0
- Monsel A, Zhu YG, Gudapati V, Lim H, Lee JW. Mesenchymal stem cell derived secretome and 43 extracellular vesicles for acute lung injury and other inflammatory lung diseases. Expert Opin Biol Ther 2016; 16: 859-871 [PMID: 27011289 DOI: 10.1517/14712598.2016.1170804]
- Liu FB, Lin Q, Liu ZW. A study on the role of apoptotic human umbilical cord mesenchymal stem cells in



bleomycin-induced acute lung injury in rat models. Eur Rev Med Pharmacol Sci 2016; 20: 969-982 [PMID: 27010158

- Chen J, Li Y, Hao H, Li C, Du Y, Hu Y, Li J, Liang Z, Li C, Liu J, Chen L. Mesenchymal Stem Cell 45 Conditioned Medium Promotes Proliferation and Migration of Alveolar Epithelial Cells under Septic Conditions In Vitro via the JNK-P38 Signaling Pathway. Cell Physiol Biochem 2015; 37: 1830-1846 [PMID: 26584283 DOI: 10.1159/000438545]
- 46 Ionescu L, Byrne RN, van Haaften T, Vadivel A, Alphonse RS, Rey-Parra GJ, Weissmann G, Hall A, Eaton F, Thébaud B. Stem cell conditioned medium improves acute lung injury in mice: in vivo evidence for stem cell paracrine action. Am J Physiol Lung Cell Mol Physiol 2012; 303: L967-L977 [PMID: 23023971 DOI: 10.1152/ajplung.00144.2011]
- 47 Zhao R, Su Z, Wu J, Ji HL. Serious adverse events of cell therapy for respiratory diseases: a systematic review and meta-analysis. Oncotarget 2017; 8: 30511-30523 [PMID: 28430622 DOI: 10.18632/oncotarget.15426]
- 48 Cheng SL, Lin CH, Yao CL. Mesenchymal Stem Cell Administration in Patients with Chronic Obstructive Pulmonary Disease: State of the Science. Stem Cells Int 2017; 2017: 8916570 [PMID: 28303154 DOI: 10.1155/2017/89165701
- Atluri S, Manchikanti L, Hirsch JA. Expanded Umbilical Cord Mesenchymal Stem Cells (UC-MSCs) as a Therapeutic Strategy in Managing Critically III COVID-19 Patients: The Case for Compassionate Use. Pain Physician 2020; 23: E71-E83 [PMID: 32214286]
- 50 Leng Z, Zhu R, Hou W, Feng Y, Yang Y, Han Q, Shan G, Meng F, Du D, Wang S, Fan J, Wang W, Deng L, Shi H, Li H, Hu Z, Zhang F, Gao J, Liu H, Li X, Zhao Y, Yin K, He X, Gao Z, Wang Y, Yang B, Jin R, Stambler I, Lim LW, Su H, Moskalev A, Cano A, Chakrabarti S, Min KJ, Ellison-Hughes G, Caruso C, Jin K, Zhao RC. Transplantation of ACE2 Mesenchymal Stem Cells Improves the Outcome of Patients with COVID-19 Pneumonia. Aging Dis 2020; 11: 216-228 [PMID: 32257537 DOI: 10.14336/AD.2020.0228]
- 51 Letko M, Marzi A, Munster V. Functional assessment of cell entry and receptor usage for SARS-CoV-2 and other lineage B betacoronaviruses. Nat Microbiol 2020; 5: 562-569 [PMID: 32094589 DOI: 10.1038/s41564-020-0688-y]
- 52 Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, Zhang L, Fan G, Xu J, Gu X, Cheng Z, Yu T, Xia J, Wei Y, Wu W, Xie X, Yin W, Li H, Liu M, Xiao Y, Gao H, Guo L, Xie J, Wang G, Jiang R, Gao Z, Jin Q, Wang J, Cao B. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. Lancet 2020; 395: 497-506 [PMID: 31986264 DOI: 10.1016/S0140-6736(20)30183-5]
- 53 Yuki K, Fujiogi M, Koutsogiannaki S. COVID-19 pathophysiology: A review. Clin Immunol 2020; 215: 108427 [PMID: 32325252 DOI: 10.1016/j.clim.2020.108427]
- 54 Xu Z, Shi L, Wang Y, Zhang J, Huang L, Zhang C, Liu S, Zhao P, Liu H, Zhu L, Tai Y, Bai C, Gao T, Song J, Xia P, Dong J, Zhao J, Wang FS. Pathological findings of COVID-19 associated with acute respiratory distress syndrome. Lancet Respir Med 2020; 8: 420-422 [PMID: 32085846 DOI: 10.1016/S2213-2600(20)30076-X
- 55 Colamartino ABL, Lemieux W, Bifsha P, Nicoletti S, Chakravarti N, Sanz J, Roméro H, Selleri S, Béland K, Guiot M, Tremblay-Laganière C, Dicaire R, Barreiro L, Lee DA, Verhoeyen E, Haddad E. Efficient and Robust NK-Cell Transduction With Baboon Envelope Pseudotyped Lentivector. Front Immunol 2019; 10: 2873 [PMID: 31921138 DOI: 10.3389/fimmu.2019.02873]
- 56 Ding Y, Wang H, Shen H, Li Z, Geng J, Han H, Cai J, Li X, Kang W, Weng D, Lu Y, Wu D, He L, Yao K. The clinical pathology of severe acute respiratory syndrome (SARS): a report from China. J Pathol 2003; 200: 282-289 [PMID: 12845623 DOI: 10.1002/path.1440]
- Ng DL, Al Hosani F, Keating MK, Gerber SI, Jones TL, Metcalfe MG, Tong S, Tao Y, Alami NN, Haynes 57 LM, Mutei MA, Abdel-Wareth L, Uyeki TM, Swerdlow DL, Barakat M, Zaki SR. Clinicopathologic, Immunohistochemical, and Ultrastructural Findings of a Fatal Case of Middle East Respiratory Syndrome Coronavirus Infection in the United Arab Emirates, April 2014. Am J Pathol 2016; 186: 652-658 [PMID: 26857507 DOI: 10.1016/j.ajpath.2015.10.024]
- 58 Tian S, Hu W, Niu L, Liu H, Xu H, Xiao SY. Pulmonary Pathology of Early-Phase 2019 Novel Coronavirus (COVID-19) Pneumonia in Two Patients With Lung Cancer. J Thorac Oncol 2020; 15: 700-704 [PMID: 32114094 DOI: 10.1016/j.jtho.2020.02.010]
- Tian S, Xiong Y, Liu H, Niu L, Guo J, Liao M, Xiao SY. Pathological study of the 2019 novel coronavirus disease (COVID-19) through postmortem core biopsies. Mod Pathol 2020; 33: 1007-1014 [PMID: 32291399 DOI: 10.1038/s41379-020-0536-x]
- Barton LM, Duval EJ, Stroberg E, Ghosh S, Mukhopadhyay S. COVID-19 Autopsies, Oklahoma, United States. Am J Clin Pathol 2020; 153: 725-733 [PMID: 32275742 DOI: 10.1093/ajcp/aqaa062]
- Magro C, Mulvey JJ, Berlin D, Nuovo G, Salvatore S, Harp J, Baxter-Stoltzfus A, Laurence J. 61 Complement associated microvascular injury and thrombosis in the pathogenesis of severe COVID-19 infection: A report of five cases. Transl Res 2020; 220: 1-13 [PMID: 32299776 DOI: 10.1016/j.trsl.2020.04.007]
- 62 Le Blanc K, Mougiakakos D. Multipotent mesenchymal stromal cells and the innate immune system. Nat Rev Immunol 2012; 12: 383-396 [PMID: 22531326 DOI: 10.1038/nri3209]
- 63 Jiang B, Yan L, Wang X, Li E, Murphy K, Vaccaro K, Li Y, Xu RH. Concise Review: Mesenchymal Stem Cells Derived from Human Pluripotent Cells, an Unlimited and Quality-Controllable Source for Therapeutic Applications. Stem Cells 2019; 37: 572-581 [PMID: 30561809 DOI: 10.1002/stem.2964]
- 64 Campagnoli C, Roberts IA, Kumar S, Bennett PR, Bellantuono I, Fisk NM. Identification of mesenchymal stem/progenitor cells in human first-trimester fetal blood, liver, and bone marrow. Blood 2001; 98: 2396-2402 [PMID: 11588036 DOI: 10.1182/blood.v98.8.2396]
- 65 Carlotti F, Zaldumbide A, Loomans CJ, van Rossenberg E, Engelse M, de Koning EJ, Hoeben RC. Isolated human islets contain a distinct population of mesenchymal stem cells. Islets 2010; 2: 164-173 [PMID: 21099310 DOI: 10.4161/isl.2.3.11449]
- Nandy SB, Mohanty S, Singh M, Behari M, Airan B. Fibroblast Growth Factor-2 alone as an efficient



inducer for differentiation of human bone marrow mesenchymal stem cells into dopaminergic neurons. J Biomed Sci 2014; 21: 83 [PMID: 25248378 DOI: 10.1186/s12929-014-0083-1]

- 67 Armiñán A, Gandía C, Bartual M, García-Verdugo JM, Lledó E, Mirabet V, Llop M, Barea J, Montero JA, Sepúlveda P. Cardiac differentiation is driven by NKX2.5 and GATA4 nuclear translocation in tissuespecific mesenchymal stem cells. Stem Cells Dev 2009; 18: 907-918 [PMID: 18983250 DOI: 10.1089/scd.2008.0292
- 68 Liu J, Wang Y, Wu Y, Ni B, Liang Z. Sodium butyrate promotes the differentiation of rat bone marrow mesenchymal stem cells to smooth muscle cells through histone acetylation. PLoS One 2014; 9: e116183 [PMID: 25548915 DOI: 10.1371/journal.pone.0116183]
- Spaggiari GM, Abdelrazik H, Becchetti F, Moretta L. MSCs inhibit monocyte-derived DC maturation and 69 function by selectively interfering with the generation of immature DCs: central role of MSCs-derived prostaglandin E2. Blood 2009; 113: 6576-6583 [PMID: 19398717 DOI: 10.1182/blood-2009-02-203943]
- 70 Ren G, Zhao X, Zhang L, Zhang J, L'Huillier A, Ling W, Roberts AI, Le AD, Shi S, Shao C, Shi Y. Inflammatory cytokine-induced intercellular adhesion molecule-1 and vascular cell adhesion molecule-1 in mesenchymal stem cells are critical for immunosuppression. J Immunol 2010; 184: 2321-2328 [PMID: 20130212 DOI: 10.4049/jimmunol.0902023]
- 71 Kriegsmann K, Kriegsmann M, Cremer M, Schmitt M, Dreger P, Goldschmidt H, Müller-Tidow C, Hundemer M. Cell-based immunotherapy approaches for multiple myeloma. Br J Cancer 2019; 120: 38-44 [PMID: 30518815 DOI: 10.1038/s41416-018-0346-9]
- 72 Patel SA, Meyer JR, Greco SJ, Corcoran KE, Bryan M, Rameshwar P. Mesenchymal stem cells protect breast cancer cells through regulatory T cells: role of mesenchymal stem cell-derived TGF-beta. J Immunol 2010; 184: 5885-5894 [PMID: 20382885 DOI: 10.4049/jimmunol.0903143]
- 73 Sotiropoulou PA, Perez SA, Gritzapis AD, Baxevanis CN, Papamichail M, Interactions between human mesenchymal stem cells and natural killer cells. Stem Cells 2006; 24: 74-85 [PMID: 16099998 DOI: 10.1634/stemcells.2004-0359]
- Djouad F, Charbonnier LM, Bouffi C, Louis-Plence P, Bony C, Apparailly F, Cantos C, Jorgensen C, Noël D. Mesenchymal stem cells inhibit the differentiation of dendritic cells through an interleukin-6-dependent mechanism. Stem Cells 2007; 25: 2025-2032 [PMID: 17510220 DOI: 10.1634/stemcells.2006-0548]
- Najar M, Rouas R, Raicevic G, Boufker HI, Lewalle P, Meuleman N, Bron D, Toungouz M, Martiat P, Lagneaux L. Mesenchymal stromal cells promote or suppress the proliferation of T lymphocytes from cord blood and peripheral blood: the importance of low cell ratio and role of interleukin-6. Cytotherapy 2009; 11: 570-583 [PMID: 19565371 DOI: 10.1080/14653240903079377]
- Chabannes D, Hill M, Merieau E, Rossignol J, Brion R, Soulillou JP, Anegon I, Cuturi MC. A role for 76 heme oxygenase-1 in the immunosuppressive effect of adult rat and human mesenchymal stem cells. Blood 2007; 110: 3691-3694 [PMID: 17684157 DOI: 10.1182/blood-2007-02-075481]
- Nasef A, Mazurier C, Bouchet S, François S, Chapel A, Thierry D, Gorin NC, Fouillard L. Leukemia inhibitory factor: Role in human mesenchymal stem cells mediated immunosuppression. Cell Immunol 2008; 253: 16-22 [PMID: 18639869 DOI: 10.1016/j.cellimm.2008.06.002]
- Rizzo R, Campioni D, Stignani M, Melchiorri L, Bagnara GP, Bonsi L, Alviano F, Lanzoni G, Moretti S, Cuneo A. Lanza F. Baricordi OR. A functional role for soluble HLA-G antigens in immune modulation mediated by mesenchymal stromal cells. Cytotherapy 2008; 10: 364-375 [PMID: 18574769 DOI: 10.1080/14653240802105299
- 79 Tyndall A, van Laar JM. Stem cell transplantation and mesenchymal cells to treat autoimmune diseases. Presse Med 2016; 45: e159-e169 [PMID: 27256975 DOI: 10.1016/j.lpm.2016.05.002]
- Akiyama K, Chen C, Wang D, Xu X, Qu C, Yamaza T, Cai T, Chen W, Sun L, Shi S. Mesenchymal-stemcell-induced immunoregulation involves FAS-ligand-/FAS-mediated T cell apoptosis. Cell Stem Cell 2012; 10: 544-555 [PMID: 22542159 DOI: 10.1016/j.stem.2012.03.007]
- Wang H, Qi F, Dai X, Tian W, Liu T, Han H, Zhang B, Li H, Zhang Z, Du C. Requirement of B7-H1 in mesenchymal stem cells for immune tolerance to cardiac allografts in combination therapy with rapamycin. Transpl Immunol 2014; 31: 65-74 [PMID: 24978830 DOI: 10.1016/j.trim.2014.06.005]
- 82 Gazdic M, Markovic BS, Arsenijevic A, Jovicic N, Acovic A, Harrell CR, Fellabaum C, Djonov V, Arsenijevic N, Lukic ML, Volarevic V. Crosstalk between mesenchymal stem cells and T regulatory cells is crucially important for the attenuation of acute liver injury. Liver Transpl 2018; 24: 687-702 [PMID: 29500914 DOI: 10.1002/lt.250491
- Luz-Crawford P, Kurte M, Bravo-Alegría J, Contreras R, Nova-Lamperti E, Tejedor G, Noël D, Jorgensen C, Figueroa F, Djouad F, Carrión F. Mesenchymal stem cells generate a CD4+CD25+Foxp3+ regulatory T cell population during the differentiation process of Th1 and Th17 cells. Stem Cell Res Ther 2013; 4:65 [PMID: 23734780 DOI: 10.1186/scrt216]
- 84 Mohammadpour H, Pourfathollah AA, Zarif MN, Tahoori MT. TNF-α modulates the immunosuppressive effects of MSCs on dendritic cells and T cells. Int Immunopharmacol 2015; 28: 1009-1017 [PMID: 26303769 DOI: 10.1016/j.intimp.2015.07.045]
- Yan Z, Zhuansun Y, Liu G, Chen R, Li J, Ran P. Mesenchymal stem cells suppress T cells by inducing 85 apoptosis and through PD-1/B7-H1 interactions. Immunol Lett 2014; 162: 248-255 [PMID: 25281059 DOI: 10.1016/j.imlet.2014.09.013]
- 86 Chinnadurai R, Copland IB, Patel SR, Galipeau J. IDO-independent suppression of T cell effector function by IFN-γ-licensed human mesenchymal stromal cells. J Immunol 2014; 192: 1491-1501 [PMID: 24403533 DOI: 10.4049/jimmunol.1301828]
- 87 Jang IK, Yoon HH, Yang MS, Lee JE, Lee DH, Lee MW, Kim DS, Park JE. B7-H1 inhibits T cell proliferation through MHC class II in human mesenchymal stem cells. Transplant Proc 2014: 46: 1638-1641 [PMID: 24935340 DOI: 10.1016/j.transproceed.2013.12.059]
- Di Nicola M. Carlo-Stella C. Magni M. Milanesi M. Longoni PD. Matteucci P. Grisanti S. Gianni AM. 88 Human bone marrow stromal cells suppress T-lymphocyte proliferation induced by cellular or nonspecific mitogenic stimuli. Blood 2002; 99: 3838-3843 [PMID: 11986244 DOI: 10.1182/blood.v99.10.3838]



- 89 Glennie S, Soeiro I, Dyson PJ, Lam EW, Dazzi F. Bone marrow mesenchymal stem cells induce division arrest anergy of activated T cells. *Blood* 2005; 105: 2821-2827 [PMID: 15591115 DOI: 10.1182/blood-2004-09-3696]
- 90 Aggarwal S, Pittenger MF. Human mesenchymal stem cells modulate allogeneic immune cell responses. Blood 2005; 105: 1815-1822 [PMID: 15494428 DOI: 10.1182/blood-2004-04-1559]
- 91 Obermajer N, Popp FC, Soeder Y, Haarer J, Geissler EK, Schlitt HJ, Dahlke MH. Conversion of Th17 into IL-17A(neg) regulatory T cells: a novel mechanism in prolonged allograft survival promoted by mesenchymal stem cell-supported minimized immunosuppressive therapy. *J Immunol* 2014; **193**: 4988-4999 [PMID: 25305313 DOI: 10.4049/jimmunol.1401776]
- 92 Lo Sicco C, Reverberi D, Balbi C, Ulivi V, Principi E, Pascucci L, Becherini P, Bosco MC, Varesio L, Franzin C, Pozzobon M, Cancedda R, Tasso R. Mesenchymal Stem Cell-Derived Extracellular Vesicles as Mediators of Anti-Inflammatory Effects: Endorsement of Macrophage Polarization. *Stem Cells Transl Med* 2017; 6: 1018-1028 [PMID: 28186708 DOI: 10.1002/sctm.16-0363]
- 93 Takizawa N, Okubo N, Kamo M, Chosa N, Mikami T, Suzuki K, Yokota S, Ibi M, Ohtsuka M, Taira M, Yaegashi T, Ishisaki A, Kyakumoto S. Bone marrow-derived mesenchymal stem cells propagate immunosuppressive/anti-inflammatory macrophages in cell-to-cell contact-independent and -dependent manners under hypoxic culture. *Exp Cell Res* 2017; 358: 411-420 [PMID: 28712928 DOI: 10.1016/j.yexcr.2017.07.014]
- 94 Spaggiari GM, Capobianco A, Becchetti S, Mingari MC, Moretta L. Mesenchymal stem cell-natural killer cell interactions: evidence that activated NK cells are capable of killing MSCs, whereas MSCs can inhibit IL-2-induced NK-cell proliferation. *Blood* 2006; 107: 1484-1490 [PMID: 16239427 DOI: 10.1182/blood-2005-07-2775]
- 95 Raffaghello L, Bianchi G, Bertolotto M, Montecucco F, Busca A, Dallegri F, Ottonello L, Pistoia V. Human mesenchymal stem cells inhibit neutrophil apoptosis: a model for neutrophil preservation in the bone marrow niche. *Stem Cells* 2008; 26: 151-162 [PMID: 17932421 DOI: 10.1634/stemcells.2007-0416]
- 96 Jiang XX, Zhang Y, Liu B, Zhang SX, Wu Y, Yu XD, Mao N. Human mesenchymal stem cells inhibit differentiation and function of monocyte-derived dendritic cells. *Blood* 2005; 105: 4120-4126 [PMID: 15692068 DOI: 10.1182/blood-2004-02-0586]
- 97 Putra A, Ridwan FB, Putridewi AI, Kustiyah AR, Wirastuti K, Sadyah NAC, Rosdiana I, Munir D. The Role of TNF-α induced MSCs on Suppressive Inflammation by Increasing TGF-β and IL-10. Open Access Maced J Med Sci 2018; 6: 1779-1783 [PMID: 30455748 DOI: 10.3889/oamjms.2018.404]
- 98 Xiao S, Huang G, Wei Z, Nie K, Liu Z, Deng C, Wang D. IL-10 Gene-Modified Human Amniotic Mesenchymal Stem Cells Augment Regenerative Wound Healing by Multiple Synergistic Effects. *Stem Cells Int* 2019; 2019: 9158016 [PMID: 31281390 DOI: 10.1155/2019/9158016]
- 99 Simonson OE, Mougiakakos D, Heldring N, Bassi G, Johansson HJ, Dalén M, Jitschin R, Rodin S, Corbascio M, El Andaloussi S, Wiklander OP, Nordin JZ, Skog J, Romain C, Koestler T, Hellgren-Johansson L, Schiller P, Joachimsson PO, Hägglund H, Mattsson M, Lehtiö J, Faridani OR, Sandberg R, Korsgren O, Krampera M, Weiss DJ, Grinnemo KH, Le Blanc K. In Vivo Effects of Mesenchymal Stromal Cells in Two Patients With Severe Acute Respiratory Distress Syndrome. *Stem Cells Transl Med* 2015; 4: 1199-1213 [PMID: 26285659 DOI: 10.5966/sctm.2015-0021]
- 100 Rubenfeld GD, Caldwell E, Peabody E, Weaver J, Martin DP, Neff M, Stern EJ, Hudson LD. Incidence and outcomes of acute lung injury. *N Engl J Med* 2005; **353**: 1685-1693 [PMID: 16236739 DOI: 10.1056/NEJMoa050333]
- 101 Golchin A, Seyedjafari E, Ardeshirylajimi A. Mesenchymal Stem Cell Therapy for COVID-19: Present or Future. Stem Cell Rev Rep 2020; 16: 427-433 [PMID: 32281052 DOI: 10.1007/s12015-020-09973-w]
- 102 Liu S, Peng D, Qiu H, Yang K, Fu Z, Zou L. Mesenchymal stem cells as a potential therapy for COVID-19. Stem Cell Res Ther 2020; 11: 169 [PMID: 32366290 DOI: 10.1186/s13287-020-01678-8]
- 103 Rogers CJ, Harman RJ, Bunnell BA, Schreiber MA, Xiang C, Wang FS, Santidrian AF, Minev BR. Rationale for the clinical use of adipose-derived mesenchymal stem cells for COVID-19 patients. *J Transl Med* 2020; 18: 203 [PMID: 32423449 DOI: 10.1186/s12967-020-02380-2]
- 104 Tsuchiya A, Takeuchi S, Iwasawa T, Kumagai M, Sato T, Motegi S, Ishii Y, Koseki Y, Tomiyoshi K, Natsui K, Takeda N, Yoshida Y, Yamazaki F, Kojima Y, Watanabe Y, Kimura N, Tominaga K, Kamimura H, Takamura M, Terai S. Therapeutic potential of mesenchymal stem cells and their exosomes in severe novel coronavirus disease 2019 (COVID-19) cases. *Inflamm Regen* 2020; 40: 14 [PMID: 32582401 DOI: 10.1186/s41232-020-00121-y]
- 105 Raza SS, Seth P, Khan MA. 'Primed' Mesenchymal Stem Cells: a Potential Novel Therapeutic for COVID19 Patients. Stem Cell Rev Rep 2020; : [PMID: 32592163 DOI: 10.1007/s12015-020-09999-0]
- 106 Qin H, Zhao A. Mesenchymal stem cell therapy for acute respiratory distress syndrome: from basic to clinics. *Protein Cell*2020 [PMID: 32519302 DOI: 10.1007/s13238-020-00738-2]
- 107 Zhang Y, Ding J, Ren S, Wang W, Yang Y, Li S, Meng M, Wu T, Liu D, Tian S, Tian H, Chen S, Zhou C. Intravenous infusion of human umbilical cord Wharton's jelly-derived mesenchymal stem cells as a potential treatment for patients with COVID-19 pneumonia. *Stem Cell Res Ther* 2020; 11: 207 [PMID: 32460839 DOI: 10.1186/s13287-020-01725-4]
- 108 Guo Z, Chen Y, Luo X, He X, Zhang Y, Wang J. Administration of umbilical cord mesenchymal stem cells in patients with severe COVID-19 pneumonia. *Crit Care* 2020; 24: 420 [PMID: 32653043 DOI: 10.1186/s13054-020-03142-8]
- 109 O'Driscoll L. Extracellular vesicles from mesenchymal stem cells as a Covid-19 treatment. Drug Discov Today 2020; 25: 1124-1125 [PMID: 32387262 DOI: 10.1016/j.drudis.2020.04.022]
- 110 Sengupta V, Sengupta S, Lazo A, Woods P, Nolan A, Bremer N. Exosomes Derived from Bone Marrow Mesenchymal Stem Cells as Treatment for Severe COVID-19. *Stem Cells Dev* 2020; 29: 747-754 [PMID: 32380908 DOI: 10.1089/scd.2020.0080]



Published by Baishideng Publishing Group Inc 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA Telephone: +1-925-3991568 E-mail: bpgoffice@wjgnet.com Help Desk: https://www.f6publishing.com/helpdesk https://www.wjgnet.com

