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ABOUT COVER

Editorial Board Member of *World Journal of Stem Cells*, Dr. Alessandra Pelagalli is a Senior Researcher of veterinary physiology in the Department of Advanced Biomedical Sciences at the University of Naples. Having completed her Pharmacy Degree from the University of Naples in 1991, Dr. Pelagalli continued her postgraduate training and received her PhD in 1996. She became a Young Researcher at the University of Naples in 1999, working in animal platelet physiology and biochemistry. Her current research interests and publications focus on the roles and behavior of bone marrow mesenchymal stem cells in the differentiation processes after stimulation, water channel proteins in cell migration, and aquaporins in various tissues, such as gut and the male reproductive tract. (L-Editor: Filopodia)

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ORIGINAL ARTICLE

Basic Study

Effect of conditioned medium from neural stem cells on glioma progression and its protein expression profile analysis

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Abstract

BACKGROUND

Emerging evidence suggests that the spread of glioma to the subventricular zone (SVZ) is closely related to glioma recurrence and patient survival. Neural stem cells (NSCs) are the main cell type in the SVZ region and exhibit tumor-homing ability.

AIM

To evaluate the effects of conditioned medium (CM) derived from SVZ NSCs on the cancer-related behaviors of glioma cells.

METHODS

The characteristics of SVZ hNSCs were identified by immunofluorescence. The normoxic-hNSC-CM and hypoxic-hNSC-CM (3% O₂, oxygen-glucose deprived [OGD] culturing) were collected from 80%-90% confluent SVZ NSCs in sterile conditions. The CCK8 and Transwell assays were used to compare and evaluate the effects of normoxic-CM and hypoxic-CM on glioma proliferation and invasion. Then proteins secreted from SVZ NSCs into the CM were investigated by mass spectrometry, and the potential effects of candidate protein NCAN in the regulation of glioma progression were examined by CCK8 and Transwell assays.

RESULTS

The CM from SVZ NSCs significantly increased the proliferation and invasion of glioma cells, particularly the CM from OGD NSCs induced under hypoxic conditions. Furthermore, the secreted protein neurocan (NCAN) in CM from OGD NSCs was identified by proteomic analysis. NCAN was expressed in glioma

Data sharing statement: The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

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cells and played regulatory roles in mediating the progression of glioma cells mainly *via* the Rho/Rho-associated protein kinase pathway.

CONCLUSION

Our study identified a potential interactive mechanism between SVZ NSCs and glioma cells, in which SVZ NSCs promote glioma progression *via* the secreted protein NCAN. These findings suggested that exploring the CM derived from cells could be a novel strategy for optimizing treatments and that NCAN derived from SVZ NSCs may be a potential new target in glioma progression.

Key Words: Neural stem cells; Glioma; Conditioned medium; NCAN; Hypoxia; Protein

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Core Tip: The cell communications or crosstalk between glioma cells and non-glioma cells in the tumor microenvironment plays dominant roles in tumor therapy. The subventricular zone (SVZ) neural stem cells (NSCs) have been shown to be closely associated with glioma progression and recurrence. Interestingly, the conditioned medium (CM) from SVZ NSCs significantly promoted glioma cell proliferation and invasion, with oxygen-glucose deprived NSC-CM showing enhanced effects. Furthermore, the secreted protein NCAN presented in oxygen-glucose deprived NSC-CM played important roles in the progression of glioma cells mainly *via* the Rho/Rho-associated protein kinase pathway. Overall, this study provided important insights into the potential therapeutic application of NCAN derived from SVZ NSCs in the treatment of glioma.

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INTRODUCTION

Glioma is a common type of primary malignant tumor found in the brain. Glioblastoma multiforme (GBM) is the most aggressive type of primary brain tumor and is associated with a high mortality rate. Less than 6% of patients with GBM survive beyond 5 years, and the median overall survival is approximately 15-23 mo^[1]. Currently, there are no effective treatment options for GBM. Despite extensive studies of novel therapeutic approaches, most strategies still lead to drug resistance or eventual tumor recurrence^[2,3]. Recent studies have suggested that GBMs are heterogeneous, characterized by genetic, epigenetic, and molecular variations among cells^[2-4]. These features are mainly affected by the tumor microenvironment (TME) surrounding the GBM. Thus, GBM growth occurs before the cells are able to communicate with or be manipulated by other cells (non-glioma cells) *via* secreted cytokines, growth factors, and chemokines in the TME, thereby promoting tumor progression and drug resistance^[4-6].

GBM cells not only interact with immune cells, including microglia/macrophages, to alter their phenotypes and support tumor growth but also communicate with normal brain cells, including neurons, astrocytes, and endothelial cells, to create a favorable microenvironment for tumor proliferation^[7-13]. Chen *et al*^[12] found that PTEN deficiency in GBM increases macrophage infiltration *via* the YAP1-LOXb1 integrin-PYK2 axis. Additionally, the infiltrated macrophages secrete SPP1 to sustain glioma cell survival and stimulate angiogenesis. Venkatesh *et al*^[11] showed that active neurons can promote glioma proliferation and growth. They used conditioned medium (CM) from optogenetically stimulated cortical slices in co-cultures with glioma cells and found that this CM significantly promoted cell proliferation. They identified synaptic protein neuroligin-3 present in active CM using a quantitative mass spectrometry approach. Moreover, Gao *et al*^[14] showed that the communication mediated by

extracellular vesicles (EVs) derived from glioma cells increased the synaptic activity of neurons. Blocking EV release caused a reduction in glioma growth *in vivo*. Henrik Heiland *et al*^[10] also reported a distinct transcriptional phenotype of reactive astrocytes from glioblastoma in the tumor environment. They revealed a complex interaction of tumor-associated astrocytes and microglia cells, which induced an immunosuppressive microenvironment. Griveau *et al*^[9] showed that distinct glioma-vascular microenvironmental interactions may regulate glial-encoded pathways. They found that targeted Wnt7a/7b deletion or pharmacologic Wnt inhibition blocked Olig²⁺ glioma single-cell vessel co-option and enhanced responses to temozolomide. Thus, these findings indicated that glioblastoma involves multiple modes of cell communication and that the multidimensional cell communications or crosstalk between glioma cells and non-glioma brain cells in the TME may play dominant roles in glioma therapy^[14,15].

Notably, the growth, invasion, and recurrence of GBM are known to be related to the subventricular zone (SVZ) of the lateral ventricles^[16-18]. GBM contacting the SVZ is thought to have a higher risk of recurrence. Moreover, the involvement of the SVZ in glioma is associated with more rapid progression and further decrease in the overall survival of patients with GBM^[16-18]. The SVZ is one of two dominating neurogenic zones located near neural stem cells (NSCs) in the adult brain. However, communication or crosstalk between neural stem cells in the SVZ (SVZ NSCs) and GBM *via* the TME remains poorly understood, especially the roles of SVZ NSCs in regulating glioma progression are still unclear.

Accordingly, in this study, we investigated the functions of SVZ NSCs in gliomas using CM derived from SVZ NSCs in cocultures with glioma cells. We also explored the potential functional molecules present in the CM by proteomics analysis. Our results provided important insights into the mechanisms of glioma invasion and suggested potential targets in the treatment of GBM.

MATERIALS AND METHODS

Culture of SVZ NSCs and glioma cells

SVZ NSCs obtained from the human fetal brain tissue were kept in our laboratory (approved by the Institutional Review Board of Zhongda Hospital Southeast University), as previously described^[19-21]. NSCs were cultured in serum-free DMEM/F12 medium (Gibco) supplemented with 2% B27 (Gibco), 20 ng/mL basic FGF (bFGF, PeproTech), 20 ng/mL EGF (PeproTech), 2 nmol/L L-glutamine (Invitrogen), 2 µg/mL heparin sodium, and 1% penicillin-streptomycin (Gibco), and incubated at 37 °C in an incubator containing 5% CO₂. Fresh medium was changed every 3 d, and cells were passaged at the 7th day. Neurospheres were detached to single cells with Accutase Dissociation Reagent (Sigma). In brief, cCM was collected from 80%-90% confluent SVZ NSCs in sterile conditions after culturing for 3 d, sequentially centrifuged to remove intact cells, and then filtered using a filter unit (Millipore) with a 0.22 µm membrane to remove cell debris. The concentration of CM was qualified by BCA method, and the same concentrations (from 1.5 µg/uL to 2.5 µg/uL) of normoxic-CM and hypoxic-CM were used for subsequent experiments.

Human glioma cell lines U87 MG and U251 MG were preserved in our laboratory, obtained from American Type Culture Collection. Glioma cells were maintained at 37 °C in 5% CO₂ in DMEM (Gibco) supplemented with 10% fetal bovine serum (FBS, Hyclone) and 1% penicillin/streptomycin (Gibco). Fresh medium was changed every 2 d, and cells were passaged at the 4th day.

NSC identification and oxygen-glucose deprivation culturing

To evaluate the characteristic of NSCs, cells were incubated in a 24 well plate coated with laminin (Sigma) at 5 × 10⁴ cells per well with complete medium. To induce cell differentiation, NSCs were dissociated with Accutase and cultured in differentiation medium (DMEM/F12 supplemented with 2% FBS and 1% Penicillin-Streptomycin, without growth factors) for 7 d. For immunofluorescence, cells were fixed in 4% paraformaldehyde, followed by Triton X-100 (Sigma), and then blocked in 10% donkey serum (Jackson). Then cells were incubated with different primary antibodies: Rabbit anti-Sox2 (1:500, Abcam), anti-Musashi1 (1:500, Abcam), mouse anti-Nestin (1:500, Abcam), anti-beta III-tubulin (Tuj1, 1:400, Abcam), goat anti-Myelin oligodendrocyte glycoprotein (1:400, Abcam), and rabbit anti-GFAP (1:400, Bioss) overnight. Next day, the secondary antibodies donkey anti-rabbit-Alexa Fluor 647/488, anti-goat-Alexa Fluor 555/488(1:600, Abcam), or donkey anti-mouse-Alexa Fluor 488 (1:600, Jackson)

were added. Finally, following staining with DAPI, the slides were cover slipped for capturing on a fluorescence microscope.

For OGD culturing, SVZ NSCs were incubated in the flask for 3 d. After new fresh medium was changed, cells were transferred into a CO₂/O₂ trigas incubator (Thermo) containing 3% O₂ and 5% CO₂ at 37 °C. Oxygen and CO₂ contents were continuously maintained at a constant level by the tri-gas incubator for 3 d. Then cells and CM were collected. To confirm the OGD-NSCs model *in vitro*, the proteins of cells were extracted and collected for Western blot analysis.

Cell viability test

The glioma cells were seeded in 96-well plates for cell viability analysis. The cells were cultured with fresh medium, different CM of SVZ NSCs, or recombinant protein NCAN of different concentrations for 24-48 h, and then 10 µL CCK-8 (KeyGen) was added into each well. After incubation for 1-4 h, absorbance value for each well was calorimetrically determined. This experiment was repeated at least three times.

Cell invasion assay

U87 MG and U251 MG cells were added into the upper chamber of the 8 µm pore polycarbonate membrane (Transwell, Corning) at 5×10^4 cells and 2×10^4 cells per well with serum-free medium in the 24-well plates, and 800 µL of complete medium was added into the lower chamber to cultivate for 12-16 h. The 8 µm pore polycarbonate membrane was pre-coated with Matrigel (BD Biosciences). Then the liquid in the upper chamber was discarded, and the pores were fixed with 4% paraformaldehyde for 30 min, followed by staining with 0.1% crystal violet for 30 min. The cells that had migrated across the membrane were observed and photographed in six independent fields for each well under an inverted microscope.

Protein analysis by liquid chromatography mass spectrometry

CM was collected from normoxic and hypoxic SVZ NSCs cultured in serum-free complete medium after incubation for 3 d. The proteins were detected by Label Free Quantitative proteomics technology (Shanghai Applied Protein Technology). The process mainly includes protein extraction, peptide digestion, chromatographic fractionation, data collection by LC-MS/MS, protein identification and quantitative analysis, screening and cluster analysis of differentially expressed proteins, functional annotation, and pathway analysis. Proteins that were up- or down- regulated more than 1.5-fold ($P < 0.05$) in hypoxic and normoxic NSC-CM were further analyzed. Blast2Go (<https://www.blast2go.com/com/>) was used for functional annotation in the Gene Ontology (GO) database (<http://www.geneontology.org>) on all proteins identified in this project, and then GO function enrichment analysis was performed on differentially expressed proteins by Fisher's exact test method. The top 20 enriched biological pathways in the GO analysis were selected.

Western blot analysis

Total protein was extracted from glioma cells with lysis buffer (SD-001, Invent), and quantified with a BCA protein assay kit (Pierce). To perform Western blot analysis, equal amount of protein (20-40 µg) of each sample was subjected to sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE, Solarbio) and transferred onto polyvinylidene difluoride membranes (PVDF, Millipore). PVDF was blocked with TBST containing 5% non-fat milk at room temperature for 1h, and then incubated with diluted primary antibodies at 4°C overnight, including anti-HIF1α (1:1000, Proteintech), anti-NCAN (1:1000, Abcam), anti-RhoA and anti-Rho-associated protein kinase (ROCK) (1:1000, Bioss), anti-β-actin(1:5000, Jackson), and anti-GAPDH (1:5000, Proteintech). The next day, the membranes were incubated with horseradish peroxidase conjugated goat anti rabbit or rabbit anti-goat secondary antibody (1:20000, Jackson) at room temperature for 1h, and then reacted with an enhanced ECL substrate (Pierce). Beta-actin was used as a control according to the protocol.

Statistical analysis

Data analyses and statistics were performed with GraphPad Prism version 8. The results are presented as the mean ± standard deviation ($x \pm s$) of at least three independent experiments. Comparisons were performed by *t*-tests or ANOVA analysis. A P value < 0.05 was considered statistically significant, with $P < 0.05$ as significant and $P < 0.01$ as very significant.

RESULTS

Optimization of CM derived from normoxic and OGD preconditioning of SVZ NSCs

SVZ NSCs were cultured in serum-free complete medium, and rapidly formed nonadherent neurospheres (Figure 1A). NSC special markers, including Nestin, SOX2, and Msil1, were identified by immunofluorescence (Figure 1B). After differentiation, NSCs were incubated in differentiation medium and showed significant expression of Tuj1 (a neuron marker), GFAP (an astrocyte marker), and myelin oligodendrocyte glycoprotein (an oligodendrocyte marker) by immunofluorescence (Figure 1C). These results identified the characteristics of NSCs. After culturing in hypoxia (3% O₂, OGD conditions), the OGD NSCs showed significant overexpression of the hypoxic marker HIF-1α (Figure 1D). Subsequently, we collected the CM derived from normoxic and OGD preconditioned SVZ NSCs *via* centrifuging and filtration to enable us to discard cells and cell debris.

NSC-derived CM promotes glioma proliferation and invasion

Next, we analyzed the effects of CM derived from SVZ NSCs on glioma proliferation and compared the results for normoxic-CM and hypoxic-CM using CCK8 assays. Our results showed that the rate of glioma cell proliferation was increased in the NSC-CM group compared with that in the control group, and the effect of OGD-CM was significantly superior to that of normoxic-CM ($P < 0.05$; Figure 2A). These findings indicated that OGD-NSC-CM further improved glioma cell activity. Moreover, Transwell invasion experiments revealed that NSC-CM promoted the invasion of tumor cells compared with that of control cells (Figure 2B-E), particularly in the OGD-CM group. These results showed that the proliferation and invasion of glioma cells were improved after co-incubation of NSC-CM with U87 and U251 glioma cells. Furthermore, hypoxia enhanced the effects of NSC-CM in glioma cells. Taken together, these findings confirmed the roles of NSC-CM in promoting glioma cell proliferation and invasion.

Identification of proteins in CM derived from SVZ NSCs by mass spectrometry

Next, proteins secreted from SVZ NSCs into the CM were investigated by protein mass spectrometry. The signal intensities of proteins from normoxic and hypoxic CM were detected and analyzed by SDS-PAGE. Proteomic analysis was performed with three pairs of biological replicates from SVZ NSC-CM, and OGD-NSC-CM was compared with normoxic NSC-CM. The experimental workflow was performed as shown in Figure 3A, and approximately 73–98 proteins were identified in each sample (Figure 3B). Proteins with a fold change of at least 1.5 and P value of less than 0.05 for the comparison between hypoxic and normoxic CM were chosen for further analysis. The results showed that two proteins were upregulated (TSPYL2 and NCAN) and eight were downregulated (PTPRZ1, HIST1H2AJ, CLPB, CST3, POPDC3, CAT, CLU, and LGALS3BP) in the CM of hypoxic NSCs compared with that in normoxic NSCs (Figure 3C).

Next, GO analysis was used to determine the top 20 enriched biological processes involving the differentially expressed proteins (Figure 3D). The biological process domain was enriched in the terms of cellular process, metabolic process, biological regulation, and regulation of biological process; the molecular function domain was enriched in the terms of binding and protein binding; moreover, the cellular component domain was enriched in the terms of cell, cell part, organelle, and organelle part (Figure 3E). Furthermore, the upregulated proteins in the hypoxic NSC-CM group were related to the progression of gliomas. The two upregulated proteins identified by MS included the nucleosome assembly protein TSPYL2, and the chondroitin sulfate proteoglycan protein NCAN. TSPYL2 is characterized by a predicted nucleosome assembly protein domain for nucleosome remodeling and gene expression regulation and functions as a tumor suppressor, cell growth regulator, and DNA damage response mediator. NCAN, which is primarily expressed in the central nervous system, accumulates in the matrix surrounding axonal bundles and neuronal cells and is involved in the modulation of cell adhesion, invasion, and migration through interactions with a variety of matrix or transmembrane molecules. Thus, we next evaluated the potential effects of the candidate protein NCAN in the regulation of glioma progression.

NCAN protein accelerates the proliferation and invasion of glioma cells

Based on the above findings, we next evaluated the potential roles of NCAN in glioma progression by incubating glioma cells with different concentrations (1–100 ng/mL) of

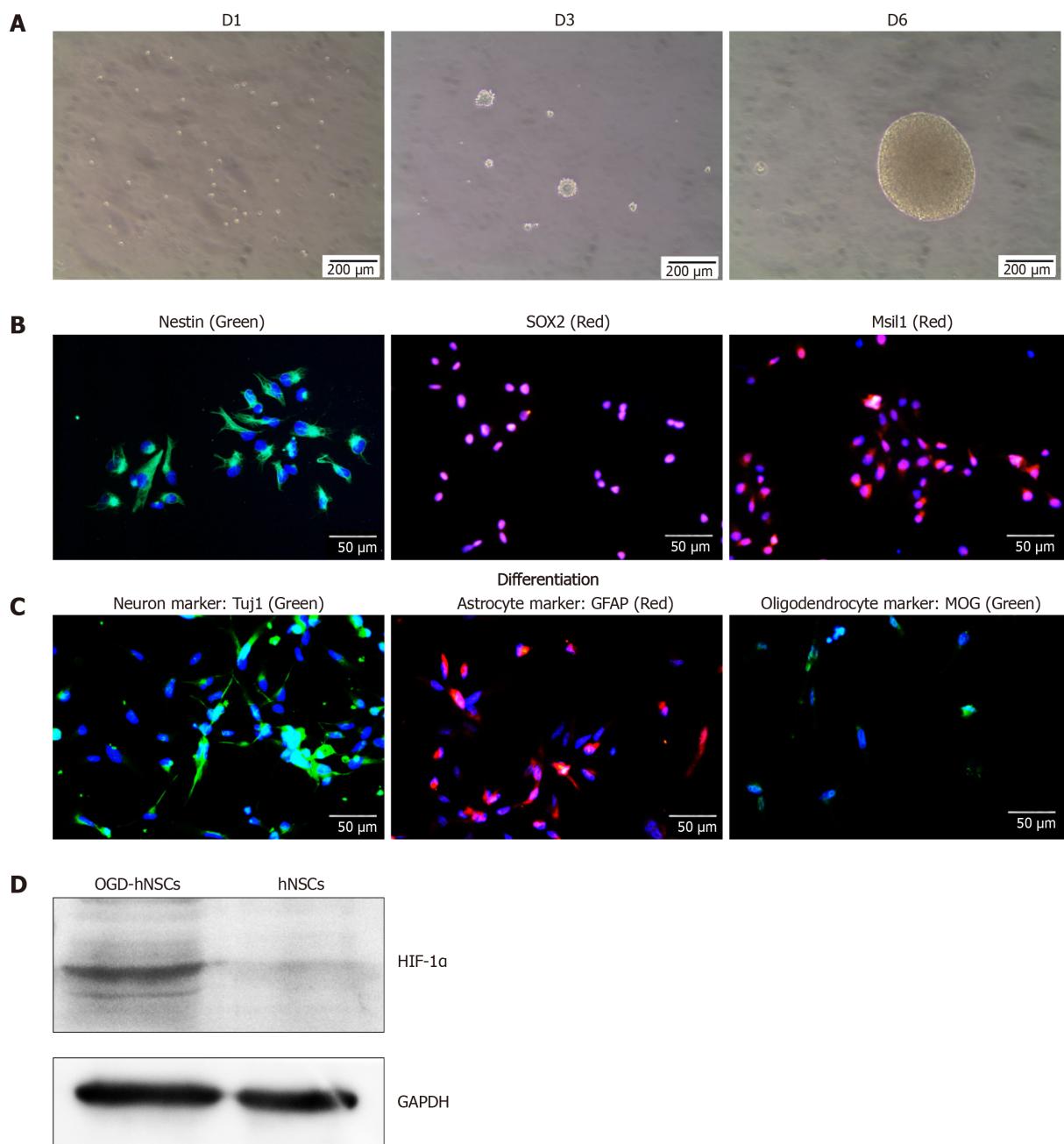


Figure 1 Characteristics of human neural stem cells. A: The proliferation morphology of human neural stem cells (hNSCs) cultured *in vitro* from Day 1 to Day 6. The hNSCs presented a neurosphere morphology; B: Immunofluorescence for hNSCs specific markers, including Nestin, SOX2, and Msil1. DAPI indicated the cell nucleus; C: The differentiation markers of hNSCs, including Tuj1 (a neuron marker), GFAP (an astrocyte marker), and MOG (an oligodendrocyte marker); D: Western blot analysis showing that after culturing in hypoxic (3% O₂) condition, oxygen-glucose deprived-hNSCs overexpressed the hypoxic marker HIF-1 α . hNSCs: Human neural stem cells; Tuj1: β -tubulin III; GFAP: Glial fibrillary acidic protein; MOG: Myelin oligodendrocyte glycoprotein.

recombinant human protein NCAN (rhNCAN). The results showed that rhNCAN increased the proliferation of glioma cells in a concentration-dependent manner, particularly at concentrations of 20 and 50 ng/mL (Figure 4A). We then examined the effects of rhNCAN on glioma cell invasion. Notably, cultivation of glioma cells with rhNCAN significantly augmented the invasion ability of glioma cells compared with that in the untreated control group (Figure 4B and C). Thus, these results suggested that NCAN had certain effects on glioma cells and mediated the proliferation and invasion of tumor cells. In addition, NCAN was also significantly expressed in hNSCs and glioma cells (Figure 4D), and treatment of glioma cells with rhNCAN resulted in dramatically increased expression levels of RhoA and ROCK compared with the control group, as demonstrated by Western blot analysis (Figure 4E). These findings suggested that the Rho-ROCK pathway may be mediated by NCAN during glioma progression and may be downstream of the NCAN signaling pathway.

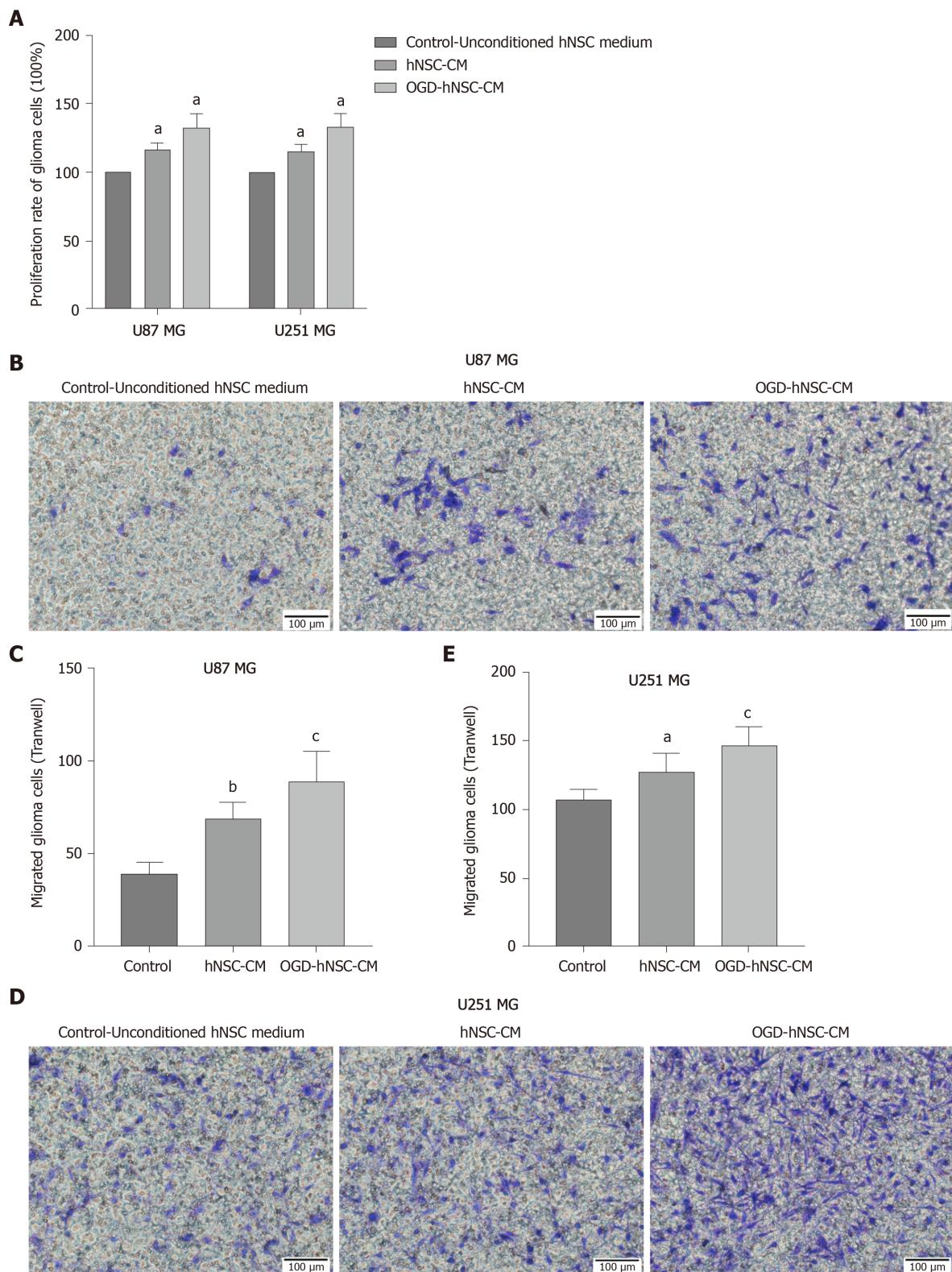


Figure 2 Oxygen-glucose deprived-human neural stem cells-condition medium further promotes glioma cell proliferation and invasion.
A: The proliferation of U87 and U251 MG cells was significantly increased in the condition medium (CM) groups, especially in the oxygen-glucose deprived-human neural stem cells (hNSC)-CM group, as measured by CCK8 assay; **B-E:** hNSC-CM promoted the invasion ability of glioma cells (U87 MG shown in B and C, U251 MG shown in D and E) compared to unconditioned hNSC medium, especially in the oxygen-glucose deprived-hNSC-CM group. Control: Unconditioned hNSC medium group. ^a $P < 0.05$ vs Control, ^b $P < 0.01$ vs Control, ^c $P < 0.05$ vs hNSC-CM. OGD: Oxygen-glucose deprived; hNSCs: Human neural stem cells; CM: Condition medium.

DISCUSSION

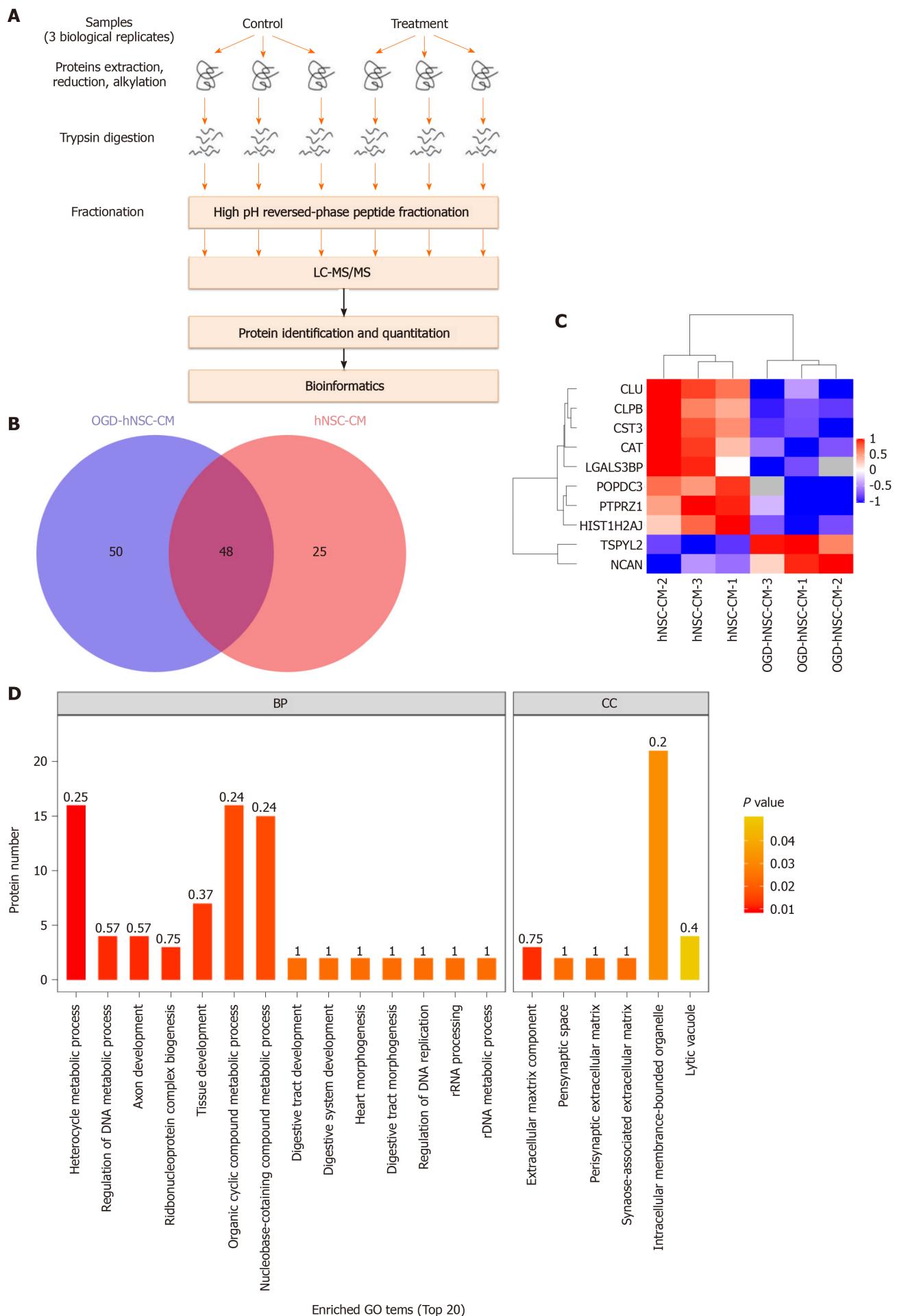
In this study, we showed that culture of glioma cells with SVZ NSC-derived CM under hypoxic conditions further increased glioma cell proliferation and invasion compared with normoxic NSC-CM. The regulatory effects induced by OGD-NSC-CM were associated with NCAN, which was secreted from OGD NSCs. Moreover, NCAN was expressed in glioma cells and played an important role in mediating the progression of tumor cells mainly *via* the Rho/ROCK pathway.

GBM is an aggressive and incurable brain tumor, which harbors many gene mutations and genetic abnormalities, such as alterations in the neurofibromin 1, *TP53*, isocitrate dehydrogenase (*IDH*), and *PTEN* genes, which are related to the proliferation of glioma cells, and the forkhead box G1 and *SOX2* genes, which affect the self-renewal of glioma cells^[22-24]. Improving our understanding of the origin or disease progression of gliomas could provide a fundamental basis for developing new treatments. Recently, NSCs in the SVZ region have been considered as the origin of gliomagenesis. Moreover, Lee *et al*^[25] found that normal SVZ tissue located at a distance from the tumor in 56.3% of patients with wild-type *IDH* GBM contained low levels of GBM driver mutations. Additionally, they suggested that astrocyte-like NSCs harboring driver mutations may have migrated from the SVZ and facilitated the development of high-grade malignant gliomas in distant brain regions. Bardella *et al*^[22] showed that the expression of *IDH1R132H* in the SVZ of the adult mouse brain caused gliomagenesis and tumor infiltration.

Emerging data have confirmed that glioma spread to the SVZ is closely associated with decreased survival of patients and increased tumor recurrence^[15,17,18,26], such as patients with GBMs contacting SVZ region have lower survival rates than those with GBMs contacting the subgranular zone, corpus callosum, or cortex. Clinical studies of radiation treatment of SVZ NSCs in patients with GBM have found that incidental radiation to the SVZ is robustly correlated with progression-free survival in patients^[17,27]. However, direct genetic evidence, which may improve prognoses in patients with GBM, is still lacking. Although glioma often spreads to the SVZ, cell communication between NSCs and gliomas have not been thoroughly evaluated.

Recently, a novel strategy using CM derived from cells has been evaluated for the treatment of various diseases. Li *et al*^[28] showed that the CM from human menstrual blood-derived endometrial stem cells had protective effects on neurotoxicant 1-methyl-4-phenylpyridinium (MPP⁺)-induced cytotoxicity and inflammation in patients with Parkinson's disease and found at least 12 neurotrophic factors in the CM by protein assays, suggesting that these factors may contribute to the protective effects against Parkinson's disease. Baez-Jurado *et al*^[29] indicated that the CM of human adipose mesenchymal stem cells improved the viability of astrocytes, reduced nuclear fragmentation of cells, and improved the mitochondrial membrane potential of astrocytes. Additionally, Bordoni *et al*^[30] cultured human CD34 cells for several weeks in hepatocyte CM and found that the expression of CD34 and CD133 markers was gradually decreased, whereas the expression of CD144 and CD14 cell markers was increased *in vitro* and *in vivo*. These findings indicated that the hematopoietic cells differentiated into endothelial cells following induction by hepatocyte CM in the cellular microenvironment. Gharaei *et al*^[31] showed that CM from human dental pulp stem cells significantly accelerated the adhesion phase (from sedimentation to attachment and spreading), the proliferation rate, and migration of human umbilical vein endothelial cells. Doeppner *et al*^[32] intravenously applied CM derived from SVZ neural progenitor cells in male C57BL6 mice with focal cerebral ischemia and found that CM dose-dependently reduced infarct volume and brain leukocyte infiltration and enhanced cell survival signaling, which was associated with increased brain glial cell line-derived neurotrophic factor and vascular endothelial growth factor. Thus, these findings suggested that neural progenitor cell-derived CM induced sustained neuroprotection and that the secreted factors derived from cells may have applications as an alternative to cell transplantation. Furthermore, Wang *et al*^[33] confirmed that NSCs were involved in the recurrence of GBM mediated by GBM-derived EVs. They suggested that GBM-derived EVs may induce NSC differentiation into tumor-promoting cells.

The biological effects of SVZ NSCs associated with GBM development may be mediated by factors derived from the CM, including cytoskeletal proteins, tumor associated-proteins, transcription factors, and growth factors. Therefore, in this study, we co-cultured glioma cells with the CM of SVZ NSCs and found significant increases in the proliferation and invasion of glioma cells, particularly under hypoxic conditions, compared with those under normoxic conditions. Furthermore, protein assays demonstrated that NCAN, which was overexpressed in the CM of OGD-NSCs,



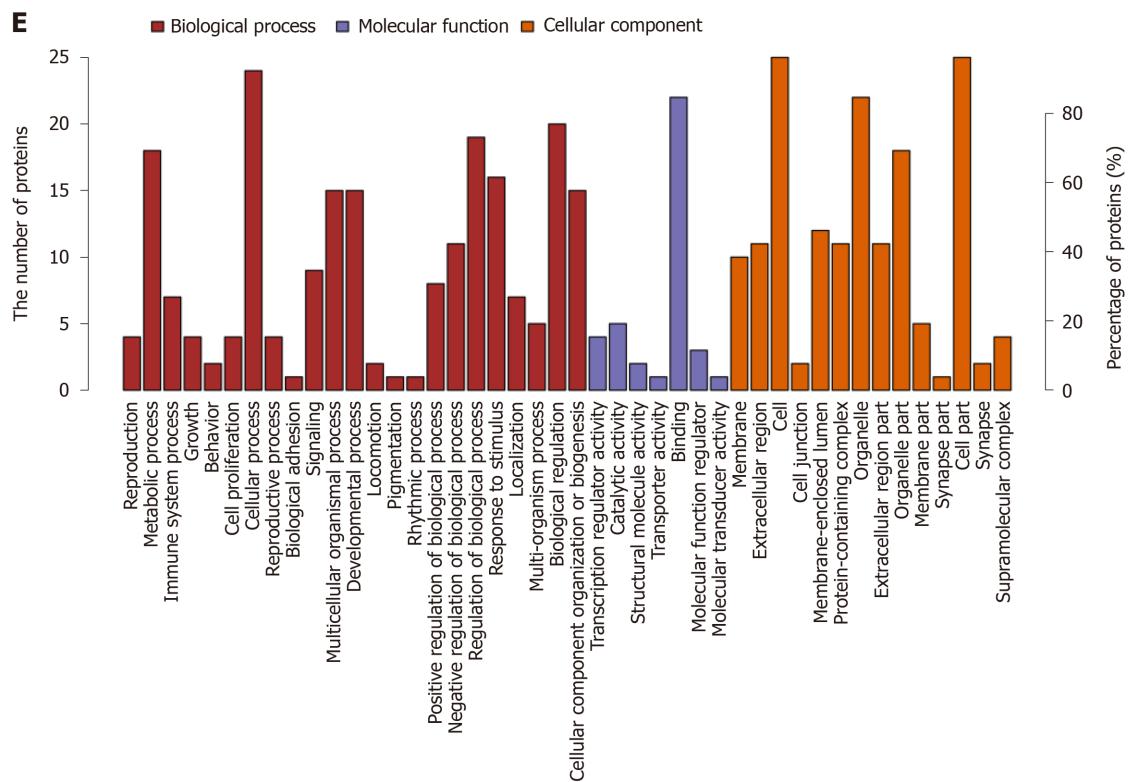
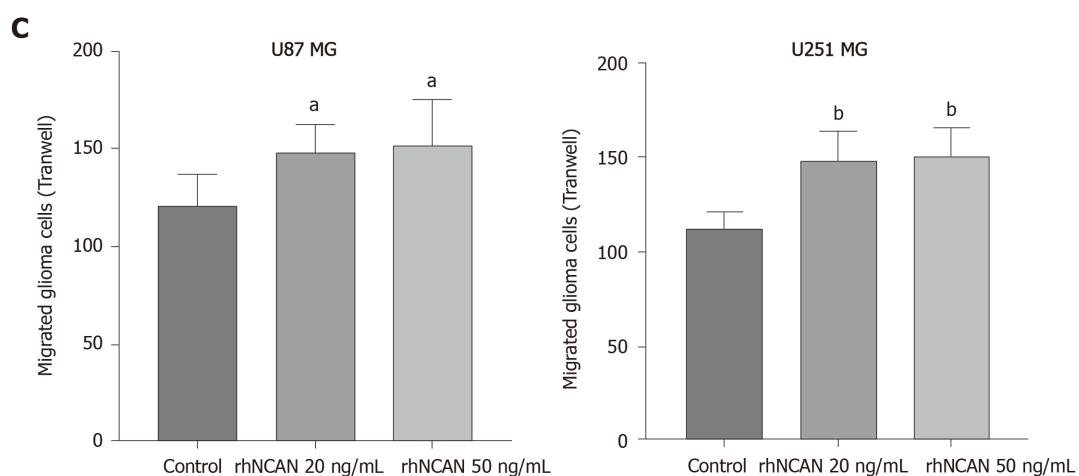
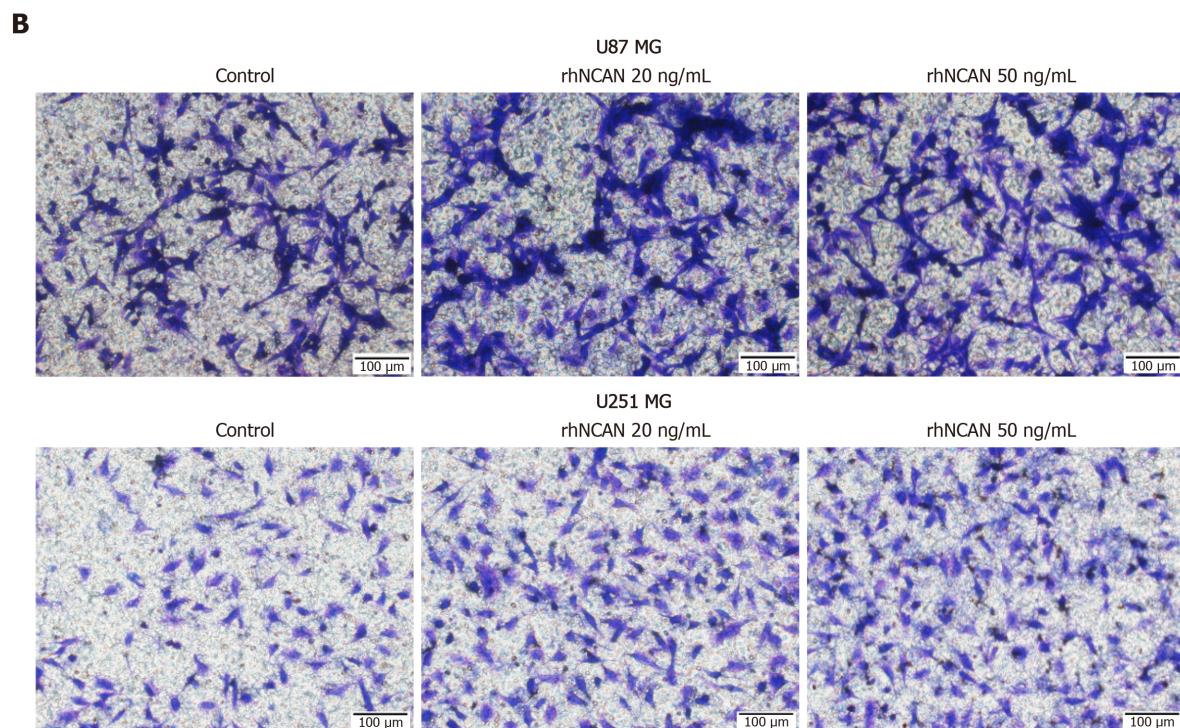
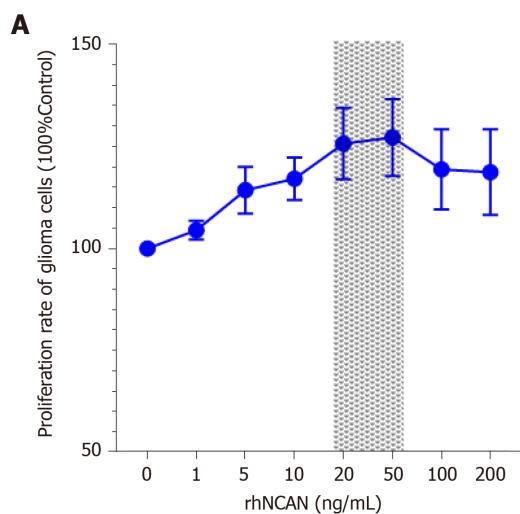


Figure 3 Protein expression analysis of condition medium derived from human neural stem cells by mass spectrometry. A: The experimental workflow of protein mass spectrometry analysis; B: Venn diagram of the proteins distribution in human neural stem cells-condition medium (hNSC-CM) and oxygen-glucose deprived (OGD)-hNSC-CM; C: Hierarchical clustering of differentially expressed proteins between hNSC-CM group and OGD-hNSC-CM group. Red color indicates higher expression, and green color indicates lower expression; D: The top 20 Gene Ontology functions of predicted targets according to the differentially expressed proteins; E: The biological process, molecular function, and cellular component of the analyzed proteins between hNSC-CM and OGD-hNSC-CM group. OGD: Oxygen-glucose deprived; hNSCs: Human neural stem cells; CM: Condition medium.

induced important regulatory effects in the proliferation and invasion of glioma cells *via* activation of the Rho/ROCK pathway. Rho/ROCK signaling is a well-established pathway in cell invasion and migration. Active ROCK inhibits myosin light chain phosphatase, which promotes actin and myosin crosslinking and thus leads to cell motion and increased cell migration^[34].

CONCLUSION

In conclusion, in this study, we have demonstrated a new role for the interaction between SVZ NSCs and glioma cells, in which SVZ NSCs promote glioma cell invasion *via* the secreted protein NCAN. Moreover, we have showed that SVZ NSC-derived CM under hypoxic conditions further increases glioma cell proliferation and invasion and that the secreted protein NCAN mediates the progression of glioma cells *via* activation of the Rho/ROCK pathway. Taken together, these findings highlight the prominent regulatory effects of SVZ NSCs in glioma progression and identify NCAN, which is secreted from NSCs, as a potential new therapeutic target for glioma treatment.



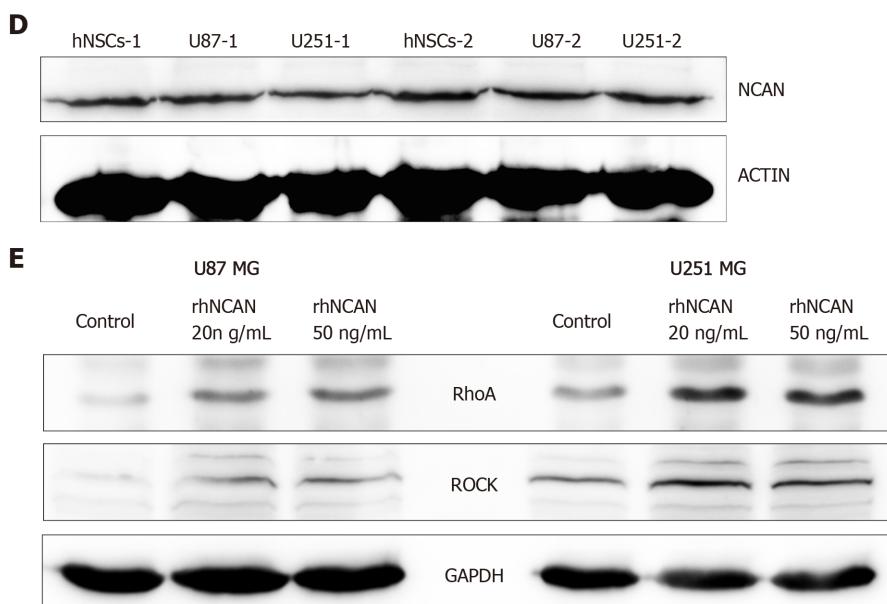


Figure 4 Recombinant human NCAN protein increases glioma cell proliferation and invasion. A: The proliferation of glioma cells was significantly increased after treatment with different concentrations of recombinant human NCAN (rhNCAN), as measured by CCK8 assay, especially at the concentrations of 20 and 50 ng/mL; B and C: rhNCAN (both 20 and 50 ng/mL) significantly improved the invasion ability of U87 and U251 glioma cells; D: Expression of NCAN in the human neural stem cells and glioma cells detected by Western blot. β -actin was used as the loading control. -1 and -2 indicated two independent samples; E: After treatment of U87 and U251 glioma cells with rhNCAN (20 and 50 ng/mL), the downstream RhoA-ROCK signaling pathway was significantly activated in both glioma cells, as measured by Western blot. β -actin was used as the loading control. Con: Control 10% FBS condition medium group; rhNCAN: Recombinant human NCAN protein. (^a $P < 0.05$ vs Control, ^b $P < 0.01$ vs Control).

ARTICLE HIGHLIGHTS

Research background

Glioma contacting with the subventricular zone (SVZ) is associated with a poor prognosis and decreased overall survival of patients. The most important cell type in the SVZ region is neural stem cells (NSCs), which can differentiate into neurons, astrocytes, and oligodendrocytes.

Research motivation

The roles of neural stem cells of the subventricular zone (SVZ NSCs) in glioma progression are still unclear.

Research objectives

To investigate the effects of SVZ NSCs in gliomas, especially the conditioned medium (CM) derived from SVZ NSCs, on the tumor-related behaviors of glioma cells.

Research methods

We used the CM derived from SVZ NSCs under normoxic and hypoxic conditions to co-culture with glioma cells, and explored the potential functional molecules in the CM by mass spectrometry analysis.

Research results

After co-cultured with glioma cells, the CM derived from hypoxic SVZ NSCs further increased glioma cell proliferation and invasion compared with normoxic CM. The MS analysis identified the candidate protein NCAN in the hypoxic CM. Furthermore, NCAN played an important role in mediating the progression of tumor cells mainly via the Rho/ROCK pathway in gliomas.

Research conclusions

SVZ NSC-derived CM under hypoxic condition further increases glioma cell proliferation and invasion, and the protein NCAN secreted from hypoxic SVZ NSCs plays prominent regulatory effects in the progression of glioma cells.

Research perspectives

The study provided important insights into the mechanisms of tumor progression associated with glioma contacting SVZ region, and suggested a new molecular target of gene therapy for gliomas.

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