World Journal of *Stem Cells*

World J Stem Cells 2020 December 26; 12(12): 1439-1690





Published by Baishideng Publishing Group Inc

W J S C World Journal of Stem Cells

Contents

Monthly Volume 12 Number 12 December 26, 2020

REVIEW

1439	Brain tumors: Cancer stem-like cells interact with tumor microenvironment
	Liu HL, Wang YN, Feng SY
1455	Effect of metformin on stem cells: Molecular mechanism and clinical prospect <i>Jiang LL, Liu L</i>
1474	Novel insights for improving the therapeutic safety and efficiency of mesenchymal stromal cells
	Najar M, Martel-Pelletier J, Pelletier JP, Fahmi H
1492	Noninvasive <i>in vivo</i> cell tracking using molecular imaging: A useful tool for developing mesenchymal stem cell-based cancer treatment
	Rajendran RL, Jogalekar MP, Gangadaran P, Ahn BC
1511	Prospects for the therapeutic development of umbilical cord blood-derived mesenchymal stem cells
	Um S, Ha J, Choi SJ, Oh W, Jin HJ
1529	Mesenchymal stem cells secretome: The cornerstone of cell-free regenerative medicine

- González-González A, García-Sánchez D, Dotta M, Rodríguez-Rey JC, Pérez-Campo FM
- Minibrain-related kinase/dual-specificity tyrosine-regulated kinase 1B implication in stem/cancer stem 1553 cells biology

Kokkorakis N, Gaitanou M

ORIGINAL ARTICLE

Basic Study

Acupuncture accelerates neural regeneration and synaptophysin production after neural stem cells 1576 transplantation in mice

Zhao L, Liu JW, Kan BH, Shi HY, Yang LP, Liu XY

- 1591 Spinal cord injury regeneration using autologous bone marrow-derived neurocytes and rat embryonic stem cells: A comparative study in rats Sadat-Ali M, Al-Dakheel DA, Ahmed A, Al-Turki HA, Al-Omran AS, Acharya S, Al-Bayat MI
- 1603 6-gingerol protects nucleus pulposus-derived mesenchymal stem cells from oxidative injury by activating autophagy

Nan LP, Wang F, Liu Y, Wu Z, Feng XM, Liu JJ, Zhang L

1623 Stem cells from human exfoliated deciduous teeth ameliorate concanavalin A-induced autoimmune hepatitis by protecting hepatocytes from apoptosis

Zhou YK, Zhu LS, Huang HM, Cui SJ, Zhang T, Zhou YH, Yang RL



Conter	World Journal of Stem Cells
conter	Monthly Volume 12 Number 12 December 26, 2020
1640	Influence of donor age on the differentiation and division capacity of human adipose-derived stem cells
	Horinouchi CD, Barisón MJ, Robert AW, Kuligovski C, Aguiar AM, Dallagiovanna B
1652	Umbilical cord-derived mesenchymal stem cells preconditioned with isorhamnetin: potential therapy for burn wounds
	Aslam S, Khan I, Jameel F, Zaidi MB, Salim A
1667	Effects of normobaric cyclic hypoxia exposure on mesenchymal stem-cell differentiation-pilot study on bone parameters in elderly
	Camacho-Cardenosa M, Quesada-Gómez JM, Camacho-Cardenosa A, Leal A, Dorado G, Torrecillas-Baena B, Casado- Díaz A



Contents

Monthly Volume 12 Number 12 December 26, 2020

ABOUT COVER

Editorial Board Member of World Journal of Stem Cells, Dr. Mohammed Grawish is a Distinguished Professor at Mansoura University and Vice-Dean for Community Services and Environmental Affairs at Delta University for Science and Technology (Egypt). Dr. Grawish received his Bachelor's degree (1990), Master's degree in Oral Biology (1998), and his PhD (2003) from the Faculty of Dentistry, Mansoura University. After, he worked as Lecturer in the Al-Gabl Al-Garby University (2005-2008; Gehrian, Libya) and as Associate Professor in the King Saud University (2011-2013; Riyadh, Saudi Arabia). His ongoing research interests focus mainly on the appropriate therapeutic use of stem cells in dentistry, the design and characterization of biomaterials as scaffold materials for loading stem cells, and the application of complementary and alternative medicine as an adjunctive treatment to traditional medicine for oral diseases. (L-Editor: Filipodia)

AIMS AND SCOPE

The primary aim of World Journal of Stem Cells (WJSC, World J Stem Cells) is to provide scholars and readers from various fields of stem cells with a platform to publish high-quality basic and clinical research articles and communicate their research findings online. WJSC publishes articles reporting research results obtained in the field of stem cell biology and regenerative medicine, related to the wide range of stem cells including embryonic stem cells, germline stem cells, tissue-specific stem cells, adult stem cells, mesenchymal stromal cells, induced pluripotent stem cells, embryonal carcinoma stem cells, hemangioblasts, lymphoid progenitor cells, etc.

INDEXING/ABSTRACTING

The WJSC is now indexed in Science Citation Index Expanded (also known as SciSearch®), Journal Citation Reports/Science Edition, Biological Abstracts, BIOSIS Previews, PubMed, and PubMed Central. The 2020 Edition of Journal Citation Reports[®] cites the 2019 impact factor (IF) for WJSC as 3.231; IF without journal self cites: 3.128; Ranking: 18 among 29 journals in cell and tissue engineering; Quartile category: Q3; Ranking: 113 among 195 journals in cell biology; and Quartile category: Q3.

RESPONSIBLE EDITORS FOR THIS ISSUE

Production Editor: Yan-Xia Xing; Production Department Director: Yun-Xiaojian Wu; Editorial Office Director: Ze-Mao Gong,

www.wjgnet.com/bpg/gerinfo/204 ELINES FOR ETHICS DOCUMENTS www.wjgnet.com/bpg/GerInfo/287 ELINES FOR NON-NATIVE SPEAKERS OF ENGLISH www.wjgnet.com/bpg/gerinfo/240 CATION ETHICS www.wjgnet.com/bpg/GerInfo/288
www.wjgnet.com/bpg/GerInfo/287 ELINES FOR NON-NATIVE SPEAKERS OF ENGLISH www.wjgnet.com/bpg/gerinfo/240 CATION ETHICS
ELINES FOR NON-NATIVE SPEAKERS OF ENGLISH www.wignet.com/bpg/gerinfo/240 CATION ETHICS
www.wjgnet.com/bpg/gerinfo/240
CATION ETHICS
www.wjgnet.com/bpg/GerInfo/288
CATION MISCONDUCT
www.wjgnet.com/bpg/gerinfo/208
LE PROCESSING CHARGE
www.wjgnet.com/bpg/gerinfo/242
FOR SUBMITTING MANUSCRIPTS
www.wjgnet.com/bpg/GerInfo/239
IE SUBMISSION
S '/

E-mail: bpgoffice@wjgnet.com https://www.wjgnet.com



W J S C World Journal of Stem Cells

Submit a Manuscript: https://www.f6publishing.com

World J Stem Cells 2020 December 26; 12(12): 1439-1454

DOI: 10.4252/wjsc.v12.i12.1439

ISSN 1948-0210 (online)

REVIEW

Brain tumors: Cancer stem-like cells interact with tumor microenvironment

Hai-Long Liu, Ya-Nan Wang, Shi-Yu Feng

ORCID number: Hai-Long Liu 0000-0002-6181-2577; Ya-Nan Wang 0000-0002-6278-3562; Shi-Yu Feng 0000-0002-6359-1124.

Author contributions: Liu HL and Wang YN contributed equally to the review; Liu HL and Feng SY contributed to the whole conception and design; Liu HL and Wang YN were responsible for the details of the review structure and interpretation; Liu HL, Wang YN, and Feng SY completed the manuscript and figures; Feng SY was responsible for the review supervision; all authors read and approved the final version of the manuscript.

Supported by The Medical Big Data Research Program of Chinese PLA General Hospital, No. 2018MBD-20 (to Feng SY); National Natural Science Foundation of China, No. 81902975 (to Liu HL); and the 65th China Postdoctoral Science Foundation, No. 2019M653940 (to Liu HL).

Conflict-of-interest statement: The authors declare no conflict of interest.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative

Hai-Long Liu, Shi-Yu Feng, Department of Neurosurgery, Chinese PLA General Hospital, Beijing 100853, China

Ya-Nan Wang, Department of Pathology, Affiliated Hospital of Hebei University, Baoding 071000, Hebei Province, China

Corresponding author: Shi-Yu Feng, MD, PhD, Doctor, Department of Neurosurgery, Chinese PLA General Hospital, No. 28 Fuxing Road, Haidian District, Beijing 100853, China. fengshiyu72123@163.com

Abstract

Cancer stem-like cells (CSCs) with potential of self-renewal drive tumorigenesis. Brain tumor microenvironment (TME) has been identified as a critical regulator of malignancy progression. Many researchers are searching new ways to characterize tumors with the goal of predicting how they respond to treatment. Here, we describe the striking parallels between normal stem cells and CSCs. We review the microenvironmental aspects of brain tumors, in particular composition and vital roles of immune cells infiltrating glioma and medulloblastoma. By highlighting that CSCs cooperate with TME via various cellular communication approaches, we discuss the recent advances in therapeutic strategies targeting the components of TME. Identification of the complex and interconnected factors can facilitate the development of promising treatments for these deadly malignancies.

Key Words: Cancer stem-like cells; Microenvironment; Brain tumor; Inflammation; Clinical application; Glioma

©The Author(s) 2020. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: To better understand the effects of interplaying between cancer stem-like cells (CSCs) and tumor microenvironment (TME) on brain tumor progression, we review the distinct characters of CSCs and the mechanisms regarding how TME regulates CSC self-renewal. Moreover, we emphasize the valuable application of singcell RNA sequencing technology in the cancer research.

Citation: Liu HL, Wang YN, Feng SY. Brain tumors: Cancer stem-like cells interact with tumor



Commons Attribution

NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Manuscript source: Invited manuscript

Specialty type: Cell and tissue engineering

Country/Territory of origin: China

Peer-review report's scientific quality classification

Grade A (Excellent): A Grade B (Very good): 0 Grade C (Good): C, C, C Grade D (Fair): 0 Grade E (Poor): 0

Received: March 29, 2020 Peer-review started: March 30, 2020 First decision: September 21, 2020 Revised: October 7, 2020 Accepted: October 27, 2020 Article in press: October 27, 2020 Published online: December 26, 2020

P-Reviewer: Hann HW, Huang AHC, Tanabe S S-Editor: Chen XF L-Editor: Wang TQ P-Editor: Ma YJ



microenvironment. World J Stem Cells 2020; 12(12): 1439-1454 URL: https://www.wjgnet.com/1948-0210/full/v12/i12/1439.htm DOI: https://dx.doi.org/10.4252/wjsc.v12.i12.1439

INTRODUCTION

Brain tumors respond poorly to existing therapies, leading to the poor prognoses. The recurrence of malignant brain tumors accounts for a majority of mortality and points a significant challenge for conventional treatment modalities^[1,2]. The available therapies including operation and chemoradiotherapy mainly focus on the bulk of tumor cells, however, they are not able to impair the subpopulation that has been identified as cancer stem-like cells (CSCs)^[3]. Actually, CSCs present the strong self-proliferation and diverse differentiation capability, which induces tumor progression and therapeutic resistance^[4,5]. However, it is noticeable that CSCs cannot maintain the stem-like properties by relying on themselves. They need to interact with tumor microenvironment (TME) to insist the stemness and protect themselves against the chemotherapeutic elements and radiation^[6]. Even when spreading along the circulation, CSCs recruit microenvironmental components, forming a cluster of metastasis niche or generating a pre-metastatic niche at the faraway organs before arriving. Previously, although bulk tumor sequence analysis can interrogate the genetic status with average expression profiles, it provides limited insight into the specific cell type, especially the immune heterogenicity^[7]. Importantly, it is difficult to identify the mechanisms of TME activities at the different stages of brain tumors through traditional methods. To address these challenges, sing-cell RNA sequencing (scRNA-seq) has characterized cancer and immune cell types at high resolution, which figures out the oncogenic signaling, proliferation, and complement/immune response^[8,9]. Therefore, to better understand the effects of interplaying between CSCs and TME on brain tumor progression, we review the distinct characters of CSCs and the mechanisms regarding how TME regulates CSC self-renewal. Moreover, we emphasize the valuable application of scRNA-seq technology in the cancer research.

CSCs

Stem cells and CSCs

Stem cells can perpetuate themselves via self-renewal and generate the particular mature cells via differentiation. Nevertheless, the rigorous ways to purification and identification of the somatic stem cells have been under debate in some conditions^[10]. Neural stem cells (NSCs), an undifferentiated cell type originating in the central nervous system (CNS), have the potential to give rise to offspring cells differentiating to multiple lineages, including neuronal and non-neuronal populations^[11]. NSCs have been isolated from mice and humans using the traditional methods as well as confirmed by the advanced scRNA-seq analysis^[12,13]. The pathways mediating the stemness of NSCs, such as Notch, WNT/ β -catenin, and Hedgehog signaling, are also critical in glioma stem-like cells to drive tumorigenicity^[14,15]. Recently, novel therapeutic approaches are established with the goal of not only reducing the tumor burden but also targeting CSCs involved^[16].

The differentiation and self-proliferation potential of NSCs and their clinical applications have been discovered in many studies^[17]. For example, the stem cell biology can provide new insights into cancer research and treatment. The intermediate filament protein, nestin, serves as a biomarker for stem cells and has been further identified to label the CSCs^[18]. We are now studying the critical role of nestin in Hedgehog signaling and revealing the proper interaction between Bergmann glia of the cerebellum and granular neuronal precursors induced by nestin. Based on these findings, our team continuously demonstrated that nestin drives sonic hedgehog (SHH)-medulloblastoma and uncovered Gli3 as a therapeutic target to treat these malignancies^[19]. Furthermore, the study was expanded to TME, indicating that tumorassociated astrocytes (TAAs)-derived SHH drives nestin expression in medulloblastoma cells through a smoothened-dependent mechanism^[20]. Therefore, the similarity between stem cells and CSCs sheds light on targeting the stemness associated profiles to control cancer progression.



TME heterogeneity mediated by CSCs

Heterogeneity has been phenotypically and functionally identified among all the malignancies^[21]. Importantly, the multiple differentiation of CSCs and microenvironmental influences provide a model for generating phenotypic and functional heterogeneity beyond the clonal evolution^[22]. The CSC models have been well established for cancer research, which differentiate into progenies with limited proliferation potential. Some cancers including medulloblastoma, neuroblastoma, and hierarchically organized cancers can arise from normal stem cells or restricted progenitors through mutations^[23,24]. Although CSCs may not address the cell of origin, this population can significantly affect the microenvironment construction. CSCs are enough "clever" to order the other populations, such as fibroblasts, astrocytes, and microglia/macrophages, to activate for serving themselves *via* cytokine or exosome secretion. Phenotypic and functional heterogeneity always occurs in cancer-associated fibroblasts (CAFs)^[25]. Differences in subtypes and functions of CAF behaviors lead to microenvironmental heterogeneity, which is mediated by augmented expression of proteolytic enzymes, deposition of extracellular matrix, and pathogenic angiogenesis derived from CSCs. In addition to the existing components, CSCs can contribute to tumor heterogeneity via differentiating to various kinds of stroma cells as occasion requires^[26]. Especially in resistance to chemoradiotherapies or recurrences, the tumor cells undergo de-differentiation to stem-like status and then differentiate to the stromal populations to override the wicked conditions^[27]. To better understand the heterogeneity of cancers, the typical flow cytometry and advanced sing-cell multiomics sequencing are the most popular technologies. The application of flow cytometry makes it possible to harvest the distinct subpopulations of malignant and non-malignant cells based on the well-known markers^[28]. Using this approach, the different kinds of cells can be separated for the following culture and experiments.

Trans-differentiation and de-differentiation

Trans-differentiation and de-differentiation have been discussed extensively in many studies. Similar to normal stem cells, CSCs can trans-differentiate into other cell lineages in addition to the original lineage arising from tumors^[29]. Trans-differentiation of CSCs provides a possible therapeutic target to control recurrence even though this molecular basis has not yet been fully recovered. A cell or tissue from one differentiated state changes to another. The de-differentiated state is unnatural and unstable, which sometimes may present during trans-differentiation^[30]. It is supposed that the cell de-differentiates to immature status and then differentiates to the other lineage^[31]. However, this routine seems to consume more energy than the direct transdifferentiation, which is mediated or affected by microenvironmental factors. CAFs have been reported to be abundant in gliomas, breast, prostate, and pancreatic cancers. The production of TGF- β 1, TGF- β 2, PDGF, IL6, and bFGF and protein kinase C in cancer cells play crucial roles in tumor-induced trans-differentiation of surrounding fibroblasts^[32]. Furthermore, TGF- β 1 or TGF- β 2 actually makes sense to the full-extent trans-differentiation, whereas the others, such as PDGF, bFGF, or IL6 (each alone), induce only partial trans-differentiation^[33]. In addition to cytokines, the cancer cellderived exosomes contain abundant and diverse signaling factors particularly under hypoxic conditions^[32], which interact with CAFs, astrocytes, and immune cells to mediate trans-differentiation.

Having identified the functions of factors on inducing trans-differentiation, the cell fusion is another approach involved in cell fate reprogramming. Somatic cells are fused with the embryonic stem cells, thereby exposing them to the reprogramming milieu of stem cells^[34,35]. The method has been confirmed in cancers using both murine and human cells. We are interested in the interaction between CSCs and microglia. We have found that microglia phagocytosed the oligodendrocyte progenitor cell like malignant cells, therefore forming de-differentiated microglia presenting both parental cell features including self-proliferation and proinflammation characters. However, the molecular mechanism regarding this two-lineage fusion induced de-differentiation remains unclear. Thus, it is important for malignant cells to achieve the ability to reprogram host body cells into stroma cells and to modulate their microenvironment and receive positive feedback for growth and drug resistance^[36].

scRNA-seq technology

The flow cytometry sorting programs are restricted to the well-established cell surface markers, thus resulting in incapable identification of new subpopulations. Additionally, a cluster of cells rather than single cells in one unit are obtained after sorting^[28,37]. On the other hand, it is difficult to accurately analyze heterogeneous



populations and status due to technical limitations of marker-based approaches. Over the past decade, the powerful scRNA-seq technology has been applied to overcome the limitations and provide an unbiased view of cell-to-cell variability with gene signatures of each subgroup^[38] (Figure 1). Both microfluidic and barcoding approaches are most commonly utilized to assay the transcriptomes from tens of thousands of single cells^[39]. Due to the exponential increase in the amounts of single-cell transcriptomic data, it is also necessary to develop computational tools to achieve the meaningful findings. To analyze the cancer heterogeneity, two bioinformatic approaches in scRNA-seq data have been developed: (1) The discrete cluster indicators for cell subtypes and status are labelled in a discrete latent variable approach; and (2) the continuous pseudo-time for differentiation trajectories is constructed in a latent variable approach^[9]. However, the double droplets and limited throughput resolution are still the major challenges.

Glioma and medulloblastoma

Glioma is the most common primary malignant brain tumor, accounting for almost 40% of primary CNS tumors, of which glioblastoma is the leading cause of mortality^[40]. Unfortunately, in spite of significant advances in diagnostic and therapeutic approaches, the median survival of glioblastoma patients remains about 14.2 mo. This could be attributed to the existing classic treatment producing limited efficacy on CSCs^[41]. Although many studies discuss pathways driving tumor initiation and progression, epigenetic reprogramming increases oncogenic potential of CSCs, which can lead to tumor growth or therapeutic resistance^[42]. Our previous study revealed that the tumor-specific maternal embryonic leucine zipper kinase (MELK) activity was essential for the EZH2/NF-KB interaction via enhancing the methyltransferase activity and maintained the stemness^[43]. NF-κB as the downstream of the MELK/EZH2 complex opens another exciting pathway to better understand the mechanism of tumorigenesis, beyond the well-established Rel/NF-κB interaction^[44]. Activation of NF-KB involves a series of sequential events including cooperation with TME via activating the inflammation associated transcriptions. On the other hand, immune microenvironment also contributes to the glioma stem-like property insistence. Zhang et al^[45] reported that C-C motif ligand 8 (CCL8) was a tumorassociated macrophage (TAM) associated factor to mediate glioblastoma stemness via ERK1/2 signaling and targeting CCL8 could provide an insight strategy for glioma treatment.

Medulloblastoma constitutes the most common malignant brain tumor in childhood^[46]. Despite the advanced therapeutic strategies, the 5-year survival rate in high-risk group is only about 40% and about half of patients suffer from metastasizing along the neuraxis^[47,48]. Recurrent or disseminated medulloblastoma accounts for the majority of pediatric brain tumor-related mortality^[49]. Previously, medulloblastoma stem-like cells (MBSCs) have been identified to drive tumorigenesis and recurrence with the potential of self-renewal and resistance to chemoradiotherapy^[50]. Among the primary medulloblastoma, MBSCs maintain stemness via activation of key pathways, such as Notch, WNT/β-catenin, and JAK2/STAT3 signaling^[51]. Nestin-expressing medulloblastoma cells are the source of medulloblastoma proliferation. MBSCs show restricted capacity to maintain stemness when undergoing metastasis, which requires the efficient cooperation of MBSC niche to protect stem-like properties. Astrocytes, the most abundance of glial cells, are reactivated to play a critical role in supporting tumor growth and inducing protection from chemotherapy^[20,52]. We have found the elevated proportion of TAAs in disseminated medulloblastoma compared with primary medulloblastoma. MBSC enrichment in recurrent medulloblastoma was attributed to an increased level of C-C motif ligand 2 (CCL2) released by TAAs undergoing necroptosis^[53]. Noticeably, no specific markers for MBSCs have been identified until now, which restricts their purification. CD133, CD15, CD34, and nestin are usually used to label or collect the MBSCs. However, the usage of CD133 to mark MBSCs has its specific drawbacks as follows: The percentage of CD133+ cells is less than that of CD15⁺ in medulloblastoma, which shows greater potential in labeling the CSCs. Only one marker is chosen to identify MBSCs with less meaningful results probably coming from two indexes, such as CD133+CD15, CD15+CD34, or CD133+nestin^[5,23,54].

TME

TME contains many non-malignant cells in addition to cancer cells, including immune cells, endothelial cells, pericytes, fibroblasts, and others^[21]. Especially, the astrocytes,



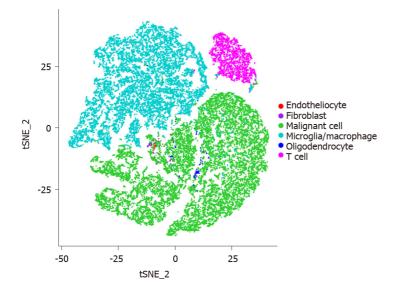


Figure 1 Sing-cell RNA sequencing charts cellular heterogeneity in gliomas. Sing-cell RNA sequence analysis showing various kinds of cells within glioma tumor microenvironment including tumor cells, microglia/macrophages, T cells, fibroblasts, and endotheliocytes.

microglia, and neurons are special tissue-resident populations in the CNS. The unique properties of the CNS require a specific framework to generate the TME-targeted interventions. As shown in Figure 2, the expression of markers for vessels, immune cells, and astroglia stroma suggested the complex compositions of malignant brain tumors. TME has been emerging as a crucial mediator for tumor progression in both primary and metastatic brain malignancies. Brain tumors respond poorly to current therapies, in which TME has been recognized to play critical roles^[55]. The CSCs may be considered as the cell origin for tumor recurrence and TME construction^[56]. However, the tumor-associated parenchymal cells also importantly function in controlling the course of pathogenesis. We focus on glioma and medulloblastoma to review how brain TME regulates tumor progression and therapeutic response *via* interacting with CSCs.

Immune microenvironment

Microglia and macrophage: The normal brain has been considered to be "immune privileged" in the whole body, which must be sheltered from immune cell entrance. The activated immune cells produce inflammatory factors that are cytotoxic to cause neurodegeneration. When dissociating the brain tumor tissues into single cells, the majority of cells are TAMs including the blood-derived macrophages and resident microglia accounting for about 35%^[57]. Some studies focus on defining context-specific microglia/macrophage activation and phenotype as a measure of functional diversity. Microglia/macrophage activation is classified into the pro-inflammatory M1 state and anti-inflammatory M2 state^[57,58]. TAMs exist along a linear M1-to-M2 phenotypic continuum. TAMs tend to be pro-tumorigenic and accumulate gradually with higher tumor grade^[59], which produce high levels of pro-inflammatory cytokines promoting tumor proliferation and stemness maintenance. However, macrophage infiltration is considered as a double-edged sword, exerting both tumor-promoting and anti-tumor effects^[60]. To support the role of macrophage-mediated inflammation in cancer induction, a previous study has discovered that genetic ablation of STAT3, an antiinflammatory transcription factor, in macrophages resulted in a chronic inflammatory response in the colon that was sufficient to induce invasive adenocarcinoma. Additionally, loss of IL10 that acts through STAT3 enhanced carcinogen-induced tumorigenesis in the intestine^[61]. Macrophage infiltration varies relying on the pathologic type or process. The immune function of macrophages can be suppressed when they are located in the glioma microenvironment^[57,62].

Microglia, the resident myeloid cell population in the CNS and a major component of brain immune system, play an essential role in neuronal homeostasis and regulate multiple pathogeneses of disorders, such as neurodegenerative diseases and brain tumors^[63]. Recently, accumulated research has reported that microglia distinguish from macrophages. Under homeostatic conditions, microglia originate from hematopoietic stem cells in the yolk sac but not from bone marrow^[64]. Although microglia share some traits with monocyte-derived macrophages, they express numbers of special genes at high levels, which may be affected by CNS environment^[65]. In glioma, microglia are



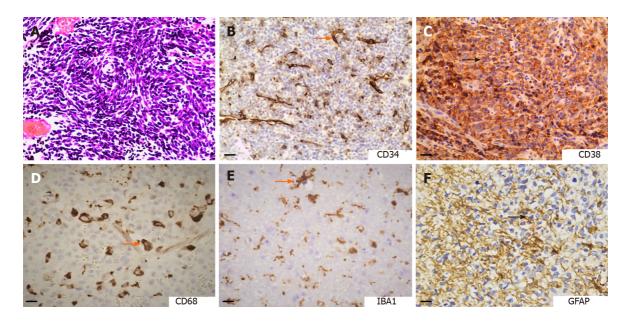


Figure 2 Hematoxylin and eosin staining and immunohistochemical staining of brain tumors. A: Hematoxylin and eosin staining displaying the pathological vessels distributed in medulloblastoma (bar, 20 µm); B: Expression of CD34 (vascular endothelial cell marker) in chordoma (bar, 20 µm); C: Expression of CD38 (T cell marker) in anaplastic diffuse astrocytoma (bar, 20 µm); D: Expression of CD68 (macrophage marker) in medulloblastoma (bar, 10 µm); E: Expression of IBA1 (microglia marker) in glioblastoma (bar, 10 µm); F: Expression of glial fibrillary acidic protein (astrocyte marker) in medulloblastoma (bar, 10 µm). GFAP: Glial fibrillary acidic protein.

reactivated within or in close proximity to masses up to half of TME, which shape the TME via releasing a wide range of cytokines for tumor proliferation and invasion^[66]. Therefore, targeting the microglia has represented a novel therapeutic approach to this malignancy.

Previously, Venteicher et al[67] detected a continuum model characterizing transition from microglia-like state to macrophage-like state in IDH1/2 mutant gliomas. Müller $et al^{[9]}$ clearly separated microglial TAMs and monocyte-derived TAMs by ontogeny in IDH1/2 wild-type glioblastoma. Both studies provided evidence that microglia vary across different grades and subtypes of gliomas. However, the underlying molecular basis involved in reactivated microglia transition and interaction with glioma cells remains poorly understood. By scRNA-seq analysis of > 50000 single cells isolated from gliomas, we are now focusing a microglial subtype associated with high-grade glioma possessing inflammasome mediated proinflammation and stem-like features, which shapes cytokine microenvironment and promotes oncogenesis. Further analysis in our study also depicts TGF-\beta1 derived from IDH1/2 wild-type glioblastoma cells with SETD2-deficiency is required for high-grade glioma associated microglia activation.

Previous studies on the differences between brain-resident microglia and bloodderived macrophages have been confounded by a lack of specific markers. Recently, the scRNA-seq study has suggested that CD11b⁺/CX3CR1⁺/P2RY12⁺ population should be the murine microglia^[68]. Based on this finding, we are purifying microglia from gliomas by utilizing CD11b and CX3CR1 markers in the scRNA-seq or fluorescent-activated cell sorting experiments. However, the expression of P2RY12 is dramatically compromised in microglia derived from glioblastoma, indicating the specific microglia activation driven by IDH1/2 wild-type cancer cells. Although much evidence supports that microglia/macrophage activation is classified into M1 and M2 state, recent scRNA-seq analysis indicates that macrophages simultaneously express gene profiles of both M1 and M2 phenotypes in an injury mouse model, raising the concern that this classification may not accurately reflect the microglia activation features^[60]. Consistent with this finding, Keren-Shaul *et al*^[69] described a novel microglia type associated with neurodegenerative diseases by using single-cell transcripts. Similarly, we established an experimental paradigm by analyzing the scRNA-seq data and histopathological staining to identify a special cohort of microglia associated with high grade glioma. We discovered the genetic program encoding a large number of risk factors, corresponding to the need for proinflammatory response and self-proliferation.

Lymphocytes: Lymphocytes consist of T cells, B cells, and natural killer cells. Mature T



cells are released to peripheral lymphoid organs where they can be primed by engaging with antigen presenting cells^[70]. In patients, whether further adaptation to specific microenvironment occurs during anti-tumor immunity remains poorly understood. Glioblastoma is a highly immunosuppressive brain tumor because of their T cell paucity^[71,72]. Recently, Garris et al^[70] revealed a specific mechanism regarding escaping immunosurveillance in brain tumors by trapping T cells in bone marrows via the deficiency of S1P receptor on T cells. The interaction between different cell types within TME also produces obvious effects on immunosuppression. The braindependent immune suppression is apparently mediated by microglial cells through TGF-β1/TβRI signaling. Pharmacological obligation of TGF-β1/TβRI signaling can partially reverse the immune suppression but cannot contribute to prolonging the survival of mice, which is due to the lack of sufficient T cells in brain tumors^[73]. Additionally, tumor associated CD8+T cells exhibit proliferation and differentiation potential within the brain, leading to enhanced retention^[74]. The researchers have detected that cooperation with brain TME reduced the population of CD8+T cells in human glioma samples. CD8+T cells in TME consist of two distinct populations of stem-like and terminally differentiated ones. The stem-like subset gives rise to more terminally differentiated, effector-expressing daughter cells. Similar to this finding, we are also interested in a subpopulation of microglia associated with high-grade glioma, which presents the stem-like property after phagocytosing the oligodendrocyte precursor cell (OPC)-like malignant cells^[75]. Therefore, these studies critically suggest that the local microenvironment can modify T cell effector functions during anti-tumor immunity^[76]. It is currently under clinical investigation that enhancing T cell activation is induced by co-stimulations through the usage of checkpoint inhibitors among the patients with brain primary or metastatic tumors. In mouse glioma models, inhibition of CTLA-4, a checkpoint molecule, leads to a prolonged survival and activity enhancement of CD4⁺ helper T cell^[77,78]. The standard care treatment is recommended to combine with these advanced clinical studies in recurrent stages. For instance, combination of temozolomide with this treatment regimen reveals an even more pronounced effect on prognosis^[79].

TAAs

Astrocytes as the specialized glial cells distribute ubiquitously throughout the CNS, which play critical roles in providing neurotransmitters and cholesterol, constructing microcirculation, producing energy metabolites, and maintaining homeostasis^[52]. Antibodies against glial fibrillary acidic protein (GFAP), S100 β , astrocyte cell surface antigen 2, and brain lipid binding protein are often used to detect astrocytes in immunohistochemistry assays^[80,81]. Specifically, astrocytes in the cerebellum are identified as Bergmann glial cells supporting proliferation and migration of granular neuronal precursors. In pathological status, such as trauma or tumor growth, astrocytes are activated with upregulated expression levels of GFAP and S100β and enhanced proliferative capability. TAAs release many cytokines to potentially develop a supportive TME for tumor growth and aggressiveness^[20]. This program is linked to the significantly elevated expression of malignancy associated genes in cancer cells, which have been proposed to protect against chemoradiotherapy. In a previous study, reactive astrocytes have been reported to mediate glioblastoma invasion through hyperactivation of matrix metalloproteinase 2[82]. Another study has demonstrated that reactive astrocytes expressing SHH were highly concentrated in the perivascular regions of glioblastoma^[83]. Consistent with this finding, our work discovered that TAAs, enriched in medulloblastoma, expressed and secreted SHH to promote medulloblastoma cell proliferation. Genetic ablation of TAAs dramatically inhibited nestin expression in medulloblastoma cells, resulting in reduced tumor growth^[20]. Furthermore, we found a higher proportion of TAAs in recurrent or disseminated medulloblastoma and TAAs within recurrent TME underwent necroptosis, releasing CCL2 to interplay with MBSCs^[53]. The fact that the CSCs are enriched more dramatically in relapsed tumors can be attributed to the dynamic variation of microenvironmental components.

CAFs

Fibrosis is a common pathophysiological response to chronic injury in many tissues. The processes of wound healing and tissue remodeling are protective mechanisms activated in response to stress and injury with the goal of maintaining functional integrity of systems^[84]. Additionally, fibrosis is the marker of chronic inflammation, which results from deregulation of normal healing and exposure to chronic injury. Chronic inflammation has been identified within TME, especially after receiving



chemoradiotherapy^[85]. Stromal fibroblasts activated by tumor cells in TME have been reported to function in angiogenesis development and metastasis formation. Fibroblasts can be activated at all stages of tumor progression and their structural and functional influences on the process work through cytokine secretion^[33]. The growth factors, chemokines and extracellular matrix derived from CAFs facilitate the angiogenic recruitment of endothelial cells and pericytes. Fibroblasts are therefore a key determinant in malignancy progression and represent an important target for cancer therapies. It is hypothesized that both CSCs and CAFs cooperate with and support each other relying on the communicating messenger or reside preferentially at the tumor-stroma interfaces^[86]. To develop the favorable niche for CSC selfproliferation, CAFs can also interact with other cells in TME, such as immune and endothelial cells^[87]. Choi et al^[88] reported that CAFs promoted cell adhesion to human brain microvascular endothelial cells *via* upregulating expression of integrin $\alpha 5\beta 1$ and $\alpha v \beta 3$, c-MET, and $\alpha 2$,6-siayltransferase. A similar role of CAFs within brain tumors has also been suggested. When coculturing human brain-derived fibroblasts and glioblastoma cells, the production and hyperactivation of matrix metalloproteinase 2/9 have been shown to be involved in tumor migration^[89].

Neurons

In addition to glial cells, neurons as a key regulator of CNS development and plasticity are the highly specialized cells contributing to tumor initiation and progression. Recently, accumulated research suggests that glioma arises from neural stem/precursor cells, specifically oligodendrocyte precursor cells (OPCs), pre-OPCs, or earlier neural precursor cells^[90]. It is known that the proliferation of neuronal cells and OPCs is stimulated by neurons via the mitogenic signals, which recalls our understanding of neuronal activity as important components of TME. The active neurons influence the proliferation, differentiation, and invasion of glioma cells^[9]. A previous study has reported that upregulation of neuroligin-3 in post-synaptic neurons promoted proliferation of cancer cells of patient-derived xenograft glioblastoma models. The mechanism involved PI3K/Akt signaling activation induced by neuronal upregulation of neuroligin-3, which subsequently elevated the expression of FOS and feedforward-upregulation of neuroligin-3 gene expression to enhance the cancerous proliferative activity^[92]. In addition to proliferation, the neuron activity affords convenient condition to malignant cell spreading. Wang et al^[93] demonstrated that CSCs were preferentially located along neuronal white matter tracts presenting a demyelinated phenotype at the invasive frontiers of glioblastoma. The Notch-induced Sox9 promoted the elevated expression of Sox2 and the methylation level of the Notch1 promoter was attenuated by the upregulation of Sox2 to reinforce Notch1 expression in CD133⁺/Notch1⁺ CSCs. Inhibition of Notch signaling attenuated the white-mattertract tropism of CSCs. For the metastases, the neoplasms could mimic neurons by activating neurotransmitter signaling via the critical elements, for example, upregulating the expression of GABA receptors and transporters^[91]. Collectively, these studies suggest that neuronal-specific processes regarding the synaptic transmission can promote brain tumor progression, which warrants further investigation to generate the indispensable roles of neurons within TME.

COMMUNICATIONS

Communication between cancer cells and non-malignant cells within TME is a twoway process involving a wide variety of stroma cells and a diverse range of mechanisms. Cells communicate in the direct and indirect manners^[94]. The essential components of cell-cell communication include the cellular junctions (chemical synapses, pannexins, connexins, and ion channels), anchoring junctions (adherence, focal adhesions, and desmosomes), and tight junctions, as well as cytokines (inflammatory factors and growth factors), exosomes, extracellular matrix, extracellular microRNAs, and different transmembrane adhesion proteins (cadherins and integrins)^[94-96]. Sharing information *via* cellular communication is mediated by different mechanisms: The direct cell-cell communication involves intracrine /autocrine and adjacent communication with nearby cells, which themselves are also regulated by other distinct patterns; and the indirect intercellular communication involves local communication over short distances (paracrine and synaptic signaling) and long distances via hormones (endocrine)^[97]. Here, we give some examples including cytokines, exosomes, and matrix.

Cytokines

The function of inflammation in cancer development has been established well. Cytokines, low-molecular-weight proteins mainly derived from immune and stromal cells, regulate proliferation, differentiation, migration, activation, and death^[98]. Within the chronic inflammatory TME, they induce malignancy transformation and affect immunotherapy based on the balance of pro- and anti-inflammatory process, relative concentrations, associated receptor expression, and surrounding cell conditions^[99]. Targeting the cytokines have delivered promising prospects on cancer therapy. Our study has revealed that inhibition of CCL2/CCR2 blocked the communication between MBSCs and TAAs and compromised the disseminated medulloblastoma stemness^[59].

Exosomes

Exosomes, transporting all the main biomolecules, perform intercellular transfer of components locally and systemically^[100]. Exosomes have emerged as new influencers in tumor progression by acting both tumor cells and tumor-associated cells. Exosomes derived from glioblastoma have been reported to induce the tumor-promoting transformation of NSCs^[101].

Matrix

Brain extracellular matrix constituents of the normal brain parenchyma, such as heparan sulfate proteoglycans and hyaluronic acid, are mainly concentrated in neural stem cell niches, modifying normal stem cell homeostasis^[102]. Dramatically increased production of heparan sulfate proteoglycans in gliomas has been identified as a reservoir for heparin-binding angiogenic growth factors^[103].

CLINICAL APPLICATION

An understanding of the contribution of TME will allow us to truly tailor therapeutic strategy for each patient. Current standard treatment for glioblastoma, for example, is resection followed by radiation and temozolomide chemotherapy^[104] as well as ifosfamide, carboplatin, vincristine, and teniposide chemotherapy for medulloblastoma^[105]. Adverse effects range in severity between individuals, such as the loss of blood-brain barrier integrity, cytokine deregulation, cognitive dysfunction, and changes in neuronal integrity. Clinical evaluation of benefit vs risk in a quantifiable manner should be considered to minimize additional unnecessary harm. Recently, several approaches to target the TME of brain tumors are ongoing in preclinical and clinical studies. Among them, targeting the vasculature through anti-angiogenic reagents, such as bevacizumab and apatinib, is relatively successful in glioblastoma patients because of highly distributed vessels^[106,107]. We have also found that apatinib exhibits efficient effects on the glioblastoma resistant to temozolomide (Figure 3). Immune checkpoint inhibitors are popular with treatment of both primary and metastatic brain tumors, such as nivolumab and/or ipilimumab vs bevacizumab in glioblastoma (NCT02017717)^[108] and ipilimumab with nivolumab/fotemustine in brain metastasis (NCT02460068)^[109]. The success of immune checkpoint inhibitors utilized in various kinds of cancers is an excellent example of addressing a TME-mediated resistance mechanism to obtain prognostic benefits. Another promising immunotherapy involves the development of T cells engineered to target proteins on the surfaces of cancer cells^[110]. Chimeric antigen receptor T-cells are constructed to target a number of different tumorous antigens. As TME provides a safe haven for cancer cells, strategies to mobilize cells from tumor niche will render the malignant cells more sensitive to therapy. However, as the engineered T cells remain to be subject to suppression by microenvironmental factors, it is necessary to illustrate the mechanisms regarding the efficacy of novel agents. In addition to targeting the TME components, oncologists should pay more attention to CSCs that have been identified as the root of tumor recurrence. Some agents targeting stemness associated genes, such as the Notch, Hedgehog, and WNT signaling, are underway in many cancers^[111]. We are now trying to treat medulloblastoma using LY3039478, an oral Notch inhibitor^[112], indicating a promising prospect (Figure 3).

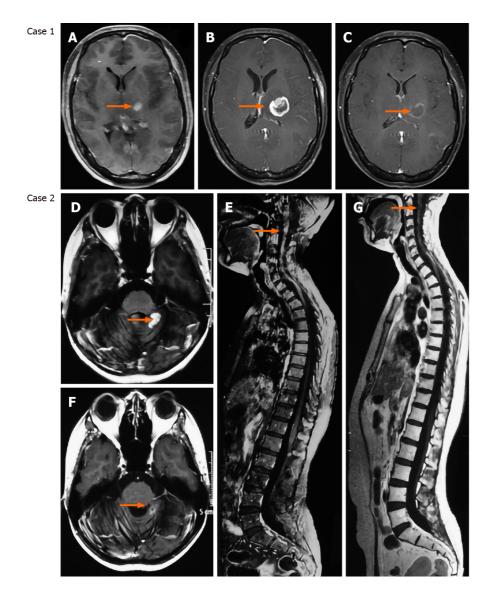


Figure 3 Targeting the vasculature presents promising effects on brain tumor therapy. Case 1 is a 29-year-old female patient who was diagnosed with IDH1/2 wild-type glioblastoma and received treatment of temozolomide combined with apatinib. Case 2 is a 31-year-old female patient who was diagnosed with recurrent medulloblastoma (local recurrence and dissemination) and received temozolomide + irinotecan + bevacizumab. A: Axial enhanced magnetic resonance imaging (MRI) showing a mass located at the left thalamus; B: Axial enhanced MRI showing that the lesion significantly progressed after 1-mo treatment of temozolomide; C: Axial enhanced MRI showing that the lesion was dramatically reduced after 1-mo treatment of combination of temozolomide and apatinib; D and E: Enhanced cerebrospinal MRI showing the local recurrent medulloblastoma and disseminated lesions along the spinal cord; F and G: Enhanced cerebrospinal MRI showing that the recurrent and disseminated lesions were significantly reduced after one cycle of chemotherapy.

CONCLUSION

Accumulating laboratory works support the thrilling concept that brain tumors rely on the interplay between CSCs and TME during progression. CSCs govern the surrounding components to maintain stem-like properties and other cells create a more permissive niche via production of extracellular substrates facilitating tumor growth and invasion. Identifying the cellular and extracellular dependent relationships unique to TME can provide exceptional opportunities to develop effective treatments targeting these symbiotic associations that support brain tumor progression.

REFERENCES

- 1 Yung WK. Imaging endpoints in brain tumor clinical trials: proceedings of the January 30, 2014 Workshop. Introduction. Neuro Oncol 2014; 16 Suppl 7: vii1 [PMID: 25313233 DOI: 10.1093/neuonc/nou2911
- 2 Aref D, Croul S. Medulloblastoma: recurrence and metastasis. CNS Oncol 2013; 2: 377-385



Zaishidena® WJSC | https://www.wjgnet.com

[PMID: 25054581 DOI: 10.2217/cns.13.30]

- 3 Magee JA, Piskounova E, Morrison SJ. Cancer stem cells: impact, heterogeneity, and uncertainty. Cancer Cell 2012; 21: 283-296 [PMID: 22439924 DOI: 10.1016/j.ccr.2012.03.003]
- 4 Peitzsch C, Tyutyunnykova A, Pantel K, Dubrovska A. Cancer stem cells: The root of tumor recurrence and metastases. Semin Cancer Biol 2017; 44: 10-24 [PMID: 28257956 DOI: 10.1016/j.semcancer.2017.02.011]
- 5 Garg N, Bakhshinyan D, Venugopal C, Mahendram S, Rosa DA, Vijayakumar T, Manoranjan B, Hallett R, McFarlane N, Delaney KH, Kwiecien JM, Arpin CC, Lai PS, Gómez-Biagi RF, Ali AM, de Araujo ED, Ajani OA, Hassell JA, Gunning PT, Singh SK. CD133⁺ brain tumor-initiating cells are dependent on STAT3 signaling to drive medulloblastoma recurrence. Oncogene 2017; 36: 606-617 [PMID: 27775079 DOI: 10.1038/onc.2016.235]
- 6 Kenda Suster N, Virant-Klun I. Presence and role of stem cells in ovarian cancer. World J Stem Cells 2019; 11: 383-397 [PMID: 31396367 DOI: 10.4252/wjsc.v11.i7.383]
- Dagogo-Jack I, Shaw AT. Tumour heterogeneity and resistance to cancer therapies. Nat Rev Clin 7 Oncol 2018; 15: 81-94 [PMID: 29115304 DOI: 10.1038/nrclinonc.2017.166]
- 8 Jaitin DA, Kenigsberg E, Keren-Shaul H, Elefant N, Paul F, Zaretsky I, Mildner A, Cohen N, Jung S, Tanay A, Amit I. Massively parallel single-cell RNA-seq for marker-free decomposition of tissues into cell types. Science 2014; 343: 776-779 [PMID: 24531970 DOI: 10.1126/science.1247651]
- 9 Müller S, Kohanbash G, Liu SJ, Alvarado B, Carrera D, Bhaduri A, Watchmaker PB, Yagnik G, Di Lullo E, Malatesta M, Amankulor NM, Kriegstein AR, Lim DA, Aghi M, Okada H, Diaz A. Singlecell profiling of human gliomas reveals macrophage ontogeny as a basis for regional differences in macrophage activation in the tumor microenvironment. Genome Biol 2017; 18: 234 [PMID: 29262845 DOI: 10.1186/s13059-017-1362-4]
- 10 Abazova N, Krijgsveld J. Advances in stem cell proteomics. Curr Opin Genet Dev 2017; 46: 149-155 [PMID: 28806595 DOI: 10.1016/j.gde.2017.07.007]
- 11 Strzyz P. Proteins clog neural stem cell activation. Nat Rev Mol Cell Biol 2018; 19: 346-347 [PMID: 29643417 DOI: 10.1038/s41580-018-0012-3]
- 12 Shang Z, Chen D, Wang Q, Wang S, Deng Q, Wu L, Liu C, Ding X, Wang S, Zhong J, Zhang D, Cai X, Zhu S, Yang H, Liu L, Fink JL, Chen F, Liu X, Gao Z, Xu X. Single-cell RNA-seq reveals dynamic transcriptome profiling in human early neural differentiation. Gigascience 2018; 7: giy117 [PMID: 30239706 DOI: 10.1093/gigascience/giy117]
- Monaco S, Baur K, Hellwig A, Hölzl-Wenig G, Mandl C, Ciccolini F. A Flow Cytometry-Based 13 Approach for the Isolation and Characterization of Neural Stem Cell Primary Cilia. Front Cell Neurosci 2018; 12: 519 [PMID: 30692915 DOI: 10.3389/fncel.2018.00519]
- 14 Bajaj J, Diaz E, Reya T. Stem cells in cancer initiation and progression. J Cell Biol 2020; 219: e201911053 [PMID: 31874116 DOI: 10.1083/jcb.201911053]
- 15 Reya T, Morrison SJ, Clarke MF, Weissman IL. Stem cells, cancer, and cancer stem cells. Nature 2001; **414**: 105-111 [PMID: 11689955 DOI: 10.1038/35102167]
- 16 Worley JR, Parker GC. Effects of environmental stressors on stem cells. World J Stem Cells 2019; 11: 565-577 [PMID: 31616535 DOI: 10.4252/wjsc.v11.i9.565]
- 17 Tuazon JP, Castelli V, Lee JY, Desideri GB, Stuppia L, Cimini AM, Borlongan CV. Neural Stem Cells. Adv Exp Med Biol 2019; 1201: 79-91 [PMID: 31898782 DOI: 10.1007/978-3-030-31206-0_4]
- 18 Lendahl U, Zimmerman LB, McKay RD. CNS stem cells express a new class of intermediate filament protein. Cell 1990; 60: 585-595 [PMID: 1689217 DOI: 10.1016/0092-8674(90)90662-x]
- Li P, Lee EH, Du F, Gordon RE, Yuelling LW, Liu Y, Ng JM, Zhang H, Wu J, Korshunov A, 19 Pfister SM, Curran T, Yang ZJ. Nestin Mediates Hedgehog Pathway Tumorigenesis. Cancer Res 2016; 76: 5573-5583 [PMID: 27496710 DOI: 10.1158/0008-5472.CAN-16-1547]
- 20 Liu Y, Yuelling LW, Wang Y, Du F, Gordon RE, O'Brien JA, Ng JMY, Robins S, Lee EH, Liu H, Curran T, Yang ZJ. Astrocytes Promote Medulloblastoma Progression through Hedgehog Secretion. Cancer Res 2017; 77: 6692-6703 [PMID: 28986380 DOI: 10.1158/0008-5472.CAN-17-1463]
- 21 Quail DF, Joyce JA. The Microenvironmental Landscape of Brain Tumors. Cancer Cell 2017; 31: 326-341 [PMID: 28292436 DOI: 10.1016/j.ccell.2017.02.009]
- 22 McGranahan N, Swanton C. Clonal Heterogeneity and Tumor Evolution: Past, Present, and the Future. Cell 2017; 168: 613-628 [PMID: 28187284 DOI: 10.1016/j.cell.2017.01.018]
- 23 Li P, Du F, Yuelling LW, Lin T, Muradimova RE, Tricarico R, Wang J, Enikolopov G, Bellacosa A, Wechsler-Reya RJ, Yang ZJ. A population of Nestin-expressing progenitors in the cerebellum exhibits increased tumorigenicity. Nat Neurosci 2013; 16: 1737-1744 [PMID: 24141309 DOI: 10.1038/nn.3553]
- Boeva V, Louis-Brennetot C, Peltier A, Durand S, Pierre-Eugène C, Raynal V, Etchevers HC, 24 Thomas S, Lermine A, Daudigeos-Dubus E, Geoerger B, Orth MF, Grünewald TGP, Diaz E, Ducos B, Surdez D, Carcaboso AM, Medvedeva I, Deller T, Combaret V, Lapouble E, Pierron G, Grossetête-Lalami S, Baulande S, Schleiermacher G, Barillot E, Rohrer H, Delattre O, Janoueix-Lerosey I. Heterogeneity of neuroblastoma cell identity defined by transcriptional circuitries. Nat Genet 2017; 49: 1408-1413 [PMID: 28740262 DOI: 10.1038/ng.3921]
- 25 Su S, Chen J, Yao H, Liu J, Yu S, Lao L, Wang M, Luo M, Xing Y, Chen F, Huang D, Zhao J, Yang L, Liao D, Su F, Li M, Liu Q, Song E. CD10⁺GPR77⁺ Cancer-Associated Fibroblasts Promote Cancer Formation and Chemoresistance by Sustaining Cancer Stemness. Cell 2018; 172: 841-856 [PMID: 29395328 DOI: 10.1016/j.cell.2018.01.009]



- Prasetyanti PR, Medema JP. Intra-tumor heterogeneity from a cancer stem cell perspective. Mol 26 Cancer 2017; 16: 41 [PMID: 28209166 DOI: 10.1186/s12943-017-0600-4]
- 27 Bai X, Ni J, Beretov J, Graham P, Li Y. Cancer stem cell in breast cancer therapeutic resistance. Cancer Treat Rev 2018; 69: 152-163 [PMID: 30029203 DOI: 10.1016/j.ctrv.2018.07.004]
- 28 Montante S, Brinkman RR. Flow cytometry data analysis: Recent tools and algorithms. Int J Lab Hematol 2019; 41 Suppl 1: 56-62 [PMID: 31069980 DOI: 10.1111/ijlh.13016]
- 29 Brown DV, Mantamadiotis T. Insights into the next generation of cancer stem cell research. Front Biosci 19: 1015-1027 [PMID: 24896333 DOI: 10.2741/4264]
- Dall'Agnese A, Caputo L, Nicoletti C, di Iulio J, Schmitt A, Gatto S, Diao Y, Ye Z, Forcato M, 30 Perera R, Bicciato S, Telenti A, Ren B, Puri PL. Transcription Factor-Directed Re-wiring of Chromatin Architecture for Somatic Cell Nuclear Reprogramming toward trans-Differentiation. Mol Cell 2019; 76: 453-472 [PMID: 31519520 DOI: 10.1016/j.molcel.2019.07.036]
- 31 Gong L, Cao L, Shen Z, Shao L, Gao S, Zhang C, Lu J, Li W. Materials for Neural Differentiation, Trans-Differentiation, and Modeling of Neurological Disease. Adv Mater 2018; 30: e1705684 [PMID: 29573284 DOI: 10.1002/adma.201705684]
- Heneberg P. Paracrine tumor signaling induces transdifferentiation of surrounding fibroblasts. Crit 32 Rev Oncol Hematol 2016; 97: 303-311 [PMID: 26467073 DOI: 10.1016/j.critrevonc.2015.09.008]
- Malanchi I, Santamaria-Martínez A, Susanto E, Peng H, Lehr HA, Delaloye JF, Huelsken J. 33 Interactions between cancer stem cells and their niche govern metastatic colonization. Nature 2011; 481: 85-89 [PMID: 22158103 DOI: 10.1038/nature10694]
- Wang R, Chen S, Li C, Ng KT, Kong CW, Cheng J, Cheng SH, Li RA, Lo CM, Man K, Sun D. 34 Fusion with stem cell makes the hepatocellular carcinoma cells similar to liver tumor-initiating cells. BMC Cancer 2016; 16: 56 [PMID: 26846780 DOI: 10.1186/s12885-016-2094-7]
- 35 Imai H, Kusakabe KT, Kiso Y, Hattori S, Kai C, Ono E, Kano K. Induction of pluripotency in mammalian fibroblasts by cell fusion with mouse embryonic stem cells. Biochem Biophys Res Commun 2020; 521: 24-30 [PMID: 31635800 DOI: 10.1016/j.bbrc.2019.10.026]
- Friedl P. Cell fusion: new mechanisms of plasticity in cancer? Lancet Oncol 2005; 6: 916-918 36 [PMID: 16321755 DOI: 10.1016/s1470-2045(05)70439-3]
- 37 Pozarowski P, Darzynkiewicz Z. Analysis of cell cycle by flow cytometry. Methods Mol Biol 2004; 281: 301-311 [PMID: 15220539 DOI: 10.1385/1-59259-811-0:301]
- Lake BB, Ai R, Kaeser GE, Salathia NS, Yung YC, Liu R, Wildberg A, Gao D, Fung HL, Chen S, 38 Vijayaraghavan R, Wong J, Chen A, Sheng X, Kaper F, Shen R, Ronaghi M, Fan JB, Wang W, Chun J, Zhang K. Neuronal subtypes and diversity revealed by single-nucleus RNA sequencing of the human brain. Science 2016; 352: 1586-1590 [PMID: 27339989 DOI: 10.1126/science.aaf1204]
- 39 Freytag S, Tian L, Lönnstedt I, Ng M, Bahlo M. Comparison of clustering tools in R for mediumsized 10x Genomics single-cell RNA-sequencing data. F1000Res 2018; 7: 1297 [PMID: 30228881 DOI: 10.12688/f1000research.15809.21
- Louis DN, Perry A, Reifenberger G, von Deimling A, Figarella-Branger D, Cavenee WK, Ohgaki H, 40 Wiestler OD, Kleihues P, Ellison DW. The 2016 World Health Organization Classification of Tumors of the Central Nervous System: a summary. Acta Neuropathol 2016; 131: 803-820 [PMID: 27157931 DOI: 10.1007/s00401-016-1545-1]
- Bao S, Wu Q, McLendon RE, Hao Y, Shi Q, Hjelmeland AB, Dewhirst MW, Bigner DD, Rich JN. 41 Glioma stem cells promote radioresistance by preferential activation of the DNA damage response. Nature 2006; 444: 756-760 [PMID: 17051156 DOI: 10.1038/nature05236]
- 42 Hu B, Wang Q, Wang YA, Hua S, Sauvé CG, Ong D, Lan ZD, Chang Q, Ho YW, Monasterio MM, Lu X, Zhong Y, Zhang J, Deng P, Tan Z, Wang G, Liao WT, Corley LJ, Yan H, Zhang J, You Y, Liu N, Cai L, Finocchiaro G, Phillips JJ, Berger MS, Spring DJ, Hu J, Sulman EP, Fuller GN, Chin L, Verhaak RGW, DePinho RA. Epigenetic Activation of WNT5A Drives Glioblastoma Stem Cell Differentiation and Invasive Growth. Cell 2016; 167: 1281-1295 [PMID: 27863244 DOI: 10.1016/j.cell.2016.10.039]
- 43 Liu H, Sun Y, Qi X, Gordon RE, O'Brien JA, Yuan H, Zhang J, Wang Z, Zhang M, Song Y, Yu C, Gu C. EZH2 Phosphorylation Promotes Self-Renewal of Glioma Stem-Like Cells Through NF-κB Methylation. Front Oncol 2019; 9: 641 [PMID: 31380279 DOI: 10.3389/fonc.2019.00641]
- Siggers T, Gilmore TD, Barron B, Penvose A. Characterizing the DNA binding site specificity of NF-KB with protein-binding microarrays (PBMs). Methods Mol Biol 2015; 1280: 609-630 [PMID: 25736775 DOI: 10.1007/978-1-4939-2422-6_36]
- 45 Zhang X, Chen L, Dang WQ, Cao MF, Xiao JF, Lv SQ, Jiang WJ, Yao XH, Lu HM, Miao JY, Wang Y, Yu SC, Ping YF, Liu XD, Cui YH, Zhang X, Bian XW. CCL8 secreted by tumorassociated macrophages promotes invasion and stemness of glioblastoma cells via ERK1/2 signaling. Lab Invest 2020; 100: 619-629 [PMID: 31748682 DOI: 10.1038/s41374-019-0345-3]
- 46 Cavalli FMG, Remke M, Rampasek L, Peacock J, Shih DJH, Luu B, Garzia L, Torchia J, Nor C, Morrissy AS, Agnihotri S, Thompson YY, Kuzan-Fischer CM, Farooq H, Isaev K, Daniels C, Cho BK, Kim SK, Wang KC, Lee JY, Grajkowska WA, Perek-Polnik M, Vasiljevic A, Faure-Conter C, Jouvet A, Giannini C, Nageswara Rao AA, Li KKW, Ng HK, Eberhart CG, Pollack IF, Hamilton RL, Gillespie GY, Olson JM, Leary S, Weiss WA, Lach B, Chambless LB, Thompson RC, Cooper MK, Vibhakar R, Hauser P, van Veelen MC, Kros JM, French PJ, Ra YS, Kumabe T, López-Aguilar E, Zitterbart K, Sterba J, Finocchiaro G, Massimino M, Van Meir EG, Osuka S, Shofuda T, Klekner A, Zollo M, Leonard JR, Rubin JB, Jabado N, Albrecht S, Mora J, Van Meter TE, Jung S, Moore AS, Hallahan AR, Chan JA, Tirapelli DPC, Carlotti CG, Fouladi M, Pimentel J, Faria CC, Saad AG,



Massimi L, Liau LM, Wheeler H, Nakamura H, Elbabaa SK, Perezpeña-Diazconti M, Chico Ponce de León F, Robinson S, Zapotocky M, Lassaletta A, Huang A, Hawkins CE, Tabori U, Bouffet E, Bartels U, Dirks PB, Rutka JT, Bader GD, Reimand J, Goldenberg A, Ramaswamy V, Taylor MD. Intertumoral Heterogeneity within Medulloblastoma Subgroups. Cancer Cell 2017; 31: 737-754 [PMID: 28609654 DOI: 10.1016/j.ccell.2017.05.005]

- 47 Liu H, Zhang J, Liu Y, Sun Y, Li C, Gu C, Wang H, Zhang H, Yu C, Zhang M. Neuraxis Metastases Of Primary Central Nervous System Tumors: A Review Of Clinicopathological And Radiographic Characters Of 198 Cases In A Single Center. Cancer Manag Res 2019; 11: 9829-9841 [PMID: 31819620 DOI: 10.2147/CMAR.S217672]
- 48 Liu H, Zhang X, Zhang M, Zhang J, Ning W, Yue A, Zhao R, Sun Y, Yu C. Skull bone tumor: a review of clinicopathological and neuroimaging characteristics of 426 cases at a single center. Cancer Commun 39: 8 [PMID: 30850028 DOI: 10.1186/s40880-019-0353-0]
- 49 Northcott PA, Robinson GW, Kratz CP, Mabbott DJ, Pomeroy SL, Clifford SC, Rutkowski S, Ellison DW, Malkin D, Taylor MD, Gajjar A, Pfister SM. Medulloblastoma. Nat Rev Dis Primers 2019; 5: 11 [PMID: 30765705 DOI: 10.1038/s41572-019-0063-6]
- Liu H, Sun Q, Sun Y, Zhang J, Yuan H, Pang S, Qi X, Wang H, Zhang M, Zhang H, Yu C, Gu C. MELK and EZH2 Cooperate to Regulate Medulloblastoma Cancer Stem-like Cell Proliferation and Differentiation. Mol Cancer Res 2017; 15: 1275-1286 [PMID: 28536141 DOI: 10.1158/1541-7786.MCR-17-0105
- Ramaswamy V, Taylor MD. Medulloblastoma: From Myth to Molecular. J Clin Oncol 2017; 35: 51 2355-2363 [PMID: 28640708 DOI: 10.1200/JCO.2017.72.7842]
- Seifert G, Schilling K, Steinhäuser C. Astrocyte dysfunction in neurological disorders: a molecular 52 perspective. Nat Rev Neurosci 2006; 7: 194-206 [PMID: 16495941 DOI: 10.1038/nrn1870]
- Liu H, Sun Y, O'Brien JA, Franco-Barraza J, Qi X, Yuan H, Jin W, Zhang J, Gu C, Zhao Z, Yu C, 53 Feng S, Yu X. Necroptotic astrocytes contribute to maintaining stemness of disseminated medulloblastoma through CCL2 secretion. Neuro Oncol 2020; 22: 625-638 [PMID: 31729527 DOI: 10.1093/neuonc/noz214]
- 54 Read TA, Fogarty MP, Markant SL, McLendon RE, Wei Z, Ellison DW, Febbo PG, Wechsler-Reya RJ. Identification of CD15 as a marker for tumor-propagating cells in a mouse model of medulloblastoma. Cancer Cell 2009; 15: 135-147 [PMID: 19185848 DOI: 10.1016/j.ccr.2008.12.016]
- Celià-Terrassa T, Kang Y. Metastatic niche functions and therapeutic opportunities. Nat Cell Biol 55 2018; 20: 868-877 [PMID: 30050120 DOI: 10.1038/s41556-018-0145-9]
- Thomas TM, Yu JS. Metabolic regulation of glioma stem-like cells in the tumor micro-56 environment. Cancer Lett 2017; 408: 174-181 [PMID: 28743531 DOI: 10.1016/j.canlet.2017.07.014]
- Gieryng A, Pszczolkowska D, Walentynowicz KA, Rajan WD, Kaminska B. Immune 57 microenvironment of gliomas. Lab Invest 2017; 97: 498-518 [PMID: 28287634 DOI: 10.1038/labinvest.2017.19]
- Herz J, Filiano AJ, Smith A, Yogev N, Kipnis J. Myeloid Cells in the Central Nervous System. 58 Immunity 2017; 46: 943-956 [PMID: 28636961 DOI: 10.1016/j.immuni.2017.06.007]
- 59 Szulzewsky F, Pelz A, Feng X, Synowitz M, Markovic D, Langmann T, Holtman IR, Wang X, Eggen BJ, Boddeke HW, Hambardzumyan D, Wolf SA, Kettenmann H. Glioma-associated microglia/macrophages display an expression profile different from M1 and M2 polarization and highly express Gpnmb and Spp1. PLoS One 2015; 10: e0116644 [PMID: 25658639 DOI: 10.1371/journal.pone.0116644]
- 60 Van Hove H, Martens L, Scheyltjens I, De Vlaminck K, Pombo Antunes AR, De Prijck S, Vandamme N, De Schepper S, Van Isterdael G, Scott CL, Aerts J, Berx G, Boeckxstaens GE, Vandenbroucke RE, Vereecke L, Moechars D, Guilliams M, Van Ginderachter JA, Saeys Y, Movahedi K. A single-cell atlas of mouse brain macrophages reveals unique transcriptional identities shaped by ontogeny and tissue environment. Nat Neurosci 2019; 22: 1021-1035 [PMID: 31061494 DOI: 10.1038/s41593-019-0393-4]
- 61 Wang Y, Shen Y, Wang S, Shen Q, Zhou X. The role of STAT3 in leading the crosstalk between human cancers and the immune system. Cancer Lett 2018; 415: 117-128 [PMID: 29222039 DOI: 10.1016/j.canlet.2017.12.003
- Ghosh A, Bhattacharya M, Sarkar P, Acharya S, Chaudhuri S. T11 target structure exerts effector 62 function by activating immune cells in CNS against glioma where cytokine modulation provide favorable microenvironment. Indian J Exp Biol 2010; 48: 879-888 [PMID: 21506495]
- 63 Prinz M, Priller J. Microglia and brain macrophages in the molecular age: from origin to neuropsychiatric disease. Nat Rev Neurosci 2014; 15: 300-312 [PMID: 24713688 DOI: 10.1038/nrn37221
- 64 Hambardzumyan D, Gutmann DH, Kettenmann H. The role of microglia and macrophages in glioma maintenance and progression. Nat Neurosci 2016; 19: 20-27 [PMID: 26713745 DOI: 10.1038/nn.4185]
- Gosselin D, Skola D, Coufal NG, Holtman IR, Schlachetzki JCM, Sajti E, Jaeger BN, O'Connor C, 65 Fitzpatrick C, Pasillas MP, Pena M, Adair A, Gonda DD, Levy ML, Ransohoff RM, Gage FH, Glass CK. An environment-dependent transcriptional network specifies human microglia identity. Science 2017; 356: eaal3222 [PMID: 28546318 DOI: 10.1126/science.aal3222]
- Bettinger I, Thanos S, Paulus W. Microglia promote glioma migration. Acta Neuropathol 2002; 66



103: 351-355 [PMID: 11904754 DOI: 10.1007/s00401-001-0472-x]

- 67 Venteicher AS, Tirosh I, Hebert C, Yizhak K, Neftel C, Filbin MG, Hovestadt V, Escalante LE, Shaw ML, Rodman C, Gillespie SM, Dionne D, Luo CC, Ravichandran H, Mylvaganam R, Mount C, Onozato ML, Nahed BV, Wakimoto H, Curry WT, Iafrate AJ, Rivera MN, Frosch MP, Golub TR, Brastianos PK, Getz G, Patel AP, Monje M, Cahill DP, Rozenblatt-Rosen O, Louis DN, Bernstein BE, Regev A, Suvà ML. Decoupling genetics, lineages, and microenvironment in IDHmutant gliomas by single-cell RNA-seq. Science 2017; 355: eaai8478 [PMID: 28360267 DOI: 10.1126/science.aai8478]
- 68 Masuda T, Sankowski R, Staszewski O, Böttcher C, Amann L, Sagar, Scheiwe C, Nessler S, Kunz P, van Loo G, Coenen VA, Reinacher PC, Michel A, Sure U, Gold R, Grün D, Priller J, Stadelmann C, Prinz M. Spatial and temporal heterogeneity of mouse and human microglia at single-cell resolution. Nature 2019; 566: 388-392 [PMID: 30760929 DOI: 10.1038/s41586-019-0924-x]
- Keren-Shaul H, Spinrad A, Weiner A, Matcovitch-Natan O, Dvir-Szternfeld R, Ulland TK, David 69 E, Baruch K, Lara-Astaiso D, Toth B, Itzkovitz S, Colonna M, Schwartz M, Amit I. A Unique Microglia Type Associated with Restricting Development of Alzheimer's Disease. Cell 2017; 169: 1276-1290 [PMID: 28602351 DOI: 10.1016/j.cell.2017.05.018]
- 70 Garris CS, Blaho VA, Hla T, Han MH. Sphingosine-1-phosphate receptor 1 signalling in T cells: trafficking and beyond. Immunology 2014; 142: 347-353 [PMID: 24597601 DOI: 10.1111/imm.12272
- 71 Vella G, Bergers G. Where Have All the T Cells Gone? Immunity 2018; 49: 592-594 [PMID: 30332627 DOI: 10.1016/j.immuni.2018.10.006]
- With GBM, T Cells May Be Stuck in Bone Marrow. Cancer Discov 2018; 8: 1203-1204 [PMID: 72 30185625 DOI: 10.1158/2159-8290.CD-NB2018-117]
- 73 Ooi YC, Tran P, Ung N, Thill K, Trang A, Fong BM, Nagasawa DT, Lim M, Yang I. The role of regulatory T-cells in glioma immunology. Clin Neurol Neurosurg 2014; 119: 125-132 [PMID: 24582432 DOI: 10.1016/j.clineuro.2013.12.004]
- 74 Jansen CS, Prokhnevska N, Master VA, Sanda MG, Carlisle JW, Bilen MA, Cardenas M, Wilkinson S, Lake R, Sowalsky AG, Valanparambil RM, Hudson WH, McGuire D, Melnick K, Khan AI, Kim K, Chang YM, Kim A, Filson CP, Alemozaffar M, Osunkoya AO, Mullane P, Ellis C, Akondy R, Im SJ, Kamphorst AO, Reyes A, Liu Y, Kissick H. An intra-tumoral niche maintains and differentiates stem-like CD8 T cells. Nature 2019; 576: 465-470 [PMID: 31827286 DOI: 10.1038/s41586-019-1836-5
- Kucharova K, Stallcup WB. NG2-proteoglycan-dependent contributions of oligodendrocyte 75 progenitors and myeloid cells to myelin damage and repair. J Neuroinflammation 2015; 12: 161 [PMID: 26338007 DOI: 10.1186/s12974-015-0385-6]
- 76 Khan AB, Carpenter B, Santos E Sousa P, Pospori C, Khorshed R, Griffin J, Velica P, Zech M, Ghorashian S, Forrest C, Thomas S, Gonzalez Anton S, Ahmadi M, Holler A, Flutter B, Ramirez-Ortiz Z, Means TK, Bennett CL, Stauss H, Morris E, Lo Celso C, Chakraverty R. Redirection to the bone marrow improves T cell persistence and antitumor functions. J Clin Invest 2018; 128: 2010-2024 [PMID: 29485974 DOI: 10.1172/JCI97454]
- 77 Bagley SJ, Desai AS, Linette GP, June CH, O'Rourke DM. CAR T-cell therapy for glioblastoma: recent clinical advances and future challenges. Neuro Oncol 2018; 20: 1429-1438 [PMID: 29509936 DOI: 10.1093/neuonc/noy032]
- 78 Rodriguez A, Brown C, Badie B. Chimeric antigen receptor T-cell therapy for glioblastoma. Transl Res 2017; 187: 93-102 [PMID: 28755873 DOI: 10.1016/j.trsl.2017.07.003]
- 79 Kleijn A, van den Bossche W, Haefner ES, Belcaid Z, Burghoorn-Maas C, Kloezeman JJ, Pas SD, Leenstra S, Debets R, de Vrij J, Dirven CMF, Lamfers MLM. The Sequence of Delta24-RGD and TMZ Administration in Malignant Glioma Affects the Role of CD8⁺T Cell Anti-tumor Activity. Mol Ther Oncolytics 2017; 5: 11-19 [PMID: 28480325 DOI: 10.1016/j.omto.2017.02.002]
- 80 Hol EM, Pekny M. Glial fibrillary acidic protein (GFAP) and the astrocyte intermediate filament system in diseases of the central nervous system. Curr Opin Cell Biol 2015; 32: 121-130 [PMID: 25726916 DOI: 10.1016/j.ceb.2015.02.004]
- 81 Ghandour MS, Labourdette G, Vincendon G, Gombos G. A biochemical and immunohistological study of S100 protein in developing rat cerebellum. Dev Neurosci 1981; 4: 98-109 [PMID: 7014178 DOI: 10.1159/000112745]
- Kegelman TP, Wu B, Das SK, Talukdar S, Beckta JM, Hu B, Emdad L, Valerie K, Sarkar D, 82 Furnari FB, Cavenee WK, Wei J, Purves A, De SK, Pellecchia M, Fisher PB. Inhibition of radiationinduced glioblastoma invasion by genetic and pharmacological targeting of MDA-9/Syntenin. Proc Natl Acad Sci USA 2017; 114: 370-375 [PMID: 28011764 DOI: 10.1073/pnas.1616100114]
- 83 Morgenroth A, Vogg AT, Ermert K, Zlatopolskiy B, Mottaghy FM. Hedgehog signaling sensitizes glioma stem cells to endogenous nano-irradiation. Oncotarget 2014; 5: 5483-5493 [PMID: 24978848 DOI: 10.18632/oncotarget.2123]
- Buechler MB, Turley SJ. A short field guide to fibroblast function in immunity. Semin Immunol 84 2018; 35: 48-58 [PMID: 29198601 DOI: 10.1016/j.smim.2017.11.001]
- 85 Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. Cell 2011; 144: 646-674 [PMID: 21376230 DOI: 10.1016/j.cell.2011.02.013]
- 86 Liu J, Chen S, Wang W, Ning BF, Chen F, Shen W, Ding J, Chen W, Xie WF, Zhang X. Cancerassociated fibroblasts promote hepatocellular carcinoma metastasis through chemokine-activated hedgehog and TGF-β pathways. Cancer Lett 2016; 379: 49-59 [PMID: 27216982 DOI:



10.1016/i.canlet.2016.05.022]

- 87 Valenti G, Quinn HM, Heynen GJJE, Lan L, Holland JD, Vogel R, Wulf-Goldenberg A, Birchmeier W. Cancer Stem Cells Regulate Cancer-Associated Fibroblasts via Activation of Hedgehog Signaling in Mammary Gland Tumors. Cancer Res 2017; 77: 2134-2147 [PMID: 28202523 DOI: 10.1158/0008-5472.CAN-15-3490]
- 88 Choi YP, Lee JH, Gao MQ, Kim BG, Kang S, Kim SH, Cho NH. Cancer-associated fibroblast promote transmigration through endothelial brain cells in three-dimensional in vitro models. Int J Cancer 2014; 135: 2024-2033 [PMID: 24643985 DOI: 10.1002/ijc.28848]
- 89 Clavreul A, Guette C, Faguer R, Tétaud C, Boissard A, Lemaire L, Rousseau A, Avril T, Henry C, Coqueret O, Menei P. Glioblastoma-associated stromal cells (GASCs) from histologically normal surgical margins have a myofibroblast phenotype and angiogenic properties. J Pathol 2014; 233: 74-88 [PMID: 24481573 DOI: 10.1002/path.4332]
- Neftel C, Laffy J, Filbin MG, Hara T, Shore ME, Rahme GJ, Richman AR, Silverbush D, Shaw ML, 90 Hebert CM, Dewitt J, Gritsch S, Perez EM, Gonzalez Castro LN, Lan X, Druck N, Rodman C, Dionne D, Kaplan A, Bertalan MS, Small J, Pelton K, Becker S, Bonal D, Nguyen QD, Servis RL, Fung JM, Mylvaganam R, Mayr L, Gojo J, Haberler C, Geyeregger R, Czech T, Slavc I, Nahed BV, Curry WT, Carter BS, Wakimoto H, Brastianos PK, Batchelor TT, Stemmer-Rachamimov A, Martinez-Lage M, Frosch MP, Stamenkovic I, Riggi N, Rheinbay E, Monje M, Rozenblatt-Rosen O, Cahill DP, Patel AP, Hunter T, Verma IM, Ligon KL, Louis DN, Regev A, Bernstein BE, Tirosh I, Suvà ML. An Integrative Model of Cellular States, Plasticity, and Genetics for Glioblastoma. Cell 2019; 178: 835-849 [PMID: 31327527 DOI: 10.1016/j.cell.2019.06.024]
- 91 Johung T, Monje M. Neuronal activity in the glioma microenvironment. Curr Opin Neurobiol 2017; 47: 156-161 [PMID: 29096244 DOI: 10.1016/j.conb.2017.10.009]
- 92 Venkatesh HS, Tam LT, Woo PJ, Lennon J, Nagaraja S, Gillespie SM, Ni J, Duveau DY, Morris PJ, Zhao JJ, Thomas CJ, Monje M. Targeting neuronal activity-regulated neuroligin-3 dependency in high-grade glioma. Nature 2017; 549: 533-537 [PMID: 28959975 DOI: 10.1038/nature24014]
- 93 Wang J, Xu SL, Duan JJ, Yi L, Guo YF, Shi Y, Li L, Yang ZY, Liao XM, Cai J, Zhang YQ, Xiao HL, Yin L, Wu H, Zhang JN, Lv SQ, Yang QK, Yang XJ, Jiang T, Zhang X, Bian XW, Yu SC. Invasion of white matter tracts by glioma stem cells is regulated by a NOTCH1-SOX2 positivefeedback loop. Nat Neurosci 2019; 22: 91-105 [PMID: 30559479 DOI: 10.1038/s41593-018-0285-z
- 94 Brücher BL, Jamall IS. Cell-cell communication in the tumor microenvironment, carcinogenesis, and anticancer treatment. Cell Physiol Biochem 2014; 34: 213-243 [PMID: 25034869 DOI: 10.1159/000362978
- 95 Maia J, Caja S, Strano Moraes MC, Couto N, Costa-Silva B. Exosome-Based Cell-Cell Communication in the Tumor Microenvironment. Front Cell Dev Biol 2018; 6: 18 [PMID: 29515996 DOI: 10.3389/fcell.2018.00018]
- 96 Sun Z, Shi K, Yang S, Liu J, Zhou Q, Wang G, Song J, Li Z, Zhang Z, Yuan W. Effect of exosomal miRNA on cancer biology and clinical applications. Mol Cancer 2018; 17: 147 [PMID: 30309355 DOI: 10.1186/s12943-018-0897-7]
- 97 Birbrair A. Stem Cell Microenvironments and Beyond. Adv Exp Med Biol 2017; 1041: 1-3 [PMID: 29204825 DOI: 10.1007/978-3-319-69194-7 1]
- Nagarsheth N, Wicha MS, Zou W. Chemokines in the cancer microenvironment and their relevance 98 in cancer immunotherapy. Nat Rev Immunol 2017; 17: 559-572 [PMID: 28555670 DOI: 10.1038/nri.2017.49
- Landskron G, De la Fuente M, Thuwajit P, Thuwajit C, Hermoso MA. Chronic inflammation and 99 cytokines in the tumor microenvironment. J Immunol Res 2014; 2014: 149185 [PMID: 24901008 DOI: 10.1155/2014/149185]
- 100 Kalluri R. The biology and function of exosomes in cancer. J Clin Invest 2016; 126: 1208-1215 [PMID: 27035812 DOI: 10.1172/JCI81135]
- 101 Wang J, Liu J, Sun G, Meng H, Wang J, Guan Y, Yin Y, Zhao Z, Dong X, Yin S, Li H, Cheng Y, Wu H, Wu A, Yu X, Chen L. Glioblastoma extracellular vesicles induce the tumour-promoting transformation of neural stem cells. Cancer Lett 2019; 466: 1-12 [PMID: 31521694 DOI: 10.1016/j.canlet.2019.09.004]
- 102 Reinhard J, Brösicke N, Theocharidis U, Faissner A. The extracellular matrix niche microenvironment of neural and cancer stem cells in the brain. Int J Biochem Cell Biol 2016; 81: 174-183 [PMID: 27157088 DOI: 10.1016/j.biocel.2016.05.002]
- 103 Harris NC, Achen MG. The proteolytic activation of angiogenic and lymphangiogenic growth factors in cancer--its potential relevance for therapeutics and diagnostics. Curr Med Chem 2014; 21: 1821-1842 [PMID: 24350854 DOI: 10.2174/0929867321666131217144550]
- Stupp R, Mason WP, van den Bent MJ, Weller M, Fisher B, Taphoorn MJ, Belanger K, Brandes 104 AA, Marosi C, Bogdahn U, Curschmann J, Janzer RC, Ludwin SK, Gorlia T, Allgeier A, Lacombe D, Cairncross JG, Eisenhauer E, Mirimanoff RO; European Organisation for Research and Treatment of Cancer Brain Tumor and Radiotherapy Groups; National Cancer Institute of Canada Clinical Trials Group. Radiotherapy plus concomitant and adjuvant temozolomide for glioblastoma. N Engl J Med 2005; 352: 987-996 [PMID: 15758009 DOI: 10.1056/NEJMoa043330]
- 105 Weller M, van den Bent M, Tonn JC, Stupp R, Preusser M, Cohen-Jonathan-Moyal E, Henriksson R, Le Rhun E, Balana C, Chinot O, Bendszus M, Reijneveld JC, Dhermain F, French P, Marosi C, Watts C, Oberg I, Pilkington G, Baumert BG, Taphoorn MJB, Hegi M, Westphal M, Reifenberger



G, Soffietti R, Wick W; European Association for Neuro-Oncology (EANO) Task Force on Gliomas. European Association for Neuro-Oncology (EANO) guideline on the diagnosis and treatment of adult astrocytic and oligodendroglial gliomas. Lancet Oncol 2017; 18: e315-e329 [PMID: 28483413 DOI: 10.1016/S1470-2045(17)30194-8]

- 106 Diaz RJ, Ali S, Qadir MG, De La Fuente MI, Ivan ME, Komotar RJ. The role of bevacizumab in the treatment of glioblastoma. J Neurooncol 2017; 133: 455-467 [PMID: 28527008 DOI: 10.1007/s11060-017-2477-x]
- Wang Y, Meng X, Zhou S, Zhu Y, Xu J, Tao R. Apatinib Plus Temozolomide for Recurrent 107 Glioblastoma: An Uncontrolled, Open-Label Study. Onco Targets Ther 2019; 12: 10579-10585 [PMID: 31819537 DOI: 10.2147/OTT.S226804]
- 108 Preusser M, Lim M, Hafler DA, Reardon DA, Sampson JH. Prospects of immune checkpoint modulators in the treatment of glioblastoma. Nat Rev Neurol 2015; 11: 504-514 [PMID: 26260659 DOI: 10.1038/nrneurol.2015.139]
- Johnson DB, Sullivan RJ, Menzies AM. Immune checkpoint inhibitors in challenging populations. 109 Cancer 2017; 123: 1904-1911 [PMID: 28241095 DOI: 10.1002/cncr.30642]
- 110 Labanieh L, Majzner RG, Mackall CL. Programming CAR-T cells to kill cancer. Nat Biomed Eng 2018; **2**: 377-391 [PMID: 31011197 DOI: 10.1038/s41551-018-0235-9]
- 111 Takebe N, Miele L, Harris PJ, Jeong W, Bando H, Kahn M, Yang SX, Ivy SP. Targeting Notch, Hedgehog, and Wnt pathways in cancer stem cells: clinical update. Nat Rev Clin Oncol 2015; 12: 445-464 [PMID: 25850553 DOI: 10.1038/nrclinonc.2015.61]
- Massard C, Azaro A, Soria JC, Lassen U, Le Tourneau C, Sarker D, Smith C, Ohnmacht U, Oakley 112 G, Patel BKR, Yuen ESM, Benhadji KA, Rodon J. First-in-human study of LY3039478, an oral Notch signaling inhibitor in advanced or metastatic cancer. Ann Oncol 2018; 29: 1911-1917 [PMID: 30060061 DOI: 10.1093/annonc/mdy244]





Published by Baishideng Publishing Group Inc 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA Telephone: +1-925-3991568 E-mail: bpgoffice@wjgnet.com Help Desk: https://www.f6publishing.com/helpdesk https://www.wjgnet.com

