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REVIEW

Communication between bone marrow mesenchymal stem cells and multiple myeloma cells: Impact on disease progression

Daniel García-Sánchez, Alberto González-González, Ana Alfonso-Fernández, Mónica Del Dujo-Gutiérrez, Flor M Pérez-Campo

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Abstract

Multiple myeloma (MM) is a hematological malignancy characterized by the accumulation of immunoglobulin-secreting clonal plasma cells at the bone marrow (BM). The interaction between MM cells and the BM microenvironment, and specifically BM mesenchymal stem cells (BM-MSCs), has a key role in the pathophysiology of this disease. Multiple data support the idea that BM-MSCs not only enhance the proliferation and survival of MM cells but are also involved in the resistance of MM cells to certain drugs, aiding the progression of this hematological tumor. The relation of MM cells with the resident BM-MSCs is a two-way interaction. MM modulate the behavior of BM-MSCs altering their expression profile, proliferation rate, osteogenic potential, and expression of senescence markers. In turn, modified BM-MSCs can produce a set of cytokines that would modulate the BM microenvironment to favor disease progression. The interaction between MM cells and BM-MSCs can be mediated by the secretion of a variety of soluble factors and extracellular vesicles carrying microRNAs, long non-coding RNAs or other molecules. However, the communication between these two types of cells could also involve a direct physical interaction through adhesion molecules or tunneling nanotubes. Thus, understanding the way this communication works and developing strategies to interfere in the process, would preclude the expansion of the MM cells and might offer alternative treatments for this incurable disease.

Key Words: Multiple myeloma; Mesenchymal stem cells; Bone marrow microenvironment; Soluble factors; Extra-cellular vesicles; Cells adhesion molecules; Tunnelling-



nanotubes

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Core Tip: Mesenchymal stem cells (MSCs), the main cell population of the bone marrow (BM) stroma, can influence BM microenvironment through their paracrine activity, involving both soluble factors and extracellular vesicles, but also through direct communication. Being the BM the predominant localization of multiple myeloma cells (MM), finding the appropriate conditions at this niche, is key for the survival and expansion of tumour cells and thus, for the progression of the disease. Since the activity of BM-MSCs could determine the fate of MM cells at BM, these cells could be interesting targets for the design of new antitumor drugs.

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INTRODUCTION

Multiple myeloma (MM) is one of the most common hematological diseases, only second to non-Hodgkin lymphoma[1]. MM affects mainly older adults, with the median age of diagnosis being around 69 years. Only in 2020, 32270 new cases and 12830 deaths in the United States were estimated by the American Cancer Society Statistics Centre. In global terms, the cases would reach 160000, accounting for 0.9% of all cancer diagnosis. Importantly, incidence of MM has risen 126% globally, and hence, there is an increasing need to find new effective treatments for this incurable disease [2,3].

Besides the initial treatments for MM, consisting in alkylating agents often combined with corticosteroids, the last couple of decades have seen an important advance in the available treatments for this disease. We first saw the introduction of proteasome inhibitors (Bortezomib), histone deacetylase inhibitors (Panobinostat) and drugs such as Selinexor, with a nuclear export inhibition activity. In recent years monoclonal antibodies such as Daratumumab (anti-CD38) or Elotuzumab (anti-SLAMF7), and more recently the use of chimeric antibody receptor (CAR) T-cell products, has introduced immunotherapy as a viable approach to MM treatment^[4]. According to data from the National Cancer Institute (Bethesa, MD, United States), all these treatments have had a deep impact on patients' survival, substantially raising the survival rate to 55% in the period between 2011 and 2017. More recently, the use of small molecules, with a molecular weight smaller than 1kDa, has also improved treatments, since it offers important advantages compared to the former therapies, as the easy cell entry, the simplicity of the molecules, and a much lower production cost than other drugs^[5]. However, despite these advancements, there are still limitations to existing treatment options. Some patients may not respond to or may develop resistance to certain medications, many patients can become refractory to treatment and thus, there is a high risk of relapse. This promotes the search for new treatments to handle relapsed or refractory MM.

MM is caused by aberrant plasma cells (PC) proliferation in the bone marrow (BM). The premalignant states, known as monoclonal gammopathy of undetermined significance (MGUS) and smoldering MM, transition under specific conditions to the malignant state of PC leukemia or extramedullary myeloma [6]. A key characteristic of MM is the infiltration into and the colonization of the BM, one of the two primary lymphoid organs^[7]. This colonization produces typical lytic bone lesions that would be present in approximately 80% of patients with newly diagnosed MM and are the major source of morbidity[8]. The bone lesions, resulting from the stimulation of bone resorption by B-cell plasmacytomas, are associated with hypercalcemia and often, severe bone pain and bone fractures[8,9].

While the initiation of a tumor mainly depends on the accumulation of genetic defects, the transition from a premalignant to a malignant state highly relies on the interaction of the tumor cell with a permissive microenvironment that would support the malignant transformation and the proliferation of the tumor cells, aiding them to evade apoptosis. The relevance of tumor microenvironment in disease progression was first discussed in the "seed and soil hypothesis" formulated by Stephen Paget in 1889, where the establishment of tumor metastatic sites is influenced by the cross-interaction between the seeds (cancer cells) and the soil (a particular microenvironment)[10]. This is not different in MM[11,12]. The progression to MM, which would occur in approximately 50% of patients diagnosed with MGUS [6], requires multiple genomic events, but also a permissive BM microenvironment[13]. MM cells proliferate almost exclusively within the BM niche, highlighting the role of this microenvironment in



supporting cancer growth. In fact, there is also mounting evidence indicating this BM microenvironment is not only key for PCs survival, but also has a crucial role in resistance to treatment and disease recurrence^[14,15]

The MM cells infiltrating the BM will encounter a complex microenvironment formed by cellular and non-cellular components. Amongst the non-cellular components influencing the BM microenvironment, it is important to consider the extracellular matrix (ECM) proteins as well as a milieu of cytokines, chemokines, and growth factors. Many of these factors can have a positive effect on MM cells, boosting their proliferation and survival and the resistance to different types of drugs. A good example of these cytokines supporting MM progression are interleukin (IL)-6 and ligands of the B-cell maturation antigen, such as a proliferation-inducing ligand and B-cell activating factor (BAFF)[16,17]. Regarding the cellular components of the BM niche, many different types of bone cells (osteoblasts, or bone forming cells, and osteoclasts, or bone resorbing cells, and osteocytes) and cells from the immune system (macrophages, natural killer cells and regulatory T-cells) share this niche. Other cells present here are fat cells (adipocytes), fibroblasts, endothelial cells and two multipotent stem cells, BM mesenchymal stem cells (BM-MSCs), which differentiate into different mesodermal cell lineages, and hematopoietic stem cells (HSCs), that would differentiate into hematological lineages, including the myeloid lineage that would give rise to osteoclasts. MM cells are likely to interact with all the cells in the BM niche and elicit mutual influence[18]. In fact, it is known that communication between MM cells and BM-MSCs is essential in the progression of MM[19]. Once MM cells infiltrate the BM, their presence in the BM niche alters the activity of many of the cells found there, including those involved in bone homeostasis such as osteoclasts[20,21] and osteoblasts[22-24]. While in normal bone homeostasis, the activities of osteoblast and osteoclasts are carefully balanced to ensure a correct bone regeneration, the influence of MM cells disrupts this balance increasing both the resorptive activity of osteoclast and their numbers, and decreasing osteoblasts numbers as well as their osteogenic capacity^[25], overall leading to an increase in bone destruction and the appearance of the aforementioned osteolytic lesions typical of this disease. Other cells at the BM niche which activity is highly influenced by MM cells are BM-MSCs. The presence of MM cells at the BM niche alters the MSCs behavior in different ways. In fact, changes in the expression of certain microRNAs (miRNAs) in BM-MSCs leading to important alterations of their secretory profile and osteogenic differentiation potential have been observed after co-cultivation of BM-MSCs and MM cells[26,27]. These changes at the BM niche upon MM invasion produce a microenvironment that would support disease progression. Indeed, there is strong evidence indicating that is precisely this interaction what leads to the formation of the lytic bone lesions^[28]. One of the characteristics of this permissive microenvironment is the high presence of pro-inflammatory cytokines that would favor the progression of neoplasia^[29]. The crosstalk between MM cells and the BM-MSCs at the BM niche is key to sustain this pro-inflammatory microenvironment and thus, to allow MM cell persistence and growth[30]. It is important to clarify, that this pro-inflammatory microenvironment would be the result of the action not only of the infiltrated MM cells but also of other cells residing at the BM niche, including BM-MSCs.

MSCs, have a key role in regulating the BM microenvironment through their paracrine activity, but also through direct cell-to-cell interaction. Regarding their paracrine activity, these cells produce a plethora of soluble biomolecules and vesicular components, known altogether as "secretome", that exert multiple actions on other cells at the BM microenvironment[31]. BM-MSCs role in MM disease development and progression has been reported as having both inhibitory[32] and supportive roles[33, 34]. Sadly, the latter is the most frequent. Once at the BM niche, MM will exert their influence on resident MSCs, altering their signaling and gene expression pattern and thus, also their secretion pattern. After interaction with MM cells, MSCs will produce a secretome rich in pro-inflammatory cytokines. In fact, it has been previously described how MSCs react to IL-1 produced by the myeloma PCs by producing large quantities of IL-6, a cytokine that would in turn stimulate the survival of the MM cells[35,36]. Therefore, the soluble part of this secretome has a key role in the progression of tumor. Moreover, in the last few years, several molecules (miRNAs) that are present in the cargo in the extracellular vesicles (EVs) produced by BM-MSCs upon MM cells stimulation also seem to have a key role in the disease promotion. Although the soluble proteins and EVs produced by the BM-MSCs are the main actors in the communication between BM-MSCs and MM cells, other ways of communication have also been implicated. This will be discussed in the following sections.

Current available treatments for MM patients mainly target MM cells but have none or limited effect on other cells in the BM or de BM microenvironment. Knowledge of the different interactions between BM-MSCs and MM cells is key to understand how MM cells behave and grow within the BM and how osteolytic lesions are formed. In this work, we will address key aspects of the different ways of communication between MSCs and MM cells as well as the outcome of this crosstalk.

SOLUBLE FACTORS IN THE COMMUNICATION BETWEEN BM-MSCs AND MM CELLS

The multiple cellular interactions taking place in the BM, make this microenvironment a dynamic compartment with a myriad of soluble factors that would affect the behavior of the various cell types



concurring at that microenvironment. Although many of those cells have paracrine activity, BM-MSCs are the ones that have a stronger impact in the BM microenvironment due to the wide variety of soluble and non-soluble factors secreted by these cells. Various constituents of the, so called, BM-MSC secretome orchestrate the fate of the MM cells, from the first step encompassing the homing of those cells to the BM, onwards.

Role of soluble factors in the homing of MM cells to the BM

A key factor in the communication between BM cells and MM cells during the first stages of BM colonization, is the cytokine stromal cell derived factor 1α (SDF1α), also known as CXCL12. This factor, produced by BM-MSCs, works as a chemoattractant, being responsible of the homing of HSCs to the BM once they abandon the fetal liver during development[37]. SDF1α activity is mediated by the binding to a specific G-protein 7-span transmembrane receptor (CXCR4) at the target cells. CXCR4 is expressed at the surface of different cells in the BM microenvironment[38], and also at the surface of MM cells and other tumor cells[39]. Thus, SDF1 α /CXCR4 interaction might have a relevant role in directing de metastasis of hematopoietic malignancies. Similar to its effect on HSCs, the interaction of $SDF1\alpha$ with its receptor at the MM cells, increases their migration, homing and adhesion towards the BM, in fact, knock down of CDCR4 in BM-MSCs or the use of the CXCR4 inhibitor AMD3100 (AnorMED), that blocks the binding of SDF1 α to its receptor[40], seems to inhibit the migration of MM towards the BM[41]. The binding of SDF1a to its receptor at the MM cells, also triggers the activation of the phosphatidylinositol 3-kinase (PI3K) and the MAPK kinase (MEK)-extracellular signal regulated kinase (ERK, MEK/ERK) pathways, inducing a rearrangement in the cytoskeleton of MM cells that facilitates BM colonization [41]. SDF1 α has also been described to act in a more indirect way, not mediated by the binding to CXCR4. SDF1a interacts with other molecules including matrix metalloproteinases (MMPs), integrins or growth factors such as hepatocyte growth factor (HGF), insulin like growth factor-1 (IGF-1) or molecules of the GTPases family. All of these effects elicited by $SDF1\alpha$, in one way or another, lead to a promotion in MM cells migration, homing or adhesion into the BM[38].

Role of soluble factors in the promotion of proliferation and MM cell survival

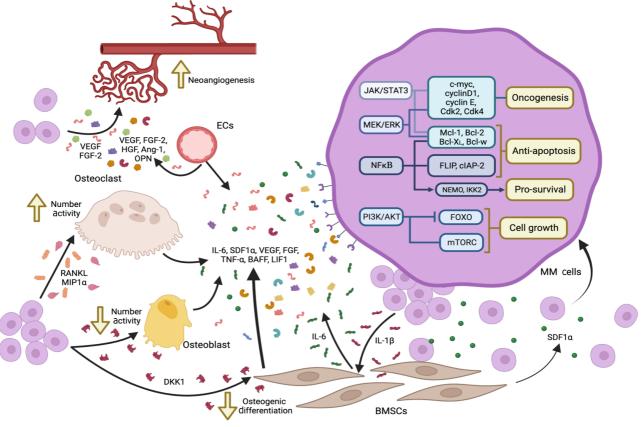
Many of the factors secreted by BM-MSCs and by other cells of the BM microenvironment, activate key signaling pathways in the MM cells that would increase their chances to survive and proliferate in the BM microenvironment. A summary of these factors as well as the signaling pathways involved in this communication are shown in Figure 1. In fact, some mutations activating those pathways have also been found in patients with MM. We will address some of those key pathways in this section.

Once in the BM, for the tumor to progress further, MM cells would need a permissive microenvironment. This microenvironment would be created by multiple soluble factors secreted by the different cell types present at the BM. The soluble factors produced by the BM-MSCs seem to be the main, but not the only, effectors of the changes elicited in the MM cells. Besides $SDF1\alpha$, BM-MSCs seem to secrete other important soluble factors such as IL-6, IL-17, vascular endothelial growth factor (VEGF), fibroblast growth factors (FGF), tumor necrosis factor- α (TNF- α), BAFF or leukemia inhibitory factor-1; osteoclasts mainly secrete IL-6 and VEGF; and vascular endothelial cells secrete cyclophilin-A[42,43]. These factors will activate specific signaling pathways in the MM cells such as PI3K/Akt, MEK/ERK, Janus kinase 2 (JAK2)-signal transducer and activator of transcription 3 (STAT3, JAK2/STAT3) pathways, related to cell survival, proliferation and drug resistance[43]. It is important to highlight that this communication is bi-directional, since MM cells would also produce cytokines such as IL-1β, VEGF, and transforming growth factor-beta (TGF- β) that would exert their effect on BM-MSCs, activating the nuclear factor kappa-B (NFkB) pathway and thus, inducing further secretion of cytokines by the BM-MSCs into the BM microenvironment, particularly IL-6[44,45].

IL-6 is the main activator of the JAK2/STAT3 pathway, known to be implicated in the pathogenicity of cancer. JAK2/STAT3 pathway activation promoted by IL-6 leads in MM cells to the expression not only of potent proto-oncogenes such as c-myc and cyclin D1, but also of anti-apoptotic genes like Mcl-1, Bcl-XL and Bcl-2. Moreover, STAT3 activation has also a immunosuppressive effect since it regulates Tcell mediated cytotoxic immune response[46], contributing to the establishment of a immunosuppressed microenvironment that would contribute to the survival and proliferation of the MM cells in the BM. On the other hand, IL-6 activation of JAK2/STAT3 pathways, also has an important role in bone destruction, a hallmark of MM. IL-6/JAK2/STAT3 axis induces the expression of the receptor activator of NFkB ligand (RANKL)[36,47] whose binding to its receptor at the surface of pre-osteoclasts, promotes their differentiation towards mature osteoclasts, activating bone resorption and thus, promoting the formation of osteolytic lesions.

It is important to highlight that the NFxB signaling pathway also has an important role in the survival of MM cells and in the maintenance of the tumorigenic microenvironment at the BM. Both canonical and non-canonical NF-kB pathways are activated by different factors present in the BM microenvironment, including IL-6, IGF-1, TNF- α or BAFF[48]. While IGF-1 is able to activate NF κ B pathway, inducing the expression of anti-apoptotic, caspase-8 inhibitors FLIP and cIAP-2[49], TNF- α has a pro-survival effect through NFxB pathway mediators such as NFxB (NEMO) and IxB kinase subunit 2[44]. On the other hand, BAFF activates NFkB non-canonical pathway upregulating the expression of antiapoptotic proteins including Mcl-1, Bcl-XL, Bcl-w and Bcl-2[50]. There is also evidence indicating that IL-6 is





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Figure 1 Schematic representation of the main factors involved in the bidirectional communication between multiple myeloma cells and cells in the bone marrow microenviroment (bone marrow mesenchymal stem cells, osteoclasts, osteoblast, etc.). The main signaling patwthays activated by these factors are also depicted (Created with Biorender.com). VEGF: Vascular endothelial growth factor; FGF: Fibroblast growth factors; HGF: Hepatocyte growth factor; OPN: Osteopontin; ECs: Endothelial cells; IL: Interleukin; SDF1a: Stromal cell derived factor 1a; TNF-a: Tumor necrosis factor-a; BAFF: Bcell activating factor; DKK-1: Dickkopf-1; MM: Multiple myeloma; BM-MSC: Bone marrow mesenchymal stem cells; JAK: Janus kinase; STAT3: Signal transducer and activator of transcription 3; NFkB: Nuclear factor kappa-B; PI3K: Phosphatidylinositol 3-kinase; RANKL: Receptor activator of NFkB ligand; Ang-1: angiopoietin-1; MEK: MAPK kinase; ERK: Extracellular signal regulated kinase; LIF1: Leukemia inhibitory factor-1. Created with BioRender.com.

> linked to the expression of VEGF in MM cells, being some of the VEFG isoform expression driven by the NFκB pathway[51,52].

> The MEK/ERK pathway is the signaling pathway most found activated in MM patients, with a prevalence in between 43% and 53% of the patients[53]. Changes in MEK/ERK pathway have important effects in cell cycle, due to the alteration in the expression of molecules such as cyclin D1, cyclin E, Cdk2 and Cdk4 and in apoptosis prevention by the induction of the phosphorylation of the pro-apoptotic protein Bim. This phosphorylation results in the release of anti-apoptotic molecules such as Mcl-1, Bcl-XL and Bcl-2, also related to Akt pathway [54]. In the absence of mutations that activate this pathway, the stimulation of the MEK/ERK pathway in the MM cells might also occur by the action of different soluble factors present in the BM microenvironment such as BAFF, IL-6, SDF1 α , VEGF or TNF- α among others[42]. As with other relevant signaling pathways that become activated in MM, the MEK/ERK is also studied as a potential therapeutic target.

> PI3K/Akt signaling pathway also has a relevant role in cell proliferation, cell cycle and apoptosis. Alteration of the PI3K/Akt/mTOR pathway due to genetic modifications or its hyper-activation contributes to carcinogenesis, metastasis, invasion, proliferation and drug resistance of tumor cells. However, no activating mutations have been described in MM cells yet. Despite this fact, PI3K/Akt/ mTOR pathway is important for MM cells survival[55,56].

Role of soluble factors in angiogenesis and bone homeostasis

Up to this point, we have mentioned some of the effects of the pro-tumorigenic microenvironment in the BM on the MM cell survival and growth. However, once modified by the BM microenvironment, MM cells will start to release different soluble factors that will not only perpetuate that tumorigenic microenvironment, but also will have a deep impact in angiogenesis and bone homeostasis.

Neovascularization in the bone is an essential feature for MM progression and the presence of high density of micro-vessels in the BM microenvironment is characteristic in MM. Cells residing at BM, such as BM-MSCs, osteoblasts, HSCs, or endothelial precursor cells, commonly express various angiogenic



factors, such as VEGF, FGF-2, TNF-a, HGF, IL-6, BAFF, SDF-1a, angiopoietin-1 or osteopontin (OPN). Also, MM cells are able to directly produce VEGF stablishing a VEGF autocrine loop where the produced VEGF would stimulate MM cells proliferation through the MEK-1/ERK pathway[57,58]. FGF-2 is another key pro-angiogenic molecule that would be produced by both MM cells and BM-MSCs[59]. However, contrary to VEGF, which is produced by all MM cells, FGF-2 production by MM does not seem to be a general feature in all MM cases [59]. Other molecules with pro-angiogenic activity such as MMPs[60,61] or OPN, also produced by MM cells, have also a relevant role in promoting micro-vessels formation in the BM microenvironment. The overall increase in the production of such angiogenic factors is elicited by the MM cells. The activation of angiogenesis linked to tumor progression is known as "angiogenic switch" [62].

Bone homeostasis is a dynamic process driven by osteoclasts, osteoblast and osteocytes. Alterations in the balance between these cell types will lead to the remodeling of the bone. The characteristic bone lesions found in MM derive from the disruption of bone homeostasis initiated by the activation of JAK2/STAT3 pathway by IL-6 and the subsequent induction of RANKL expression by MM cells. Not only this but, as will be discussed later, cell-to-cell interaction of MM cells with BMSCs also induce the expression of the macrophage inflammatory protein (MIP)-1 α [63]. Both RANKL and MIP-1 α are mediators in the bone destruction driven by MM as they have an both in the number of osteoclasts and in their activity. MIP-1 α is a chemoattractant for osteoclasts and stimulates osteoclast formation[64], while RANKL after being recognized by its receptor RANK, will induce the commitment of the macrophage/monocyte precursor cells to the osteoclast lineage[65].

Secreted by MM cell in response to the activation of the JNK pathway, Dickkopf-1 (DKK-1) is also a disruptor in bone homeostasis[66]. DKK-1 is an extracellular inhibitor of the Wnt pathway. DKK-1 interacts with membrane receptors as transmembrane proteins Kremen 1/2 and the human low-density lipoprotein receptor-related protein 5/6, thus competing with Wnt[67]. As one of the main regulatory pathways for osteogenic differentiation of BM-MSCs into osteoblasts[68], the inhibition of the Wnt/ β catenin pathway by DKK-1 will result in a reduced number of osteoblasts. By the action of these factors, RANKL, MIP-1 α and DKK-1, the balance between bone formation and bone resorption driven by osteoblasts and osteoclasts is disrupted, resulting in the characteristic bone lesions present in MM patients.

A table summarizing the latest scientific evidence regarding key factors involved in MM/BM-MSCs communication and their effect is shown (Table 1).

EVs-MEDIATED COMMUNICATION BETWEEN BM-MSCs AND MM CELLS

Under non-pathological conditions, BM homeostasis is maintained by cell-to-cell contact, soluble molecules, and EVs. Whereas, over the years solid evidence has accumulated about the relevance of the first two, the involvement of EVs-mediated communication in the maintenance of BM homeostasis has started to be contemplated only in the last few decades[69]. Despite being a fairly new field, important advances have been made in the knowledge of EVs, such as their classification, in terms of their size and biogenesis, into three major categories (exosomes, micro-vesicles, and apoptotic bodies) and the fact that its content varies according to the state of their parental cells[31].

As we have previously discussed, MM cells have the capacity to alter the environment in which they reside[70] as well as the characteristics of cells present in that microenvironment. Thus, it is not surprising that the EVs produced by MM cells also play a key role in disease progression. In fact, it has recently been shown that, exosomes (a particular class of EVs) produced by both BM-MSCs and MM cells are largely responsible for MM pathogenesis[71]. This recent demonstration of the relevance of EVs in MM progression has resulted in several studies in the lats few years, however, the multitude of agents and interactions involved in the development and progression of this disease has made it difficult to fully understand the molecular mechanisms involved. In this section we aim to gather the available information so far.

Effect of MM-EVs on the BM-MSCs and bone homeostasis

As previously mentioned, osteolysis, one of the main hallmarks of MM disease, is linked to the negative effect of MM cells on cells responsible for bone homeostasis, such as MSCs, osteoblasts and osteoclasts [72]. In particular, myeloma bone disease (MBD) has a unique feature compared to other diseases that encompass bone destruction, since in MBD osteoblast activity is also severely impaired [24]. Several authors have suggested that an essential part of this bone damage is related to EVs directly produced by MM cells (MM-EVs). Zhang et al^[73] demonstrated that the cargo of MM-EVs was enriched in various molecules which negatively regulate osteogenesis. They confirmed that MM-EVs induced high expression of miR-103a-3p in BM-MSCs, which led to impaired osteogenesis in vitro. Moreover, they showed that injection of MM-EVs in mouse tibia resulted in defective bone formation. Interestingly, in vitro assays also revealed that MM-EVs were also able to influence MM cells increasing viability and IL-6 production, known to regulate MM cell proliferation thus, establishing an autocrine feedback. MM-EVs also increased miR103a-3p expression in MM cells however, in those cells the increased prolif-



Table 1 Summary for the key soluble factors involved in multiple myeloma/bone marrow mesenchymal stem cells communication						
Soluble factors	Origin	Function	Ref.			
SDF1a	BMSCs	Chemoattractant of MM cell towards the BM microenvironment	[38]			
IL-1β	MM cells	Act over BMSCs inducing the secretion of soluble factors, mainlyIL-6	[45]			
IL-6	BMSCs	Closely related with cancer pathogenicity due to it proto-oncogenic and anti-apoptotic effect over MM cells	[46,47]			
		Immunosuppressive effect over T cells				
		Also related with bone destruction by inducing the expression of RANKL by the MM cells				
VEGF	BM cells, MM cells	Promotes bone neovascularization, essential for tumour progression	[58]			
RANKL	MM cells	Induce the commitment of the macrophage/monocyte precursor cells to the osteoclast lineage. Promoting indirectly bone destruction	[<mark>63</mark>]			
DKK-1	MM cells	Disruptor of bone homeostasis by inhibiting BMSCs differentiation into osteoblasts	[66,67]			

SDF1a: Stromal cell derived factor 1a; BM-MSC: Bone marrow mesenchymal stem cells; MM: Multiple myeloma; IL: Interleukin; RANKL: Receptor activator of NFkB ligand; VEGF: Vascular endothelial growth factor; DKK-1: Dickkopf-1.

> eration of MM cells after exposures to MM-EVs does not seem to be related to miR103a-3p but to other miRNAs also present in the MM-EVs cargo, such as miR107 and miR181a-3p[24].

> Among the different biomolecules found as part of the exosome cargo, long non-coding RNAs (lncRNAs) and miRNAs have been the focus of attention due to their key regulatory roles. Various miRNAs found in MM-EVs have been studied for their involvement in the disruption of osteogenesis. miR-129-5p was identified as a player in vesicle-mediated bone disease[74]. In particular, miR-129-5p seemed to inhibit the transcription factor specificity protein 1, leading to a reduction of ALPL, both at the mRNA and protein levels, during the early osteogenic differentiation of MSCs. On the other hand, the long non coding RNA Long Intergenic Non-Protein Coding RNA 461, found as part of the MMexosomes cargo, has also been found to inhibit osteoblast differentiation by reducing the activity of Wnt/ β -Catenin pathways, responsible for osteoblast proliferation, differentiation and activity [75]. Other molecules, such as soluble proteins present in the MM-EVs cargo also showed anti-osteogenic activity Faict *et al*[72] revealed that Wnt/ β -Catenin inhibitor DKK-1 is present in MM-EVs and observed a lower expression of Osterix (OSX), Collagen 1A1 and alkaline phosphatase in differentiated MC3T3-E1 cells after MM-EVs treatment.

> Runx2 is the master regulator of early osteogenic differentiation, and therefore a possible target for the anti-osteogenic effect of MM-EVs. In fact, IncRNA RUNX2-AS1 present in the MM-EVs cargo was identified as a bioactive molecule able to reach MSCs and form a transcriptionally repressed RNA duplex with RUNX2 premRNA, reducing the osteogenic activity[76]. In addition, a MM-EVs impact in osteoblastic differentiation through reduction of Runx2, together with OSX and OCN, has been described by Liu et al [77]. These authors also record increased levels of IL-6 secretion via APE1/NF-kB which, as aforementioned, is an important survival factor of MM cells.

> Once the EVs produced by MM cells reach the BM-MSCs, their cargo modifies the BM-MSCs behaviour in the benefit of MM cells. A clear example of this is miR-146a which acts in a positive loop to favor disease progression[19]. Once this miRNA targets BM-MSCs, it produces an increase in the secretion of several cytokines and chemokines from those cells, including CXCL1, IL-6, IL-8, inducible protein 10 (IP-10), monocyte chemoattractant protein 1 (MCP-1), and CCL-5, which, in turn, once released into the BM microenvironment, would favor MM cell viability and migration. In addition, MM-EVs cargo miR-146a and miR-21, participate in proliferation and transformation of MSCs into cancer associated fibroblasts (CAFs). This is a type of cell which could contribute to a tumour-supportive microenvironment through secretion of cytokines, including IL-6 and TGF- β [78].

> Interestingly, it has been shown that conventional chemotherapeutic agents including melphalan, and anti-proteases such as bortezomib and carfilzomib can stimulate a considerable MM-EVs release. The EVs produced under these circumstances are called "chemoexosomes". These chemoexosomes are characterized by the high presence of the heparanase enzyme in their surface. This heparanase is implied in several cellular changes leading to chemoresistance and the subsequent relapse of the patient. Heparanase EVs content is delivered in MM cells and activate ERK pathway as well as $TNF-\alpha$ production by macrophages, matrix degradation and migration promotion[71].

Effect of EVs produced by BM-MSCs on MM cells and MM progression

So far, we have analyzed the influence of MM-EVs on BM-MSCs, however, this communication, as previously mentioned, is bidirectional. In 2016, Wang et al[69] showed that BM-MSC-EVs from MM patients contained a lower level of the tumor suppressor miR-15a, and higher levels of oncogenic



proteins, cytokines, and adhesion molecules, when compared to EVs from healthy BM-MSCs. Cytokines such as IL-1ra, interferon-IP-10, MCP-1, MIP-1a, MIP-1β, and SDF1a were detected in murine BM-MSC-EVs. They confirmed that BM-MSC-EVs from MM patients act on MM cells activating proliferation, survival, and migration, as well as drug resistance to bortezomib, a widely used clinical drug for MM treatment.

In a similar study, a reduction of mir-15a levels in the cargo of BM-MSCs-EVs from MM patients was also detected. This change was shown to promote cell proliferation and dissemination or metastasis to other niches, which is a hallmark of MM. The same authors also revealed the importance of some of the proteins present in BM-MSCs-EVs cargo, as they detected higher content levels of IL-6, CCL2, γ-catenin and fibronectin, which are key to MM pathogenesis^[70]. Other miRNAs cargo were also implicated in these processes. miR-483-5p was found packed in BM-MSCs-EVs and was responsible for promoting MM cell proliferation and reduced apoptosis via the miR-483-5p/TIMP2 axis[79]. Umezu et al[80] highlighted the role of miR-10a in MM disease since its transference via BM-MSC-EVs promoted cell proliferation in several MM cell lines (RPMI 8226, KMS-11, and U266) compared to BM-MSC-EVs with miR-10a blocked. Moreover, Gao et al [81] studied miR-155 present in BM-MSC-EVs cargo, which turned out to be involved in viability, stemness and drug resistance in MM cells. The role of miR-155 was underscored by the fact that incubation of the MM cells line mitochondrial pyruvate carrier 11 (MPC-11) with miR-155-mimics for 24 h resulted in a significantly reduced cell apoptosis in vitro and augmented expression of stemness maintenance markers OCT-4 and Nanog and drug resistance-associated proteins MRP1, ABCG2 and P-g.

As in the previous section, a table summarizing the main works referred to the relevance of communication between MM cells and BM-MSCs through EVs and the role of their cargo is shown (Table 2).

The resistance to treatment is precisely one of the major problems in MM at the clinical level, as this is directly responsible for the relapses. Some studies investigating the mechanisms behind this resistance have highlighted the implication of the activation of several signaling pathways, including p38, p53, c-Jun N-terminal kinases and Akt through the assessment of bortezomib treatment. The role of BM-MSC-EVs in interfering with the antitumor effect developed by bortezomib in MM was confirmed through different experiments. BM-MSC-EVs were able to alter apoptosis-related proteins Bcl-2, Bax, caspase-8, caspase-9, and caspase-3 promoting an antiapoptotic profile in both murine and human cells. These EVs blocked the significant reduction of Bcl-2 expression caused by bortezomib and reduced cleaved caspase-9, caspase-3, and PARP either in the absence or presence of bortezomib. Moreover, the use of GW4869, a neutral sphingomyelinase inhibitor of the formation of exosomes by the ceramide pathway, in combination with bortezomib treatment led to a significant effect on tumor load reduction[71,82].

In conclusion, the two-way communication between MM cells and BM-MSCs mediated by EVs is extremely intricate and plays a pivotal role in the progression of the disease. Since BM-MSCs-EVs have a key role in supporting MM development, this could become a key target to develop new therapies for the treatment of this hematological disease.

COMMUNICATION THROUGH CONTACT DEPENDENT MECHANISMS

As well as the already described interactions through paracrine secretion of different cytokines and EVs, MM cells also interact with BM-MSCs by direct cell-to-cell contact. These cell-to-cell interactions are not restricted to MM and BM-MSCs since MM cells also interact with other cells of the BM microenvironment such as osteoclasts and osteoblasts, endothelial cells, and lymphocytes. It is known that these contacts are also key to protect MM cells against chemotherapy, helping them to accumulate inside the BM[83], to adhere to endothelium, and to spread into the bloodstream[84], although the detailed mechanisms involved in those processes have not been completely elucidated[85].

Cell adhesion molecules in MM/BM-MSCs communication

Direct cell-to-cell adhesion and communication mechanisms have been known for more than 40 years [86,87]. These cell-to-cell communication is mediated by Cell Adhesion Molecules (CAMs), a subcategory of adhesion proteins located at the cell surface, involved in binding either to other cells, or in attaching cells to proteins of the ECM[88], suchas fibronectin, laminin or collagen (Figure 2). While it has been well documented that the ECM promotes the survival of different types of tumors, much less is known about the influence of the direct contact of BM-MSCs in their progression.

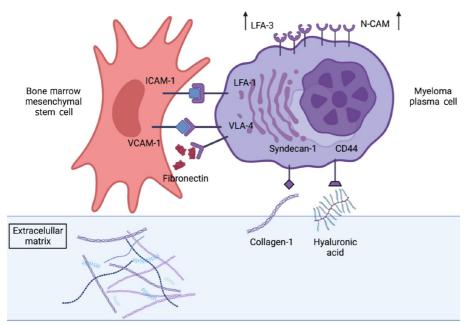
CAMs play a central role in cell communication and the maintenance of tissue homeostasis[89]. There are different superfamilies or groups of CAMs with different specificities and distributions. These families would include the Immunoglobulin superfamily CAMs (IgCAMs), integrins, cadherins and one superfamily of proteins that contain a C-type lectin-like domain (C-type lectin domain proteins or CTLDs)[89]. Following other criteria, CAMs can be classified into calcium-independent or calciumdependent molecules [90], meaning that these molecules would need Ca²⁺ ions binding to different domains of the protein in order to rigidify their extracellular domains and enable interaction[91]. Integrins and IgCAMs belong to the calcium independent group whereas CLTDs and selectins belong to



Table 2 Summary of evidence about the relevance of different cargo molecules in the extracellular vesicles of multiple myeloma cells and bone marrow mesenchymal stem cells s to the progression of multiple myeloma

	Function	Ref.
MM-EVs cargo		
lncRNA RUNX2-AS1	Form a RNA duplex with RUNX2 premRNA, reducing the osteogenic activity in MSCs	[76]
miR-146a	Increase the secretion of several cytokines in BM-MSCs that favor MM cell viability and migration and induce CAF transformation	[78]
DKK-1	Lower expression of OSX, COL1A1 and ALP in osteoblast precusor cell line (MC3t3-E1)	[72]
MSC-EVs cargo		
mir-15a	Promote MM cell proliferation and dissemination to other niches	[70]
miR-483-5p	Induce MM cell proliferation and reduced apoptosis	[79]
miR-155	Reduce MM cell apoptosis and augment expression of stemness maintenance and drug resistance markers	[81]

EVs: Extracellular vesicles; MM: Multiple myeloma; BM-MSC: Bone marrow mesenchymal stem cells; CAF: Cancer associated fibroblast; DKK-1: Dickkopf-1; OSX: Osterix; COL1A1: Collagen 1A1; ALP: Alkaline phosphatase.



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Figure 2 Schematic representation of the main cell adhesion molecules in multiple myeloma cells and bone marrow mesenchymal stem cells. The main interactions between cell adhesion molecules (CAMs) of these two types of cells as well as the interactions of these CAMs with proteins of the extracellular matrix are displayed (Created with Biorender.com). ICAM-1: Intercellular adhesion molecule 1; VCAM-1: Vascular cell adhesion molecule-1; LFA-1: Leukocyte function-associated antigen 1; N-CAM: Neural cell adhesion molecule; VLA: Very late antigen. Created with BioRender.com.

> calcium dependent group[92]. Cell adhesion molecules bind to different ligands. Cadherins, selectins and IgCAMs are associated with the cell-to-cell contact, while integrins are involved in the attachment of MM cells to the ECM[93]. All these molecules are integral in modeling cellular mechanisms such as growth, contact inhibition and apoptosis. In fact, changes in cell adhesion, involving these molecules, can be the defining event in a wide range of diseases, including cancer development[94], as lower intercellular adhesiveness allows malignant cells to scape from their site, thus, destroying the architecture of the original tissue, commonly the first step leading to cancer [94].

> As well as the already described interactions through paracrine secretion of different cytokines and EVs, MM cells also interact with BM-MSCs by direct cell-to-cell contact. In fact, recent studies revealed that many of the changes undergone by BM-MSCs supporting the progression of MM, are acquirable by physical contact with MM cells[95]. In MM, this cell-to-cell interactions are not restricted to MM and BM-MSCs since MM cells also interact with other cells of the BM microenvironment such as osteoclasts and osteoblasts, endothelial cells, and lymphocytes. All these interactions are regulated by CAMs. It is

known that these contacts are key to protect MM cells against chemotherapy, helping them to accumulate inside the BM[83], to adhere to endothelium, and to spread into the bloodstream[84], although the detailed mechanisms involved in those processes have not been completely elucidated[85].

The most relevant role of CAMs in MM pathophysiology is related to the homing of malignant PCs to the BM. To complete the process of homing, mediated by CXCL12, MM cells need to adhere to either ECM proteins or BM-MSCs. This is mediated by CAMs such as very late antigen (VLA)4, VLA5, CD44, leukocyte function-associated antigen 1 (LFA-1), intercellular adhesion molecule 1 (ICAM-1), MPC-1 and syndecan (Figure 2).

One way of ensuring adhesion of MM to the ECM is the binding of its integrin VLA-4 to fibronectin, a common component of the ECM. VLA-4, which is in fact a heterodimer of two integrins CD49d(a4) and CD29(b1), also mediates the interaction of MM cells with BM-MSCs, through the vascular cell adhesion molecule-1 (VCAM-1), located at the BM-MSCs[96]. This interaction activates the secretion of MIP-1 α and MIP-1 β in MM cells, leading to an increase of osteoclastogenic activity [97]. Moreover, the direct contact of these two types of cells through VLA-4 also induces the production of DKK-1 by MM cells, which inhibits osteoblastic differentiation of BM-MSCs. Thus, these two actions, promotion of osteoclastogenesis and inhibition of osteogenesis, would have a detrimental effect on bone structure, contributing to the typical osteolytic lesions in MM. In addition, BM-MSCs unable to undergo osteoblastic differentiation would produce higher levels of IL-6, a cytokine that would stimulate the proliferation of DKK-1secreting MM cells[25]. Moreover, it has been observed that VLA-4-fibronectin binding is an essential step that supports the IL-6-mediated induction of PCs in normal BM, since antibodies against VLA-4 were found to inhibit the secretion of IL-6 in co-cultures of MM cells and BM-MSCs cells[96,98,99].

The interaction between MM cells and BM-MSCs is also mediated by ICAM-1 (CD54) and LFA-1 (CD11a/CD18) expressed in BM-MSCs and MM respectively. The glycoprotein ICAM-1 is the main ligand for b2 integrins and its expression is induced in response to an inflammatory microenvironment [100], such as the one resulting in the BM following the colonization by MM cells. The ICAM-1/LFA-1 interaction seems to have a key role in the progression of MM since the blocking of LFA-1 through the use of monoclonal antibodies, inhibits the production of IL-6 by BM-MSCs. Thus, this interaction is focus of various studies aimed to the development of treatments for MM[101].

Syndecan (CD138) is the principal transmembrane proteoglycan expressed in the surface of MM cells and has in fact been used as a marker for the detection of this pathology. Syndecan has multiple functions in MM. This molecule mediates de adhesion of MM cells to the collagen in the ECM through direct interaction with collagen molecules but can also mediate myeloma cell-cell adhesion[102]. Syndecan-1 also plays a broad role in cells signaling since heparan sulfate chains on syndecan-1 can bind to and sequester growth factors and cytokines, regulating their availability to cells. Also, a recent study has shown that syndecan contributes to the survival of mature MM cells by enhancing IL-6 signaling[103]. Finally, the binding of syndecan to VEGF and other angiogenic factors, has been shown to promote angiogenesis in MM[104].

Finally, CD44, a transmembrane glycoprotein, interacts mainly although not exclusively with hyaluronic acid in the ECM[105]. CD44 signaling has been shown to activate various signaling pathways in different types of cancer including PI3k/AKT, MAPK/ERK and NF-kB[106], which, as we have seen, promote MM cell survival.

Although normal PC and MM cells express basically the same set of CAMS, some of these molecules were found to be more significantly overexpressed in MM cells when compared to healthy patients. In this group we can include, leukocyte adhesion molecule LFA-3 (CD58)[107] and neural cell adhesion molecule (CD56)[108]. MM cells can also express the lymphocyte function-associated antigen LFA-1 $(CD11\alpha/CD18)$ which was associated with tumor growth and homotypic tumor cell adhesion or aggregation^[109]. It is also worth mentioning that some homing molecules could not be detected on MPCs: Selectin molecule L-selectin and collagen receptor VLA-2[89]. Although this study provides relevant information, for this information to be biologically relevant, ligands of these receptors had to be available within the tumor environment.

Overall, given the importance of some of these CAMs in the process of MM cells homing, these molecules could be important targets for designing antitumoral treatments. Several approaches have been explored, including antibodies specifically targeting these molecules on the cell surface, as well as small molecule inhibitors that interfere with the binding of the CAMs to their ligands. Moreover, receptor-blocking antibodies against most of these CAMs (VLA-4, CD56, MPC-1, CD21) were found to partially block MM cells adhesion to the BM stroma. This partial effect could be attributed to an additional adhesion mechanism yet to be discovered[110,111].

In MM a specific type of drug resistance seems to be mediated by CAMs, the so called, CAMs mediated drug resistance[112]. CAMs can activate intracellular pathways that promote cell survival, promote cancer cell adhesion to the ECM and regulate the expression of drug transporters that could pump chemotherapy drug out of cancer cells and reduce their efficacy. It is also important to highlight that MM spreading in the last stages of the diseases also involve important changes in cell adhesion. MM can abandon de BM microenvironment and become stroma independent because of different processes involving changes in the expression levels of CAM and certain cytokines. Once this happens, cells can be found to spread extramedullary at different sites such as lungs, liver, or pleural fluid [113, 114].



Role of tunneling nanotubes in MM/BM-MSCs communication

As we have seen, communication between MM and BM-MSCs cells can take place through mechanisms that can be classified as contact-dependent and/or contact-independent mechanisms[115]. While the previous section has been dedicated to direct communication mediated by cell adhesion molecules, in this last section we will briefly discuss transport via tunneling nanotubes (TNTs), another form of contact-dependent interaction.

TNTs are transient intercellular structures formed by the polymerization of F-actin which provide an important and general mechanism of cell-to-cell communication[116,117] and constitute a reliable infrastructure for vesicle and protein trafficking[118]. Numerous examples of communication between MSCs and malignant hematological cells such as B cells, MM and chronic lymphocytic leukemia, are already known, as well as the effects of this communication, such as increased drug resistance. This has already been demonstrated in acute myeloid leukemia (AML), B-cell precursor acute lymphocytic leukemia (ALL) or CML[119,120]. Therefore, TNTs are considered one of the key pharmacological targets in current research.

The role of TNTs is to deliver autophagosomes, mitochondria and other lipophiles to MSCs. This induces the secretion of specific cytokines, including interferon-γ-IP-10, CXCL10, IL-8, MCP-1 and CCL2 and other growth factors which, in turn, induce tumor cell survival, enhanced growth and even drug resistance[121]. This has been checked in AML, where increased survival of cells against chemotherapy treatments is observed by means of mitochondrial transfer from MSCs routed by TNT. In this case, the mitochondrial transfer translates into an increase of up to 14% in mitochondrial mass in co-cultures of tumor cells with MSCs and a 1.5-fold increase in mitochondrial adenosine triphosphate production (ATP), making them less prone to mitochondrial depolarization and thus resulting in increased survival against chemotherapy treatments[122].

Numerous lines of treatment are currently under development for various hematological diseases that reduce the formation of TNT by blocking actin polymerization. This inhibits the cellular communication that promotes disease progression. Those treatments include cytochalasin D, cytarabine, latrunculin A and B, daunorubicin, everolimus, metformin, nocodazole CK-666, ML-141 or 6-thio-GTP [123]. In addition, vinca alkaloids or taxanes are also being targeted because of their role in the polymerization of microtubules[124].

Although the fate of mitochondria transferred into tumor cells remains unclear, there is evidence indicating that MSCs play a key role in the progression of AML, ALL, MM and mitochondrial transfer chemoresistance. It is well known that the initiation of cancer requires metabolic adjustments, since rapid proliferation cancer cells have high metabolic requirements. This mitochondrial and/or mitochondrial DNA transfer to cancer cells increases mitochondrial content and enhances the mitochondrial process of oxidative phosphorylation (Oxphos), which generates a larger quantity of ATP than glycolysis, thus, promoting cell proliferation and invasion[125]. Therefore, targeting mitochondrial respiration and Oxphos is also a treatment option, FOXM1 is known to regulate myeloma cell metabolism by increasing glycolysis and Oxphos. NB73 is a FOXM1 inhibitor that promotes FOXM1 degradation and thus growth of MM cells, making it a potential drug targeting Oxphos[126].

Studies to date have elucidated that mitochondrial transfer dynamically induced resistance occurs between MM cells and other cells in the BM microenvironment via TNT, providing a starting point for the development of new targeted therapies [127]. An example of this line of treatment for MM is the use of anti-CD38 monoclonal antibodies [128]. This antibodies have different mechanisms of action, including cell apoptosis[129]. Moreover, their administration in mice has shown inhibition of mitochondrial transfer, a reduction in tumor volume and, in general, increased survival[1]. However, it should be noted that, although patients who have received this treatment show increased survival, it has been observed that resistance to these treatments can be acquired in the long term.

CONCLUSION

Conditions at the BM microenvironment are essential for the establishment and progression of MM. The complex BM microenvironment encompasses hematopoietic cells, immune cells, and cells involved in bone homeostasis such as osteoclasts, osteoblasts and BM-MSCs. Thus, it is understandable that the disruption of microenvironment homeostasis by MM cells results in angiogenesis, osteolysis, immune suppression and anemia[69].

As key regulators of this microenvironment, BM-MSCs play an important role in the progression of the disease. The crosstalk between MM cells and BM-MSCs takes place at different levels, through soluble cytokines, EVs, and direct cell-to-cell contact.

The interaction between these two cell types can have both positive and negative effects on the proliferation and survival of MM cells. The communication between MM cells and BM-MSCs can promote tumor growth. The survival and proliferation of MM cells once they reach the BM is associated with immune suppression, eliminating the possibility of an effective antitumor response. Although it is the interaction between all cells in the BM what produces this immunosuppressive microenvironment, BM-MSCs have a relevant role in the construction of this particular microenvironment due not only to their



important paracrine activity, but also to their ability to establish direct communication with other cells in that microenvironment. All these direct or indirect interactions activate a pleiotropic proliferative and antiapoptotic cascades favoring disease progression.

On the other hand, the communication between MM cells and BM-MSCs can also have a negative impact on cancer cell growth and survival. BM-MSCs can secrete factors that inhibit the growth and survival of MM cells.

Currently, therapeutic advances in the treatment of this disease are based on targeted therapies using monoclonal antibodies or CAR-T. These treatments have improved patient prognosis, although longterm resistance is still observed, and further research is needed into the specific mechanisms by which cells acquire this resistance. In the quest for new effective treatments for MM, the importance of communication between MM cells and BM-MSCs cannot be overstated. Understanding the molecular mechanisms involved in this two-way communication can provide valuable insights into MM pathogenesis and help identify key targets involved in the survival and proliferation of MM cells in the BM microenvironment and thus, opening new opportunities for the design of targeted therapies to avoid disease progression.

FOOTNOTES

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REFERENCES

- Siegel RL, Miller KD, Jemal A. Cancer statistics, 2019. CA Cancer J Clin 2019; 69: 7-34 [PMID: 30620402 DOI: 10.3322/caac.21551]
- Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, Bray F. Global Cancer Statistics 2020: 2 GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. CA Cancer J Clin 2021; 71: 209-249 [PMID: 33538338 DOI: 10.3322/caac.21660]
- Padala SA, Barsouk A, Rawla P, Vakiti A, Kolhe R, Kota V, Ajebo GH. Epidemiology, Staging, and Management of 3 Multiple Myeloma. Med Sci (Basel) 2021; 9 [PMID: 33498356 DOI: 10.3390/medsci9010003]
- Abramson HN. Recent Advances in the Applications of Small Molecules in the Treatment of Multiple Myeloma. Int J 4 Mol Sci 2023; 24 [PMID: 36768967 DOI: 10.3390/ijms24032645]
- Yamamoto C, Minakata D, Koyama S, Sekiguchi K, Fukui Y, Murahashi R, Nakashima H, Matsuoka S, Ikeda T, 5 Kawaguchi SI, Toda Y, Ito S, Nagayama T, Umino K, Nakano H, Morita K, Yamasaki R, Ashizawa M, Ueda M, Hatano K, Sato K, Ohmine K, Fujiwara SI, Kanda Y. Daratumumab in first-line therapy is cost-effective in transplant-eligible patients with newly diagnosed myeloma. Blood 2022; 140: 594-607 [PMID: 35580269 DOI: 10.1182/blood.2021015220]



- Kumar SK, Rajkumar V, Kyle RA, van Duin M, Sonneveld P, Mateos MV, Gay F, Anderson KC. Multiple myeloma. Nat 6 Rev Dis Primers 2017; 3: 17046 [PMID: 28726797 DOI: 10.1038/nrdp.2017.46]
- Kyle RA. Multiple myeloma: review of 869 cases. Mayo Clin Proc 1975; 50: 29-40 [PMID: 1110582]
- Bataille R, Chappard D, Marcelli C, Dessauw P, Baldet P, Sany J, Alexandre C. Recruitment of new osteoblasts and 8 osteoclasts is the earliest critical event in the pathogenesis of human multiple myeloma. J Clin Invest 1991; 88: 62-66 [PMID: 2056131 DOI: 10.1172/JCI115305]
- Mundy GR, Raisz LG, Cooper RA, Schechter GP, Salmon SE. Evidence for the secretion of an osteoclast stimulating factor in myeloma. N Engl J Med 1974; 291: 1041-1046 [PMID: 4413338 DOI: 10.1056/NEJM197411142912001]
- Paget S. The distribution of secondary growths in cancer of the breast. 1889. Cancer Metastasis Rev 1989; 8: 98-101 10 [PMID: 2673568]
- Lomas OC, Tahri S, Ghobrial IM. The microenvironment in myeloma. Curr Opin Oncol 2020; 32: 170-175 [PMID: 11 31895122 DOI: 10.1097/CCO.000000000000615]
- Moschetta M, Kawano Y, Sacco A, Belotti A, Ribolla R, Chiarini M, Giustini V, Bertoli D, Sottini A, Valotti M, Ghidini 12 C, Serana F, Malagola M, Imberti L, Russo D, Montanelli A, Rossi G, Reagan MR, Maiso P, Paiva B, Ghobrial IM, Roccaro AM. Bone Marrow Stroma and Vascular Contributions to Myeloma Bone Homing. Curr Osteoporos Rep 2017; 15: 499-506 [PMID: 28889371 DOI: 10.1007/s11914-017-0399-3]
- 13 Hoang PH, Cornish AJ, Dobbins SE, Kaiser M, Houlston RS. Mutational processes contributing to the development of multiple myeloma. Blood Cancer J 2019; 9: 60 [PMID: 31387987 DOI: 10.1038/s41408-019-0221-9]
- Visram A, Dasari S, Anderson E, Kumar S, Kourelis TV. Relapsed multiple myeloma demonstrates distinct patterns of 14 immune microenvironment and malignant cell-mediated immunosuppression. Blood Cancer J 2021; 11: 45 [PMID: 33649314 DOI: 10.1038/s41408-021-00440-4]
- García-Ortiz A, Rodríguez-García Y, Encinas J, Maroto-Martín E, Castellano E, Teixidó J, Martínez-López J. The Role 15 of Tumor Microenvironment in Multiple Myeloma Development and Progression. Cancers (Basel) 2021; 13 [PMID: 33435306 DOI: 10.3390/cancers13020217]
- Chu VT, Berek C. The establishment of the plasma cell survival niche in the bone marrow. Immunol Rev 2013; 251: 177-16 188 [PMID: 23278749 DOI: 10.1111/imr.12011]
- Lindquist RL, Niesner RA, Hauser AE. In the Right Place, at the Right Time: Spatiotemporal Conditions Determining 17 Plasma Cell Survival and Function. Front Immunol 2019; 10: 788 [PMID: 31068930 DOI: 10.3389/fimmu.2019.00788]
- Tsukasaki M, Takayanagi H. Osteoimmunology: evolving concepts in bone-immune interactions in health and disease. 18 Nat Rev Immunol 2019; 19: 626-642 [PMID: 31186549 DOI: 10.1038/s41577-019-0178-8]
- De Veirman K, Wang J, Xu S, Leleu X, Himpe E, Maes K, De Bruyne E, Van Valckenborgh E, Vanderkerken K, Menu 19 E, Van Riet I. Induction of miR-146a by multiple myeloma cells in mesenchymal stromal cells stimulates their pro-tumoral activity. Cancer Lett 2016; 377: 17-24 [PMID: 27102001 DOI: 10.1016/j.canlet.2016.04.024]
- 20 Fu J, Li S, Feng R, Ma H, Sabeh F, Roodman GD, Wang J, Robinson S, Guo XE, Lund T, Normolle D, Mapara MY, Weiss SJ, Lentzsch S. Multiple myeloma-derived MMP-13 mediates osteoclast fusogenesis and osteolytic disease. J Clin Invest 2016; 126: 1759-1772 [PMID: 27043283 DOI: 10.1172/JCI80276]
- Colombo M, Thümmler K, Mirandola L, Garavelli S, Todoerti K, Apicella L, Lazzari E, Lancellotti M, Platonova N, 21 Akbar M, Chiriva-Internati M, Soutar R, Neri A, Goodyear CS, Chiaramonte R. Notch signaling drives multiple myeloma induced osteoclastogenesis. Oncotarget 2014; 5: 10393-10406 [PMID: 25257302 DOI: 10.18632/oncotarget.2084]
- Roodman GD. Osteoblast function in myeloma. Bone 2011; 48: 135-140 [PMID: 20601285 DOI: 22 10.1016/j.bone.2010.06.016]
- 23 Yaccoby S. Osteoblastogenesis and tumor growth in myeloma. Leuk Lymphoma 2010; 51: 213-220 [PMID: 20038269 DOI: 10.3109/10428190903503438]
- Reagan MR, Liaw L, Rosen CJ, Ghobrial IM. Dynamic interplay between bone and multiple myeloma: emerging roles of 24 the osteoblast. Bone 2015; 75: 161-169 [PMID: 25725265 DOI: 10.1016/j.bone.2015.02.021]
- Gunn WG, Conley A, Deininger L, Olson SD, Prockop DJ, Gregory CA. A crosstalk between myeloma cells and marrow 25 stromal cells stimulates production of DKK1 and interleukin-6: a potential role in the development of lytic bone disease and tumor progression in multiple myeloma. Stem Cells 2006; 24: 986-991 [PMID: 16293576 DOI: 10.1634/stemcells.2005-0220
- Berenstein R, Blau O, Nogai A, Waechter M, Slonova E, Schmidt-Hieber M, Kunitz A, Pezzutto A, Doerken B, Blau IW. 26 Multiple myeloma cells alter the senescence phenotype of bone marrow mesenchymal stromal cells under participation of the DLK1-DIO3 genomic region. BMC Cancer 2015; 15: 68 [PMID: 25886144 DOI: 10.1186/s12885-015-1078-3]
- Berenstein R, Nogai A, Waechter M, Blau O, Kuehnel A, Schmidt-Hieber M, Kunitz A, Pezzutto A, Dörken B, Blau IW. 27 Multiple myeloma cells modify VEGF/IL-6 levels and osteogenic potential of bone marrow stromal cells via Notch/miR-223. Mol Carcinog 2016; 55: 1927-1939 [PMID: 27023728 DOI: 10.1002/mc.22440]
- Terpos E, Ntanasis-Stathopoulos I, Gavriatopoulou M, Dimopoulos MA. Pathogenesis of bone disease in multiple 28 myeloma: from bench to bedside. Blood Cancer J 2018; 8: 7 [PMID: 29330358 DOI: 10.1038/s41408-017-0037-4]
- 29 Dewald JH, Colomb F, Bobowski-Gerard M, Groux-Degroote S, Delannoy P. Role of Cytokine-Induced Glycosylation Changes in Regulating Cell Interactions and Cell Signaling in Inflammatory Diseases and Cancer. Cells 2016; 5 [PMID: 27916834 DOI: 10.3390/cells5040043]
- Kyle RA, Rajkumar SV. Multiple myeloma. N Engl J Med 2004; 351: 1860-1873 [PMID: 15509819 DOI: 30 10.1056/NEJMra041875
- González-González A, García-Sánchez D, Dotta M, Rodríguez-Rey JC, Pérez-Campo FM. Mesenchymal stem cells 31 secretome: The cornerstone of cell-free regenerative medicine. World J Stem Cells 2020; 12: 1529-1552 [PMID: 33505599 DOI: 10.4252/wjsc.v12.i12.1529]
- Atsuta I, Liu S, Miura Y, Akiyama K, Chen C, An Y, Shi S, Chen FM. Mesenchymal stem cells inhibit multiple myeloma 32 cells via the Fas/Fas ligand pathway. Stem Cell Res Ther 2013; 4: 111 [PMID: 24025590 DOI: 10.1186/scrt322]
- Kumar S, Witzig TE, Timm M, Haug J, Wellik L, Kimlinger TK, Greipp PR, Rajkumar SV. Bone marrow angiogenic 33



ability and expression of angiogenic cytokines in myeloma: evidence favoring loss of marrow angiogenesis inhibitory activity with disease progression. Blood 2004; 104: 1159-1165 [PMID: 15130943 DOI: 10.1182/blood-2003-11-3811]

- Gupta D, Treon SP, Shima Y, Hideshima T, Podar K, Tai YT, Lin B, Lentzsch S, Davies FE, Chauhan D, Schlossman 34 RL, Richardson P, Ralph P, Wu L, Payvandi F, Muller G, Stirling DI, Anderson KC. Adherence of multiple myeloma cells to bone marrow stromal cells upregulates vascular endothelial growth factor secretion: therapeutic applications. Leukemia 2001; 15: 1950-1961 [PMID: 11753617 DOI: 10.1038/sj.leu.2402295]
- Xiong Y, Donovan KA, Kline MP, Gornet MK, Moon-Tasson LL, Lacy MQ, Dispenzieri A, Gertz MA, Greipp PR, Lust 35 JA. Identification of two groups of smoldering multiple myeloma patients who are either high or low producers of interleukin-1. J Interferon Cytokine Res 2006; 26: 83-95 [PMID: 16487028 DOI: 10.1089/jir.2006.26.83]
- Harmer D, Falank C, Reagan MR. Interleukin-6 Interweaves the Bone Marrow Microenvironment, Bone Loss, and 36 Multiple Myeloma. Front Endocrinol (Lausanne) 2018; 9: 788 [PMID: 30671025 DOI: 10.3389/fendo.2018.00788]
- Teicher BA, Fricker SP. CXCL12 (SDF-1)/CXCR4 pathway in cancer. Clin Cancer Res 2010; 16: 2927-2931 [PMID: 37 20484021 DOI: 10.1158/1078-0432.CCR-09-2329]
- 38 Bouyssou JM, Ghobrial IM, Roccaro AM. Targeting SDF-1 in multiple myeloma tumor microenvironment. Cancer Lett 2016; 380: 315-318 [PMID: 26655999 DOI: 10.1016/j.canlet.2015.11.028]
- Chatterjee S, Behnam Azad B, Nimmagadda S. The intricate role of CXCR4 in cancer. Adv Cancer Res 2014; 124: 31-82 39 [PMID: 25287686 DOI: 10.1016/B978-0-12-411638-2.00002-1]
- De Clercq E. Potential clinical applications of the CXCR4 antagonist bicyclam AMD3100. Mini Rev Med Chem 2005; 5: 40 805-824 [PMID: 16178723 DOI: 10.2174/1389557054867075]
- 41 Alsayed Y, Ngo H, Runnels J, Leleu X, Singha UK, Pitsillides CM, Spencer JA, Kimlinger T, Ghobrial JM, Jia X, Lu G, Timm M, Kumar A, Côté D, Veilleux I, Hedin KE, Roodman GD, Witzig TE, Kung AL, Hideshima T, Anderson KC, Lin CP, Ghobrial IM. Mechanisms of regulation of CXCR4/SDF-1 (CXCL12)-dependent migration and homing in multiple myeloma. Blood 2007; 109: 2708-2717 [PMID: 17119115 DOI: 10.1182/blood-2006-07-035857]
- 42 Hideshima T, Anderson KC. Signaling Pathway Mediating Myeloma Cell Growth and Survival. Cancers (Basel) 2021; 13 [PMID: 33435632 DOI: 10.3390/cancers13020216]
- 43 Hideshima T, Mitsiades C, Tonon G, Richardson PG, Anderson KC. Understanding multiple myeloma pathogenesis in the bone marrow to identify new therapeutic targets. Nat Rev Cancer 2007; 7: 585-598 [PMID: 17646864 DOI: 10.1038/nrc2189]
- Musolino C, Allegra A, Innao V, Allegra AG, Pioggia G, Gangemi S. Inflammatory and Anti-Inflammatory Equilibrium, 44 Proliferative and Antiproliferative Balance: The Role of Cytokines in Multiple Myeloma. Mediators Inflamm 2017; 2017: 1852517 [PMID: 29089667 DOI: 10.1155/2017/1852517]
- Lust JA, Lacy MQ, Zeldenrust SR, Witzig TE, Moon-Tasson LL, Dinarello CA, Donovan KA. Reduction in C-reactive 45 protein indicates successful targeting of the IL-1/IL-6 axis resulting in improved survival in early stage multiple myeloma. Am J Hematol 2016; 91: 571-574 [PMID: 26945843 DOI: 10.1002/ajh.24352]
- Chong PSY, Chng WJ, de Mel S. STAT3: A Promising Therapeutic Target in Multiple Myeloma. Cancers (Basel) 2019; 46 11 [PMID: 31130718 DOI: 10.3390/cancers11050731]
- Hashizume M, Hayakawa N, Mihara M. IL-6 trans-signalling directly induces RANKL on fibroblast-like synovial cells 47 and is involved in RANKL induction by TNF-alpha and IL-17. Rheumatology (Oxford) 2008; 47: 1635-1640 [PMID: 18786965 DOI: 10.1093/rheumatology/ken363]
- Roy P, Sarkar UA, Basak S. The NF-KB Activating Pathways in Multiple Myeloma. Biomedicines 2018; 6 [PMID: 48 29772694 DOI: 10.3390/biomedicines6020059]
- Mitsiades CS, Mitsiades N, Poulaki V, Schlossman R, Akiyama M, Chauhan D, Hideshima T, Treon SP, Munshi NC, 49 Richardson PG, Anderson KC. Activation of NF-kappaB and upregulation of intracellular anti-apoptotic proteins via the IGF-1/Akt signaling in human multiple myeloma cells: therapeutic implications. Oncogene 2002; 21: 5673-5683 [PMID: 12173037 DOI: 10.1038/sj.onc.1205664]
- Hengeveld PJ, Kersten MJ. B-cell activating factor in the pathophysiology of multiple myeloma: a target for therapy? 50 Blood Cancer J 2015; 5: e282 [PMID: 25723853 DOI: 10.1038/bcj.2015.3]
- 51 Chilov D, Kukk E, Taira S, Jeltsch M, Kaukonen J, Palotie A, Joukov V, Alitalo K. Genomic organization of human and mouse genes for vascular endothelial growth factor C. J Biol Chem 1997; 272: 25176-25183 [PMID: 9312130 DOI: 10.1074/jbc.272.40.25176]
- Kumar S, Witzig TE, Timm M, Haug J, Wellik L, Fonseca R, Greipp PR, Rajkumar SV. Expression of VEGF and its 52 receptors by myeloma cells. Leukemia 2003; 17: 2025-2031 [PMID: 14513053 DOI: 10.1038/sj.leu.2403084]
- 53 John L, Krauth MT, Podar K, Raab MS. Pathway-Directed Therapy in Multiple Myeloma. Cancers (Basel) 2021; 13 [PMID: 33916289 DOI: 10.3390/cancers13071668]
- 54 McCubrey JA, Steelman LS, Chappell WH, Abrams SL, Wong EW, Chang F, Lehmann B, Terrian DM, Milella M, Tafuri A, Stivala F, Libra M, Basecke J, Evangelisti C, Martelli AM, Franklin RA. Roles of the Raf/MEK/ERK pathway in cell growth, malignant transformation and drug resistance. Biochim Biophys Acta 2007; 1773: 1263-1284 [PMID: 17126425 DOI: 10.1016/j.bbamcr.2006.10.001]
- Ramakrishnan V, Kumar S. PI3K/AKT/mTOR pathway in multiple myeloma: from basic biology to clinical promise. 55 Leuk Lymphoma 2018; 59: 2524-2534 [PMID: 29322846 DOI: 10.1080/10428194.2017.1421760]
- Rascio F, Spadaccino F, Rocchetti MT, Castellano G, Stallone G, Netti GS, Ranieri E. The Pathogenic Role of PI3K/AKT 56 Pathway in Cancer Onset and Drug Resistance: An Updated Review. Cancers (Basel) 2021; 13 [PMID: 34439105 DOI: 10.3390/cancers13163949]
- Ria R, Vacca A, Russo F, Cirulli T, Massaia M, Tosi P, Cavo M, Guidolin D, Ribatti D, Dammacco F. A VEGF-57 dependent autocrine loop mediates proliferation and capillarogenesis in bone marrow endothelial cells of patients with multiple myeloma. Thromb Haemost 2004; 92: 1438-1445 [PMID: 15583754 DOI: 10.1160/TH04-06-0334]
- Ribatti D, Vacca A. New Insights in Anti-Angiogenesis in Multiple Myeloma. Int J Mol Sci 2018; 19 [PMID: 30002349 DOI: 10.3390/ijms19072031]



- Colla S, Morandi F, Lazzaretti M, Polistena P, Svaldi M, Coser P, Bonomini S, Hojden M, Martella E, Chisesi T, Rizzoli 59 V, Giuliani N. Do human myeloma cells directly produce basic FGF? Blood 2003; 102: 3071-2; author reply 3072 [PMID: 14527891 DOI: 10.1182/blood-2003-06-1883]
- 60 Rundhaug JE. Matrix metalloproteinases and angiogenesis. J Cell Mol Med 2005; 9: 267-285 [PMID: 15963249 DOI: 10.1111/j.1582-4934.2005.tb00355.x]
- Terpos E, Anargyrou K, Katodritou E, Kastritis E, Papatheodorou A, Christoulas D, Pouli A, Michalis E, Delimpasi S, 61 Gkotzamanidou M, Nikitas N, Koumoustiotis V, Margaritis D, Tsionos K, Stefanoudaki E, Meletis J, Zervas K, Dimopoulos MA; Greek Myeloma Study Group, Greece. Circulating angiopoietin-1 to angiopoietin-2 ratio is an independent prognostic factor for survival in newly diagnosed patients with multiple myeloma who received therapy with novel antimyeloma agents. Int J Cancer 2012; 130: 735-742 [PMID: 21484787 DOI: 10.1002/ijc.26062]
- 62 Bergers G, Benjamin LE. Tumorigenesis and the angiogenic switch. Nat Rev Cancer 2003; 3: 401-410 [PMID: 12778130 DOI: 10.1038/nrc1093]
- Abe M, Hiura K, Ozaki S, Kido S, Matsumoto T. Vicious cycle between myeloma cell binding to bone marrow stromal 63 cells via VLA-4-VCAM-1 adhesion and macrophage inflammatory protein-1alpha and MIP-1beta production. J Bone *Miner Metab* 2009; **27**: 16-23 [PMID: 19057841 DOI: 10.1007/s00774-008-0012-z]
- Oyajobi BO, Franchin G, Williams PJ, Pulkrabek D, Gupta A, Munoz S, Grubbs B, Zhao M, Chen D, Sherry B, Mundy 64 GR. Dual effects of macrophage inflammatory protein-lalpha on osteolysis and tumor burden in the murine 5TGM1 model of myeloma bone disease. Blood 2003; 102: 311-319 [PMID: 12649140 DOI: 10.1182/blood-2002-12-3905]
- Park JH, Lee NK, Lee SY. Current Understanding of RANK Signaling in Osteoclast Differentiation and Maturation. Mol 65 Cells 2017; 40: 706-713 [PMID: 29047262 DOI: 10.14348/molcells.2017.0225]
- 66 Colla S, Zhan F, Xiong W, Wu X, Xu H, Stephens O, Yaccoby S, Epstein J, Barlogie B, Shaughnessy JD Jr. The oxidative stress response regulates DKK1 expression through the JNK signaling cascade in multiple myeloma plasma cells. Blood 2007; 109: 4470-4477 [PMID: 17255354 DOI: 10.1182/blood-2006-11-056747]
- Dun X, Jiang H, Zou J, Shi J, Zhou L, Zhu R, Hou J. Differential expression of DKK-1 binding receptors on stromal cells and myeloma cells results in their distinct response to secreted DKK-1 in myeloma. Mol Cancer 2010; 9: 247 [PMID: 20846389 DOI: 10.1186/1476-4598-9-247]
- Houschyar KS, Tapking C, Borrelli MR, Popp D, Duscher D, Maan ZN, Chelliah MP, Li J, Harati K, Wallner C, Rein S, 68 Pförringer D, Reumuth G, Grieb G, Mouraret S, Dadras M, Wagner JM, Cha JY, Siemers F, Lehnhardt M, Behr B. Wnt Pathway in Bone Repair and Regeneration - What Do We Know So Far. Front Cell Dev Biol 2018; 6: 170 [PMID: 30666305 DOI: 10.3389/fcell.2018.00170]
- Wang J, Faict S, Maes K, De Bruyne E, Van Valckenborgh E, Schots R, Vanderkerken K, Menu E. Extracellular vesicle 69 cross-talk in the bone marrow microenvironment: implications in multiple myeloma. Oncotarget 2016; 7: 38927-38945 [PMID: 26950273 DOI: 10.18632/oncotarget.7792]
- Roccaro AM, Sacco A, Maiso P, Azab AK, Tai YT, Reagan M, Azab F, Flores LM, Campigotto F, Weller E, Anderson 70 KC, Scadden DT, Ghobrial IM. BM mesenchymal stromal cell-derived exosomes facilitate multiple myeloma progression. J Clin Invest 2013; 123: 1542-1555 [PMID: 23454749 DOI: 10.1172/JCI66517]
- 71 Moloudizargari M, Abdollahi M, Asghari MH, Zimta AA, Neagoe IB, Nabavi SM. The emerging role of exosomes in multiple myeloma. Blood Rev 2019; 38: 100595 [PMID: 31445775 DOI: 10.1016/j.blre.2019.100595]
- Faict S, Muller J, De Veirman K, De Bruyne E, Maes K, Vrancken L, Heusschen R, De Raeve H, Schots R, Vanderkerken 72 K, Caers J, Menu E. Exosomes play a role in multiple myeloma bone disease and tumor development by targeting osteoclasts and osteoblasts. *Blood Cancer J* 2018; 8: 105 [PMID: 30409995 DOI: 10.1038/s41408-018-0139-7]
- Zhang L, Lei Q, Wang H, Xu C, Liu T, Kong F, Yang C, Yan G, Sun L, Zhao A, Chen W, Hu Y, Xie H, Cao Y, Fu F, 73 Yuan G, Chen Z, Guo AY, Li Q. Tumor-derived extracellular vesicles inhibit osteogenesis and exacerbate myeloma bone disease. Theranostics 2019; 9: 196-209 [PMID: 30662562 DOI: 10.7150/thno.27550]
- 74 Raimondo S, Urzì O, Conigliaro A, Bosco GL, Parisi S, Carlisi M, Siragusa S, Raimondi L, Luca A, Giavaresi G, Alessandro R. Extracellular Vesicle microRNAs Contribute to the Osteogenic Inhibition of Mesenchymal Stem Cells in Multiple Myeloma. Cancers (Basel) 2020; 12 [PMID: 32075123 DOI: 10.3390/cancers12020449]
- Wu Y, Zhang Z, Wu J, Hou J, Ding G. The Exosomes Containing LINC00461 Originated from Multiple Myeloma Inhibit 75 the Osteoblast Differentiation of Bone Mesenchymal Stem Cells via Sponging miR-324-3p. J Healthc Eng 2022; 2022: 3282860 [PMID: 35126917 DOI: 10.1155/2022/3282860]
- Li B, Xu H, Han H, Song S, Zhang X, Ouyang L, Qian C, Hong Y, Qiu Y, Zhou W, Huang M, Zhuang W. Exosome-76 mediated transfer of lncRUNX2-AS1 from multiple myeloma cells to MSCs contributes to osteogenesis. Oncogene 2018: 37: 5508-5519 [PMID: 29895968 DOI: 10.1038/s41388-018-0359-0]
- Liu Z, Liu H, Li Y, Shao Q, Chen J, Song J, Fu R. Multiple myeloma-derived exosomes inhibit osteoblastic differentiation 77 and improve il-6 secretion of bmscs from multiple myeloma. J Investig Med 2020; 68: 45-51 [PMID: 31784427 DOI: 10.1136/jim-2019-001010]
- Cheng Q, Li X, Liu J, Ye Q, Chen Y, Tan S. Multiple Myeloma-Derived Exosomes Regulate the Functions of 78 Mesenchymal Stem Cells Partially via Modulating miR-21 and miR-146a. Stem Cells Int 2017; 2017: 9012152 [PMID: 29333170 DOI: 10.1155/2017/9012152]
- Gu J, Wang M, Wang X, Li J, Liu H, Lin Z, Yang X, Zhang X. Exosomal miR-483-5p in Bone Marrow Mesenchymal 79 Stem Cells Promotes Malignant Progression of Multiple Myeloma by Targeting TIMP2. Front Cell Dev Biol 2022; 10: 862524 [PMID: 35300408 DOI: 10.3389/fcell.2022.862524]
- Umezu T, Imanishi S, Yoshizawa S, Kawana C, Ohyashiki JH, Ohyashiki K. Induction of multiple myeloma bone marrow 80 stromal cell apoptosis by inhibiting extracellular vesicle miR-10a secretion. Blood Adv 2019; 3: 3228-3240 [PMID: 31698453 DOI: 10.1182/bloodadvances.2019000403]
- 81 Gao X, Zhou J, Wang J, Dong X, Chang Y, Jin Y. Mechanism of exosomal miR-155 derived from bone marrow mesenchymal stem cells on stemness maintenance and drug resistance in myeloma cells. J Orthop Surg Res 2021; 16: 637 [PMID: 34689803 DOI: 10.1186/s13018-021-02793-9]



- Wang J, Hendrix A, Hernot S, Lemaire M, De Bruyne E, Van Valckenborgh E, Lahoutte T, De Wever O, Vanderkerken K, Menu E. Bone marrow stromal cell-derived exosomes as communicators in drug resistance in multiple myeloma cells. Blood 2014; 124: 555-566 [PMID: 24928860 DOI: 10.1182/blood-2014-03-562439]
- 83 Katz BZ. Adhesion molecules--The lifelines of multiple myeloma cells. Semin Cancer Biol 2010; 20: 186-195 [PMID: 20416379 DOI: 10.1016/j.semcancer.2010.04.003]
- Butcher EC, Picker LJ. Lymphocyte homing and homeostasis. Science 1996; 272: 60-66 [PMID: 8600538 DOI: 84 10.1126/science.272.5258.60
- Zhang X, Sun Y, Wang Z, Huang Z, Li B, Fu J. Up-regulation of connexin-43 expression in bone marrow mesenchymal 85 stem cells plays a crucial role in adhesion and migration of multiple myeloma cells. Leuk Lymphoma 2015; 56: 211-218 [PMID: 24724781 DOI: 10.3109/10428194.2014.913289]
- Cunningham BA, Hemperly JJ, Murray BA, Prediger EA, Brackenbury R, Edelman GM. Neural cell adhesion molecule: 86 structure, immunoglobulin-like domains, cell surface modulation, and alternative RNA splicing. Science 1987; 236: 799-806 [PMID: 3576199 DOI: 10.1126/science.3576199]
- Takeichi M. Functional correlation between cell adhesive properties and some cell surface proteins. J Cell Biol 1977; 75: 87 464-474 [PMID: 264120 DOI: 10.1083/jcb.75.2.464]
- 88 Edelman GM. Cell adhesion molecules. Science 1983; 219: 450-457 [PMID: 6823544 DOI: 10.1126/science.6823544]
- Bou Zerdan M, Nasr L, Kassab J, Saba L, Ghossein M, Yaghi M, Dominguez B, Chaulagain CP. Adhesion molecules in 89 multiple myeloma oncogenesis and targeted therapy. Int J Hematol Oncol 2022; 11: IJH39 [PMID: 35663420 DOI: 10.2217/ijh-2021-0017]
- Brackenbury R, Rutishauser U, Edelman GM. Distinct calcium-independent and calcium-dependent adhesion systems of 90 chicken embryo cells. Proc Natl Acad Sci U S A 1981; 78: 387-391 [PMID: 6165990 DOI: 10.1073/pnas.78.1.387]
- 91 Nagar B. Overduin M. Ikura M. Rini JM. Structural basis of calcium-induced E-cadherin rigidification and dimerization. Nature 1996; 380: 360-364 [PMID: 8598933 DOI: 10.1038/380360a0]
- Hale JS, Li M, Lathia JD. The malignant social network: cell-cell adhesion and communication in cancer stem cells. Cell 92 Adh Migr 2012; 6: 346-355 [PMID: 22796941 DOI: 10.4161/cam.21294]
- Samanta D, Almo SC. Nectin family of cell-adhesion molecules: structural and molecular aspects of function and specificity. Cell Mol Life Sci 2015; 72: 645-658 [PMID: 25326769 DOI: 10.1007/s00018-014-1763-4]
- Hirohashi S, Kanai Y. Cell adhesion system and human cancer morphogenesis. Cancer Sci 2003; 94: 575-581 [PMID: 94 12841864 DOI: 10.1111/j.1349-7006.2003.tb01485.x]
- Garcia-Gomez A, De Las Rivas J, Ocio EM, Díaz-Rodríguez E, Montero JC, Martín M, Blanco JF, Sanchez-Guijo FM, 95 Pandiella A, San Miguel JF, Garayoa M. Transcriptomic profile induced in bone marrow mesenchymal stromal cells after interaction with multiple myeloma cells: implications in myeloma progression and myeloma bone disease. Oncotarget 2014; 5: 8284-8305 [PMID: 25268740 DOI: 10.18632/oncotarget.2058]
- Roldán E, García-Pardo A, Brieva JA. VLA-4-fibronectin interaction is required for the terminal differentiation of human 96 bone marrow cells capable of spontaneous and high rate immunoglobulin secretion. J Exp Med 1992; 175: 1739-1747 [PMID: 1588291 DOI: 10.1084/jem.175.6.1739]
- Abe M, Hiura K, Wilde J, Moriyama K, Hashimoto T, Ozaki S, Wakatsuki S, Kosaka M, Kido S, Inoue D, Matsumoto T. 97 Role for macrophage inflammatory protein (MIP)-1alpha and MIP-1beta in the development of osteolytic lesions in multiple myeloma. Blood 2002; 100: 2195-2202 [PMID: 12200385]
- 98 Asosingh K, Vankerkhove V, Van Riet I, Van Camp B, Vanderkerken K. Selective in vivo growth of lymphocyte function- associated antigen-1-positive murine myeloma cells. Involvement of function-associated antigen-1-mediated homotypic cell-cell adhesion. Exp Hematol 2003; 31: 48-55 [PMID: 12543106 DOI: 10.1016/s0301-472x(02)00970-0]
- Lokhorst HM, Lamme T, de Smet M, Klein S, de Weger RA, van Oers R, Bloem AC. Primary tumor cells of myeloma 99 patients induce interleukin-6 secretion in long-term bone marrow cultures. Blood 1994; 84: 2269-2277 [PMID: 7919345]
- Hubbard AK, Rothlein R. Intercellular adhesion molecule-1 (ICAM-1) expression and cell signaling cascades. Free 100 Radic Biol Med 2000; 28: 1379-1386 [PMID: 10924857 DOI: 10.1016/s0891-5849(00)00223-9]
- Sherbenou DW, Su Y, Behrens CR, Aftab BT, Perez de Acha O, Murnane M, Bearrows SC, Hann BC, Wolf JL, Martin 101 TG, Liu B. Potent Activity of an Anti-ICAM1 Antibody-Drug Conjugate against Multiple Myeloma. Clin Cancer Res 2020; 26: 6028-6038 [PMID: 32917735 DOI: 10.1158/1078-0432.CCR-20-0400]
- 102 Ridley RC, Xiao H, Hata H, Woodliff J, Epstein J, Sanderson RD. Expression of syndecan regulates human myeloma plasma cell adhesion to type I collagen. Blood 1993; 81: 767-774 [PMID: 8427968]
- McCarron MJ, Park PW, Fooksman DR. CD138 mediates selection of mature plasma cells by regulating their survival. Blood 2017; 129: 2749-2759 [PMID: 28381397 DOI: 10.1182/blood-2017-01-761643]
- Andersen NF, Kristensen IB, Preiss BS, Christensen JH, Abildgaard N. Upregulation of Syndecan-1 in the bone marrow microenvironment in multiple myeloma is associated with angiogenesis. Eur J Haematol 2015; 95: 211-217 [PMID: 25353275 DOI: 10.1111/ejh.12473]
- 105 van Riet I, de Greef C, del Favero H, Demanet C, Van Camp B. Production of fibronectin and adherence to fibronectin by human myeloma cell lines. Br J Haematol 1994; 87: 258-265 [PMID: 7947265 DOI: 10.1111/j.1365-2141.1994.tb04907.x]
- Jordan AR, Racine RR, Hennig MJ, Lokeshwar VB. The Role of CD44 in Disease Pathophysiology and Targeted 106 Treatment. Front Immunol 2015; 6: 182 [PMID: 25954275 DOI: 10.3389/fimmu.2015.00182]
- 107 Barker HF, Hamilton MS, Ball J, Drew M, Franklin IM. Expression of adhesion molecules LFA-3 and N-CAM on normal and malignant human plasma cells. Br J Haematol 1992; 81: 331-335 [PMID: 1382543 DOI: 10.1111/j.1365-2141.1992.tb08236.x
- Van Camp B, Durie BG, Spier C, De Waele M, Van Riet I, Vela E, Frutiger Y, Richter L, Grogan TM. Plasma cells in 108 multiple myeloma express a natural killer cell-associated antigen: CD56 (NKH-1; Leu-19). Blood 1990; 76: 377-382 [PMID: 1695113]
- Ahsmann EJ, Lokhorst HM, Dekker AW, Bloem AC. Lymphocyte function-associated antigen-1 expression on plasma



cells correlates with tumor growth in multiple myeloma. Blood 1992; 79: 2068-2075 [PMID: 1562732]

- 110 Huang N, Kawano MM, Harada H, Harada Y, Sakai A, Kuramoto A, Niwa O. Heterogeneous expression of a novel MPC-1 antigen on myeloma cells: possible involvement of MPC-1 antigen in the adhesion of mature myeloma cells to bone marrow stromal cells. Blood 1993; 82: 3721-3729 [PMID: 8260709]
- 111 Huang N, Kawano MM, Mahmoud MS, Mihara K, Tsujimoto T, Niwa O, Kuramoto A. Expression of CD21 antigen on myeloma cells and its involvement in their adhesion to bone marrow stromal cells. Blood 1995; 85: 3704-3712 [PMID: 7780154]
- Damiano JS, Cress AE, Hazlehurst LA, Shtil AA, Dalton WS. Cell adhesion mediated drug resistance (CAM-DR): role of 112 integrins and resistance to apoptosis in human myeloma cell lines. Blood 1999; 93: 1658-1667 [PMID: 10029595]
- Klimienė I, Radzevičius M, Matuzevičienė R, Sinkevič-Belliot K, Kučinskienė ZA, Pečeliūnas V. Adhesion molecule 113 immunophenotype of bone marrow multiple myeloma plasma cells impacts the presence of malignant circulating plasma cells in peripheral blood. Int J Lab Hematol 2021; 43: 403-408 [PMID: 33185981 DOI: 10.1111/ijlh.13387]
- Pellat-Deceunynck C, Barillé S, Puthier D, Rapp MJ, Harousseau JL, Bataille R, Amiot M. Adhesion molecules on 114 human myeloma cells: significant changes in expression related to malignancy, tumor spreading, and immortalization. Cancer Res 1995; 55: 3647-3653 [PMID: 7543019]
- 115 Goodarzi A, Valikhani M, Amiri F, Safari A. The mechanisms of mutual relationship between malignant hematologic cells and mesenchymal stem cells: Does it contradict the nursing role of mesenchymal stem cells? Cell Commun Signal 2022; 20: 21 [PMID: 35236376 DOI: 10.1186/s12964-022-00822-6]
- 116 Gerdes HH, Rustom A, Wang X. Tunneling nanotubes, an emerging intercellular communication route in development. Mech Dev 2013; 130: 381-387 [PMID: 23246917 DOI: 10.1016/j.mod.2012.11.006]
- 117 Gurke S, Barroso JF, Gerdes HH. The art of cellular communication: tunneling nanotubes bridge the divide. Histochem Cell Biol 2008; 129: 539-550 [PMID: 18386044 DOI: 10.1007/s00418-008-0412-0]
- 118 Kolba MD, Dudka W, Zaręba-Kozioł M, Kominek A, Ronchi P, Turos L, Chroscicki P, Włodarczyk J, Schwab Y, Klejman A, Cysewski D, Srpan K, Davis DM, Piwocka K. Tunneling nanotube-mediated intercellular vesicle and protein transfer in the stroma-provided imatinib resistance in chronic myeloid leukemia cells. Cell Death Dis 2019; 10: 817 [PMID: 31659149 DOI: 10.1038/s41419-019-2045-8]
- 119 Mangolini M, Ringshausen I. Bone Marrow Stromal Cells Drive Key Hallmarks of B Cell Malignancies. Int J Mol Sci 2020; 21 [PMID: 32098106 DOI: 10.3390/ijms21041466]
- 120 de Rooij B, Polak R, van den Berk LCJ, Stalpers F, Pieters R, den Boer ML. Acute lymphoblastic leukemia cells create a leukemic niche without affecting the CXCR4/CXCL12 axis. Haematologica 2017; 102: e389-e393 [PMID: 28619846 DOI: 10.3324/haematol.2016.159517]
- Polak R, de Rooij B, Pieters R, den Boer ML. B-cell precursor acute lymphoblastic leukemia cells use tunneling 121 nanotubes to orchestrate their microenvironment. Blood 2015; 126: 2404-2414 [PMID: 26297738 DOI: 10.1182/blood-2015-03-634238]
- 122 Moschoi R, Imbert V, Nebout M, Chiche J, Mary D, Prebet T, Saland E, Castellano R, Pouyet L, Collette Y, Vey N, Chabannon C, Recher C, Sarry JE, Alcor D, Peyron JF, Griessinger E. Protective mitochondrial transfer from bone marrow stromal cells to acute myeloid leukemic cells during chemotherapy. Blood 2016; 128: 253-264 [PMID: 27257182 DOI: 10.1182/blood-2015-07-655860]
- 123 Mittal R, Karhu E, Wang JS, Delgado S, Zukerman R, Mittal J, Jhaveri VM. Cell communication by tunneling nanotubes: Implications in disease and therapeutic applications. J Cell Physiol 2019; 234: 1130-1146 [PMID: 30206931 DOI: 10.1002/jcp.27072
- 124 Zampieri LX, Silva-Almeida C, Rondeau JD, Sonveaux P. Mitochondrial Transfer in Cancer: A Comprehensive Review. Int J Mol Sci 2021; 22 [PMID: 33806730 DOI: 10.3390/ijms22063245]
- Suzuki R, Ogiya D, Ogawa Y, Kawada H, Ando K. Targeting CAM-DR and Mitochondrial Transfer for the Treatment of Multiple Myeloma. Curr Oncol 2022; 29: 8529-8539 [PMID: 36354732 DOI: 10.3390/curroncol29110672]
- 126 Cheng Y, Sun F, Thornton K, Jing X, Dong J, Yun G, Pisano M, Zhan F, Kim SH, Katzenellenbogen JA, Katzenellenbogen BS, Hari P, Janz S. FOXM1 regulates glycolysis and energy production in multiple myeloma. Oncogene 2022; 41: 3899-3911 [PMID: 35794249 DOI: 10.1038/s41388-022-02398-4]
- 127 Wang J, Liu X, Qiu Y, Shi Y, Cai J, Wang B, Wei X, Ke Q, Sui X, Wang Y, Huang Y, Li H, Wang T, Lin R, Liu Q, Xiang AP. Cell adhesion-mediated mitochondria transfer contributes to mesenchymal stem cell-induced chemoresistance on T cell acute lymphoblastic leukemia cells. J Hematol Oncol 2018; 11: 11 [PMID: 29357914 DOI: 10.1186/s13045-018-0554-z]
- Leleu X, Martin T, Weisel K, Schjesvold F, Iida S, Malavasi F, Manier S, Chang-Ki Min, Ocio EM, Pawlyn C, Perrot A, 128 Quach H, Richter J, Spicka I, Yong K, Richardson PG. Anti-CD38 antibody therapy for patients with relapsed/refractory multiple myeloma: differential mechanisms of action and recent clinical trial outcomes. Ann Hematol 2022; 101: 2123-2137 [PMID: 35943588 DOI: 10.1007/s00277-022-04917-5]
- D'Agostino M, Mina R, Gay F. Anti-CD38 monoclonal antibodies in multiple myeloma: another cook in the kitchen? 129 Lancet Haematol 2020; 7: e355-e357 [PMID: 32171060 DOI: 10.1016/S2352-3026(19)30254-6]





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