

# World Journal of *Hepatology*

*World J Hepatol* 2018 November 27; 10(11): 785-891



### EDITORIAL

- 785 Exosomal microRNAs as a potential therapeutic strategy in hepatocellular carcinoma  
*Gougelet A*
- 790 Treating nonalcoholic steatohepatitis with antidiabetic drugs: Will GLP-1 agonists end the struggle?  
*Kalogirou M, Sinakos E*
- 795 Novel insights in the prevention of perinatal transmission of hepatitis B  
*Tziomalos K, Neokosmidis G, Mavromatidis G, Dinas K*

### REVIEW

- 799 Role of traditional Chinese medicine in the management of patients with hepatocellular carcinoma  
*Xi SY, Minuk GY*
- 807 Genetic diversity of hepatitis viruses in West-African countries from 1996 to 2018  
*Assih M, Ouattara AK, Diarra B, Yonli AT, Compaore TR, Obiri-Yeboah D, Djigma FW, Karou S, Simpore J*
- 822 Bioengineered functional humanized livers: An emerging supportive modality to bridge the gap of organ transplantation for management of end-stage liver diseases  
*Vishwakarma SK, Lakkireddy C, Bardia A, Paspala SAB, Tripura C, Habeeb MA, Khan AA*

### MINIREVIEWS

- 837 Decision modelling for economic evaluation of liver transplantation  
*Qu Z, Krauth C, Amelung VE, Kaltenborn A, Gwiasda J, Harries L, Beneke J, Schrem H, Liersch S*

### ORIGINAL ARTICLE

#### Retrospective Study

- 849 African Americans are less likely to receive curative treatment for hepatocellular carcinoma  
*Sobotka LA, Hinton A, Conteh LF*

#### Observational Study

- 856 Factors associated with DAA virological treatment failure and resistance-associated substitutions description in HIV/HCV coinfecting patients  
*Salmon D, Trimoulet P, Gilbert C, Solas C, Lafourcade E, Chas J, Piroth L, Lacombe K, Katlama C, Peytavin G, Aumaitre H, Alric L, Boué F, Morlat P, Poizot-Martin I, Billaud E, Rosenthal E, Naqvi A, Miaillhes P, Bani-Sadr F, Esterle L, Carrieri P, Dabis F, Sogni P, Wittkop L; ANRS CO13 Hepaviv study group*



- 867 Cross-sectional study to determine viral hepatitis knowledge in different urban populations in Brazil  
*Cruz HM, Barbosa JR, Baima Colares JK, de Moraes Neto AH, Alencar MF, Bastos FI, da Mota JC, Carvalho-Costa FA, Ivantes CA, Lewis-Ximenez LL, Villar LM*

**META-ANALYSIS**

- 877 Cardiac stress testing and coronary artery disease in liver transplantation candidates: Meta-analysis  
*Soldera J, Camazzola F, Rodríguez S, Brandão A*

**CASE REPORT**

- 887 Trapped vessel of abdominal pain with hepatomegaly: A case report  
*Grandhe S, Lee JA, Chandra A, Marsh C, Frenette CT*

**ABOUT COVER**

Editorial Board Member of *World Journal of Hepatology*, Hie-Won Hann, MD, Professor, Department of Medicine, Division of Gastroenterology and Hepatology, Jefferson Medical College of Thomas Jefferson University, Philadelphia, PA 19107, United States

**AIM AND SCOPE**

*World Journal of Hepatology* (*World J Hepatol*, *WJH*, online ISSN 1948-5182, DOI: 10.4254), is a peer-reviewed open access academic journal that aims to guide clinical practice and improve diagnostic and therapeutic skills of clinicians.

*WJH* covers topics concerning liver biology/pathology, cirrhosis and its complications, liver fibrosis, liver failure, portal hypertension, hepatitis B and C and inflammatory disorders, steatohepatitis and metabolic liver disease, hepatocellular carcinoma, biliary tract disease, autoimmune disease, cholestatic and biliary disease, transplantation, genetics, epidemiology, microbiology, molecular and cell biology, nutrition, geriatric and pediatric hepatology, diagnosis and screening, endoscopy, imaging, and advanced technology. Priority publication will be given to articles concerning diagnosis and treatment of hepatology diseases. The following aspects are covered: Clinical diagnosis, laboratory diagnosis, differential diagnosis, imaging tests, pathological diagnosis, molecular biological diagnosis, immunological diagnosis, genetic diagnosis, functional diagnostics, and physical diagnosis; and comprehensive therapy, drug therapy, surgical therapy, interventional treatment, minimally invasive therapy, and robot-assisted therapy.

We encourage authors to submit their manuscripts to *WJH*. We will give priority to manuscripts that are supported by major national and international foundations and those that are of great basic and clinical significance.

**INDEXING/ABSTRACTING**

*World Journal of Hepatology* (*WJH*) is now abstracted and indexed in PubMed, PubMed Central, Emerging Sources Citation Index (Web of Science), Scopus, China National Knowledge Infrastructure (CNKI), and Superstar Journals Database.

**EDITORS FOR THIS ISSUE**

**Responsible Assistant Editor:** *Xiang Li*  
**Responsible Electronic Editor:** *Wen-Wen Tan*  
**Proofing Editor-in-Chief:** *Lian-Sheng Ma*

**Responsible Science Editor:** *Ying Dou*  
**Proofing Editorial Office Director:** *Jin-Lei Wang*

**NAME OF JOURNAL**  
*World Journal of Hepatology*

**ISSN**  
 ISSN 1948-5182 (online)

**LAUNCH DATE**  
 October 31, 2009

**FREQUENCY**  
 Monthly

**EDITORIAL BOARD MEMBERS**  
 All editorial board members resources online at <http://www.wjgnet.com/1948-5182/editorialboard.htm>

**EDITORIAL OFFICE**  
 Jin-Lei Wang, Director  
*World Journal of Hepatology*  
 Baishideng Publishing Group Inc  
 7901 Stoneridge Drive, Suite 501,

Pleasanton, CA 94588, USA  
 Telephone: +1-925-2238242  
 Fax: +1-925-2238243  
 E-mail: [editorialoffice@wjgnet.com](mailto:editorialoffice@wjgnet.com)  
 Help Desk: <http://www.f6publishing.com/helpdesk>  
<http://www.wjgnet.com>

**PUBLISHER**  
 Baishideng Publishing Group Inc  
 7901 Stoneridge Drive, Suite 501,  
 Pleasanton, CA 94588, USA  
 Telephone: +1-925-2238242  
 Fax: +1-925-2238243  
 E-mail: [bpgoffice@wjgnet.com](mailto:bpgoffice@wjgnet.com)  
 Help Desk: <http://www.f6publishing.com/helpdesk>  
<http://www.wjgnet.com>

**PUBLICATION DATE**  
 November 27, 2018

**COPYRIGHT**  
 © 2018 Baishideng Publishing Group Inc. Articles published by this Open Access journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non commercial and is otherwise in compliance with the license.

**SPECIAL STATEMENT**  
 All articles published in journals owned by the Baishideng Publishing Group (BPG) represent the views and opinions of their authors, and not the views, opinions or policies of the BPG, except where otherwise explicitly indicated.

**INSTRUCTIONS TO AUTHORS**  
<http://www.wjgnet.com/bpg/gerinfo/204>

**ONLINE SUBMISSION**  
<http://www.f6publishing.com>

## Observational Study

**Factors associated with DAA virological treatment failure and resistance-associated substitutions description in HIV/HCV coinfecting patients**

Dominique Salmon, Pascale Trimoulet, Camille Gilbert, Caroline Solas, Eva Lafourcade, Julie Chas, Lionel Piroth, Karine Lacombe, Christine Katlama, Gilles Peytavin, Hugues Aumaitre, Laurent Alric, François Boué, Philippe Morlat, Isabelle Poizot-Martin, Eric Billaud, Eric Rosenthal, Alissa Naqvi, Patrick Mialhes, Firouzé Bani-Sadr, Laure Esterle, Patrizia Carrieri, François Dabis, Philippe Sogni, Linda Wittkop; ANRS CO13 Hepaviv study group

Dominique Salmon, Assistance Publique des Hôpitaux de Paris, Hôpitaux Universitaires Paris Centre, Hôpital Hôtel Dieu, Unité des Maladies infectieuses et tropicales, Paris 75004, France

Karine Lacombe, Université Pierre et Marie Curie, UMR S1136, Institut Pierre Louis d'Epidémiologie et de Santé Publique, Paris 75646, France

Dominique Salmon, Université Paris Descartes, Sorbonne Paris Cité, Paris 75006, France

Christine Katlama, Université Paris-Sorbonne, Paris 75005, France

Pascale Trimoulet, CHU de Bordeaux, Hôpital Pellegrin, Laboratoire de Virologie, Bordeaux 33000, France

Christine Katlama, Assistance Publique des Hôpitaux de Paris Hôpital Pitié Salpêtrière, Services Maladies infectieuses et tropicales, Paris 75013, France

Pascale Trimoulet, CNRS-UMR 5234, Microbiologie fondamentale et Pathogénicité, Université de Bordeaux, Bordeaux 3000, France

Gilles Peytavin, Assistance Publique des Hôpitaux de Paris, Hôpital Bichat-Claude Bernard, Laboratoire de Pharmacologie, Paris 75877, France

Camille Gilbert, Eva Lafourcade, Philippe Morlat, Laure Esterle, François Dabis, Linda Wittkop, Univ. Bordeaux, ISPED, Inserm, Bordeaux Population Health Research Center, team MORPH3EUS, UMR 1219, CIC-EC 1401, Bordeaux F-33000, France

Gilles Peytavin, IAME, UMR 1137, Sorbonne Paris Cité, INSERM, Université Paris Diderot, Paris 75890, France

Caroline Solas, APHM, Hôpital La Timone, Laboratoire de Pharmacocinétique et Toxicologie, Marseille 13005, France

Hugues Aumaitre, Centre Hospitalier de Perpignan, Service Maladies infectieuses et tropicales, Perpignan 66000, France

Julie Chas, Assistance Publique des Hôpitaux de Paris, Hôpital Tenon, Service Maladies infectieuses et tropicales, Paris 75020, France

Laurent Alric, Centre Hospitalier Universitaire de Toulouse, Hôpital Purpan, Service Médecine interne-Pôle Digestif, Toulouse 31300, France

Lionel Piroth, Centre Hospitalier Universitaire de Dijon, Département d'Infectiologie, Dijon cedex 21079, France

Laurent Alric, UMR 152 IRD Université Toulouse III, Paul Sabatier, Toulouse 31330, France

Lionel Piroth, INSERM-CIC 1342 Université de Bourgogne, Dijon 21000, France

François Boué, Hôpital Antoine-Béclère, Assistance Publique des Hôpitaux de Paris, Université Paris Sud, Service Médecine interne et immunologie, Clamart 92140, France

Karine Lacombe, Assistance Publique des Hôpitaux de Paris, GHUEP site Saint-Antoine, Services Maladies infectieuses et tropicales, Paris 75011, France

Philippe Morlat, Centre Hospitalier Universitaire de Bordeaux, Service de médecine interne, Hôpital Saint-André, Bordeaux 33000, France

Isabelle Poizot-Martin, Aix-Marseille Univ, APHM Sainte-

Marguerite, Service d'Immuno-hématologie clinique, Marseille 13274, France

Isabelle Poizot-Martin, Patrizia Carrieri, Sciences Economiques and Sociales de la Santé and Traitement de l'Information Médicale, UMR912 INSERM, Aix-Marseille Université, IRD, Marseille 13009, France

Eric Billaud, Department of Infectious Diseases, CHU de Nantes and CIC 1413, Inserm, Nantes 44000, France

Eric Rosenthal, Centre Hospitalier Universitaire de Nice, Service de Médecine Interne, Hôpital l'Archet, Nice 06202, France

Eric Rosenthal, Université de Nice-Sophia Antipolis, Nice 06100, France

Alissa Naqvi, Centre Hospitalier Universitaire de Nice, Service d'Infectiologie, Hôpital l'Archet, Nice 06100, France

Patrick Miaillhes, Service des Maladies Infectieuses et Tropicales, Hospices Civils de Lyon, Hôpital de la Croix Rousse, Lyon 69004, France

Firouzé Bani-Sadr, Centre Hospitalier Universitaire de Reims, Service de Médecine Interne, Maladies Infectieuses et Immunologie Clinique, Reims 51100, France

Firouzé Bani-Sadr, Faculté de Médecine EA-4684/SFR CAP-SANTE, Université de Reims, Champagne-Ardenne, Reims 51100, France

Philippe Sogni, Assistance Publique des Hôpitaux de Paris, Hôpital Cochin, Service d'Hépatologie, Paris 75014, France

Philippe Sogni, Inserm U-1223 - Institut Pasteur, Paris 75015, France

Linda Wittkop, CHU de Bordeaux, Pôle de santé Publique, Service d'information médicale, Bordeaux F-33000, France

ORCID number: Dominique Salmon (0000-0002-6817-8951); Pascale Trimoulet (0000-0002-8371-381X); Camille Gilbert (0000-0003-3959-6174); Caroline Solas (0000-0002-0943-9648); Eva Lafourcade (0000-0001-8537-4201); Julie Chas (0000-0002-1001-9229); Lionel Piroth (0000-0003-4478-1032); Karine Lacombe (0000-0001-8772-9029); Christine Katlama (0000-0002-5862-3863); Gilles Peytavin (0000-0002-4359-537X); Hugues Aumaitre (0000-0002-0023-7652); Laurent Alric (0000-0003-0676-7539); François Boué (0000-0003-0161-4533); Philippe Morlat (0000-0001-6474-383X); Isabelle Poizot-Martin (0000-0002-5676-5411); Eric Billaud (0000-0002-3420-1228); Eric Rosenthal (0000-0003-1010-0964); Alissa Naqvi (0000-0001-6474-383X); Patrick Miaillhes (0000-0002-7979-3829); Firouzé Bani-Sadr (0000-0001-8268-866X); Laure Esterle (0000-0002-1017-1327); Patrizia Carrieri (0000-0002-6794-4837); François Dabis (0000-0002-1614-8857); Philippe Sogni (0000-0003-3316-8785); Linda Wittkop (0000-0003-2403-0960).

Author contributions: All the authors contributed to this work

Supported by Inserm-ANRS (French National Institute for Health and Medical Research - ANRS/France REcherche Nord and Sud Sida-hiv Hépatites).

**Institutional review board statement:** The study was approved by the Institutional Review Board Ile de France III, Paris, France

**Informed consent statement:** A written informed consent was obtained from each participant to the study.

**Conflict-of-interest statement:** Dominique Salmon has been speaker and received invitation to conferences by Gilead, Abott, and MSD. Laurent Alric received grant and personal fees from MSD, Gilead, Abbvie, Janssen and BMS outside the submitted work. Christine Katlama received consultancy fees and/or travel grants from MSD, Janssen, ViiV outside the submitted work. Karine Lacombe personal fees from Gilead, personal fees from Janssen, personal fees from Abbvie, personal fees from Merck outside the submitted work. Philippe Morlat received personal fees and non-financial support from GILEAD, Janssen, MSD and ViiV Health Care outside the submitted work. Gilles Peytavin received travel grants, consultancy fees or study grants from pharmaceutical companies including Abbvie, Bristol-Myers Squibb, Gilead sciences, Janssen, Merck and ViiV Healthcare outside the submitted work. Eric Rosenthal received personal fees from Gilead and Abbvie and travel grants, consultancy fees from Gilead, Abbvie, MSD and BMS outside the submitted work. Philippe Sogni received personal fees and non-financial support from Gilead, BMS, MSD Abvie outside the submitted work. Caroline Solas received personal fees from Gilead, Abbvie, Janssen, MSD and ViiV Healthcare outside the submitted work. Linda Wittkop reports grants from ANRS during the conduct of the study; personal fees from Janssen, Gilead, MSD, outside the submitted work. Other authors had nothing to declare.

**STROBE statement:** The guidelines of the STROBE Statement have been adopted. The authors have read the STROBE Statement-checklist of items, and the manuscript was prepared and revised according to the STROBE Statement-checklist of items.

**Open-Access:** This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

**Manuscript source:** Unsolicited manuscript

**Corresponding author to:** Dominique Salmon, MD, PhD, Professor, Assistance Publique des Hôpitaux de Paris, Hôpitaux Universitaires Paris Centre, Hôpital Hôtel Dieu, Unité des Maladies infectieuses et tropicales, Sorbonne Paris Cité, 1 place du Parvis Notre-Dame, Paris 75004, France. [dominique.salmon@aphp.fr](mailto:dominique.salmon@aphp.fr)  
Telephone: +33-1-42347956  
Fax: +33-1-42348852

Received: June 15, 2018

Peer-review started: June 15, 2018

First decision: July 9, 2018

Revised: September 10, 2018

Accepted: October 10, 2018

Article in press: October 10, 2018

Published online: November 27, 2018

## Abstract

### AIM

To describe factors associated with treatment failure and frequency of resistance-associated substitutions (RAS).

### METHODS

Human immunodeficiency virus (HIV)/hepatitis C virus (HCV) coinfecting patients starting a first direct-acting antiviral (DAA) regimen before February 2016 and included in the French ANRS CO13 HEPAVIH cohort were eligible. Failure was defined as: (1) non-response [HCV-RNA remained detectable during treatment, at end of treatment (EOT)]; and (2) relapse (HCV-RNA suppressed at EOT but detectable thereafter). Sequencing analysis was performed to describe prevalence of drug class-specific RAS. Factors associated with failure were determined using logistic regression models.

### RESULTS

Among 559 patients, 77% had suppressed plasma HIV-RNA < 50 copies/mL at DAA treatment initiation, 41% were cirrhotic, and 68% were HCV treatment-experienced. Virological treatment failures occurred in 22 patients and were mainly relapses (17, 77%) then undefined failures (3, 14%) and non-responses (2, 9%). Mean treatment duration was 16 wk overall. Post-treatment NS3, NS5A or NS5B RAS were detected in 10/14 patients with samples available for sequencing analysis. After adjustment for age, sex, ribavirin use, HCV genotype and treatment duration, low platelet count was the only factor significantly associated with a higher risk of failure (OR: 6.5; 95%CI: 1.8-22.6).

### CONCLUSION

Only 3.9% HIV-HCV coinfecting patients failed DAA regimens and RAS were found in 70% of those failing. Low platelet count was independently associated with virological failure.

**Key words:** Human immunodeficiency virus; Hepatitis C virus; Direct-acting antiviral; Treatment virological failure; Resistant associated mutations

© **The Author(s) 2018.** Published by Baishideng Publishing Group Inc. All rights reserved.

**Core tip:** In co-infected human immunodeficiency virus-hepatitis C virus (HCV) patients, after adjustment for age, sex, ribavirin use, HCV genotype and treatment duration, low platelets count was the only factor significantly associated with a higher risk of failure.

Salmon D, Trimoulet P, Gilbert C, Solas C, Lafourcade E, Chas J, Piroth L, Lacombe K, Katlama C, Peytavin G, Aumaitre H, Alric L, Boué F, Morlat P, Poizot-Martin I, Billaud E, Rosenthal E, Naqvi A, Miaillhes P, Bani-Sadr F, Esterle L, Carrieri P, Dabis F, Sogni P, Wittkop L; ANRS CO13 Hepaviv study group. Factors associated with DAA virological treatment failure and resistance-

associated substitutions description in HIV/HCV coinfecting patients. *World J Hepatol* 2018; 10(11): 856-866 Available from: URL: <http://www.wjgnet.com/1948-5182/full/v10/i11/856.htm> DOI: <http://dx.doi.org/10.4254/wjh.v10.i11.856>

## INTRODUCTION

The treatment of hepatitis C virus (HCV) infection had been revolutionized with the recent development of direct-acting antiviral (DAA) combinations. Cure rates of over 90%, similar to those in HCV mono-infected patients, can now be achieved in human immunodeficiency virus (HIV)/HCV coinfecting patients. This has been documented in clinical trials<sup>[1-5]</sup> as well as in real-life cohorts<sup>[6-9]</sup>. For the few patients failing treatment, resistance-associated substitutions (RAS) can emerge and emerging resistant strains appearing at viral rebound are a consequence rather than a cause of failure<sup>[10,11]</sup>.

The real causes of failure to all-oral DAA regimens can be multiple. Several social and medical factors can jeopardize treatment adherence. Some first-generation regimens may not be optimal to treat difficult cases of hepatitis C, such as decompensated cirrhosis or genotype 3 HCV infection. In rare circumstances, especially for genotype 1a viruses, baseline mutations in the non-structural-5A (NS5A) gene can preexist in the viral species before treatment introduction and may have a potentially deleterious impact on sustained virological response (SVR)<sup>[12]</sup>. Drug-drug interactions between DAA and ARV therapy or other commonly prescribed medications in HIV/HCV coinfecting patients are frequent and can decrease drug levels, thereby reducing the efficacy of therapy. Finally, adverse events, although rare with new DAA combinations, can occur and lead to treatment interruption and thus to treatment failure.

We aimed to describe the characteristics of patients failing first-line DAA treatment in the real-life French nationwide ANRS CO13 HEPAVIH cohort of HIV/HCV coinfecting patients. Furthermore, we described the emergence of clinically relevant RAS to DAA classes upon DAA treatment failure, and report pharmacological drug monitoring results. Finally, we identified factors associated with the occurrence of virological treatment failure.

## MATERIALS AND METHODS

### Study population

The ANRS CO13 HEPAVIH cohort (ClinicalTrials.gov Identifier: NCT03324633) is a national multi-centre prospective hospital-based observational study of patients coinfecting with HIV and viral hepatitis C that received approval by an Institutional Review board [Comité de Protection des Personnes (CPP) Ile de France III, Paris, France].

All patients included in the cohort gave their consent

for study participation. In addition, patients from the 29 centers participating in the ANRS CO13 HEPAVIH cohort, who weren't included in the cohort but who gave their consent for specific follow-up during and after DAA treatment, were also eligible. For this sub-study, patients were included if they had started an all-oral DAA-based regimen before January 2016 (3 mo treatment), February 2016 (2 mo treatment) or October 2015 (6 mo treatment). Patients who participated in completed and published clinical trials were included in the analysis. We did not include patients who were participating in an ongoing clinical trial (including those completed but not yet published), patients who were treated with combinations including Peg-interferon (PegIFN), or with the sofosbuvir (SOF) + ribavirin (RBV) combination. Patients with premature treatment interruption for intolerance or death were also excluded because we were specifically interested in a virological outcome. The DAA regimen was at the discretion of the patient's physician<sup>[13-15]</sup>.

### Data collection and definitions

The following data were collected prospectively by each participating center, using an eCRF: Age, sex, risk factors for both HIV and HCV infections, HCV genotype, previous anti-HCV treatment, HIV-related characteristics, start and end dates of DAA treatment, initial doses of anti-HCV and anti-HIV drugs, any changes during follow-up, and HCV-RNA at each time point [baseline, week (W)2, W4, W8, W12 if treatment duration was 24 wk, EOT, follow-up W4 (FU-W4) and FU-W12]. Virological treatment failures were categorized as: (1) Non-response: HCV-RNA never undetectable during treatment; (2) Relapse: HCV-RNA undetectable at EOT and then detectable within the following 12 wk; and (3) Undefined failure: HCV-RNA unknown at end of treatment (EOT) and positive thereafter, without premature discontinuation of treatment. Cirrhotic status was based on liver biopsy (METAVIR fibrosis stage F4), liver stiffness  $\geq 12.5$  kPa (FibroScan®; Echosens, France), a FibroTest® value  $\geq 0.75$  (Biopredictive, France) or physical and biological signs of end-stage liver disease, as previously published<sup>[16,17]</sup>.

### Sequencing analysis

Patients with virological treatment failure, who provided specific consent for HCV genotype testing and who had HCV-RNA  $> 1000$  IU/mL at the sequencing time point were included for HCV testing. Prevalence of drug class-specific RAS was evaluated at failure. The HCV NS3, NS5A and/or NS5B domains were amplified by reverse transcriptase nested polymerase chain reaction (PCR) using genotype and subtype-specific PCR primers to ensure successful amplification of the target gene(s). PCR products were purified and analyzed by population sequencing using an automated sequencer (ABI-3500xL Dx). The cutoff frequency for detecting variants with Sanger sequencing was approximately 15%. Sanger-derived sequences were aligned with Clustal\_W, version

1.74 (Conway Institute UCD, Dublin, Ireland). NS3, NS5A and NS5B RAS were defined as clinically relevant when inducing  $> 10$ -fold resistance to DAA<sup>[13,18-20]</sup>.

### Drug concentrations

Plasma drug concentrations for DAA and RBV were collected, when available, for patients included in the cohort as part of routine therapeutic drug monitoring performed in several centers. Drug concentrations were measured using liquid chromatography coupled with the tandem mass spectrometry method<sup>[21]</sup>. Data were considered interpretable if concentrations were determined at steady-state and information regarding the time of the last drug intake was available.

A suboptimal concentration was defined as below the  $2 \mu\text{g/mL}$  threshold for RBV<sup>[22,23]</sup>, and when concentrations were below the reported expected range for DAA<sup>[24-27]</sup>.

### Statistical analysis

We included all patients who met the inclusion criteria, as described in the study population section. Variables are described as number and percentages, or median and IQR [or mean (SD)], as appropriate. Patient characteristics are reported upon initiation of DAA treatment. The Wilcoxon-Mann-Whitney test and Fisher's exact test were used to compare quantitative and qualitative variables between groups, respectively. Factors associated with virological treatment failure were determined using logistic regression models. In order to identify new independent predictors of virological treatment failure, we systematically adjusted for a fixed set of potential confounders based on literature reports. The following variables were thus forced in all models: age, sex, RBV use, and prescribed treatment duration<sup>[28]</sup>. We then tested the following variables in the model containing the forced variables: HCV genotype (3 vs others), cirrhosis (Yes vs No), severe cirrhosis (Yes vs No), and defined by a B or C or an elastometry value  $\geq 20$  kPa), plasma HIV-RNA (detectable vs undetectable), and platelet count ( $< 100$  Giga/L vs  $\geq 100$  Giga/L). The effect of RBV on virological treatment failure and other potential factors was assessed by a marginal structural model (MSM) in order to consider a potential indication bias for the prescription of RBV. Sensitivity analyses, including patients with premature treatment discontinuations for intolerance/death, were also performed. The statistical methods of this study were reviewed by Linda Wittkop from Bordeaux Population Health Research Center, Bordeaux. SAS software version 9.4 (SAS Institute Inc., Cary, North Carolina) was used for all analyses.

## RESULTS

### General characteristics at DAA initiation

Among 877 patients treated with DAA-combination, 559 met the inclusion criteria and were included in the analysis (318 were not included for the following reasons: Treatment with PegIFN ( $n = 30$ ), inclusion

**Table 1 Patient characteristics at treatment initiation according to virological response**

	Overall ( <i>n</i> = 559)	SVR ( <i>n</i> = 537)	Virological treatment failure ( <i>n</i> = 22)	<i>P</i> value
Male sex	431 (77)	414 (77)	17 (77)	0.985
Age (yr)	52 (49-56)	52 (49-56)	53 (51-57)	0.586
CD4 (/mm <sup>3</sup> ) ( <i>n</i> = 557)	618 (426-850)	619 (429-861)	527 (346-704)	0.040
Undetectable HIV-RNA ( <i>n</i> = 558)	486 (87)	469 (88)	17 (77)	0.186
ARV treatment	549 (98)	527 (98)	22 (100)	1.000
PI <sup>1</sup>	127 (23)	122 (23)	5 (23)	
NNRTI <sup>2</sup>	98 (18)	95 (18)	3 (14)	
II <sup>3</sup>	204 (37)	197 (37)	7 (32)	
Others	120 (22)	113 (21)	7 (32)	
Active tobacco consumption ( <i>n</i> = 263)	153 (58)	148 (58)	5 (71)	0.703
Active alcohol consumption ( <i>n</i> = 266)	135 (51)	132 (51)	3 (43)	0.719
Active drug consumption ( <i>n</i> = 257)	7 (3)	7 (3)	0 (0)	1.000
HCV genotype ( <i>n</i> = 558)				0.475
1 without precision	26 (5)	24 (5)	2 (9)	
1a	232 (42)	221 (41)	11 (50)	
1b	64 (12)	64 (12)	0 (0)	
2	6 (1)	6 (1)	0 (0)	
3	62 (11)	60 (11)	2 (9)	
4	165 (30)	158 (30)	7 (32)	
5	1 (0)	1 (0)	0 (0)	
6	2 (0)	2 (0)	0 (0)	
Cirrhosis ( <i>n</i> = 555)	209 (38)	200 (38)	9 (41)	0.748
Child Pugh, if cirrhosis ( <i>n</i> = 189)				0.537
A	172 (91)	165 (91)	7 (88)	
B/C	17 (9)	16 (9)	1 (12)	
FIB-4 ( <i>n</i> = 405)	2.1 (1.4-3.7)	2.1 (1.4-3.7)	3.3 (1.9-7.3)	0.313
FIB-4 > 3.25 ( <i>n</i> = 405)	120 (30)	113 (29)	7 (50)	0.132
Elastometry (kPa) ( <i>n</i> = 115)	9 (6-14)	9 (6-14)	10 (6-17)	0.942
Elastometry ≥ 12.5 kPa ( <i>n</i> = 115)	32 (28)	30 (27)	2 (50)	0.309
Elastometry ≥ 20 kPa ( <i>n</i> = 115)	17 (15)	16 (14)	1 (25)	0.478
HCV treatment history				0.570
Naïve	210 (38)	203 (38)	7 (32)	
Pretreated	349 (62)	334 (62)	15 (68)	
HCV viral load (log <sub>10</sub> IU/mL) ( <i>n</i> = 558)	6.09 (5.59-6.51)	6.09 (5.59-6.51)	6.04 (5.72-6.49)	0.886
Prothrombin rate ( <i>n</i> = 298)	99 (89-100)	99 (89-100)	92 (82-100)	0.116
Prothrombin rate < 85% ( <i>n</i> = 298)	54 (18)	50 (17)	4 (40)	0.087
Platelets (Giga/L) ( <i>n</i> = 408)	171 (131-219)	171 (133-219)	148 (97-184)	0.168
Platelets < 100 Giga/L ( <i>n</i> = 408)	57 (14)	51 (13)	6 (43)	0.007
Albumin (g/L) ( <i>n</i> = 301)	41 (38-44)	41 (38-44)	42 (37-45)	0.939
Albumin < 35 g/L ( <i>n</i> = 301)	26 (9)	24 (8)	2 (25)	0.146
DAA-combination				NA <sup>5</sup>
SOF + DCV ± RBV <sup>4</sup>	240 (43)	231 (43)	9 (41)	
SOF/LDV ± RBV	271 (49)	261 (49)	10 (46)	
SOF + SMV ± RBV	26 (4)	23 (4)	3 (14)	
Others <sup>4</sup>	22 (4)	22 (4)	0 (0)	
Mean (SD) DAA treatment duration	16 (6)	15 (5)	16 (6)	

Results are presented as number (as percentages in brackets) or median (IQR in brackets) unless stated otherwise. <sup>1</sup>PI was boosted in 98 patients with SVR and in five patients with treatment failure; <sup>2</sup>NNRTI molecule was rilpivirine in 60 patients with SVR and three with failure, and was efavirenz in 25 patients with SVR; <sup>3</sup>II molecule was raltegravir in 153 patients with SVR and four patients with failure, and was dolutegravir in 38 patients with SVR and two with treatment failure; <sup>4</sup>Initial doses of DCV were 30, 60, 90 mg/d in respectively 57, 159 and 21 patients. The dose was unknown for the five other patients; <sup>5</sup>NA: not applicable, no formal statistical comparison was performed as the prescription of the DAA regimen was chosen by each patient's physician. SVR: Sustained virological response; ARV: Antiretroviral; PI: Protease inhibitor; NNRTI: Non-nucleoside reverse-transcriptase inhibitor; II: Integrase inhibitor; DAA: All-oral direct-acting antiviral; SOF: Sofosbuvir; RBV: Ribavirin; DCV: Daclatasvir; LDV: Ledipasvir; SMV: Simeprevir.

in an ongoing clinical trial (*n* = 2), treatment after the period of analysis (*n* = 190), no available treatment result (*n* = 32), treatment with SOF + RBV (*n* = 60), premature treatment interruption for intolerance (*n* = 3), and one patient died while on treatment). Mean treatment duration was 16 wk overall (15 wk in patients who failed DAA therapy and 16 wk in those with SVR). The characteristics of the 559 patients are summarized in

Table 1.

### Virological treatment failure

The virological treatment failure rate was 3.9% (95%CI: 2.5-5.9). Overall, 22 virological treatment failures were observed: Two non-responses, 17 relapses and three undefined virological treatment failures (HCV-RNA unknown at EOT). By univariate analysis (Table

**Table 2** Adjusted logistic regression for factors associated with virological treatment failure

Covariables	Model 1		Model 2		Model 3		Model 4	
	OR (95%CI)	P value	OR (95% CI)	P value	OR (95%CI)	P value	OR (95%CI)	P value
	n = 538		n = 538		n = 526		n = 395	
Age at treatment initiation (per 10 yr)	1.2 (0.6-2.4)	0.58	1.3 (0.7-2.5)	0.48	1.2 (0.6-2.4)	0.53	1.6 (0.7-4.0)	0.29
Ribavirin <i>vs</i> no ribavirin	1.0 (0.3-3.0)	0.97	1.1 (0.3-3.2)	0.93	1.0 (0.3-3.0)	0.97	1.4 (0.4-5.5)	0.61
Male sex <i>vs</i> female	1.0 (0.4-2.8)	0.98	0.9 (0.3-2.7)	0.92	1.0 (0.3-2.8)	0.97	0.8 (0.2-2.7)	0.69
Treatment duration 24 wk <i>vs</i> 12 wk	0.4 (0.1-1.4)	0.15	0.5 (0.2-1.5)	0.21	0.4 (0.1-1.4)	0.16	0.2 (0.0-1.0)	0.05
Platelet count < 100 Giga/L <i>vs</i> $\geq$ 100							6.5 (1.8-22.6)	0.004
Cirrhosis <i>vs</i> no cirrhosis	1.4 (0.5-3.9)	0.51						
HIV-RNA detectable <i>vs</i> undetectable			2.1 (0.7-5.9)	0.17				
Severe cirrhosis <i>vs</i> no severe cirrhosis					2.1 (0.4-10.3)	0.35		
HCV genotype 3 <i>vs</i> others							0.9 (0.1-7.5)	0.91

HCV: Hepatitis C virus; HIV: Human immunodeficiency virus.

1), patients with virological treatment failure had a significantly lower CD4 cell count (median 527 cells/mm<sup>3</sup>) compared to patients with SVR (619 cells/mm<sup>3</sup>;  $P = 0.040$ ). They also more frequently had a platelet count below 100 Giga/L ( $P = 0.007$ ) and a trend for more frequently having a prothrombin time < 85% (40% *vs* 17%,  $P = 0.087$ ) and albumin < 35 g/L (25% *vs* 8%,  $P = 0.146$ ). They also had a non-significant trend for less frequent HIV-RNA suppression (77% *vs* 88%,  $P = 0.186$ ).

#### Factors associated with treatment failure

In adjusted models (Table 2), platelet count < 100 Giga/L was significantly associated with a higher probability of virological treatment failure (Model 4). However, clinical cirrhosis status (Model 1), severe cirrhosis status (Model 3) or blood albumin (data not shown) were not associated with a higher probability of failure. Neither HIV-RNA (Model 2) nor CD4 cell count (data not shown) were associated with virological treatment failure. In addition, in the model containing platelet count, a prescribed treatment duration of 24 wk was associated with a lower risk of virological treatment failure (Model 4). RBV use was not associated with outcome in adjusted logistic regression models, and this result was confirmed by an analysis using MSMs (data not shown).

Sensitivity analyses, including patients with premature treatment discontinuations for intolerance/death, showed similar results (data not shown).

#### HCV resistance at virological treatment failure

The results of RAS analysis in the 14 patients with virological treatment failure, in whom either mutation NS3, NS5A or NS5B could be sequenced, are presented in Table 3. Almost three quarters of patients with available data (10/14; 71%) had at least one detectable RAS at the time of virological treatment failure. In patients receiving an NS5A inhibitor-based regimen, 55% (6/11) had NS5A RAS upon virological treatment failure. Common substitutions detected at failure included Q30R/H, 30E, 58D and/or Y93C/N, all found in patients with HCV genotype 1. In patients treated with daclatasvir (DCV) and an available NS5A RAS result ( $n = 5$ ), four patients developed resistance to DCV. Furthermore,

among six treated with ledipasvir (LDV) with available NS5A RAS result, two developed resistance to LDV. Overall, in all patients with available genotype ( $n = 14$ ), six (43%) presented at least one NS5A RAS, leading to a high level of resistance to NS5A inhibitors (> 10-fold resistance). In patients receiving NS3 protease inhibitors, 2/2 patients with available data had NS3 RAS upon virological treatment failure. The substitutions detected at failure were 80K, 170T, 174N and 168V, leading to a high level of resistance to most protease inhibitors.

Multiple RAS conferring a higher level of resistance were detected in three (21%) patients, including two with NS3 + NS5A RAS and one with NS3 + NS5A + NS5B RAS. These three patients were previously treated with PegIFN + RBV and were exposed to NS5A inhibitors but not NS3 inhibitors.

#### Pharmacological data

Nine of the 22 (41%) patients who had DAA therapy failure had measurements of DAA and/or RBV concentration at W2 or W4 of treatment, seven of which were interpretable. Among these seven patients, suboptimal concentrations were reported in two (29%). These low concentrations concerned either DCV (in a patient treated with SOF + DCV, whose ARV treatment was rilpivirine + raltegravir), or RBV (in a patient treated with SOF/LDV + RBV who was taking rilpivirine + dolutegravir).

## DISCUSSION

In this cohort of HIV/HCV coinfecting patients, who were treated with an interferon-free DAA regimen with or without RBV, we report a low virological treatment failure rate of 3.9%. Our results are similar to those observed in clinical trials<sup>[28]</sup> or previous real-world studies of HIV/HCV coinfection<sup>[6,7]</sup>. Most of these virological treatment failures were due to relapse (77%) followed by non-response (9%), while 14% were due to undefined virological treatment failures (HCV-RNA unknown at EOT).

Due to very high rates of SVR, it has been difficult to identify factors associated with virological treatment failure of DAA in real-world studies, and no study to date has focused on HIV coinfection. In studies of HCV

**Table 3** Resistance-associated substitution results in 14 patients with virological treatment failure for whom sequencing was performed in routine care

Pat	HCV treatment history	Treatment received	HCV genotype		Cirrhosis	ARV treatment	RAS		
			Before treatment	After treatment			NS3	NS5A	NS5B
A	Pretreated	SOF + SMV 12 wk	1a	1a	Yes	II	Q80K, I170T, S174N	Abs	Abs
B	Pretreated	SOF + SMV 12 wk	1a	1a	Yes	II	D168V	Abs	Abs
C	Pretreated	SOF/LDV 12 wk	4	4a	No	PI	Abs	Abs	Abs
G	Pretreated	SOF/LDV + RBV 12 wk	4	4d	No	Others	Abs	Abs	Abs
H	Pretreated	SOF + DCV 10 wk <sup>3</sup>	4	4	No	Others	Abs	ND	Abs
I	Naive	SOF/LDV 12 wk	1a	1a	No	PI	Abs	Abs	Abs
J	Pretreated	SOF/LDV 12 wk	1a	1a	No	Others	ND	Y93C	Abs
L	Naive	SOF + DCV 13 wk <sup>2</sup>	1a	1a	Yes	NNRTI	Q80K	Abs	Abs
M	Pretreated	SOF/LDV 12 wk	4	4a	No	PI	ND	Abs	A421V, M414L
N	Pretreated	SOF/LDV + RBV 12 wk	1a	1a	Yes	II	A168V	30E, 58D	Abs
P	Pretreated	SOF + DCV + RBV 12 wk <sup>1</sup>	1a	1a	No	PI	Abs	Y93N	Abs
Q	Pretreated	SOF + DCV + RBV 24 wk <sup>1</sup>	1a	1a	No	Others	T54S	Q30R	Abs
R	Pretreated	SOF + DCV 24 wk <sup>2</sup>	1a	1a	Yes	II	Q80K	Y93C	Y448H
W	Pretreated	SOF + DCV 24 wk <sup>1</sup>	1	1a	Yes	II	Abs	Q30H	Abs

<sup>1</sup>Initial dose of DCV: 30 mg/d; <sup>2</sup>Initial dose of DCV: 60 mg/d; <sup>3</sup>Initial dose of DCV: 90 mg/d. Pat: Patient; ARV: Antiretroviral; RAS: Resistance-associated substitution; NS3: Non-structural-3; NS5A: Non-structural-5A; NS5B: Non-structural-5B; SOF: Sofosbuvir; RBV: Ribavirin; DCV: Daclatasvir; LDV: Ledipasvir; SMV: Simeprevir; ND: Not done; PI: Protease inhibitor; NNRTI: Non-nucleoside reverse-transcriptase inhibitor; II: Integrase inhibitor; Abs: No RAS found.

monoinfected patients, however, several factors have been found to be associated with virological treatment failure: severity of cirrhosis (assessed by presence of ascites), low albumin, low platelet count/high total bilirubin<sup>[29-35]</sup>, male sex<sup>[30,31]</sup>, and the preexistence of baseline RAS<sup>[34,36]</sup>.

In our study, we found that low platelet count was significantly associated with a higher rate of virological treatment failure. It is likely that low platelet count is a surrogate marker of cirrhosis, since we found an association between low albumin levels and low PT time by univariate analysis. However, we failed to observe a significant relationship between severe cirrhosis and failure. This might be due to the fact that in cases of severe cirrhosis, physicians adapted the treatment to each complex situation by extending the duration or by adding RBV (76% of the patients with Child Pugh B or C cirrhosis received treatment of 24 wk duration vs 29% of the other patients in our study), and this might be explained by unreported events of decompensation.

In the first randomized phase 3 clinical trials, which assessed the efficacy and safety of DAA, decompensated cirrhosis was an exclusion criterion, which precluded the possibility of assessing this factor as a potential predictor of failure. More recently, several trials have clearly demonstrated that patients with Child Pugh B or C cirrhosis and those with genotype 3 infection have a lower rate of SVR with DAA alone and need the addition of RBV. This was the case for the SOF/LDV combination and for a combination<sup>[37,38]</sup> of velpatasvir/SOF<sup>[39]</sup>.

We observed a trend (by univariate analysis only) toward a higher rate of detectable HIV-RNA in patients with virological treatment failure vs in those with SVR ( $P = 0.19$ ). This might reflect suboptimal adherence, with

patients who are non-compliant for their HIV treatment while possibly also non-adherent to their HCV treatment. Nonetheless, this result did not remain significant by multivariable analysis and thus may also simply reflect a biased estimate.

Moreover, among seven patients with failure and interpretable pharmacological data, suboptimal blood concentrations of DAA were measured in two of them. These results could reflect different situations (drug interactions, suboptimal dosing errors, suboptimal adherence) and warrant both further investigation and wider-scale assessment of pharmacological data. Regarding RAS in our study, we did not determine pretreatment RAS and we cannot exclude the possibility that some failures may be due to pre-existing RAS. However, at a population level, the effects of baseline RAS in NS5A, although not rare, are minimal<sup>[10,36,40,41]</sup>. This prompted EASL experts<sup>[18,20]</sup> to recommend that genotyping should not be performed for naïve patients but instead considered when retreatment is anticipated with a NS5A inhibitor regimen in patients who have previously failed NS5A treatment.

In most of our patients who failed DAA-treatment, RAS was investigated. We found RAS in 50% of those failing NS5A-based therapy and in the two patients failing NS3, but no major RAS S282T to NS5B. This high prevalence of NS5A and -3 RAS failure in our study confirms the EASL recommendation to evaluate HCV resistance to NS5A inhibitors (spanning amino acids 24 to 93) if resistance testing is available, as these analyses can guide decisions for further treatment<sup>[18,20]</sup>.

There are several limitations to this study. Firstly, since the study was an observational cohort, our results must be interpreted with caution, since treatment

prescriptions were dependent on drug availability (with variations over time) and known efficacy with regards to HCV genotypes. Those results were obtained with second generation DAA (LDV, DCV, elbasvir/grazoprevir), and those results may not be entirely applicable to the newer, pangenotypic regimens such as velpatasvir/SOF or pibentavir/glecaprevir. Our analysis is limited by the small number of subjects with virological treatment failure, and thus likely has limited power to identify all potential risk factors. All patients with virological treatment failure could not be explored by genotyping to investigate the emergence of RAS due to the need to obtain patient consent. Furthermore, baseline genotyping was not available routinely, since this test is not recommended in France for treatment-naïve patients. Finally, Sanger sequencing was used for the detection of RAS, which may not be sensitive enough to detect minor populations of RAS (< 15%). The strengths of our study include prospective data collection with regular monitoring and high quality data.

In conclusion, our study identified that low platelet count is associated with a higher probability of DAA failure. This parameter likely reflects hepatic insufficiency, and our results are concordant with previously published findings on HCV mono-infected patients. We also speculate that some degree of low adherence could explain some cases of failure, since suboptimal drug levels were observed in 29% of the cases that could be explored, and HIV viral load was often detectable in patients with virological treatment failure to DAA. This study confirms the very low rate of treatment failure with all-oral DAA in HIV/HCV coinfecting patients, as well as the high risk of the emergence of non-structural NS3 or NS5A RAS in patients with virological DAA failure.

## ARTICLE HIGHLIGHTS

### Research background

In human immunodeficiency virus (HIV)/hepatitis C virus (HCV) coinfection, all-oral direct-acting antiviral (DAA) regimens achieve virological cures in > 95% of patients.

### Research motivation

Risk factors for failure are mainly related to severity of cirrhosis in HCV mono-infected patients, but are unknown in the population of HIV HCV coinfecting patients. We wanted to know whether additional factors related to non-adherence or HIV status could be involved in the occurrence of failures. We believed that identifying the risk factors for failure would allow for the adaptation of treatment to patients with higher risk of failure.

### Research objectives

The main objectives were to determine the risk factors for virological treatment failure to DAA in HIV/HCV coinfecting patients and to describe the frequency of RAS.

### Research methods

HIV/HCV coinfecting patients who started the first DAA regimen before February 2016 and who were included in the French ANRS CO13 HEPAVIH cohort were eligible. Failure was defined as: i) Non-response (HCV-RNA remained detectable during treatment, at end of treatment (EOT)), ii) relapse (HCV-RNA suppressed at EOT but detectable thereafter). Sequencing analysis

was performed to describe prevalence of drug class specific RAS. Factors associated with failure were determined using logistic regression models.

### Research results

Research findings: Among 559 patients, 77% had suppressed plasma HIV-RNA < 50 copies/mL at DAA treatment initiation, 41% were cirrhotic, and 68% were HCV treatment-experienced. Virological treatment failures occurred in 22 patients and were mainly relapses (17, 77%) then undefined failure (3, 14%) and non-responses (2, 9%). Mean treatment duration was 16 wk overall. Post-treatment NS3, NS5A or NS5B RAS were detected in 10/14 patients with samples available for sequencing analysis. After adjustment for age, sex, RBV use, HCV genotype and treatment duration, low platelet count was the only factor significantly associated with a higher risk of failure (OR: 6.5; 95%CI: 1.8-22.6); Contributions to the field: In HIV/HCV coinfecting patients, the risk factors of failure were more related to the severity of cirrhosis than to HIV immunovirological status or non-adherence issues. Problems that remain to be solved: It remains to be determined whether the low platelet count associated with a higher probability of failure reflects the severity of cirrhosis.

### Research conclusions

In our study of HIV/HCV patients receiving all-oral DAA, only 3.9% HIV-HCV coinfecting patients failed DAA regimens. RAS were found in 70% of those failing. Low platelet count was independently associated with virological failure. We think that this low platelet count reflects the severity of cirrhosis.

### Research perspectives

As the treatment failure number is low, it would be useful to build international collaborations and gather data for several cohorts in order to gain significance power. The results obtained with first generation all-oral DAA could be compared with the newer, pangenotypic drug regimen.

## ACKNOWLEDGMENTS

Patients of the ANRS CO13 HEPAVIH Cohort. Scientific Committee: Salmon D (co-Principal Investigator), Wittkop L (co-Principal Investigator), Sogni P (co-Principal Investigator), Esterle L (project manager), Trimoulet V, Izopet J, Serfaty L, Paradis V, Spire B, Carrier P, Valantin MA, Pialoux G, Chas J, Poizot-Martin I, Barangue K, Naqvi A, Rosenthal E, Bicart-See A, Bouchaud O, Gervais A, Lascoux-Combe C, Goujard V, Lacombe K, Duvalier C, Vittecoq D, Neau D, Morlat P, Bani-Sadr F, Meyer L, Boufassa F, Dominguez S, Autran B, Roque AM, Solas C, Fontaine H, Costagliola D, Piroth L, Simon A, Zucman D, Boué F, Mialhes P, Billaud E, Aumaître H, Rey D, Peytavin G, Petrov-Sanchez V, Pailhe A. Clinical Centres: APHP Cochin, Paris (Médecine Interne et Maladies Infectieuses: Salmon D, Usubillaga R; Hépatogastro-entérologie: Sogni P; Anatomopathologie: Terris B; Virologie: Tremeaux P); APHP Pitié-Salpêtrière, Paris (Maladies Infectieuses et Tropicales: Katlama C, Valantin MA, Stitou H; Hépatogastro-entérologie: Benhamou Y; Anatomopathologie: Charlotte F; Virologie: Fourati S); APHP Pitié-Salpêtrière, Paris (Médecine Interne: Simon A, Cacoub P, Nafissa S); APHM Sainte-Marguerite, Marseille (Service d'Immuno-Hématologie Clinique: Poizot-Martin I, Zaegel O, Laroche H; Virologie: Tamalet C); APHP Tenon, Paris (Maladies Infectieuses et Tropicales: Pialoux G, Chas J; Anatomopathologie: Callard P, Bendjaballah F; Virologie: Le Pendevan C); CHU Purpan, Toulouse (Maladies Infectieuses et

Tropicales: Marchou B; Hépatogastro-entérologie: Alric L, Barange K, Metivier S; Anatomopathologie: Selves J; Virologie: Larroquette F); CHU Archet, Nice (Médecine Interne: Rosenthal E; Infectiologie: Naqvi A, Rio V; Anatomopathologie: Haudebourg J, Saint-Paul MC; Virologie: Partouche C); APHP Avicenne, Bobigny (Médecine Interne - Unité VIH: Bouchaud O; Anatomopathologie: Ziou M; Virologie: Baazia Y); Hôpital Joseph Ducuing, Toulouse (Médecine Interne: Uzan M, Bicart-See A, Garipuy D, Ferro-Collados MJ; Anatomopathologie: Selves J; Virologie: Nicot F); APHP Bichat - Claude-Bernard, Paris (Maladies Infectieuses: Gervais A, Yazdanpanah Y; Anatomopathologie: Adle-Biassette H; Virologie: Alexandre G); APHP Saint-Louis, Paris (Maladies Infectieuses: Lascoux-Combe C, Molina JM; Anatomopathologie: Bertheau P; Virologie: Chaix ML, Delaugerre C, Maylin S); APHP Saint-Antoine (Maladies Infectieuses et Tropicales: Lacombe K, Bottero J, Krause J, Girard PM, Anatomopathologie: Wendum D, Cervera P, Adam J; Virologie: Viala C); APHP Bicêtre, Paris (Médecine Interne: Goujard C, Quertainmont Y, Teicher E; Virologie: Pallier C; Maladies Infectieuses: Vittecoq D); APHP Necker, Paris (Maladies Infectieuses et Tropicales: Lortholary O, Duvivier C, Rouzaud C, Lourenco J, Touam F, Louisin C; Virologie: Avettand-Fenoel V, Mélard A); CHU Pellegrin, Bordeaux (Maladies Infectieuses et Tropicales: Neau D, Ochoa A, Blanchard E, Castet-Lafarie S, Cazanave C, Malvy D, Dupon M, Dutronc H, Dauchy F, Lacaze-Buzy L; Anatomopathologie: Bioulac-Sage P; Virologie: Trimoulet P, Reigadas S); Hôpital Saint-André, Bordeaux (Médecine Interne et Maladies Infectieuses: Médecine Interne et Maladies Infectieuses: Morlat P, Lacoste D, Bonnet F, Bernard N, Hessamfar M, Paccalin JF, Martell C, Pertusa MC, Vandenhende M, Mercier P, Malvy D, Pistone T, Receveur MC, Méchain M, Duffau P, Rivoisy C, Faure I, Caldato S; Anatomopathologie: Bioulac-Sage P; Virologie: Trimoulet P, Reigadas S); Hôpital Haut-Levêque, Bordeaux (Médecine Interne: Pellegrin JL, Viallard JF, Lazzaro E, Greib C; Anatomopathologie: Bioulac-Sage P; Virologie: Trimoulet P, Reigadas S); Hôpital FOCH, Suresnes (Médecine Interne: Zucman D, Majerholc C; Virologie: Farfour E); APHP Antoine Béclère, Clamart (Médecine Interne: Boué F, Polo Devoto P, Kansau I, Chambrin C, Pignon C, Berroukeche L, Fior R, Martinez V; Virologie: Deback C); CHU Henri Mondor, Créteil (Immunologie Clinique: Lévy Y, Dominguez S, Lelièvre JD, Lascaux AS, Melica G); CHU Hôtel Dieu, Nantes (Maladies Infectieuses et Tropicales: Billaud E, Raffi F, Allavena C, Reliquet V, Boutoille D, Biron C; Virologie: Rodallec A, Le Guen L); Hôpital de la Croix Rousse, Lyon (Maladies Infectieuses et Tropicales: Miaillhes P, Peyramond D, Chidiac C, Ader F, Biron F, Boibieux A, Cotte L, Ferry T, Perpoint T, Koffi J, Zoulim F, Bailly F, Lack P, Maynard M, Radenne S, Amiri M; Virologie: Scholtes C, Le-Thi TT); CHU Dijon, Dijon (Département d'infectiologie: Piroth L, Chavanet P, Duong Van Huyen M, Buisson M, Waldner-Comberoux A, Mahy S, Binois R, Simonet-Lann AL, Croisier-Bertin

D); CH Perpignan, Perpignan (Maladies infectieuses et tropicales: Aumaître H); CHU Robert Debré, Reims (Médecine interne, maladies infectieuses et immunologie clinique: Bani-Sadr F, Lambert D, Nguyen Y, Berger JL); CHRU Strasbourg (Le Trait d'Union: Rey D, Partisani M, Batard ML, Cheneau C, Priester M, Bernard-Henry C, de Mautort E; Virologie: Gantner et P, Fafi-Kremer S), APHP Bichat-Claude Bernard (Pharmacologie: Peytavin G). Data collection: Roustant F, Kmiec I, Traore L, Lepuil S, Parlier S, Sicart-Payssan V, Bedel E, Touam F, Louisin C, Mole M, Bolliot C, Mebarki M, Adda-Lievin A, Makhoukhi F-Z, Braik O, Bayoud R, Pietri M-P, Le Baut V, Bornarel D, Chesnel C, Beniken D, Pauchard M, Akel S, Caldato S, Lions C, Chalal L, Julia Z, Hue H, Soria A, Cavellec M, Breau S, Joulie A, Fisher P, Ondo Eyene C, Ogoudjobi S, Brochier C, Thoirain-Galvan V. Management, statistical analyses: Boerg E, Carrieri P, Conte V, Dequae-Merchadou L, Desvallees M, Douiri N, Esterle L, Gilbert C, Gillet S, Knight R, Marcellin F, Michel L, Mora M, Nordmann S, Protopopescu C, Roux P, Spire B, Tezkratt S, Vilotitch A, Yaya I, Wittkop L.

## REFERENCES

- 1 **Osinusi A**, Townsend K, Kohli A, Nelson A, Seamon C, Meissner EG, Bon D, Silk R, Gross C, Price A, Sajadi M, Sidharthan S, Sims Z, Herrmann E, Hogan J, Teferi G, Talwani R, Proschan M, Jenkins V, Kleiner DE, Wood BJ, Subramanian GM, Pang PS, McHutchison JG, Polis MA, Fauci AS, Masur H, Kottlilil S. Virologic response following combined ledipasvir and sofosbuvir administration in patients with HCV genotype 1 and HIV co-infection. *JAMA* 2015; **313**: 1232-1239 [PMID: 25706232 DOI: 10.1001/jama.2015.1373]
- 2 **Rockstroh JK**, Nelson M, Katlama C, Lalezari J, Mallolas J, Bloch M, Matthews GV, Saag MS, Zamor PJ, Orkin C, Gress J, Klopfer S, Shaughnessy M, Wahl J, Nguyen BY, Barr E, Platt HL, Robertson MN, Sulkowski M. Efficacy and safety of grazoprevir (MK-5172) and elbasvir (MK-8742) in patients with hepatitis C virus and HIV co-infection (C-EDGE CO-INFECTION): a non-randomised, open-label trial. *Lancet HIV* 2015; **2**: e319-e327 [PMID: 26423374 DOI: 10.1016/S2352-3018(15)00114-9]
- 3 **Sulkowski MS**, Eron JJ, Wyles D, Trinh R, Lalezari J, Wang C, Slim J, Bhatti L, Gathe J, Ruane PJ, Elion R, Bredeek F, Brennan R, Blick G, Khatri A, Gibbons K, Hu YB, Fredrick L, Schnell G, Pilot-Matias T, Tripathi R, Da Silva-Tillmann B, McGovern B, Campbell AL, Podsadecki T. Ombitasvir, paritaprevir co-dosed with ritonavir, dasabuvir, and ribavirin for hepatitis C in patients co-infected with HIV-1: a randomized trial. *JAMA* 2015; **313**: 1223-1231 [PMID: 25706092 DOI: 10.1001/jama.2015.1328]
- 4 **Wyles D**, Bräu N, Kottlilil S, Daar ES, Ruane P, Workowski K, Luetkemeyer A, Adeyemi O, Kim AY, Doehle B, Huang KC, Mogalian E, Osinusi A, McNally J, Brainard DM, McHutchison JG, Naggie S, Sulkowski M; ASTRAL-5 Investigators. Sofosbuvir and Velpatasvir for the Treatment of Hepatitis C Virus in Patients Coinfected With Human Immunodeficiency Virus Type 1: An Open-Label, Phase 3 Study. *Clin Infect Dis* 2017; **65**: 6-12 [PMID: 28369210 DOI: 10.1093/cid/cix260]
- 5 **Wyles DL**, Ruane PJ, Sulkowski MS, Dieterich D, Luetkemeyer A, Morgan TR, Sherman KE, Dretler R, Fishbein D, Gathe JC Jr, Henn S, Hinesrososa F, Huynh C, McDonald C, Mills A, Overton ET, Ramgopal M, Rashbaum B, Ray G, Scarsella A, Yozviak J, McPhee F, Liu Z, Hughes E, Yin PD, Noviello S, Ackerman P; ALLY-2 Investigators. Daclatasvir plus Sofosbuvir for HCV in Patients Coinfected with HIV-1. *N Engl J Med* 2015; **373**: 714-725

- [PMID: 26196502 DOI: 10.1056/NEJMoa1503153]
- 6 **d'Arminio Monforte A**, Cozzi-Lepri A, Ceccherini-Silberstein F, De Luca A, Lo Caputo S, Castagna A, Mussini C, Cingolani A, Tavelli A, Shanyinde M, Gori A, Girardi E, Andreoni M, Antinori A, Puoti M; Ico Foundation and Hepalco Study Group. Access and response to direct antiviral agents (DAA) in HIV-HCV coinfecting patients in Italy: Data from the Ico cohort. *PLoS One* 2017; **12**: e0177402 [PMID: 28520749 DOI: 10.1371/journal.pone.0177402]
  - 7 **Ingiliz P**, Christensen S, Kimhofer T, Hueppe D, Lutz T, Schewe K, Busch H, Schmutz G, Wehmeyer MH, Boesecke C, Simon KG, Berger F, Rockstroh JK, Schulze zur Wiesch J, Baumgarten A, Mauss S. Sofosbuvir and Ledipasvir for 8 Weeks for the Treatment of Chronic Hepatitis C Virus (HCV) Infection in HCV-Monoinfected and HIV-HCV-Coinfected Individuals: Results From the German Hepatitis C Cohort (GECCO-01). *Clin Infect Dis* 2016; **63**: 1320-1324 [PMID: 27535952 DOI: 10.1093/cid/ciw567]
  - 8 **Piroth L**, Wittkop L, Lacombe K, Rosenthal E, Gilbert C, Miallhes P, Carrieri P, Chas J, Poizot-Martin I, Gervais A, Dominguez S, Neau D, Zucman D, Billaud E, Morlat P, Aumaitre H, Lascoux-Combe C, Simon A, Bouchaud O, Teicher E, Bani-Sadr F, Alric L, Vittecoq D, Boué F, Duvalier C, Valantin MA, Esterle L, Dabis F, Sogni P, Salmon D; ANRS CO13 HEPAVIH study group. Efficacy and safety of direct-acting antiviral regimens in HIV/HCV-coinfecting patients - French ANRS CO13 HEPAVIH cohort. *J Hepatol* 2017; **67**: 23-31 [PMID: 28235612 DOI: 10.1016/j.jhep.2017.02.012]
  - 9 **Sogni P**, Gilbert C, Lacombe K, Piroth L, Rosenthal E, Miallhes P, Gervais A, Esterle L, Chas J, Poizot-Martin I, Dominguez S, Simon A, Morlat P, Neau D, Zucman D, Bouchaud O, Lascoux-Combe C, Bani-Sadr F, Alric L, Goujard C, Vittecoq D, Billaud E, Aumaitre H, Boué F, Valantin MA, Dabis F, Salmon D, Wittkop L. All-oral Direct-acting Antiviral Regimens in HIV/Hepatitis C Virus-coinfecting Patients With Cirrhosis Are Efficient and Safe: Real-life Results From the Prospective ANRS CO13-HEPAVICH Cohort. *Clin Infect Dis* 2016; **63**: 763-770 [PMID: 27317796 DOI: 10.1093/cid/ciw379]
  - 10 **Di Maio VC**, Cento V, Lenci I, Aragri M, Rossi P, Barbaliscia S, Melis M, Verucchi G, Magni CF, Teti E, Bertoli A, Antonucci F, Bellocchi MC, Micheli V, Masetti C, Landonio S, Francioso S, Santopaolo F, Pellicelli AM, Calvaruso V, Gianserra L, Siciliano M, Romagnoli D, Cozzolongo R, Grieco A, Vecchiet J, Morisco F, Merli M, Brancaccio G, Di Biagio A, Loggi E, Mastroianni CM, Pace Palitti V, Tarquini P, Puoti M, Taliani G, Sarmati L, Picciotto A, Vullo V, Caporaso N, Paoloni M, Pasquazzi C, Rizzardini G, Parruti G, Craxi A, Babudieri S, Andreoni M, Angelico M, Perno CF, Ceccherini-Silberstein F; HCV Italian Resistance Network Study Group. Multiclass HCV resistance to direct-acting antiviral failure in real-life patients advocates for tailored second-line therapies. *Liver Int* 2017; **37**: 514-528 [PMID: 28105744 DOI: 10.1111/liv.13327]
  - 11 **Wyles D**, Dvory-Sobol H, Svarovskaia ES, Doehle BP, Martin R, Afdhal NH, Kowdley KV, Lawitz E, Brainard DM, Miller MD, Mo H, Gane EJ. Post-treatment resistance analysis of hepatitis C virus from phase II and III clinical trials of ledipasvir/sofosbuvir. *J Hepatol* 2017; **66**: 703-710 [PMID: 27923693 DOI: 10.1016/j.jhep.2016.11.022]
  - 12 **Zeuzem S**, Mizokami M, Pianko S, Mangia A, Han KH, Martin R, Svarovskaia E, Dvory-Sobol H, Doehle B, Hedskog C, Yun C, Brainard DM, Knox S, McHutchison JG, Miller MD, Mo H, Chuang WL, Jacobson I, Dore GJ, Sulkowski M. NS5A resistance-associated substitutions in patients with genotype 1 hepatitis C virus: Prevalence and effect on treatment outcome. *J Hepatol* 2017; **66**: 910-918 [PMID: 28108232 DOI: 10.1016/j.jhep.2017.01.007]
  - 13 **American Association for the Study of Liver Diseases**. HCV Guidance: Recommendations for Testing, Managing, and Treating Hepatitis C. 2016. Available from: URL: <https://www.hcvguidelines.org/>
  - 14 **AFEF**. Recommandations AFEF sur la prise en charge des hépatites virales C. 2017. Available from: URL: <http://www.afef.asso.fr/ckfinder/userfiles/files/recommandations-textes-officiels/recommandations/RecommandationsAFEFmars2017.pdf>
  - 15 **European AIDS Clinical Society**. Guidelines version 8.0, 2015. Available from: URL: [http://www.eacsociety.org/files/guidelines\\_8\\_0-english\\_web.pdf](http://www.eacsociety.org/files/guidelines_8_0-english_web.pdf)
  - 16 **Loko MA**, Salmon D, Carrieri P, Wincock M, Mora M, Merchadou L, Gillet S, Pambrun E, Delaune J, Valantin MA, Poizot-Martin I, Neau D, Bonnard P, Rosenthal E, Barange K, Morlat P, Lacombe K, Gervais A, Rouges F, See AB, Lascoux-Combe C, Vittecoq D, Goujard C, Duvalier C, Spire B, Izopet J, Sogni P, Serfaty L, Benhamou Y, Bani-Sadr F, Dabis F; ANRS CO 13 HEPAVIH Study Group. The French national prospective cohort of patients co-infected with HIV and HCV (ANRS CO13 HEPAVIH): early findings, 2006-2010. *BMC Infect Dis* 2010; **10**: 303 [PMID: 20969743 DOI: 10.1186/1471-2334-10-303]
  - 17 **Miallhes P**, Gilbert C, Lacombe K, Arends JE, Puoti M, Rockstroh JK, Sogni P, Fontaine H, Rosenthal E, Wincock M, Loko MA, Wittkop L, Dabis F, Salmon D; ESCMID European Study Group on Viral Hepatitis. Triple therapy with boceprevir or telaprevir in a European cohort of cirrhotic HIV/HCV genotype 1-coinfecting patients. *Liver Int* 2015; **35**: 2090-2099 [PMID: 25650873 DOI: 10.1111/liv.12799]
  - 18 **European Association for the Study of the Liver**. EASL Recommendations on Treatment of Hepatitis C 2016. *J Hepatol* 2017; **66**: 153-194 [PMID: 27667367 DOI: 10.1016/j.jhep.2016.09.001]
  - 19 **Leroy V**, Angus P, Bronowicki JP, Dore GJ, Hezode C, Pianko S, Pol S, Stuart K, Tse E, McPhee F, Bhore R, Jimenez-Exposito MJ, Thompson AJ. Daclatasvir, sofosbuvir, and ribavirin for hepatitis C virus genotype 3 and advanced liver disease: A randomized phase III study (ALLY-3+). *Hepatology* 2016; **63**: 1430-1441 [PMID: 26822022 DOI: 10.1002/hep.28473]
  - 20 **European Association for the Study of the Liver**. EASL Recommendations on Treatment of Hepatitis C 2018. *J Hepatol* 2018; **69**: 461-511 [PMID: 29650333 DOI: 10.1016/j.jhep.2018.03.026]
  - 21 **Rezk MR**, Bendas ER, Basalious EB, Karim IA. Development and validation of sensitive and rapid UPLC-MS/MS method for quantitative determination of daclatasvir in human plasma: Application to a bioequivalence study. *J Pharm Biomed Anal* 2016; **128**: 61-66 [PMID: 27232152 DOI: 10.1016/j.jpba.2016.05.016]
  - 22 **Solas C**, Paré M, Quaranta S, Stanke-Labesque F; pour le groupe Suivi Thérapeutique Pharmacologique de la Société Française de Pharmacologie et de Thérapeutique. [Not Available]. *Thérapie* 2011; **66**: 221-230 [PMID: 27393202 DOI: 10.2515/therapie/2011036]
  - 23 **Dominguez S**, Ghosn J, Cassard B, Melica G, Poizot-Martin I, Solas C, Lascaux AS, Bouvier-Alias M, Katlama C, Lévy Y, Peytavin G. Erythrocyte and plasma ribavirin concentrations in the assessment of early and sustained virological responses to pegylated interferon-alpha 2a and ribavirin in patients coinfecting with hepatitis C virus and HIV. *J Antimicrob Chemother* 2012; **67**: 1449-1452 [PMID: 22396433 DOI: 10.1093/jac/dks045]
  - 24 **Food and Drug Administration**. Clinical Pharmacology and Biopharmaceutics Reviews. Copegus, 2002: 21-511. Available from: URL: [https://www.accessdata.fda.gov/drugsatfda\\_docs/nda/2002/21-411\\_Strattera\\_biopharmr\\_P3.pdf](https://www.accessdata.fda.gov/drugsatfda_docs/nda/2002/21-411_Strattera_biopharmr_P3.pdf)
  - 25 **Food and Drug Administration**. Clinical Pharmacology and Biopharmaceutics Reviews. Daklinza 206843 Orig1s000, 2014. Available from: URL: [https://www.accessdata.fda.gov/drugsatfda\\_docs/nda/2015/206843Orig1s000ClinPharmR.pdf](https://www.accessdata.fda.gov/drugsatfda_docs/nda/2015/206843Orig1s000ClinPharmR.pdf)
  - 26 **Food and Drug Administration**. Clinical Pharmacology and Biopharmaceutics Review(s). Harvoni 205834 Orig1s000, 2014. Available from: URL: [https://www.accessdata.fda.gov/drugsatfda\\_docs/nda/2014/205834Orig1s000MedR.pdf](https://www.accessdata.fda.gov/drugsatfda_docs/nda/2014/205834Orig1s000MedR.pdf)
  - 27 **Food and Drug Administration**. Clinical Pharmacology and Biopharmaceutics Reviews. Sofosbuvir (GS-7977) 204671Orig1s000, 2013. Available from: URL: [https://www.accessdata.fda.gov/drugsatfda\\_docs/nda/2013/204671Orig1s000ClinPharmR.pdf](https://www.accessdata.fda.gov/drugsatfda_docs/nda/2013/204671Orig1s000ClinPharmR.pdf)
  - 28 **Welzel TM**, Petersen J, Herzer K, Ferenci P, Gschwantler M, Wedemeyer H, Berg T, Spengler U, Weiland O, van der Valk M, Rockstroh J, Peck-Radosavljevic M, Zhao Y, Jimenez-Exposito

- MJ, Zeuzem S. Daclatasvir plus sofosbuvir, with or without ribavirin, achieved high sustained virological response rates in patients with HCV infection and advanced liver disease in a real-world cohort. *Gut* 2016; **65**: 1861-1870 [PMID: 27605539 DOI: 10.1136/gutjnl-2016-312444]
- 29 **Chang CY**, Nguyen P, Le A, Zhao C, Ahmed A, Daugherty T, Garcia G, Lutchman G, Kumari R, Nguyen MH. Real-world experience with interferon-free, direct acting antiviral therapies in Asian Americans with chronic hepatitis C and advanced liver disease. *Medicine* (Baltimore) 2017; **96**: e6128 [PMID: 28178174 DOI: 10.1097/MD.00000000000006128]
- 30 **Dalgard O**, Weiland O, Noraberg G, Karlsen L, Heggelund L, Färkkilä M, Balslev U, Belard E, Øvrehus A, Skalshei Kjør M, Krarup H, Thorup Røge B, Hallager S, Madsen LG, Lund Laursen A, Lagging M, Weis N. Sofosbuvir based treatment of chronic hepatitis C genotype 3 infections-A Scandinavian real-life study. *PLoS One* 2017; **12**: e0179764 [PMID: 28704381 DOI: 10.1371/journal.pone.0179764]
- 31 **Ioannou GN**, Beste LA, Chang MF, Green PK, Lowy E, Tsui JI, Su F, Berry K. Effectiveness of Sofosbuvir, Ledipasvir/Sofosbuvir, or Paritaprevir/Ritonavir/Ombitasvir and Dasabuvir Regimens for Treatment of Patients With Hepatitis C in the Veterans Affairs National Health Care System. *Gastroenterology* 2016; **151**: 457-471.e5 [PMID: 27267053 DOI: 10.1053/j.gastro.2016.05.049]
- 32 **Ippolito AM**, Milella M, Messina V, Conti F, Cozzolongo R, Morisco F, Brancaccio G, Barone M, Santantonio T, Masetti C, Tundo P, Smedile A, Carretta V, Gatti P, Termine AP, Valvano MR, Bruno G, Fabrizio C, Andreone P, Zappimulso M, Gaeta GB, Napoli N, Fontanella L, Lauletta G, Cuccorese G, Metrangola A, Francavilla R, Ciraci E, Rizzo S, Andriulli A. HCV clearance after direct-acting antivirals in patients with cirrhosis by stages of liver impairment: The ITAL-C network study. *Dig Liver Dis* 2017; **49**: 1022-1028 [PMID: 28487083 DOI: 10.1016/j.dld.2017.03.025]
- 33 **Jiménez-Macías FM**, Cabanillas-Casafranca M, Maraver-Zamora M, Romero-Herrera G, García-García F, Correia-Varela-Almeida A, Cabello-Fernández A, Ramos-Lora M. Experience in real clinical practice with new direct acting antivirals in chronic hepatitis C. *Med Clin* (Barc) 2017; **149**: 375-382 [PMID: 28416232 DOI: 10.1016/j.medcli.2017.03.007]
- 34 **Kan H**, Imamura M, Kawakami Y, Daijo K, Teraoka Y, Honda F, Nakamura Y, Morio K, Kobayashi T, Nakahara T, Nagaoki Y, Kawaoka T, Tsuge M, Aikata H, Hayes CN, Miki D, Ochi H, Honda Y, Mori N, Takaki S, Tsuji K, Chayama K. Emergence of drug resistance-associated variants and changes in serum lipid profiles in sofosbuvir plus ledipasvir-treated chronic hepatitis C patients. *J Med Virol* 2017; **89**: 1963-1972 [PMID: 28657143 DOI: 10.1002/jmv.24885]
- 35 **Terrault NA**, Zeuzem S, Di Bisceglie AM, Lim JK, Pockros PJ, Frazier LM, Kuo A, Lok AS, Shiffman ML, Ben Ari Z, Akushevich L, Vainorius M, Sulkowski MS, Fried MW, Nelson DR; HCV-TARGET Study Group. Effectiveness of Ledipasvir-Sofosbuvir Combination in Patients With Hepatitis C Virus Infection and Factors Associated With Sustained Virologic Response. *Gastroenterology* 2016; **151**: 1131-1140.e5 [PMID: 27565882 DOI: 10.1053/j.gastro.2016.08.004]
- 36 **Sarrazin C**, Dvory-Sobol H, Svarovskaia ES, Doehle BP, Pang PS, Chuang SM, Ma J, Ding X, Afdhal NH, Kowdley KV, Gane EJ, Lawitz E, Brainard DM, McHutchison JG, Miller MD, Mo H. Prevalence of Resistance-Associated Substitutions in HCV NS5A, NS5B, or NS3 and Outcomes of Treatment With Ledipasvir and Sofosbuvir. *Gastroenterology* 2016; **151**: 501-512.e1 [PMID: 27296509 DOI: 10.1053/j.gastro.2016.06.002]
- 37 **Charlton M**, Gane E, Manns MP, Brown RS Jr, Curry MP, Kwo PY, Fontana RJ, Gilroy R, Teperman L, Muir AJ, McHutchison JG, Symonds WT, Brainard D, Kirby B, Dvory-Sobol H, Denning J, Arterburn S, Samuel D, Forns X, Terrault NA. Sofosbuvir and ribavirin for treatment of compensated recurrent hepatitis C virus infection after liver transplantation. *Gastroenterology* 2015; **148**: 108-117 [PMID: 25304641 DOI: 10.1053/j.gastro.2014.10.001]
- 38 **Rockstroh JK**, Peters L, Grint D, Soriano V, Reiss P, Monforte Ad, Beniowski M, Losso MH, Kirk O, Kupfer B, Mocroft A; EuroSIDA in EuroCoord. Does hepatitis C viremia or genotype predict the risk of mortality in individuals co-infected with HIV? *J Hepatol* 2013; **59**: 213-220 [PMID: 23583272 DOI: 10.1016/j.jhep.2013.04.005]
- 39 **Curry MP**, O'Leary JG, Bzowej N, Muir AJ, Korenblat KM, Fenkel JM, Reddy KR, Lawitz E, Flamm SL, Schiano T, Teperman L, Fontana R, Schiff E, Fried M, Doehle B, An D, McNally J, Osinusi A, Brainard DM, McHutchison JG, Brown RS Jr, Charlton M; ASTRAL-4 Investigators. Sofosbuvir and Velpatasvir for HCV in Patients with Decompensated Cirrhosis. *N Engl J Med* 2015; **373**: 2618-2628 [PMID: 26569658 DOI: 10.1056/NEJMoa1512614]
- 40 **Bartolini B**, Giombini E, Taibi C, Lionetti R, Montalbano M, Visco-Comandini U, D'Offizi G, Capobianchi MR, McPhee F, Garbuglia AR. Characterization of Naturally Occurring NS5A and NS5B Polymorphisms in Patients Infected with HCV Genotype 3a Treated with Direct-Acting Antiviral Agents. *Viruses* 2017; **9**: pii: E212 [PMID: 28783119 DOI: 10.3390/v9080212]
- 41 **Halfon P**, Scholtès C, Izopet J, Larrat S, Trimoulet P, Zoulim F, Alric L, Métivier S, Leroy V, Ouzan D, de Lédighen V, Mohamed S, Pénaranda G, Khiri H, Thélu MA, Plauzolles A, Chiche L, Bourlière M, Abravanel F. Baseline and post-treatment hepatitis C NS5A resistance in relapsed patients from a multicentric real-life cohort. *Antivir Ther* 2018; **23**: 307-314 [PMID: 28730994 DOI: 10.3851/IMP3184]

**P- Reviewer:** Abushady EAE, Bouare N, Lee GH, Milovanovic T

**S- Editor:** Ji FF **L- Editor:** Filipodia **E- Editor:** Bian YN





Published by **Baishideng Publishing Group Inc**  
7901 Stoneridge Drive, Suite 501, Pleasanton, CA 94588, USA  
Telephone: +1-925-223-8242  
Fax: +1-925-223-8243  
E-mail: [bpgoffice@wjgnet.com](mailto:bpgoffice@wjgnet.com)  
Help Desk: <http://www.f6publishing.com/helpdesk>  
<http://www.wjgnet.com>

