World Journal of *Hepatology*

World J Hepatol 2023 February 27; 15(2): 123-320





Published by Baishideng Publishing Group Inc

World Journal of Hepatology

Contents

Monthly Volume 15 Number 2 February 27, 2023

EDITORIAL

123 Metabolic-associated fatty liver disease: New nomenclature and approach with hot debate Fouad Y

REVIEW

- 129 Current status and prospect of treatments for recurrent hepatocellular carcinoma Yang YQ, Wen ZY, Liu XY, Ma ZH, Liu YE, Cao XY, Hou L, Hui X
- 151 Bioengineering liver tissue by repopulation of decellularised scaffolds Afzal Z, Huguet EL
- 180 Antioxidant and anti-inflammatory agents in chronic liver diseases: Molecular mechanisms and therapy Zhang CY, Liu S, Yang M

MINIREVIEWS

- 201 Galectin-3 inhibition as a potential therapeutic target in non-alcoholic steatohepatitis liver fibrosis Kram M
- 208 Clostridioides difficile infection in patients with nonalcoholic fatty liver disease-current status Kiseleva YV, Maslennikov RV, Gadzhiakhmedova AN, Zharikova TS, Kalinin DV, Zharikov YO
- 216 Sonographic gallbladder wall thickness measurement and the prediction of esophageal varices among cirrhotics

Emara MH, Zaghloul M, Amer IF, Mahros AM, Ahmed MH, Elkerdawy MA, Elshenawy E, Rasheda AMA, Zaher TI, Haseeb MT, Emara EH, Elbatae H

ORIGINAL ARTICLE

Clinical and Translational Research

225 Progressive changes in platelet counts and Fib-4 scores precede the diagnosis of advanced fibrosis in NASH patients

Zijlstra MK, Gampa A, Joseph N, Sonnenberg A, Fimmel CJ

Retrospective Cohort Study

237 Baseline hepatocyte ballooning is a risk factor for adverse events in patients with chronic hepatitis B complicated with nonalcoholic fatty liver disease

Tan YW, Wang JM, Zhou XB

Extended criteria brain-dead organ donors: Prevalence and impact on the utilisation of livers for 255 transplantation in Brazil

Braga VS, Boteon APCS, Paglione HB, Pecora RAA, Boteon YL



World Journal of Hepatology

Monthly Volume 15 Number 2 February 27, 2023

265 Prevalence of non-alcoholic fatty liver disease in patients with nephrotic syndrome: A population-based study

Onwuzo SS, Hitawala AA, Boustany A, Kumar P, Almomani A, Onwuzo C, Monteiro JM, Asaad I

Retrospective Study

Contents

274 Diabetes mellitus is not associated with worse short term outcome in patients older than 65 years old postliver transplantation

Alghamdi S, Alamro S, Alobaid D, Soliman E, Albenmousa A, Bzeizi KI, Alabbad S, Alqahtani SA, Broering D, Al-Hamoudi W

282 Hospitalizations for alcoholic liver disease during the COVID-19 pandemic increased more for women, especially young women, compared to men

Campbell JP, Jahagirdar V, Muhanna A, Kennedy KF, Helzberg JH

289 Racial and gender-based disparities and trends in common psychiatric conditions in liver cirrhosis hospitalizations: A ten-year United States study

Patel P, Ali H, Inayat F, Pamarthy R, Giammarino A, Ilyas F, Smith-Martinez LA, Satapathy SK

Observational Study

303 Outcomes of gout in patients with cirrhosis: A national inpatient sample-based study

Khrais A, Kahlam A, Tahir A, Shaikh A, Ahlawat S

CASE REPORT

311 Autoimmune hepatitis and eosinophilia: A rare case report Garrido I, Lopes S, Fonseca E, Carneiro F, Macedo G

LETTER TO THE EDITOR

318 Glecaprevir/pibrentasvir + sofosbuvir for post-liver transplant recurrent hepatitis C virus treatment Arora R, Martin MT, Boike J, Patel S



Contents

Monthly Volume 15 Number 2 February 27, 2023

ABOUT COVER

Editorial Board Member of World Journal of Hepatology, Hend M El Tayebi, PhD, Associate Professor, Pharmacist, Senior Scientist, Clinical Pharmacology and Pharmacogenomics Research Group, Department of Pharmacology and Toxicology, Faculty of Pharmacy and Biotechnology, German University in Cairo, Cairo 11835, Egypt. hend.saber@guc.edu.eg

AIMS AND SCOPE

The primary aim of World Journal of Hepatology (WJH, World J Hepatol) is to provide scholars and readers from various fields of hepatology with a platform to publish high-quality basic and clinical research articles and communicate their research findings online.

WJH mainly publishes articles reporting research results and findings obtained in the field of hepatology and covering a wide range of topics including chronic cholestatic liver diseases, cirrhosis and its complications, clinical alcoholic liver disease, drug induced liver disease autoimmune, fatty liver disease, genetic and pediatric liver diseases, hepatocellular carcinoma, hepatic stellate cells and fibrosis, liver immunology, liver regeneration, hepatic surgery, liver transplantation, biliary tract pathophysiology, non-invasive markers of liver fibrosis, viral hepatitis.

INDEXING/ABSTRACTING

The WJH is now abstracted and indexed in PubMed, PubMed Central, Emerging Sources Citation Index (Web of Science), Scopus, Reference Citation Analysis, China National Knowledge Infrastructure, China Science and Technology Journal Database, and Superstar Journals Database. The 2022 edition of Journal Citation Reports® cites the 2021 Journal Citation Indicator (JCI) for WJH as 0.52. The WJH's CiteScore for 2021 is 3.6 and Scopus CiteScore rank 2021: Hepatology is 42/70.

RESPONSIBLE EDITORS FOR THIS ISSUE

Production Editor: Yi-Xuan Cai; Production Department Director: Xiang Li; Editorial Office Director: Xiang Li.

NAME OF JOURNAL	INSTRUCTIONS TO AUTHORS
World Journal of Hepatology	https://www.wjgnet.com/bpg/gerinfo/204
ISSN 1948-5182 (online)	GUIDELINES FOR ETHICS DOCUMENTS
LAUNCH DATE	GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH
October 31, 2009	https://www.wignet.com/bpg/gerinfo/240
FREQUENCY	PUBLICATION ETHICS
Monthly	https://www.wjgnet.com/bpg/GerInfo/288
EDITORS-IN-CHIEF	PUBLICATION MISCONDUCT
Nikolaos Pyrsopoulos, Ke-Qin Hu, Koo Jeong Kang	https://www.wjgnet.com/bpg/gerinfo/208
EDITORIAL BOARD MEMBERS https://www.wignet.com/1948-5182/editorialboard.htm	ARTICLE PROCESSING CHARGE
PUBLICATION DATE February 27, 2023	STEPS FOR SUBMITTING MANUSCRIPTS
COPYRIGHT	ONLINE SUBMISSION
© 2023 Baishideng Publishing Group Inc	https://www.f6publishing.com

© 2023 Baishideng Publishing Group Inc. All rights reserved. 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA E-mail: bpgoffice@wjgnet.com https://www.wjgnet.com



W J H World Journal of Henatology Hepatology

Submit a Manuscript: https://www.f6publishing.com

World J Hepatol 2023 February 27; 15(2): 180-200

DOI: 10.4254/wjh.v15.i2.180

ISSN 1948-5182 (online)

REVIEW

Antioxidant and anti-inflammatory agents in chronic liver diseases: Molecular mechanisms and therapy

Chun-Ye Zhang, Shuai Liu, Ming Yang

Specialty type: Gastroenterology and hepatology

Provenance and peer review: Invited article; Externally peer reviewed.

Peer-review model: Single blind

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): B Grade C (Good): C Grade D (Fair): 0 Grade E (Poor): 0

P-Reviewer: Ban Q, China; Prikhodko V, Russia

Received: November 9, 2022 Peer-review started: November 9, 2022 First decision: November 23, 2022 Revised: November 30, 2022 Accepted: February 7, 2023 Article in press: February 7, 2023 Published online: February 27, 2023



Chun-Ye Zhang, Christopher S. Bond Life Sciences Center, University of Missouri, Columbia, MO 65211, United States

Shuai Liu, The First Affiliated Hospital, Zhejiang University, Hangzhou 310006, Zhejiang Province, China

Ming Yang, Department of Surgery, University of Missouri, Columbia, MO 65211, United States

Corresponding author: Ming Yang, DVM, PhD, Postdoctoral Fellow, Department of Surgery, University of Missouri, Room 2203, NexGen Precision Building, 1030 Hitt Street, Columbia, MO 65211, United States. yangmin@health.missouri.edu

Abstract

Chronic liver disease (CLD) is a continuous process that causes a reduction of liver function lasting more than six months. CLD includes alcoholic liver disease (ALD), non-alcoholic fatty liver disease (NAFLD), chronic viral infection, and autoimmune hepatitis, which can lead to liver fibrosis, cirrhosis, and cancer. Liver inflammation and oxidative stress are commonly associated with the development and progression of CLD. Molecular signaling pathways such as AMPactivated protein kinase (AMPK), C-Jun N-terminal kinase, and peroxisome proliferator-activated receptors (PPARs) are implicated in the pathogenesis of CLD. Therefore, antioxidant and anti-inflammatory agents from natural products are new potent therapies for ALD, NAFLD, and hepatocellular carcinoma (HCC). In this review, we summarize some powerful products that can be potential applied in all the stages of CLD, from ALD/NAFLD to HCC. The selected agents such as β -sitosterol, curcumin, genistein, and silymarin can regulate the activation of several important molecules, including AMPK, Farnesoid X receptor, nuclear factor erythroid 2-related factor-2, PPARs, phosphatidylinositol-3-kinase, and lysyl oxidase-like proteins. In addition, clinical trials are undergoing to evaluate their efficacy and safety.

Key Words: Chronic liver disease; Alcoholic liver disease; Non-alcoholic fatty liver disease; Hepatocellular carcinoma; Natural products; Inflammation; Oxidative stress; Treatment; Clinical trials

©The Author(s) 2023. Published by Baishideng Publishing Group Inc. All rights reserved.



Core Tip: Chronic liver disease (CLD) is a continuous process that causes a reduction of liver function lasting more than six months. CLD can be subclassified into alcoholic liver disease, non-alcoholic fatty liver disease, chronic viral infection, and autoimmune hepatitis, which can lead to liver fibrosis, cirrhosis, and cancer. Liver inflammation and oxidative stress are commonly associated with the development and progression of CLD. Therefore, anti-inflammatory and antioxidant agents are promising drugs for CLD treatment. Clinical trials are undergoing to evaluate their efficacy and safety.

Citation: Zhang CY, Liu S, Yang M. Antioxidant and anti-inflammatory agents in chronic liver diseases: Molecular mechanisms and therapy. World J Hepatol 2023; 15(2): 180-200 URL: https://www.wjgnet.com/1948-5182/full/v15/i2/180.htm DOI: https://dx.doi.org/10.4254/wjh.v15.i2.180

INTRODUCTION

Chronic liver disease (CLD) is a continuous process of inflammation, destruction, and regeneration of liver parenchyma, with a reduction of liver function that lasts more than six months[1]. According to the spectrum of etiologies of CLD, it can be subclassified into alcoholic liver disease (ALD), non-alcoholic fatty liver disease (NAFLD), chronic viral infection, and autoimmune hepatitis, which can lead to liver fibrosis, cirrhosis, and cancer[2-4].

The spectrum of ALD includes alcoholic fatty liver, alcoholic hepatitis, fibrosis, and cirrhosis[5]. Alcohol drinking history and volume are direct causing factors for ALD, which can progress into hepatocellular carcinoma (HCC, Figure 1), the most common type of primary liver cancer[3]. In addition, factors such as age, gender, genetic variants, chronic virus infection, and smoking contribute to the development and progression of ALD[6,7]. Development of transgenic mouse models of ALD has provided a powerful tool to understand the disease pathogenesis[8]. Cellular and molecular mechanism studies have advanced our knowledge of the pathogenesis of ALD[8,9]. Multiple processes including excessive accumulation of lipids, reactive oxygen species (ROS) production, mitochondrial dysfunction, and cell inflammation and death are involved in ALD pathogenesis^[10]. Despite all these efforts, there are no Food and Drug Administration-approved therapies for ALD[11].

NAFLD is the most common CLD with a broad spectrum, ranging from non-alcohol fatty liver (NAFL) to non-alcoholic steatohepatitis (NASH) with the progression of liver inflammation and different degrees of fibrosis[12]. NASH also can progression to HCC (Figure 1)[13]. The global prevalence of NAFLD was estimated to be 29.8% [95% confidence interval (CI): 28.6%-31.1%] in 2019[14], and the prevalence is estimated to be 32.4% (95%CI: 29.9-34.9) in 2022[15]. It affects more than 30% of people in the United States [16]. NAFLD is closely associated with other metabolic disorders, including obesity, diabetes, chronic kidney disease, and cardiovascular disease [17,18]. A new nomenclature for NAFLD has been suggested by a group of experts, namely metabolic dysfunction-associated fatty liver disease (MAFLD), which is based on the evidence of hepatic steatosis plus one of the following three criteria, including the presence of overweight or obesity, or presence of type 2 diabetes mellitus (T2DM), or evidence of metabolic dysregulation [19,20]. However, there are no currently approved medicines for NAFLD or MAFLD treatment[12].

Oxidative stress and inflammation are commonly associated with CLD independent of disease types [21,22]. For example, ethanol consumption can induce alcohol liver steatosis, inflammation, and production of ROS, resulting in the development of ALD with liver inflammation and oxidative stress [23]. In addition to hepatocyte injury, both innate and adaptive immune cells including macrophages, dendritic cells, neutrophils, and lymphocytes are involved in the development of CLD[24,25]. Production of ROS and inflammatory cytokines produced by immune cells under the stimuli of alcohol and diet metabolites, such as cholesterol and acetaldehyde, can further trigger liver oxidative stress, inflammation, and cell apoptosis or death to cause the progression of CLD[26,27].

Treatments, such as lifestyle intervention [28,29], gene editing [30,31], and pharmaceutical therapies [32], can ameliorate or cure CLD at the early stages. However, server condition of CLD requires liver transplantation, which lacks donor availability. Here, the roles of antioxidants and anti-inflammatory agents in CLD treatment, especially for ALD, NAFLD, and HCC, are reviewed. Examples of clinical trials for evaluating the potential efficacies of potential treatment agents are summarized.

DATABASE SEARCHING

The databases of PubMed, Cochrane Library (Wiley), Embase, Web of Science, and Google Scholar from the last five years (from July 2020) were searched for studies by keywords of CLD, ALD, NAFLD, or HCC, and their treatments with anti-oxidative and anti-inflammatory agents. Papers written in English



Zhang CY et al. Natural products in liver disease



Figure 1 The development of hepatocellular carcinoma from non-alcoholic fatty liver disease and alcoholic fatty liver disease. The prevalence (20%-30%) of non-alcoholic fatty liver (NAFL) in the world population and the following percentages of NAFL into non-alcoholic steatohepatitis (NASH) (15%-25%), NASH into cirrhosis (5%-10%), and cirrhosis into hepatocellular carcinoma (HCC) (2%-5%) are labeled. Around 90%-100% of heavy drinkers can develop alcoholic liver disease (ALD), then the percentages of progression from simple ALD into alcohol liver steatohepatitis (10%-35%), cirrhosis (8%-20%), and HCC (2%) are shown in the graphic. This cartoon was created using Biorender online tools (https://biorender.com). NASH: Non-alcoholic steatohepatitis.

were studied. When reviewing oxidative stress and/or inflammation-related molecules in CLD, the time restriction of the published data was removed.

INFLAMMATION AND OXIDATIVE STRESS IN CLD AND UNDERLYING MOLECULAR MECHANISMS

Inflammation and oxidative stress are commonly associated with each other in the pathogenesis of CLD [33], including ALD, NAFLD, and HCC. Several common signaling pathways are involved in liver inflammation and oxidative stress, such as Toll-like receptor (TLR)/nuclear factor kappa B (NF-κB) and heme oxygenase-1 (HO-1) signaling pathways [34,35]. Dysregulation of lipid metabolism contributes to the pathogenesis of CLD[36,37], which is commonly associated with liver oxidative stress and inflammation. Molecules such as peroxisome proliferator-activated receptors (PPARs) are involved in alcohol or non-alcohol factors-induced lipid metabolism dysregulation and hepatic steatosis[38,39]. In this section, we review some important signaling pathways involved in liver inflammation and oxidative stress during CLD.

AMP-activated protein kinase

AMP-activated protein kinase (AMPK) as a crucial energy sensor plays an important role in energy metabolism in multiple tissues, including the liver[40]. Activation of AMPK by metformin can reduce induced triglyceride accumulation in the livers of mice treated with ethanol compared to control groups [41]. Activation of sirtuin 1 (SIRT1)/Liver kinase B1/AMPK signaling with botulin (a triterpene) treatment reduces serum aminotransferase and triglyceride levels in mice with chronic-binge ethanol [42]. Activation of the AMPK signaling pathway with plant sterol ester of α -linolenic acid can also attenuate endoplasmic reticulum (ER) stress-induced hepatocyte apoptosis in mice with NAFLD[43]. Similarly, stimulating the activation of AMPK by an activator PXL770 reduces de novo lipogenesis in primary mice and human hepatocytes, which can result in the suppression of hepatic steatosis, inflammation, and fibrogenesis in mice with NASH. In addition, PXL770 has a direct inhibitory effect on the production of proinflammatory cytokines and activation of hepatic stellate cells[44].

C-Jun N-terminal kinase

Activation of C-Jun N-terminal kinase (JNK) signaling pathway is involved in lipotoxicity, inflammation, ER stress, and mitochondrial dysfunction. Palmitic acid (PA)-induced activation of JNK/Sab (SH3 domain-binding protein 5) signaling contributes to NASH progression, which is associated with mitochondrial dysfunction, oxidative stress, hepatic steatosis, and inflammation[45].

Deficiency of hypoxia-induced gene domain protein- 1α (Higd- 1α), a mitochondrial inner membrane protein, promotes free fatty acids (FFAs)-induced apoptosis and oxidative stress in hepatocytes [46]. In this process, the production of cytosolic oxidized mitochondrial DNA (ox-mtDNA) is increased, which induces activation of NOD-like receptor family pyrin domain containing 3 (NLRP3) inflammasomes and JNK signaling but decreases fatty acid oxidation (FAO). In contrast, exercise can increase the expression





DOI: 10.4254/wjh.v15.i2.180 Copyright ©The Author(s) 2023.

Figure 2 Molecular signaling pathway in liver inflammation and oxidative stress. Inflammation and oxidative stress are involved in the development of chronic liver diseases such as alcoholic liver disease, non-alcoholic fatty liver disease, non-alcoholic steatohepatitis, fibrosis, and cirrhosis into hepatocellular carcinoma. Many factors including cytokines, growth factors, and gut microbiota-derived products such as lipopolysaccharide can activate their receptors such as peroxisome proliferator-activated receptor-α and toll-like receptor 4, resulting in upregulation or inhibition of downstream genes to induce or prevent inflammatory cytokines and production of reactive oxygen species. This cartoon was created using Biorender online tools (https://biorender.com). LPS: Lipopolysaccharide; TLR4: Toll-like receptor 4; ALD: Alcoholic liver disease; NAFLD: Non-alcoholic fatty liver disease; NASH: Non-alcoholic steatohepatitis; PPAR-α: Peroxisome proliferator-activated receptor-α; SIRT1: Sirtuin 1; SREBP-1c: Sterol regulatory element binding protein 1c; PI3K: Phosphatidylinositol-3-kinase; AKT: Protein kinase B; mTOR: Mammalian target of rapamycin; FAO: Fatty acid oxidation; NLRP3: NOD-like receptor family pyrin domain containing 3; NF-κB: Nuclear factor kappa B; IL: Interleukin; TNF-α: Tumor necrosis factor-α; NLRP3: NOD-like receptor family pyrin domain containing 3; ROS: Reactive oxygen species; NOS: Nitric oxide synthase.

of Higd-1 α in the liver to ameliorate hepatic steatosis and inflammation by suppressing ox-mtDNA/NLRP3/JNK pathway[46].

Farnesoid X receptor

Farnesoid X receptor (FXR) is a nuclear receptor that metabolically regulates glucose, bile acid, and lipid metabolism[47,48]. Treatment of Lactobacillus reuteri can ameliorate lipid accumulation in mice with ALD by upregulating FXR expression, which is associated with the upregulation of carbohydrate response element binding protein and downregulation of sterol regulatory element binding transcription factor 1 and cluster of differentiation (CD36)[49]. In addition, the FXR/fibroblast growth factors (FGFs) axis (FGF-15 and FGF-19) also plays a key in the regulation of hepatic inflammation, lipid metabolism, and fibrosis[50,51]. Clinically, treatment of FXR agonist vonafexor also shows anti-fibrotic effects in patients with NASH[52].

Nuclear factor erythroid 2-related factor-2/HO-1

Nuclear factor erythroid 2-related factor-2 (Nrf2) is a key transcription factor that plays a critical role in oxidative stress and inflammatory responses. For example, Nrf2 expression is positively associated with oyster peptide-mediated suppression of inflammation mediated by upregulation of NF- κ B signaling and upregulation of antioxidant response in mice with ALD[53]. Activation of Nrf2 is involved in the protective effect of diallyl disulfide against chemical (CCl₄)-induced liver injury and oxidative stress [54]. HO-1, an inducible form of antioxidant zyme HO isoforms that regulates heme group degradation, plays an essential role in liver inflammation and oxidative stress[55]. Nrf2 can regulate HO-1 to suppress liver oxidative stress, ER stress, and inflammation[56].

Nrf2 also plays an important role in the pathogenesis of NASH. Activation of Nrf2 can ameliorate liver inflammation, ER stress, iron overload, and lipotoxicity to suppress NASH and oxidative stress, which can be suppressed by transforming growth factor-beta (TGF- β)[57]. Activation of Nrf2 can suppress the expression of ROS and NLRP3 and inhibit Caspase 1/interleukin (IL)-1 β and IL-18-mediated inflammation[58]. In addition, pharmacologic activation of Nrf2 by TBE-31, acetylenic tricyclic bis(cyano enone), decreases insulin resistance and liver fat accumulation, inflammation, fibrosis, and



oxidative stress in mice with a high-fat plus fructose diet. However, the TBR-31-mediated effect was abolished in Nrf2-null mice[59].

PPARs

PPARs are a group of nuclear receptor proteins that function as ligand-activated receptors to regulate genes in energy metabolism and inflammation. PPARs comprise three subtypes, PPAR- α , PPAR- β/δ , and PPAR- γ , which are pharmaceutical targets for disease treatments[60,61]. These PPARs play important roles in ALD[62], NAFLD[63], hepatitis virus-mediated liver injury[64], and HCC[65].

Activation of PPAR-α by agonist WY-14643 (Pirinixic Acid, Figure 2) ameliorates ethanol-induced liver fat accumulation by increasing FAO[66]. Sustained activation of PPAR- α can decrease obesity and improve insulin resistance to rebuild glucose homeostasis. However, it increases the risk of HCC development due to liver ER stress[67]. Treatment with GW9662, an antagonist of PPAR- γ , significantly decreased lipopolysaccharide (LPS)/TLR4-mediated expression of IL-1β, IL-6, inducible nitric oxide synthase, and nitrite (NO_2^{-}) concentration[68].

Treatment with a dual PPAR- α/γ agonist Saroglitazar is able to reduce serum transaminases and 63% of overweight patients with NALFD reduced bodyweight (> 5%)[69]. In addition, many clinical trials have been performed to evaluate the effects of PPARs in ALD. For example, pemafibrate can improve liver function and glucose metabolism in patients with hypertriglyceridemia[70] and decrease liver stiffness in patients with NAFLD measured by magnetic resonance elastography (ClinicalTrials.gov, number: NCT03350165)[71]. Treatments that target PPAR- α such as pemafibrate[71], PPAR- β/δ such as seladelpar^[72], and PPAR-y such as pioglitazone^[73,74] show promising efficacy in the clinic for CLD treatment (Figure 3). Meanwhile, a dual PPAR- α/δ agonist elafibranor and a pan-PPAR regulator lanifibranor show promising efficacy for CLD treatment in the clinic [75,76]. For example, a phase 2b clinical trial reveals that treatment of lanifibranor (1200 mg) compared with the placebo can decrease at least 2 points of steatosis, activity, and fibrosis score that incorporates scores for ballooning and inflammation^[76].

Phosphatidylinositol-3-kinase/protein kinase B/mammalian target of rapamycin

The phosphatidylinositol-3-kinase (PI3K)/protein kinase B (PKB or AKT)/mammalian target of rapamycin (mTOR) signaling pathway is implicated in the pathogenesis of liver disease and therapy [77, 78]. For example, this signaling pathway is involved in the anti-steatosis effect of D-mannose in ALD [79]. Activation of PI3K/AKT/mTOR signaling pathway by arecoline (2.5 µM), an alkaloid ester found in the betel nut palm seeds, promotes the proliferation and migration of HepG2 cells[80]. Acid-sensitive ion channel 1α can upregulate the activation of PI3K/AKT/mTOR signaling pathway to enhance the expression of matrix metalloproteinase (MMP)2 and MMP9 to promote liver cancer cell (HepG2 and SK-Hep1 cells) migration and invasion[81]. One human study also indicates that PI3K is more strongly expressed in tumors than that in cirrhotic livers but not AKT and mTOR, and the expression of PI3K in tumor tissues is independent of etiology [82]. In addition, activation of growth factor receptor protein tyrosine kinases (Figure 2) can result in autophosphorylation on tyrosine residues and subsequent binding and activation of PI3K[83], playing an important role in cancer development. Inhibition or blockade of this signaling pathway can suppress liver fibrosis[84,85] and cancer progression[86,87].

Furthermore, lysyl oxidase family members (LOX) and LOX-like proteins (LOXL1-4) play important roles in liver fibrosis and cancer[88]. Insulin resistance can promote extracellular matrix stabilization by upregulating hepatic production of LOXL2 through upregulation of the expression of Forkhead box protein O1 in NAFLD[89]. In addition, galectins such as galectin-3 also play an essential role in CLD[90-92], including liver fibrosis and cancer. Overall, these molecular signaling pathways are involved in liver inflammation and oxidative stress to promote the development of CLD to HCC (Figure 2).

ANTIOXIDANT AND ANTI-INFLAMMATORY AGENTS IN ALD

Many ingredients from natural products or plants have both antioxidant and anti-inflammatory functions, which are good candidates for CLD treatment. Some of these products may have preventive effects on hepatic steatosis in ALD and NAFLD. For example, diallyl trisulfide (DATS) is a bioactive compound isolated from garlic and can reduce serum levels of aspartate transaminase (AST) and alanine aminotransferase (ALT) and decrease alcohol-induced liver injury [93]. DATS can upregulate PPAR-α expression and down-regulate sterol regulatory element binding protein 1c (SREBP-1c) expression to inhibit hepatic steatosis. Meanwhile, it can reduce liver oxidative stress by increasing antioxidant products and reducing ROS and malondialdehyde (MDA) production in the fatty liver[93]. In this section, we review some promising agents in ALD treatments either in animal models or clinical trials.

β-sitosterol

 β -sitosterol is isolated from the roots of *Panax ginseng*[94]. As a plant sterol, β -sitosterol can reduce alcohol-induced liver injury and oxidative stress via restoration of erythrocyte membrane fluidity,



upregulation of glutathione (GSH) activity, and reduction of MDA production. In addition, β -sitosterol can suppress apoptosis-related gene expression by increasing the phosphorylation of PI3K and AKT[95].

Curcumin

Curcumin is an orange-yellow component of turmeric or curry powder isolated from the rhizome of *Curcuma longa*[96,97]. Supplementation of curcumin can significantly increase the activities of superoxide dismutase (SOD), catalase, and glutathione peroxidase (GPx) to reduce swimming-induced oxidative stress in mice, by activating Nrf2 signaling pathway[98]. Treatment of curcumin significantly decreases serum levels of ALT, AST, alkaline phosphatase (ALP), gamma-glutamyl transferase, Arginase I, and blood urea nitrogen, while it increases serum levels of Albumin and total protein in ethanol-treated rats compared to the control group[99]. Development of self-assembled micelles of curcumin can be administered by oral delivery to enhance its anti-oxidative stress ability to prevent ALD and gastric mucosa damage[100]. Encapsulation enables to improve the adsorption of curcumin in intestinal epithelial cells and enhance its hepatoprotective effects in rats, *via* increasing the activity of GPx and decreasing high levels of MDA in the liver[101]. Furthermore, a combined treatment of curcumin and bacicalin shows more protective effects on ALD in rats by reducing liver oxidative damage through activation of the Nrf2/HO-1 signaling pathway[102].

Empagliflozin

Empagliflozin (EMPA) has benefits in cardiovascular, renal, and cerebral diseases, which is potentially mediated through its antioxidant and anti-inflammatory activities. Treatment with EMPA can decrease serum levels of ALT, AST, and ALP. It also increases the activities of GSH and SOD in the liver homogenates and decreases the liver content of MDA and nitric oxide (NO)[103]. Moreover, EMPA can downregulate NF- κ B signaling to suppress the expression of proinflammatory cytokines, including tumor necrosis factor-alpha (TNF- α), IL-1 β , and IL-6, which is associated with the upregulation of PPAR- γ , Nrf2, and their target gene HO-1[103].

Gastrodin

Gastrodin is the main bioactive component of *Gastrodia elata Blume* and displays anti-inflammatory and antioxidant properties. For example, administration of gastrodin (50 or 100 mg/kg) in mice significantly inhibits concanavalin A (ConA)-induced acute hepatitis, partly by suppressing IL-6/Janus Kinase 2/ signal transducer and activator of transcription 3 signaling pathway[104]. In addition, treatment with gastrodin ameliorated acetaminophen-induced liver injury in mice. The anti-inflammatory and anti-oxidative stress functions of gastrodin are mediated through the inhibition of signal-regulated kinase/JNK/mitogen-activated protein kinase signaling pathways and hepatic MDA activity, as well as activation of Nrf2 expression and SOD activity[105].

Genistein

Genistein is an isoflavone first isolated from the brooming plant Dyer's *Genista tinctoria*, which is widely distributed in the Fabaceae family[106-109]. Treatment of genistein at a dose of 0.3 mmol/kg of bodyweight can ameliorate liver fibrosis and apoptosis in mice by suppressing the expression of proinflammatory cytokines such as TNF- α , IL-6, profibrotic cytokines such as TGF- β 1, and cell caspase 3 [110]. In contrast, another study shows that supplementation of soy proteins significantly decreases serum ALT concentrations and hepatic TNF- α and CD-14 expression and decreases NF- κ B protein in casein-based 35% high-fat ethanol liquid diet (EtOH)-treated mice by inhibiting β -catenin signaling [111]. More functional studies of genistein have been performed in NAFLD models, which are discussed in the following section.

Lactoferrin

Lactoferrin (LF) is an iron-binding protein found at relatively high concentrations in mammalian milk [112]. LF displays multiple functions, including antioxidant, anti-cancer, and anti-inflammatory activities. For example, LF treatment can decrease the levels of liver superoxide and suppress liver inflammation in male mice with alcoholic-induced liver injury (ALI) by upregulating the expression of aldehyde dehydrogenase-2 and suppressing overexpression of cytochrome P450 2E1 (CYP2E1)[113]. LF treatment also displays a protective effect in female mice with acute ALI by regulating redox-stress response capacity[114]. The protective effect of LF on ALI is associated with the manipulation of gut microbiota and the modulation of hepatic alcohol metabolism[113].

Selenium

Selenium plays an essential role against oxidation, which is part of the catalytic center of different antioxidant selenoproteins including GPxs and selenoprotein P[115]. The serum levels of selenium are decreased in adult patients with acute and chronic alcoholic-related diseases, accompanied by liver damage and the severity of oxidation[115,116].

Silymarin

Silymarin is an active compound from the extracts of milk thistle (Silybum marianum)[117]. Silymarin displays antioxidant, antifibrotic, anti-inflammatory, and hepatoprotective properties in different types of CLD[118,119], such as ALD. Simultaneous supplementation of silymarin with alcohol treatment can reduce the ethanol-induced increase of serum ALT levels and hepatic microvesicular steatosis and TNF- α expression [120]. Another study on non-human primates also shows that silymarin can prevent the development of alcohol-induced liver fibrosis by decreasing the production of type I collagens[121].

Taraxasterol

Taraxasterol (TAS) is an active ingredient of Taraxacum officinale, which has protective effects on the liver and kidneys by reducing serum levels of ALT and AST, increasing serum and liver SOD and GPx, and maintaining the balance of ion homeostasis[122]. TAS also displays anti-inflammatory function in cultured mouse primary lymphocytes stimulated with Con A and in mice with Con A-induced acute hepatitis[123]. Mechanism studies reveal that TAS inhibits T cell activation and proliferation by suppressing IL-2/IL-2 receptor-mediated downstream signaling pathways[123].

Telmisartan

Telmisartan (TEL) exhibits similar effects with EMPA on ALD. Treatment of TEL (10 mg/kg/day) decreased serum levels of ALT, AST, and ALP in mice with ALD[124]. In addition, TEL displays antiinflammatory and antioxidant properties in mice with ALD by increasing the activity of SOD and GPx to reduce liver contents of NO and MDA, upregulating the expression of Nrf-2, PPAR-γ, and Hmox-1, and downregulating NF- κ B expression[124].

ANTIOXIDANT AND ANTI-INFLAMMATORY AGENTS IN NAFLD

Hepatic inflammation and oxidative stress are also associated with NAFLD pathogenesis[125]. Therefore, many above-discussed products also display similar bioactive functions against NAFLD.

β-sitosterol

Treatment with β-sitosterol can prevent high-fructose diet-induced macrovesicular hepatic steatosis and inhibit the progression of NAFL to NASH in male rats[126]. Meanwhile, it is also able to inhibit highfructose diet-induced visceral obesity, hypertriglyceridemia, plasma insulin concentration, and homeostatic model assessment of insulin resistance (HOMA-IR) but increase plasma levels of adiponectin in female rats[127]. Another study shows that in combination with stigmasterol, a dietary phytosterol, β-sitosterol can alleviate a high-fat western-style diet-induced NAFLD in mice post-17-wk treatment, by decreasing hepatic di- and tri-acylglycerols and circulating ceramide levels [128].

Curcumin

Curcumin is a natural polyphenol, which shows anti-inflammatory and antioxidant activities. It can improve insulin resistance and reduce hepatic fat accumulation in dietary obese rat models[129]. Accumulating evidence identifies that curcumin can attenuate hepatic steatosis by suppressing hepatic expression of CD36, PPAR-γ, SREBP-1c, and fatty acid synthase (FAS) in NAFLD mice, through upregulation of Nrf2 and FXR expression and downregulation of liver X receptor α expression[130,131]. In addition, curcumin can induce activation of AMPK and upregulation of PPAR-a, and suppress the highfat diet (HFD)-induced increase in the expression of SREBP-1, acetyl-CoA carboxylase 1, FAS, and CD36 [132]. Meanwhile, curcumin is able to prevent intestinal permeability and suppress LPS/TLR4/NF-κBmediated inflammatory response to protect against diet-induced hepatic steatosis and inflammation [133]. In addition, curcumin can also suppress NLRP3 inflammasome (Figure 2) and pro-IL-1 β synthesis by suppressing LPS-mediated activation of NF-KB signaling pathway[134].

Ex vivo studies also show that treatment of curcumin decreases linoleic acid-induced ROS production and leptin-induced TNF- α expression in human peripheral blood mononuclear cells[135]. A randomized controlled trial in Iran demonstrates that supplementation with curcumin in a phytosomal form (1000 mg/day) significantly reduces body mass index (BMI), waist circumference, and serum levels of AST and ALT[136]. This dose was safe and well tolerated in NAFLD patients[136]. Another double-blind, randomized, placebo-controlled trial displays that daily supplementation of low-dose phospholipid curcumin (250 mg) for 2 mo can significantly decrease hepatic steatosis and serum AST levels in NAFLD patients compared to placebo[137]. In addition, a combined therapy of curcumin (500 mg/day) with piperine, an alkaloid in black pepper with many pharmacological effects on chronic diseases[138], also decreases the severity of NAFLD and serum ALP levels[139]. Large clinical trials are needed for further evaluation of the efficacy of curcumin and its synergistic treatments.

EMPA

EMPA is an inhibitor of sodium-glucose co-transporter 2 (SGLT2), which plays an important role in



NAFLD. EMPA treatment can inhibit PA-induced lipid deposition in hepatocytes (HepG2 cells) and HFD-induced hepatic lipid accumulation and inflammation in mice by upregulating the expression of a stress-inducible protein Sestrin2 and activating AMPK-mTOR signaling pathway[140]. Another study demonstrates that EMPA can upregulate the expression of medium-chain acyl-CoA dehydrogenase in NASH liver and PA and glucose-treated hepatocytes by activating AMPK/forkhead box A2 signaling pathway, resulting in a reduction of hepatic lipid deposition *in vivo* and *in vitro*[141]. A meta-analysis shows that EMPA can significantly reduce BMI, HOMA-IR, AST, and liver fibrosis in patients with NAFLD[142].

In addition, other SGLT2 inhibitors or gliflozins, such as licogliflozin[143,144] and dapagliflozin[145, 146], also can control glycemic production and bodyweight, normalize serum ALT levels, and reduce Fibrosis-4 NAFLD patients with T2DM.

Gastrodin

Gastrodin has been shown to significantly decrease lipid accumulation and inflammatory response in primary mice and human hepatocytes treated with 0.5 mmol/L PA along with 1.0 mmol/L oleic acid. In addition, it ameliorates diet-induced hepatic steatosis and inflammation in mice by activating the AMPK signaling pathway^[147]. Gastrodin can also regulate lipid metabolism and display antioxidant effects in larval zebrafish with high-cholesterol diet-induced NAFLD[148].

Genistein

Genistein has been shown to play an important role in NAFLD and NASH treatment. Treatment of genistein reduces the levels of $TNF-\alpha$ and reduces TLR4 mRNA and protein expression and inflammation in the livers of rats with NASH[149]. A combination of genistein with metformin (0.2% + 0.23%) for 3 mo shows a synergistic effect on the reduction of AST, ALT, and TG, liver TG and number of macrophages, and NAFLD activity score (NAS) in HFD-fed mice[150]. The reduction of hepatic steatosis is associated with decreased mRNA levels of lipogenic-related genes SREBP-1c and FAS and upregulated mRNA expression of FAO-related gene carnitine palmitoyl transferase 1[150]. Genistein treatment (16 mg/kg BW/day) for 5 wk can significantly decrease hepatic steatosis, inflammation, and hepatocyte ballooning in ovariectomized rats with high-fat and high-fructose diet-induced NASH[151].

Consumption of dietary isoflavones including genistein is reversely associated with NAFLD, hypertension, and hyperlipidemia in a study on Chinese adults[152]. Molecular mechanism studies show that genistein can suppress the activation of SREBP-1c in FFA-induced fat accumulation in primary human hepatocytes, whereas genistein-mediated upregulation of PPAR- α proteins in normal hepatocytes is abolished in steatotic hepatocytes[153].

LF

LF is an iron-binding protein in mammalian milk and displays multiple functions, including antioxidant, anti-cancer, and anti-inflammatory activities. During NASH progression, LF treatment can inhibit NF-KB activation to downregulate a high-fat diet and chemical dimethylnitrosamine-induced liver injury, inflammation, and fibrosis[154]. Treatment with LF improves insulin sensitivity and reduces hepatic steatosis in ob/ob mice by downregulating SREBP-2. It also regulates hepatocellular iron transport by controlling the hepcidin-ferroportin axis to maintain liver oxidative balance and suppress hepatocyte death[155].

Mastiha

Mastiha is a natural and aromatic resin isolated from the trunk and brunches of mastic trees with antioxidant and anti-inflammatory properties [156]. Mice with diet-induced NASH fed with 0.2% (w/w) Mastiha supplementation for 8 wk can reduce the circulating ALT levels, NAS, hepatic steatosis, and liver collagen production[157]. This study also identifies that Mastiha supplementation changes NASHinduced gut microbiota profile to the diversity and composition of healthy mice. A randomized clinical trial (NCT03135873, www.clinicaltrials.gov) shows that supplementation of Mastiha improves the total antioxidant status (TAS) levels in NAFLD patients with severe obesity compared to that in the corresponding placebo group[158]. The anti-inflammatory function of Mastiha is associated with the expression of microRNA-155 in the plasma of NAFLD patients, which may regulate the differentiation and function of T helper-17 cells[159].

Selenium

Treatment with selenium-enriched green tea extract (200 mg/kg body weight) for 15 wk can significantly reduce body weight gain and visceral fat accumulation in mice with obesity and NAFLD [160]. Reduced serum levels of selenium are independently associated with hepatic fibrosis in NAFLD patients[161]. Another study reveals that selenium deficiency induces hepatic inflammation in pigs by activating the NF-kB signaling pathway, decreasing antioxidant capacity, and increasing ROS levels [162]. Selenium-enriched Lactobacillus acidophilus SNZ 86 (probiotic) can decrease western-style dietinduced hepatic steatosis in mice with NAFLD, by activating autophagy through the upregulation of AMPK/SIRT1 signaling pathway[163]. Co-supplementation of selenium with vitamin B6 can reduce



Zhang CY et al. Natural products in liver disease



Figure 3 Structures of peroxisome proliferator-activated receptor agonists or modulators applied for the treatment of chronic liver disease. Many peroxisome proliferator-activated receptor regulators have been evaluated in the clinic, showing promising effects in patients with chronic liver disease. All the chemical structures were collected online from the Chemical Book (https://www.chemicalbook.com, accessed on August 10, 2022). PPAR: Peroxisome proliferator-activated receptor.

liver lipid synthesis and deposition by increasing the expression of SIRT1 to downregulate SREBP-1c expression (Figure 2) and upregulate PPAR- α expression in HFD-fed rats[164].

Silymarin

The major active compound of silymarin is silybin. Treatment with silybin can significantly decrease lipid accumulation in mice with NAFLD by activating PPAR- α [165]. Since it can partially inhibit the effect of PPAR- α agonist fenofibrate, it is not suggested to be simultaneously applied with PPAR- α agonists. Silymarin also displays a synergistic effect with quercetin on the reduction of lipid accumulation in rat hepatocytes[166]. Silymarin treatment significantly ameliorates high fructose-induced oxidative stress and hepatic steatosis in rats[167]. Silymarin supplementation (560 mg daily) for 8 wk significantly improves serum AST/ALT ratio, ultrasound fatty liver grading, and BMI in patients with morbid obesity and NAFLD[168].

TEL

Treatment with TEL significantly improves fibrosis scores and reduces the levels of serum leptin and its expression in liver tissue[169]. As an angiotensin receptor blocker, it significantly decreases fasting serum-FFA levels and triglyceride-glucose index in patients with NAFLD[170]. TEL displays a similar effect as vitamin E on the reduction of NAS, and improvement of hepatic steatosis, but it has a better effect on the reduction of liver lobular inflammation and hepatocyte ballooning[171]. It can function as a PPAR- γ/α dual agonist to simultaneously improve insulin-sensitivity *via* activating PPAR- γ and improve lipid metabolism by activating PPAR- α [172].

Delta-tocotrienol

Tocotrienols are natural compounds that belong to one part of two vitamin E components (Tocopherols as another part), including α , β , γ , and δ tocotrienols[173]. Among them, δ -tocotrienol shows strongly anti-inflammatory activity, which can decrease insulin resistance, hepatic steatosis, and serum trigly-ceride concentrations in rats with diet-induced obesity[174]. Recent studies also show that δ -tocotrienol has anti-cancer properties by regulating angiogenesis and cell proliferation and apoptosis[175].

A human study indicates that oral supplementation of δ -tocotrienol (300 mg, twice daily) for 12 wk significantly decreases serum aminotransferases, high sensitivity C-reactive protein (hs-CRP), and MDA, and fatty liver index (FLI) score compared to placebo[176]. Clinical trials reveal that δ -tocotrienol supplementation results in a significant reduction in plasma glucose, insulin, glycosylated hemoglobin, MDA, high sensitive C-reactive protein, and proinflammatory cytokines (TNF- α and IL-6), and HOMA-IR in pre-diabetic and diabetic patients[177,178]. Another trial also demonstrates that treatment of δ -tocotrienol (300 mg, twice daily) for 24 wk further significantly reduces FLI score, HOMA-IR, and hepatic steatosis than placebo, except decreased serum levels of hs-CRP, MDA, ALT, and AST, without causing adverse events[179].

ANTIOXIDANT AND ANTI-INFLAMMATORY AGENTS IN LIVER CANCER

Both ALD and NAFLD are major contributors to HCC initiation and progression. Therefore, the abovediscussed biomolecules may also exhibit anti-HCC effects. For example, treatment of β-sitosterol niosomes, a form of β -sitosterol with polyethylene glycol modification, shows cytotoxicity to HepG2 cells due to increased cellular uptake and displays in vivo anti-HCC ability in Wistar albino rats[180]. Treatment of β -sitosterol-assisted silver nanoparticles (BSS-SNPs) significantly inhibits the proliferation of HepG2 cells and their production of ROS and Nrf2, resulting in the regulation of pro-apoptotic genes such as Bcl-2 Associated X-protein and caspases 3 and 9[181]. Similarly, compounds including curcumin [182], EMPA[183], gastrodin[184], genistein[185], LF[186], selenium[187], silymarin[188], TAS[189], TEL [190], and delta-tocotrienol[191] display anti-HCC effects either *in vitro* or *in vivo*, or both (Table 1).

CLINICAL TRIALS

Clinical trials have been started to evaluate the efficacy of these molecules in CLD (Table 2), such as EMPA[192] and silymarin[193,194]. For example, treatment with EMPA can improve liver steatosis in patients with NAFLD without T2DM[192]. Another trial shows that oral supplementation of genistein (250 mg) for 8 wk can decrease insulin resistance, oxidative stress, and inflammation and improve lipid metabolism in patients with NAFLD[195].

CONCLUSION

CLD is a continuous process that causes a reduction of liver function that lasts more than six months. CLD has a broad spectrum with complex cellular and molecular mechanisms. It can be subclassified into ALD, NAFLD or MAFLD, chronic viral infection, and autoimmune hepatitis, which can lead to liver fibrosis, cirrhosis, and cancer. However, there are no currently available treatments for ALD, NAFLD, and liver fibrosis, except the preventive strategies, such as changes in exercise, diet, and alcohol use. Early preventive strategies predict good outcomes. Patients with advanced ALD and NAFLD require liver transplantation, but without enough donor organs. Liver inflammation and oxidative stress are ubiquitously associated with the development and progression of CLD. Molecular signaling pathways such as AMPK, JNK, and PPAR-mediated signaling pathways are implicated in liver inflammation, oxidative stress, and lipid metabolism. Accumulating studies have demonstrated that natural products with antioxidant and anti-inflammatory functions display therapeutic effects against inflammation, fibrosis, and metabolic disorders, including ALD and NAFLD. These products such as β -sitosterol, curcumin, EMPA, gastrodin, and genistein have shown potential application at all the stages of CLD, from ALD/NAFLD to HCC. In addition, clinical trials that are undergoing to evaluate their efficacy and safety are reviewed. Overall, pre-clinical studies in cell and animal models reveal the protective effects of these agents in CLD. However, more clinical trials are required to evaluate their efficacy and safety.

Natural products, especially antioxidant and anti-inflammatory products, show potent therapeutic alternatives for CLD treatment with their efficacy and low side effects. Remarkably, these products also display anti-HCC functions. However, many pharmaceutical dynamic assays have not been tested, and the potential adverse effects of long-term use of these products are not available. In the future, the synergistic effects of different drugs should be evaluated to treat CLD, due to its complex pathogenic factors.

Molecules	Model	Function	Ref.
β-sitosterol	HepG2 cells; Rat HCC	Treatment of β -sitosterol niosomes displays direct cytotoxicity to HepG2 cells in vitro and anti-HCC ability in rats	[1 <mark>82</mark>]
Curcumin	HepG2 and SK-Hep-1 cells. A nude mouse xenograft model bearing HepG2 cells	It can inhibit cell proliferation and increase cell apoptosis and cell cycle arrest at the G0/G1 phase of cancer cells by downregulating the expression of BCLAF1 and inhibiting the activation of the PI3K/AKT/GSK-3 β pathway	[183]
Empagliflozin	DENA-induced HCC in mice	It shows a synergistic effect on the control of angiogenesis, invasion, and metastasis of tumor cells in mice with DENA-induced HCC by inhibiting the expression of MAPKs and reducing liver injury enzymes	[184]
Gastrodin	Subcutaneous H22 cells-induced tumor	It can specifically increase the expression of NF-ĸB	[185]

Table 1 Antioxidant and anti-inflammatory agents for the treatment of hepatocellular carcinoma



Zhang CY et al. Natural products in liver disease

	in mice	downstream genes such as Bcl-xL, Bcl-2, and IL-2 in CD4 but not CD8 T cells $% \left(\mathcal{L}^{2}\right) =0$	
Genistein	TAA-induced HCC in rats	It displays antioxidant and anti-HCC effects by suppressing the versican/PDGF bidirectional axis and protein expression of PKC and ERK-1	[186]
Lactoferrin	DEN-induced HCC in rats	It shows a chemopreventive effect against DEN-induced HCC in rats in a dose-dependent manner by suppressing the expression and activation of AKT	[187]
Selenium	TAA-induced HCC in rats	Selenium nanoparticles improve the tumor suppressive effect of sorafenib and overcome drug resistance in rat HCC by inducing apoptosis and targeting AKT/mTOR and NF- κ B signaling pathways, as well as epigenetic regulation	[188]
Silymarin	$\text{DEN}/\text{AAF}/\text{CCl}_4$ induced HCC in rats	It suppresses cancer cell growth in rats with DEN/AAF/CCl ₄ -induced tumors by inhibiting the expression of Ki-67 and HGF/c-Met, Wnt/ β -catenin, and Pl3K/Akt/mTOR signaling pathways	[189]
Taraxasterol	HepG2 and Huh7H22 bearing mice	It can suppress tumor cell growth by suppressing Ki67 expression and inducing cell apoptosis <i>via</i> suppressing IL-6/STAT3 signaling pathway, as well as promoting T cell infiltration in tumor tissue	[190]
Telmisartan	NDEA-induced HCC in mice	It exerts an anti-HCC effect and increases tumor cell sensitivity to sorafenib treatment by suppressing phosphorylation-induced activation of TAK1 and the ERK1/2 and NF-κB signaling pathways	[<mark>191</mark>]
Delta-tocotrienol	HCC cell lines SK Hep-1 and Huh7	It promotes the anti-HCC cell activity of IFN- α by increasing ROS and increasing cell apoptosis together with an increased Bax/Bcl-xL ratio. In addition, it can activate Notch1 signaling pathway	[192]

AKT: Protein kinase B; Bax: Bcl-2-like protein 4; Bcl-2: B-cell lymphoma 2; Bcl-xL: B-cell lymphoma extra-large; BCLAF1: BCL-2-associated transcription factor 1; CD4: Cluster of differentiation 4; c-Met: Tyrosine-protein kinase Met; ERK-1/2: Extracellular signal-regulated kinases 1/2; GSK-3β: Glycogen synthase kinase-3β; HCC: Hepatocellular carcinoma; HGF: Hepatocyte growth factor; IL-2: Interleukin 2; Ki-67: Marker of proliferation Ki-67; MAPK: Mitogen-activated protein kinase; mTOR: Mammalian target of rapamycin; NF-KB: Nuclear factor KB; PI3K: Phosphatidylinositol-3-kinase; PDGF: Plateletderived growth factor; SIRT1: Sirtuin 1; SREBP-1c: Sterol regulatory element binding protein 1c; STAT3: Signal transducer and activator of transcription 3; DENA Diethylnitrosamine; TAA: Thioacetamide; ROS: Reactive oxygen species; NDEA: N-Nitrosodiethylamine; AAF: 2-acetylaminofluorene; CCl₄: Carbon tetrachloride.

Table 2 Clinical trials for evaluating the efficacy of compounds in liver disease			
Treatment	Trial number	Phase	Aims or results
Curcumin	NCT02908152	2-3	To investigate the effects of curcumin supplements on metabolic factors and hepatic fibrosis in NAFLD patients with T2DM
	NCT04109742	2	To test the effect of curcumin in pediatric patients with NAFLD
Empagliflozin	NCT03867487	2	To evaluate the preliminary feasibility, initial efficacy, and safety of empagliflozin as a SGLT2 inhibitor for treating NAFLD in adolescents with obesity
	NCT04642261	4	To test the effects of empagliflozin on reducing hepatic fat content as measured by MRI-PDFF in NAFLD patients without DM
Gastrodin	NCT04035824	4	To treat hypertension together with Uncaria
Genistein	IRCT201312132480N5	3	Oral supplementation of genistein (250 mg) for 8 wk can decrease insulin resistance, oxidative stress, and inflammation and improve lipid metabolism in patients with NAFLD
Lactoferrin	NCT04335058	None	To test the effect of lactoferrin with iron versus iron alone in the treatment of anemia in CLD
Selenium	NCT00271245	None	To test the effect of selenium in patients with cirrhosis
	NCT01650181	4	To test the impacts using siliphos-selenium-methionine-alpha lipoic acid plus metformin versus metformin in patients with fatty liver and NASH
Silymarin	NCT00389376	1	An increase in silymarin is observed in NAFLD patients, compared to that in patients with HCV
	NCT00680407	2	The effect of silymarin on NASH patients remains inconclusive due to the lack of a



			substantial number of patients
Telmisartan	NCT02213224	4	To evaluate the therapeutic effects of telmisartan and perindopril for NAFLD patients with hypertension

T2DM: Type 2 diabetes mellitus; NAFLD: Non-alcoholic fatty liver disease; SGLT2: Sodium-glucose cotransporter-2; MRI-PDFF: Magnetic resonance imaging-derived proton density fat fraction; CLD: Chronic liver disease; NASH: Non-alcoholic steatohepatitis; HCV: Hepatitis C virus.

FOOTNOTES

Author contributions: Zhang CY, Liu S, and Yang M designed, collected data, wrote, revised, and finalized the manuscript, contributed equally, and shared the first authorship.

Conflict-of-interest statement: All the authors report no relevant conflicts of interest for this article.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is noncommercial. See: https://creativecommons.org/Licenses/by-nc/4.0/

Country/Territory of origin: United States

ORCID number: Chun-Ye Zhang 0000-0003-2567-029X; Shuai Liu 0000-0001-9695-2492; Ming Yang 0000-0002-4895-5864.

S-Editor: Fan JR L-Editor: A P-Editor: Fan JR

REFERENCES

- 1 Sharma A, Nagalli S. Chronic Liver Disease. StatPearls Publishing LLC., 2022. [cited 10 December 2022]. Available from: https://www.ncbi.nlm.nih.gov/books/NBK554597
- Embade N, Millet O. Molecular Determinants of Chronic Liver Disease as Studied by NMR-Metabolomics. Curr Top 2 Med Chem 2017; 17: 2752-2766 [PMID: 28685692 DOI: 10.2174/1568026617666170707124539]
- 3 Seitz HK, Bataller R, Cortez-Pinto H, Gao B, Gual A, Lackner C, Mathurin P, Mueller S, Szabo G, Tsukamoto H. Alcoholic liver disease. Nat Rev Dis Primers 2018; 4: 16 [PMID: 30115921 DOI: 10.1038/s41572-018-0014-7]
- Mantovani A, Beatrice G, Dalbeni A. Coronavirus disease 2019 and prevalence of chronic liver disease: A meta-analysis. 4 Liver Int 2020; 40: 1316-1320 [PMID: 32329563 DOI: 10.1111/liv.14465]
- Sharma P, Arora A. Clinical presentation of alcoholic liver disease and non-alcoholic fatty liver disease: spectrum and 5 diagnosis. Transl Gastroenterol Hepatol 2020; 5: 19 [PMID: 32258523 DOI: 10.21037/tgh.2019.10.02]
- 6 Wang WJ, Xiao P, Xu HO, Niu JO, Gao YH. Growing burden of alcoholic liver disease in China: A review. World J Gastroenterol 2019; 25: 1445-1456 [PMID: 30948908 DOI: 10.3748/wjg.v25.i12.1445]
- 7 Axley PD, Richardson CT, Singal AK. Epidemiology of Alcohol Consumption and Societal Burden of Alcoholism and Alcoholic Liver Disease. Clin Liver Dis 2019; 23: 39-50 [PMID: 30454831 DOI: 10.1016/j.cld.2018.09.011]
- 8 Ferdouse A, Clugston RD. Pathogenesis of Alcohol-Associated Fatty Liver: Lessons From Transgenic Mice. Front Physiol 2022; 13: 940974 [PMID: 35864895 DOI: 10.3389/fphys.2022.940974]
- 9 Nagy LE, Ding WX, Cresci G, Saikia P, Shah VH. Linking Pathogenic Mechanisms of Alcoholic Liver Disease With Clinical Phenotypes. Gastroenterology 2016; 150: 1756-1768 [PMID: 26919968 DOI: 10.1053/j.gastro.2016.02.035]
- Zhao X, Wang C, Dai S, Liu Y, Zhang F, Peng C, Li Y. Quercetin Protects Ethanol-Induced Hepatocyte Pyroptosis via 10 Scavenging Mitochondrial ROS and Promoting PGC-1α-Regulated Mitochondrial Homeostasis in L02 Cells. Oxid Med Cell Longev 2022; 2022: 4591134 [PMID: 35879991 DOI: 10.1155/2022/4591134]
- 11 Patel F, Parwani K, Patel D, Mandal P. Metformin and Probiotics Interplay in Amelioration of Ethanol-Induced Oxidative Stress and Inflammatory Response in an In Vitro and In Vivo Model of Hepatic Injury. Mediators Inflamm 2021; 2021: 6636152 [PMID: 33953643 DOI: 10.1155/2021/6636152]
- 12 Zhang C, Yang M. Current Options and Future Directions for NAFLD and NASH Treatment. Int J Mol Sci 2021; 22 [PMID: 34299189 DOI: 10.3390/ijms22147571]
- Chen YH, Wu WK, Wu MS. Microbiota-Associated Therapy for Non-Alcoholic Steatohepatitis-Induced Liver Cancer: A 13 Review. Int J Mol Sci 2020; 21 [PMID: 32825440 DOI: 10.3390/ijms21175999]
- Le MH, Yeo YH, Li X, Li J, Zou B, Wu Y, Ye Q, Huang DQ, Zhao C, Zhang J, Liu C, Chang N, Xing F, Yan S, Wan 14 ZH, Tang NSY, Mayumi M, Liu X, Rui F, Yang H, Yang Y, Jin R, Le RHX, Xu Y, Le DM, Barnett S, Stave CD, Cheung R, Zhu Q, Nguyen MH. 2019 Global NAFLD Prevalence: A Systematic Review and Meta-analysis. Clin Gastroenterol Hepatol 2022; 20: 2809-2817.e28 [PMID: 34890795 DOI: 10.1016/j.cgh.2021.12.002]
- 15 Riazi K, Azhari H, Charette JH, Underwood FE, King JA, Afshar EE, Swain MG, Congly SE, Kaplan GG, Shaheen AA.



The prevalence and incidence of NAFLD worldwide: a systematic review and meta-analysis. Lancet Gastroenterol Hepatol 2022; 7: 851-861 [PMID: 35798021 DOI: 10.1016/S2468-1253(22)00165-0]

- 16 Atsawarungruangkit A, Laoveeravat P, Promrat K. Machine learning models for predicting non-alcoholic fatty liver disease in the general United States population: NHANES database. World J Hepatol 2021; 13: 1417-1427 [PMID: 34786176 DOI: 10.4254/wjh.v13.i10.1417]
- 17 Perdomo CM, Garcia-Fernandez N, Escalada J. Diabetic Kidney Disease, Cardiovascular Disease and Non-Alcoholic Fatty Liver Disease: A New Triumvirate? J Clin Med 2021; 10 [PMID: 34068699 DOI: 10.3390/jcm10092040]
- 18 Bonora E, Targher G. Increased risk of cardiovascular disease and chronic kidney disease in NAFLD. Nat Rev Gastroenterol Hepatol 2012; 9: 372-381 [PMID: 22565095 DOI: 10.1038/nrgastro.2012.79]
- 19 Eslam M, Newsome PN, Sarin SK, Anstee QM, Targher G, Romero-Gomez M, Zelber-Sagi S, Wai-Sun Wong V, Dufour JF, Schattenberg JM, Kawaguchi T, Arrese M, Valenti L, Shiha G, Tiribelli C, Yki-Järvinen H, Fan JG, Grønbæk H, Yilmaz Y, Cortez-Pinto H, Oliveira CP, Bedossa P, Adams LA, Zheng MH, Fouad Y, Chan WK, Mendez-Sanchez N, Ahn SH, Castera L, Bugianesi E, Ratziu V, George J. A new definition for metabolic dysfunction-associated fatty liver disease: An international expert consensus statement. J Hepatol 2020; 73: 202-209 [PMID: 32278004 DOI: 10.1016/j.jhep.2020.03.039]
- 20 Eslam M, Sanyal AJ, George J; International Consensus Panel. MAFLD: A Consensus-Driven Proposed Nomenclature for Metabolic Associated Fatty Liver Disease. Gastroenterology 2020; 158: 1999-2014.e1 [PMID: 32044314 DOI: 10.1053/i.gastro.2019.11.312
- Pohl R, Feder S, Haberl EM, Rein-Fischboeck L, Weiss TS, Spirk M, Bruckmann A, McMullen N, Sinal CJ, Buechler C. 21 Chemerin Overexpression in the Liver Protects against Inflammation in Experimental Non-Alcoholic Steatohepatitis. Biomedicines 2022; 10 [PMID: 35052810 DOI: 10.3390/biomedicines10010132]
- 22 Gabbia D, Cannella L, De Martin S. The Role of Oxidative Stress in NAFLD-NASH-HCC Transition-Focus on NADPH Oxidases. Biomedicines 2021; 9 [PMID: 34204571 DOI: 10.3390/biomedicines9060687]
- Mathur M, Yeh YT, Arya RK, Jiang L, Pornour M, Chen W, Ma Y, Gao B, He L, Ying Z, Xue B, Shi H, Choi Y, Yu L. 23 Adipose lipolysis is important for ethanol to induce fatty liver in the National Institute on Alcohol Abuse and Alcoholism murine model of chronic and binge ethanol feeding. Hepatology 2022 [PMID: 35844150 DOI: 10.1002/hep.32675]
- Dallio M, Sangineto M, Romeo M, Villani R, Romano AD, Loguercio C, Serviddio G, Federico A. Immunity as 24 Cornerstone of Non-Alcoholic Fatty Liver Disease: The Contribution of Oxidative Stress in the Disease Progression. Int J Mol Sci 2021; 22 [PMID: 33406763 DOI: 10.3390/ijms22010436]
- Gu Y, Lian Y, Zheng Q, Huang Z, Gu L, Bi Y, Li J, Huang Y, Wu Y, Chen L. Association among cytokine profiles of 25 innate and adaptive immune responses and clinical-virological features in untreated patients with chronic hepatitis B. BMC Infect Dis 2020; 20: 509 [PMID: 32664850 DOI: 10.1186/s12879-020-05233-x]
- 26 Petagine L, Zariwala MG, Patel VB. Alcoholic liver disease: Current insights into cellular mechanisms. World J Biol Chem 2021; 12: 87-103 [PMID: 34630912 DOI: 10.4331/wjbc.v12.i5.87]
- 27 Lai Y, Tan Q, Xv S, Huang S, Wang Y, Li Y, Zeng T, Mo C, Chen Y, Zhou C, Gao L, Lv Z. Ginsenoside Rb1 Alleviates Alcohol-Induced Liver Injury by Inhibiting Steatosis, Oxidative Stress, and Inflammation. Front Pharmacol 2021; 12: 616409 [PMID: 33716743 DOI: 10.3389/fphar.2021.616409]
- Monserrat-Mesquida M, Quetglas-Llabrés M, Bouzas C, Montemayor S, Mascaró CM, Casares M, Llompart I, Gámez 28 JM, Tejada S, Martínez JA, Tur JA, Sureda A. A Greater Improvement of Intrahepatic Fat Contents after 6 Months of Lifestyle Intervention Is Related to a Better Oxidative Stress and Inflammatory Status in Non-Alcoholic Fatty Liver Disease. Antioxidants (Basel) 2022; 11 [PMID: 35883758 DOI: 10.3390/antiox11071266]
- 29 Franco I, Bianco A, Mirizzi A, Campanella A, Bonfiglio C, Sorino P, Notarnicola M, Tutino V, Cozzolongo R, Giannuzzi V, Aballay LR, Buongiorno C, Bruno I, Osella AR. Physical Activity and Low Glycemic Index Mediterranean Diet: Main and Modification Effects on NAFLD Score. Results from a Randomized Clinical Trial. Nutrients 2020; 13 [PMID: 33379253 DOI: 10.3390/nu13010066]
- Zabaleta N, Torella L, Weber ND, Gonzalez-Aseguinolaza G. mRNA and gene editing: Late breaking therapies in liver 30 diseases. Hepatology 2022; 76: 869-887 [PMID: 35243655 DOI: 10.1002/hep.32441]
- 31 Aravalli RN, Steer CJ. CRISPR/Cas9 therapeutics for liver diseases. J Cell Biochem 2018; 119: 4265-4278 [PMID: 29266637 DOI: 10.1002/jcb.26627]
- 32 Harrison SA, Neff G, Guy CD, Bashir MR, Paredes AH, Frias JP, Younes Z, Trotter JF, Gunn NT, Moussa SE, Kohli A, Nelson K, Gottwald M, Chang WCG, Yan AZ, DePaoli AM, Ling L, Lieu HD. Efficacy and Safety of Aldafermin, an Engineered FGF19 Analog, in a Randomized, Double-Blind, Placebo-Controlled Trial of Patients With Nonalcoholic Steatohepatitis. Gastroenterology 2021; 160: 219-231.e1 [PMID: 32781086 DOI: 10.1053/j.gastro.2020.08.004]
- 33 Xu JJ, Li HD, Wu MF, Zhu L, Du XS, Li JJ, Li Z, Meng XM, Huang C, Li J. 3-B-RUT, a derivative of RUT, protected against alcohol-induced liver injury by attenuating inflammation and oxidative stress. Int Immunopharmacol 2021; 95: 107471 [PMID: 33756231 DOI: 10.1016/j.intimp.2021.107471]
- 34 Yue SR, Tan YY, Zhang L, Zhang BJ, Jiang FY, Ji G, Liu BC, Wang RR. Gynostemma pentaphyllum polysaccharides ameliorate non-alcoholic steatohepatitis in mice associated with gut microbiota and the TLR2/NLRP3 pathway. Front Endocrinol (Lausanne) 2022; 13: 885039 [PMID: 35937847 DOI: 10.3389/fendo.2022.885039]
- Ai G, Wu X, Dou Y, Huang R, Zhong L, Liu Y, Xian Y, Lin Z, Li Y, Su Z, Chen J, Qu C. Oxyberberine, a novel HO-1 35 agonist, effectively ameliorates oxidative stress and inflammatory response in LPS/D-GalN induced acute liver injury mice via coactivating erythrocyte metabolism and Nrf2 signaling pathway. Food Chem Toxicol 2022; 166: 113215 [PMID: 35691465 DOI: 10.1016/j.fct.2022.113215]
- 36 Perez-Matos MC, Sandhu B, Bonder A, Jiang ZG. Lipoprotein metabolism in liver diseases. Curr Opin Lipidol 2019; 30: 30-36 [PMID: 30550414 DOI: 10.1097/MOL.00000000000569]
- 37 Heeren J, Scheja L. Metabolic-associated fatty liver disease and lipoprotein metabolism. Mol Metab 2021; 50: 101238 [PMID: 33892169 DOI: 10.1016/j.molmet.2021.101238]
- 38 Zhao Z, Deng ZT, Huang S, Ning M, Feng Y, Shen Y, Zhao QS, Leng Y. Alisol B Alleviates Hepatocyte Lipid



Accumulation and Lipotoxicity via Regulating RARa-PPARy-CD36 Cascade and Attenuates Non-Alcoholic Steatohepatitis in Mice. Nutrients 2022; 14 [PMID: 35745142 DOI: 10.3390/nu14122411]

- 39 Xu Y, Denning KL, Lu Y. PPARα agonist WY-14,643 induces adipose atrophy and fails to blunt chronic ethanol-induced hepatic fat accumulation in mice lacking adipose FGFR1. Biochem Pharmacol 2021; 192: 114678 [PMID: 34265279 DOI: 10.1016/j.bcp.2021.114678]
- Pham TH, Lee GH, Jin SW, Lee SY, Han EH, Kim ND, Jeong HG. Puerarin attenuates hepatic steatosis via G-protein-40 coupled estrogen receptor-mediated calcium and SIRT1 signaling pathways. Phytother Res 2022; 36: 3601-3618 [PMID: 35871535 DOI: 10.1002/ptr.7526]
- Xie F, Zhong Y, Wang D, So KF, Xiao J, Lv Y. Metformin protects against ethanol-induced liver triglyceride 41 accumulation by the LKB1/AMPK/ACC pathway. Mol Biol Rep 2022; 49: 7837-7848 [PMID: 35733070 DOI: 10.1007/s11033-022-07610-y]
- Bai T, Yang Y, Yao YL, Sun P, Lian LH, Wu YL, Nan JX. Betulin alleviated ethanol-induced alcoholic liver injury via 42 SIRT1/AMPK signaling pathway. Pharmacol Res 2016; 105: 1-12 [PMID: 26776965 DOI: 10.1016/j.phrs.2015.12.022]
- 43 Han H, Xue T, Li J, Guo Y, Li X, Wang L, Pei L, Zheng M. Plant sterol ester of α-linolenic acid improved non-alcoholic fatty liver disease by attenuating endoplasmic reticulum stress-triggered apoptosis via activation of the AMPK. J Nutr Biochem 2022; 107: 109072 [PMID: 35660097 DOI: 10.1016/j.jnutbio.2022.109072]
- Gluais-Dagorn P, Foretz M, Steinberg GR, Batchuluun B, Zawistowska-Deniziak A, Lambooij JM, Guigas B, Carling D, 44 Monternier PA, Moller DE, Bolze S, Hallakou-Bozec S. Direct AMPK Activation Corrects NASH in Rodents Through Metabolic Effects and Direct Action on Inflammation and Fibrogenesis. Hepatol Commun 2022; 6: 101-119 [PMID: 34494384 DOI: 10.1002/hep4.1799]
- Jiang Y, Xu J, Huang P, Yang L, Liu Y, Li Y, Wang J, Song H, Zheng P. Scoparone Improves Nonalcoholic 45 Steatohepatitis Through Alleviating JNK/Sab Signaling Pathway-Mediated Mitochondrial Dysfunction. Front Pharmacol 2022; 13: 863756 [PMID: 35592421 DOI: 10.3389/fphar.2022.863756]
- Zhu JY, Chen M, Mu WJ, Luo HY, Guo L. Higd1a facilitates exercise-mediated alleviation of fatty liver in diet-induced 46 obese mice. Metabolism 2022; 134: 155241 [PMID: 35750235 DOI: 10.1016/j.metabol.2022.155241]
- 47 Panzitt K, Wagner M. FXR in liver physiology: Multiple faces to regulate liver metabolism. Biochim Biophys Acta Mol Basis Dis 2021; 1867: 166133 [PMID: 33771667 DOI: 10.1016/j.bbadis.2021.166133]
- Jiao Y, Lu Y, Li XY. Farnesoid X receptor: a master regulator of hepatic triglyceride and glucose homeostasis. Acta 48 Pharmacol Sin 2015; 36: 44-50 [PMID: 25500875 DOI: 10.1038/aps.2014.116]
- Cheng Y, Xiang X, Liu C, Cai T, Li T, Chen Y, Bai J, Shi H, Zheng T, Huang M, Fu W. Transcriptomic Analysis Reveals 49 Lactobacillus reuteri Alleviating Alcohol-Induced Liver Injury in Mice by Enhancing the Farnesoid X Receptor Signaling Pathway. J Agric Food Chem 2022; 70: 12550-12564 [PMID: 36154116 DOI: 10.1021/acs.jafc.2c05591]
- Liu Y, Kang W, Liu S, Li J, Liu J, Chen X, Gan F, Huang K. Gut microbiota-bile acid-intestinal Farnesoid X receptor 50 signaling axis orchestrates cadmium-induced liver injury. Sci Total Environ 2022; 849: 157861 [PMID: 35934034 DOI: 10.1016/j.scitotenv.2022.157861]
- Hartmann P, Hochrath K, Horvath A, Chen P, Seebauer CT, Llorente C, Wang L, Alnouti Y, Fouts DE, Stärkel P, 51 Loomba R, Coulter S, Liddle C, Yu RT, Ling L, Rossi SJ, DePaoli AM, Downes M, Evans RM, Brenner DA, Schnabl B. Modulation of the intestinal bile acid/farnesoid X receptor/fibroblast growth factor 15 axis improves alcoholic liver disease in mice. Hepatology 2018; 67: 2150-2166 [PMID: 29159825 DOI: 10.1002/hep.29676]
- 52 Ratziu V, Harrison SA, Loustaud-Ratti V, Bureau C, Lawitz E, Abdelmalek M, Alkhouri N, Francque S, Girma H, Darteil R, Couchoux H, Wolf M, Sanyal A, Vonderscher J, Scalfaro P. Hepatic and renal improvements with FXR agonist vonafexor in individuals with suspected fibrotic NASH. J Hepatol 2022 [PMID: 36334688 DOI: 10.1016/j.jhep.2022.10.023
- 53 Wang X, Yu H, Xing R, Li P. Hepatoprotective Effect of Oyster Peptide on Alcohol-Induced Liver Disease in Mice. Int J Mol Sci 2022; 23 [PMID: 35897657 DOI: 10.3390/ijms23158081]
- 54 Lee IC, Kim SH, Baek HS, Moon C, Kang SS, Kim YB, Shin IS, Kim JC. The involvement of Nrf2 in the protective effects of diallyl disulfide on carbon tetrachloride-induced hepatic oxidative damage and inflammatory response in rats. Food Chem Toxicol 2014; 63: 174-185 [PMID: 24246655 DOI: 10.1016/j.fct.2013.11.006]
- Origassa CS, Câmara NO. Cytoprotective role of heme oxygenase-1 and heme degradation derived end products in liver 55 injury. World J Hepatol 2013; 5: 541-549 [PMID: 24179613 DOI: 10.4254/wjh.v5.i10.541]
- 56 Chen Y, Guan W, Zhang N, Wang Y, Tian Y, Sun H, Li X, Liu J. Lactobacillus plantarum Lp2 improved LPS-induced liver injury through the TLR-4/MAPK/NFxB and Nrf2-HO-1/CYP2E1 pathways in mice. Food Nutr Res 2022; 66 [PMID: 35903291 DOI: 10.29219/fnr.v66.5459]
- 57 Bathish B, Robertson H, Dillon JF, Dinkova-Kostova AT, Hayes JD. Nonalcoholic steatohepatitis and mechanisms by which it is ameliorated by activation of the CNC-bZIP transcription factor Nrf2. Free Radic Biol Med 2022; 188: 221-261 [PMID: 35728768 DOI: 10.1016/j.freeradbiomed.2022.06.226]
- Biao Y, Chen J, Liu C, Wang R, Han X, Li L, Zhang Y. Protective Effect of Danshen Zexie Decoction Against Non-58 Alcoholic Fatty Liver Disease Through Inhibition of ROS/NLRP3/IL-1β Pathway by Nrf2 Signaling Activation. Front Pharmacol 2022; 13: 877924 [PMID: 35800450 DOI: 10.3389/fphar.2022.877924]
- Sharma RS, Harrison DJ, Kisielewski D, Cassidy DM, McNeilly AD, Gallagher JR, Walsh SV, Honda T, McCrimmon 59 RJ, Dinkova-Kostova AT, Ashford MLJ, Dillon JF, Hayes JD. Experimental Nonalcoholic Steatohepatitis and Liver Fibrosis Are Ameliorated by Pharmacologic Activation of Nrf2 (NF-E2 p45-Related Factor 2). Cell Mol Gastroenterol Hepatol 2018; 5: 367-398 [PMID: 29552625 DOI: 10.1016/j.jcmgh.2017.11.016]
- 60 Wagner N, Wagner KD. The Role of PPARs in Disease. Cells 2020; 9 [PMID: 33126411 DOI: 10.3390/cells9112367]
- 61 Decara J, Rivera P, López-Gambero AJ, Serrano A, Pavón FJ, Baixeras E, Rodríguez de Fonseca F, Suárez J. Peroxisome Proliferator-Activated Receptors: Experimental Targeting for the Treatment of Inflammatory Bowel Diseases. Front Pharmacol 2020; 11: 730 [PMID: 32536865 DOI: 10.3389/fphar.2020.00730]
- 62 Xu Y, Lu Y. Alcoholic fatty liver is blunted by rFGF21 administration in mice lacking adipose FGFR1: The role of FGF21 in PPARα-mediated regulation of adipose tissue mass. Biochem Biophys Res Commun 2022; 619: 84-89 [PMID:



35749940 DOI: 10.1016/j.bbrc.2022.05.099]

- 63 Pan J, Zhou W, Xu R, Xing L, Ji G, Dang Y. Natural PPARs agonists for the treatment of nonalcoholic fatty liver disease. Biomed Pharmacother 2022; 151: 113127 [PMID: 35598367 DOI: 10.1016/j.biopha.2022.113127]
- 64 Wang Y, Che Y, Wang S, Wang J, Liu X, Kou B, Guan Y, Chen D, Shi Y. ASPP2 reduction attenuates HBV induced chronic liver damage: A hybrid mouse model study. Biochem Biophys Res Commun 2022; 610: 61-69 [PMID: 35436632 DOI: 10.1016/j.bbrc.2022.03.109]
- 65 Ning Z, Guo X, Liu X, Lu C, Wang A, Wang X, Wang W, Chen H, Qin W, Zhou L, Ma C, Du J, Lin Z, Luo H, Otkur W, Qi H, Chen D, Xia T, Liu J, Tan G, Xu G, Piao HL. USP22 regulates lipidome accumulation by stabilizing PPARy in hepatocellular carcinoma. Nat Commun 2022; 13: 2187 [PMID: 35449157 DOI: 10.1038/s41467-022-29846-9]
- Xu Y, Denning KL, Lu Y. PPARa agonist WY-14,643 induces the PLA2/COX-2/ACOX1 pathway to enhance 66 peroxisomal lipid metabolism and ameliorate alcoholic fatty liver in mice. Biochem Biophys Res Commun 2022; 613: 47-52 [PMID: 35526488 DOI: 10.1016/j.bbrc.2022.04.132]
- 67 Huang J, Jia Y, Fu T, Viswakarma N, Bai L, Rao MS, Zhu Y, Borensztajn J, Reddy JK. Sustained activation of PPARa by endogenous ligands increases hepatic fatty acid oxidation and prevents obesity in ob/ob mice. FASEB J 2012; 26: 628-638 [PMID: 22009939 DOI: 10.1096/fj.11-194019]
- Baumann A, Burger K, Brandt A, Staltner R, Jung F, Rajcic D, Lorenzo Pisarello MJ, Bergheim I. GW9662, a 68 peroxisome proliferator-activated receptor gamma antagonist, attenuates the development of non-alcoholic fatty liver disease. Metabolism 2022; 133: 155233 [PMID: 35654114 DOI: 10.1016/j.metabol.2022.155233]
- Padole P, Arora A, Sharma P, Chand P, Verma N, Kumar A. Saroglitazar for Nonalcoholic Fatty Liver Disease: A Single Centre Experience in 91 Patients. J Clin Exp Hepatol 2022; 12: 435-439 [PMID: 35535066 DOI: 10.1016/j.jceh.2021.06.015]
- 70 Yokote K, Yamashita S, Arai H, Araki E, Matsushita M, Nojima T, Suganami H, Ishibashi S. Effects of pemafibrate on glucose metabolism markers and liver function tests in patients with hypertriglyceridemia: a pooled analysis of six phase 2 and phase 3 randomized double-blind placebo-controlled clinical trials. Cardiovasc Diabetol 2021; 20: 96 [PMID: 33947390 DOI: 10.1186/s12933-021-01291-w]
- 71 Nakajima A, Eguchi Y, Yoneda M, Imajo K, Tamaki N, Suganami H, Nojima T, Tanigawa R, Iizuka M, Iida Y, Loomba R. Randomised clinical trial: Pemafibrate, a novel selective peroxisome proliferator-activated receptor a modulator (SPPARMa), versus placebo in patients with non-alcoholic fatty liver disease. Aliment Pharmacol Ther 2021; 54: 1263-1277 [PMID: 34528723 DOI: 10.1111/apt.16596]
- Jones D, Boudes PF, Swain MG, Bowlus CL, Galambos MR, Bacon BR, Doerffel Y, Gitlin N, Gordon SC, Odin JA, 72 Sheridan D, Wörns MA, Clark V, Corless L, Hartmann H, Jonas ME, Kremer AE, Mells GF, Buggisch P, Freilich BL, Levy C, Vierling JM, Bernstein DE, Hartleb M, Janczewska E, Rochling F, Shah H, Shiffman ML, Smith JH, Choi YJ, Steinberg A, Varga M, Chera H, Martin R, McWherter CA, Hirschfield GM. Seladelpar (MBX-8025), a selective PPAR-o agonist, in patients with primary biliary cholangitis with an inadequate response to ursodeoxycholic acid: a double-blind, randomised, placebo-controlled, phase 2, proof-of-concept study. Lancet Gastroenterol Hepatol 2017; 2: 716-726 [PMID: 28818518 DOI: 10.1016/S2468-1253(17)30246-7]
- 73 Gastaldelli A, Sabatini S, Carli F, Gaggini M, Bril F, Belfort-DeAguiar R, Positano V, Barb D, Kadiyala S, Harrison S, Cusi K. PPAR-y-induced changes in visceral fat and adiponectin levels are associated with improvement of steatohepatitis in patients with NASH. Liver Int 2021; 41: 2659-2670 [PMID: 34219361 DOI: 10.1111/liv.15005]
- Della Pepa G, Russo M, Vitale M, Carli F, Vetrani C, Masulli M, Riccardi G, Vaccaro O, Gastaldelli A, Rivellese AA, Bozzetto L. Pioglitazone even at low dosage improves NAFLD in type 2 diabetes: clinical and pathophysiological insights from a subgroup of the TOSCA.IT randomised trial. *Diabetes Res Clin Pract* 2021; **178**: 108984 [PMID: 34311022 DOI: 10.1016/j.diabres.2021.108984]
- Sven M F, Pierre B, Manal F A, Quentin M A, Elisabetta B, Vlad R, Philippe HM, Bruno S, Jean-Louis J, Jean-Louis A. A randomised, double-blind, placebo-controlled, multi-centre, dose-range, proof-of-concept, 24-week treatment study of lanifibranor in adult subjects with non-alcoholic steatohepatitis: Design of the NATIVE study. Contemp Clin Trials 2020; 98: 106170 [PMID: 33038502 DOI: 10.1016/j.cct.2020.106170]
- 76 Francque SM, Bedossa P, Ratziu V, Anstee QM, Bugianesi E, Sanyal AJ, Loomba R, Harrison SA, Balabanska R, Mateva L, Lanthier N, Alkhouri N, Moreno C, Schattenberg JM, Stefanova-Petrova D, Vonghia L, Rouzier R, Guillaume M, Hodge A, Romero-Gómez M, Huot-Marchand P, Baudin M, Richard MP, Abitbol JL, Broqua P, Junien JL, Abdelmalek MF; NATIVE Study Group. A Randomized, Controlled Trial of the Pan-PPAR Agonist Lanifibranor in NASH. N Engl J Med 2021; 385: 1547-1558 [PMID: 34670042 DOI: 10.1056/NEJMoa2036205]
- Sun EJ, Wankell M, Palamuthusingam P, McFarlane C, Hebbard L. Targeting the PI3K/Akt/mTOR Pathway in 77 Hepatocellular Carcinoma. Biomedicines 2021; 9 [PMID: 34829868 DOI: 10.3390/biomedicines9111639]
- 78 Zhang C, Liu S, Yang M. Hepatocellular Carcinoma and Obesity, Type 2 Diabetes Mellitus, Cardiovascular Disease: Causing Factors, Molecular Links, and Treatment Options. Front Endocrinol (Lausanne) 2021; 12: 808526 [PMID: 35002979 DOI: 10.3389/fendo.2021.808526]
- Hu M, Chen Y, Deng F, Chang B, Luo J, Dong L, Lu X, Zhang Y, Chen Z, Zhou J. D-Mannose Regulates Hepatocyte 79 Lipid Metabolism via PI3K/Akt/mTOR Signaling Pathway and Ameliorates Hepatic Steatosis in Alcoholic Liver Disease. Front Immunol 2022; 13: 877650 [PMID: 35464439 DOI: 10.3389/fimmu.2022.877650]
- Xie H, Jing R, Liao X, Chen H, Xie X, Dai H, Pan L. Arecoline promotes proliferation and migration of human HepG2 80 cells through activation of the PI3K/AKT/mTOR pathway. Hereditas 2022; 159: 29 [PMID: 35836300 DOI: 10.1186/s41065-022-00241-0
- Zhang Y, Liang J, Cao N, Gao J, Xie Y, Zhou S, Tang X. ASIC1a up-regulates MMP-2/9 expression to enhance mobility 81 and proliferation of liver cancer cells via the PI3K/AKT/mTOR pathway. BMC Cancer 2022; 22: 778 [PMID: 35840921 DOI: 10.1186/s12885-022-09874-w]
- 82 Diniz PHC, Silva SDC, Vidigal PVT, Xavier MAP, Lima CX, Faria LC, Ferrari TCA. Expression of MAPK and PI3K/ AKT/mTOR Proteins according to the Chronic Liver Disease Etiology in Hepatocellular Carcinoma. J Oncol 2020; 2020: 4609360 [PMID: 33178273 DOI: 10.1155/2020/4609360]



- Fresno Vara JA, Casado E, de Castro J, Cejas P, Belda-Iniesta C, González-Barón M. PI3K/Akt signalling pathway and 83 cancer. Cancer Treat Rev 2004; 30: 193-204 [PMID: 15023437 DOI: 10.1016/j.ctrv.2003.07.007]
- 84 Yuan Z, He J, Xie T, Zhou M, Chen TT, Shi LP, He Y, Wang J, Shao M, Che JY. Effects and mechanisms of ziqi ruangan decoction on hepatic fibrosis. Pak J Pharm Sci 2021; 34: 2101-2107 [PMID: 35034870]
- 85 Li HG, You PT, Xia Y, Cai Y, Tu YJ, Wang MH, Song WC, Quan TM, Ren HY, Liu YW, Dan HX, Xu SQ. Yu Gan Long Ameliorates Hepatic Fibrosis by Inhibiting PI3K/AKT, Ras/ERK and JAK1/STAT3 Signaling Pathways in CCl(4)induced Liver Fibrosis Rats. Curr Med Sci 2020; 40: 539-547 [PMID: 32681257 DOI: 10.1007/s11596-020-2211-3]
- 86 Wang S, Wu Y, Liu M, Zhao Q, Jian L. DHW-208, A Novel Phosphatidylinositol 3-Kinase (PI3K) Inhibitor, Has Anti-Hepatocellular Carcinoma Activity Through Promoting Apoptosis and Inhibiting Angiogenesis. Front Oncol 2022; 12: 955729 [PMID: 35903690 DOI: 10.3389/fonc.2022.955729]
- Jung YY, Um JY, Sethi G, Ahn KS. Fangchinoline abrogates growth and survival of hepatocellular carcinoma by 87 negative regulation of c-met/HGF and its associated downstream signaling pathways. Phytother Res 2022; 36: 4542-4557 [PMID: 35867025 DOI: 10.1002/ptr.7573]
- 88 Chen W, Yang A, Jia J, Popov YV, Schuppan D, You H. Lysyl Oxidase (LOX) Family Members: Rationale and Their Potential as Therapeutic Targets for Liver Fibrosis. Hepatology 2020; 72: 729-741 [PMID: 32176358 DOI: 10.1002/hep.31236
- Dongiovanni P, Meroni M, Baselli GA, Bassani GA, Rametta R, Pietrelli A, Maggioni M, Facciotti F, Trunzo V, Badiali S, Fargion S, Gatti S, Valenti L. Insulin resistance promotes Lysyl Oxidase Like 2 induction and fibrosis accumulation in non-alcoholic fatty liver disease. Clin Sci (Lond) 2017; 131: 1301-1315 [PMID: 28468951 DOI: 10.1042/CS20170175]
- 90 Zetterberg FR, MacKinnon A, Brimert T, Gravelle L, Johnsson RE, Kahl-Knutson B, Leffler H, Nilsson UJ, Pedersen A, Peterson K, Roper JA, Schambye H, Slack RJ, Tantawi S. Discovery and Optimization of the First Highly Effective and Orally Available Galectin-3 Inhibitors for Treatment of Fibrotic Disease. J Med Chem 2022; 65: 12626-12638 [PMID: 36154172 DOI: 10.1021/acs.jmedchem.2c00660]
- 91 Herrera-Marcos LV, Martínez-Beamonte R, Macías-Herranz M, Arnal C, Barranquero C, Puente-Lanzarote JJ, Gascón S, Herrero-Continente T, Gonzalo-Romeo G, Alastrué-Vera V, Gutiérrez-Blázquez D, Lou-Bonafonte JM, Surra JC, Rodríguez-Yoldi MJ, García-Gil A, Güemes A, Osada J. Hepatic galectin-3 is associated with lipid droplet area in nonalcoholic steatohepatitis in a new swine model. Sci Rep 2022; 12: 1024 [PMID: 35046474 DOI: 10.1038/s41598-022-04971-z]
- 92 Sideras K, de Man RA, Harrington SM, Polak WG, Zhou G, Schutz HM, Pedroza-Gonzalez A, Biermann K, Mancham S, Hansen BE, Bart Takkenberg R, van Vuuren AJ, Pan Q, Ijzermans JNM, Sleijfer S, Sprengers D, Dong H, Kwekkeboom J, Bruno MJ. Circulating levels of PD-L1 and Galectin-9 are associated with patient survival in surgically treated Hepatocellular Carcinoma independent of their intra-tumoral expression levels. Sci Rep 2019; 9: 10677 [PMID: 31337865 DOI: 10.1038/s41598-019-47235-z]
- 93 Chen LY, Chen Q, Cheng YF, Jin HH, Kong DS, Zhang F, Wu L, Shao JJ, Zheng SZ. Diallyl trisulfide attenuates ethanol-induced hepatic steatosis by inhibiting oxidative stress and apoptosis. Biomed Pharmacother 2016; 79: 35-43 [PMID: 27044810 DOI: 10.1016/j.biopha.2016.01.009]
- 94 Lee DG, Lee J, Kim KT, Lee SW, Kim YO, Cho IH, Kim HJ, Park CG, Lee S. High-performance liquid chromatography analysis of phytosterols in Panax ginseng root grown under different conditions. J Ginseng Res 2018; 42: 16-20 [PMID: 29348717 DOI: 10.1016/j.jgr.2016.10.004]
- 95 Chen Z, Wu A, Jin H, Liu F. β-Sitosterol attenuates liver injury in a rat model of chronic alcohol intake. Arch Pharm Res 2020; **43**: 1197-1206 [PMID: 33155166 DOI: 10.1007/s12272-020-01271-w]
- 96 Lu W, Khatibi Shahidi F, Khorsandi K, Hosseinzadeh R, Gul A, Balick V, An update on molecular mechanisms of curcumin effect on diabetes. J Food Biochem 2022; 46: e14358 [PMID: 35945662 DOI: 10.1111/jfbc.14358]
- Song X, Zhang M, Dai E, Luo Y. Molecular targets of curcumin in breast cancer (Review). Mol Med Rep 2019; 19: 23-29 [PMID: 30483727 DOI: 10.3892/mmr.2018.9665]
- Chen Y, Wang J, Jing Z, Ordovas JM, Shen L. Anti-fatigue and anti-oxidant effects of curcumin supplementation in 98 exhaustive swimming mice via Nrf2/Keap1 signal pathway. Curr Res Food Sci 2022; 5: 1148-1157 [PMID: 35875345 DOI: 10.1016/j.crfs.2022.07.006]
- 99 Farashbandi AL, Shariati M, Mokhtari M. Comparing the Protective Effects of Curcumin and Ursodeoxycholic Acid after Ethanol-Induced Hepatotoxicity in Rat Liver. Ethiop J Health Sci 2021; 31: 673-682 [PMID: 34483625 DOI: 10.4314/ejhs.v31i3.25
- Bao S, Zhang Y, Ye J, Zhu Y, Li R, Xu X, Zhang Q. Self-assembled micelles enhance the oral delivery of curcumin for 100 the management of alcohol-induced tissue injury. *Pharm Dev Technol* 2021; 26: 880-889 [PMID: 34238120 DOI: 10.1080/10837450.2021.1950185]
- Kim SG, Suh HJ, Han SH, Lee HS, Kim HW, Kim H. Encapsulated Curcumin Enhances Intestinal Absorption and 101 Improves Hepatic Damage in Alcoholic Liver Disease-Induced Rats. Prev Nutr Food Sci 2019; 24: 410-417 [PMID: 31915636 DOI: 10.3746/pnf.2019.24.4.410]
- 102 Wang X, Chang X, Zhan H, Zhang Q, Li C, Gao Q, Yang M, Luo Z, Li S, Sun Y. Curcumin and Baicalin ameliorate ethanol-induced liver oxidative damage via the Nrf2/HO-1 pathway. J Food Biochem 2020; e13425 [PMID: 32770697 DOI: 10.1111/jfbc.13425]
- 103 Abdelhamid AM, Elsheakh AR, Abdelaziz RR, Suddek GM. Empagliflozin ameliorates ethanol-induced liver injury by modulating NF-κB/Nrf-2/PPAR-γ interplay in mice. Life Sci 2020; 256: 117908 [PMID: 32512011 DOI: 10.1016/j.lfs.2020.117908
- 104 Zhou Y, Chen J, Yao Z, Gu X. Gastrodin ameliorates Concanavalin A-induced acute hepatitis via the IL6/JAK2/STAT3 pathway. Immunopharmacol Immunotoxicol 2022; 44: 925-934 [PMID: 35881007 DOI: 10.1080/08923973.2022.2093741]
- Liao CC, Yu HP, Chou AH, Lee HC, Hu LM, Liu FC. Gastrodin Alleviates Acetaminophen-Induced Liver Injury in a 105 Mouse Model Through Inhibiting MAPK and Enhancing Nrf2 Pathways. Inflammation 2022; 45: 1450-1462 [PMID: 35474551 DOI: 10.1007/s10753-021-01557-1]



- 106 Sharifi-Rad J, Quispe C, Imran M, Rauf A, Nadeem M, Gondal TA, Ahmad B, Atif M, Mubarak MS, Sytar O, Zhilina OM, Garsiya ER, Smeriglio A, Trombetta D, Pons DG, Martorell M, Cardoso SM, Razis AFA, Sunusi U, Kamal RM, Rotariu LS, Butnariu M, Docea AO, Calina D. Genistein: An Integrative Overview of Its Mode of Action, Pharmacological Properties, and Health Benefits. Oxid Med Cell Longev 2021; 2021: 3268136 [PMID: 34336089 DOI: 10.1155/2021/3268136
- 107 Suksri K, Semprasert N, Limjindaporn T, Yenchitsomanus PT, Kooptiwoot S, Kooptiwut S. Cytoprotective effect of genistein against dexamethasone-induced pancreatic β-cell apoptosis. Sci Rep 2022; 12: 12950 [PMID: 35902739 DOI: 10.1038/s41598-022-17372-z]
- 108 Zhang W, Zhang L, Zhang X. Anti-atherosclerotic effects of genistein in preventing ox-low-density lipoprotein-induced smooth muscle-derived foam cell formation via inhibiting SRC expression and L-Ca channel currents. Ann Transl Med 2022; 10: 700 [PMID: 35845495 DOI: 10.21037/atm-22-2113]
- Jafari A, Esmaeilzadeh Z, Khezri MR, Ghasemnejad-Berenji H, Pashapour S, Sadeghpour S, Ghasemnejad-Berenji M. 109 An overview of possible pivotal mechanisms of Genistein as a potential phytochemical against SARS-CoV-2 infection: A hypothesis. J Food Biochem 2022; 46: e14345 [PMID: 35866873 DOI: 10.1111/jfbc.14345]
- 110 Zhao L, Zhang N, Yang D, Yang M, Guo X, He J, Wu W, Ji B, Cheng Q, Zhou F. Protective Effects of Five Structurally Diverse Flavonoid Subgroups against Chronic Alcohol-Induced Hepatic Damage in a Mouse Model. Nutrients 2018; 10 [PMID: 30441755 DOI: 10.3390/nu10111754]
- Mercer KE, Pulliam CF, Hennings L, Cleves MA, Jones EE, Drake RR, Ronis MJJ. Diet Supplementation with Soy 111 Protein Isolate, but Not the Isoflavone Genistein, Protects Against Alcohol-Induced Tumor Progression in DEN-Treated Male Mice. Adv Exp Med Biol 2018; 1032: 115-126 [PMID: 30362095 DOI: 10.1007/978-3-319-98788-0 9]
- 112 Soliman SA, Emeish WFA, Abdel-Hafeez HH. Lactoferrin improves the immune response and resistance of silver carp, a hematological, light (histochemical and immunohistochemical), fluorescent, and scanning electron microscopic study. Microsc Res Tech 2022; 85: 3565-3581 [PMID: 35876377 DOI: 10.1002/jemt.24208]
- 113 Li D, He Q, Yang H, Du Y, Yu K, Yang J, Tong X, Guo Y, Xu J, Qin L. Daily Dose of Bovine Lactoferrin Prevents Ethanol-Induced Liver Injury and Death in Male Mice by Regulating Hepatic Alcohol Metabolism and Modulating Gut Microbiota. Mol Nutr Food Res 2021; 65: e2100253 [PMID: 34331394 DOI: 10.1002/mnfr.202100253]
- 114 Li D, Hu Z, He Q, Guo Y, Chong Y, Xu J, Qin L. Lactoferrin Alleviates Acute Alcoholic Liver Injury by Improving Redox-Stress Response Capacity in Female C57BL/6J Mice. J Agric Food Chem 2021; 69: 14856-14867 [PMID: 34873911 DOI: 10.1021/acs.jafc.1c06813]
- Ojeda ML, Nogales F, Del Carmen Gallego-López M, Carreras O. Binge drinking during the adolescence period causes 115 oxidative damage-induced cardiometabolic disorders: A possible ameliorative approach with selenium supplementation. Life Sci 2022; 301: 120618 [PMID: 35533761 DOI: 10.1016/j.lfs.2022.120618]
- 116 Ojeda ML, Rua RM, Murillo ML, Carreras O, Nogales F. Binge drinking during adolescence disrupts Se homeostasis and its main hepatic selenoprotein expression. Alcohol Clin Exp Res 2015; 39: 818-826 [PMID: 25864381 DOI: 10.1111/acer.12707]
- Federico A, Dallio M, Loguercio C. Silymarin/Silybin and Chronic Liver Disease: A Marriage of Many Years. Molecules 117 2017; 22 [PMID: 28125040 DOI: 10.3390/molecules22020191]
- 118 Said ES, Mohammed AH, Ali HM, Babiker AY, Alnughaymishi R, Althaqeel NZ, Ahmed AS. Evaluation of hepatoprotective effect of Nebivolol and sodium copper Chlorophyllin on CCL4-induced hepatotoxicity in mice. Eur Rev Med Pharmacol Sci 2022; 26: 1717-1728 [PMID: 35302221 DOI: 10.26355/eurrev_202203_28241]
- Aghemo A, Alekseeva OP, Angelico F, Bakulin IG, Bakulina NV, Bordin D, Bueverov AO, Drapkina OM, Gillessen A, 119 Kagarmanova EM, Korochanskaya NV, Kucheryavii UA, Lazebnik LB, Livzan MA, Maev IV, Martynov AI, Osipenko MF, Sas EI, Starodubova A, Uspensky YP, Vinnitskaya EV, Yakovenko EP, Yakovlev AA. Role of silymarin as antioxidant in clinical management of chronic liver diseases: a narrative review. Ann Med 2022; 54: 1548-1560 [PMID: 35635048 DOI: 10.1080/07853890.2022.2069854]
- 120 Song Z, Deaciuc I, Song M, Lee DY, Liu Y, Ji X, McClain C. Silymarin protects against acute ethanol-induced hepatotoxicity in mice. Alcohol Clin Exp Res 2006; 30: 407-413 [PMID: 16499481 DOI: 10.1111/j.1530-0277.2006.00063.x]
- Lieber CS, Leo MA, Cao Q, Ren C, DeCarli LM. Silymarin retards the progression of alcohol-induced hepatic fibrosis in 121 baboons. J Clin Gastroenterol 2003; 37: 336-339 [PMID: 14506392 DOI: 10.1097/00004836-200310000-00013]
- 122 Yousefi Ghale-Salimi M, Eidi M, Ghaemi N, Khavari-Nejad RA. Antiurolithiatic effect of the taraxasterol on ethylene glycol induced kidney calculi in male rats. Urolithiasis 2018; 46: 419-428 [PMID: 29189886 DOI: 10.1007/s00240-017-1023-9
- Ye XJ, Xu R, Liu SY, Hu B, Shi ZJ, Shi FL, Zeng B, Xu LH, Huang YT, Chen MY, Zha QB, He XH, Ouyang DY. 123 Taraxasterol mitigates Con A-induced hepatitis in mice by suppressing interleukin-2 expression and its signaling in T lymphocytes. Int Immunopharmacol 2022; 102: 108380 [PMID: 34848154 DOI: 10.1016/j.intimp.2021.108380]
- 124 Abdelhamid AM, Elsheakh AR, Suddek GM, Abdelaziz RR. Telmisartan alleviates alcohol-induced liver injury by activation of PPAR-γ/ Nrf-2 crosstalk in mice. Int Immunopharmacol 2021; 99: 107963 [PMID: 34273638 DOI: 10.1016/j.intimp.2021.107963]
- Yang M, Kimchi ET, Staveley-O'Carroll KF, Li G. Astaxanthin Prevents Diet-Induced NASH Progression by Shaping 125 Intrahepatic Immunity. Int J Mol Sci 2021; 22 [PMID: 34681695 DOI: 10.3390/ijms222011037]
- 126 Gumede NM, Lembede BW, Nkomozepi P, Brooksbank RL, Erlwanger KH, Chivandi E. β-Sitosterol mitigates the development of high-fructose diet-induced nonalcoholic fatty liver disease in growing male Sprague-Dawley rats. Can J Physiol Pharmacol 2020; 98: 44-50 [PMID: 31560861 DOI: 10.1139/cjpp-2019-0295]
- 127 Gumede NM, Lembede BW, Brooksbank RL, Erlwanger KH, Chivandi E. β-Sitosterol Shows Potential to Protect Against the Development of High-Fructose Diet-Induced Metabolic Dysfunction in Female Rats. J Med Food 2020; 23: 367-374 [PMID: 31517568 DOI: 10.1089/jmf.2019.0120]
- Feng S, Dai Z, Liu AB, Huang J, Narsipur N, Guo G, Kong B, Reuhl K, Lu W, Luo Z, Yang CS. Intake of stigmasterol 128 and β-sitosterol alters lipid metabolism and alleviates NAFLD in mice fed a high-fat western-style diet. Biochim Biophys



Acta Mol Cell Biol Lipids 2018; 1863: 1274-1284 [PMID: 30305244 DOI: 10.1016/j.bbalip.2018.08.004]

- Maithilikarpagaselvi N, Sridhar MG, Swaminathan RP, Sripradha R. Preventive effect of curcumin on inflammation, 129 oxidative stress and insulin resistance in high-fat fed obese rats. J Complement Integr Med 2016; 13: 137-143 [PMID: 26845728 DOI: 10.1515/jcim-2015-0070]
- 130 Yan C, Zhang Y, Zhang X, Aa J, Wang G, Xie Y. Curcumin regulates endogenous and exogenous metabolism via Nrf2-FXR-LXR pathway in NAFLD mice. Biomed Pharmacother 2018; 105: 274-281 [PMID: 29860219 DOI: 10.1016/j.biopha.2018.05.135]
- 131 Liu Y, Cheng F, Luo Y, Zhan Z, Hu P, Ren H, Tang H, Peng M. PEGylated Curcumin Derivative Attenuates Hepatic Steatosis via CREB/PPAR-y/CD36 Pathway. Biomed Res Int 2017; 2017: 8234507 [PMID: 28770225 DOI: 10.1155/2017/8234507
- 132 Um MY, Hwang KH, Ahn J, Ha TY. Curcumin attenuates diet-induced hepatic steatosis by activating AMP-activated protein kinase. Basic Clin Pharmacol Toxicol 2013; 113: 152-157 [PMID: 23574662 DOI: 10.1111/bcpt.12076]
- Feng D, Zou J, Su D, Mai H, Zhang S, Li P, Zheng X. Curcumin prevents high-fat diet-induced hepatic steatosis in 133 ApoE(-/-) mice by improving intestinal barrier function and reducing endotoxin and liver TLR4/NF-KB inflammation. Nutr Metab (Lond) 2019; 16: 79 [PMID: 31788011 DOI: 10.1186/s12986-019-0410-3]
- 134 Hasanzadeh S, Read MI, Bland AR, Majeed M, Jamialahmadi T, Sahebkar A. Curcumin: an inflammasome silencer. Pharmacol Res 2020; 159: 104921 [PMID: 32464325 DOI: 10.1016/j.phrs.2020.104921]
- 135 Inzaugarat ME, De Matteo E, Baz P, Lucero D, García CC, Gonzalez Ballerga E, Daruich J, Sorda JA, Wald MR, Cherñavsky AC. New evidence for the therapeutic potential of curcumin to treat nonalcoholic fatty liver disease in humans. PLoS One 2017; 12: e0172900 [PMID: 28257515 DOI: 10.1371/journal.pone.0172900]
- 136 Panahi Y, Kianpour P, Mohtashami R, Jafari R, Simental-Mendía LE, Sahebkar A. Efficacy and Safety of Phytosomal Curcumin in Non-Alcoholic Fatty Liver Disease: A Randomized Controlled Trial. Drug Res (Stuttg) 2017; 67: 244-251 [PMID: 28158893 DOI: 10.1055/s-0043-100019]
- 137 Mirhafez SR, Azimi-Nezhad M, Dehabeh M, Hariri M, Naderan RD, Movahedi A, Abdalla M, Sathyapalan T, Sahebkar A. The Effect of Curcumin Phytosome on the Treatment of Patients with Non-alcoholic Fatty Liver Disease: A Double-Blind, Randomized, Placebo-Controlled Trial. Adv Exp Med Biol 2021; 1308: 25-35 [PMID: 33861434 DOI: 10.1007/978-3-030-64872-5_3]
- 138 Derosa G, Maffioli P, Sahebkar A. Piperine and Its Role in Chronic Diseases. Adv Exp Med Biol 2016; 928: 173-184 [PMID: 27671817 DOI: 10.1007/978-3-319-41334-1 8]
- 139 Mirhafez SR, Dehabeh M, Hariri M, Farimani AR, Movahedi A, Naderan RD, Jamialahmadi T, Simental-Mendía LE, Sahebkar A. Curcumin and Piperine Combination for the Treatment of Patients with Non-alcoholic Fatty Liver Disease: A Double-Blind Randomized Placebo-Controlled Trial. Adv Exp Med Biol 2021; 1328: 11-19 [PMID: 34981468 DOI: 10.1007/978-3-030-73234-9 21
- Ma Y, Zhang G, Kuang Z, Xu Q, Ye T, Li X, Qu N, Han F, Kan C, Sun X. Empagliflozin activates Sestrin2-mediated 140 AMPK/mTOR pathway and ameliorates lipid accumulation in obesity-related nonalcoholic fatty liver disease. Front Pharmacol 2022; 13: 944886 [PMID: 36133815 DOI: 10.3389/fphar.2022.944886]
- 141 Wang Y, Shen QL, Xin Q, Sun B, Zhang S, Fang QH, Shi YX, Niu WY, Lin JN, Li CJ. MCAD activation by empagliflozin promotes fatty acid oxidation and reduces lipid deposition in NASH. J Mol Endocrinol 2022; 69: 415-430 [PMID: 35900373 DOI: 10.1530/JME-22-0022]
- Zhang Y, Liu X, Zhang H, Wang X. Efficacy and Safety of Empagliflozin on Nonalcoholic Fatty Liver Disease: A 142 Systematic Review and Meta-Analysis. Front Endocrinol (Lausanne) 2022; 13: 836455 [PMID: 35282455 DOI: 10.3389/fendo.2022.836455]
- 143 Ohki T, Isogawa A, Toda N, Tagawa K. Effectiveness of Ipragliflozin, a Sodium-Glucose Co-transporter 2 Inhibitor, as a Second-line Treatment for Non-Alcoholic Fatty Liver Disease Patients with Type 2 Diabetes Mellitus Who Do Not Respond to Incretin-Based Therapies Including Glucagon-like Peptide-1 Analogs and Dipeptidyl Peptidase-4 Inhibitors. Clin Drug Investig 2016; 36: 313-319 [PMID: 26914659 DOI: 10.1007/s40261-016-0383-1]
- 144 Harrison SA, Manghi FP, Smith WB, Alpenidze D, Aizenberg D, Klarenbeek N, Chen CY, Zuckerman E, Ravussin E, Charatcharoenwitthaya P, Cheng PN, Katchman H, Klein S, Ben-Ari Z, Mendonza AE, Zhang Y, Martic M, Ma S, Kao S, Tanner S, Pachori A, Badman MK, He Y, Ukomadu C, Sicard E. Licogliflozin for nonalcoholic steatohepatitis: a randomized, double-blind, placebo-controlled, phase 2a study. Nat Med 2022; 28: 1432-1438 [PMID: 35725922 DOI: 10.1038/s41591-022-01861-9]
- 145 Qiao P, Jia Y, Ma A, He J, Shao C, Li X, Wang S, Yang B, Zhou H. Dapagliflozin protects against nonalcoholic steatohepatitis in db/db mice. Front Pharmacol 2022; 13: 934136 [PMID: 36059948 DOI: 10.3389/fphar.2022.934136]
- 146 He K, Li J, Xi W, Ge J, Sun J, Jing Z. Dapagliflozin for nonalcoholic fatty liver disease: A systematic review and metaanalysis. Diabetes Res Clin Pract 2022; 185: 109791 [PMID: 35202771 DOI: 10.1016/j.diabres.2022.109791]
- 147 Wan J, Zhang Y, Yang D, Liang Y, Yang L, Hu S, Liu Z, Fang Q, Tian S, Ding Y. Gastrodin Improves Nonalcoholic Fatty Liver Disease Through Activation of the Adenosine Monophosphate-Activated Protein Kinase Signaling Pathway. Hepatology 2021; 74: 3074-3090 [PMID: 34297426 DOI: 10.1002/hep.32068]
- 148 Ahmad O, Wang B, Ma K, Deng Y, Li M, Yang L, Yang Y, Zhao J, Cheng L, Zhou Q, Shang J. Lipid Modulating Antioxidant Stress Activity of Gastrodin on Nonalcoholic Fatty Liver Disease Larval Zebrafish Model. Int J Mol Sci 2019; 20 [PMID: 31018538 DOI: 10.3390/ijms20081984]
- 149 Yin Y, Liu H, Zheng Z, Lu R, Jiang Z. Genistein can ameliorate hepatic inflammatory reaction in nonalcoholic steatohepatitis rats. Biomed Pharmacother 2019; 111: 1290-1296 [PMID: 30841442 DOI: 10.1016/j.biopha.2019.01.004]
- Zamani-Garmsiri F, Hashemnia SMR, Shabani M, Bagherieh M, Emamgholipour S, Meshkani R. Combination of 150 metformin and genistein alleviates non-alcoholic fatty liver disease in high-fat diet-fed mice. J Nutr Biochem 2021; 87: 108505 [PMID: 32956824 DOI: 10.1016/j.jnutbio.2020.108505]
- 151 Witayavanitkul N, Werawatganon D, Chayanupatkul M, Klaikeaw N, Sanguanrungsirikul S, Siriviriyakul P. Genistein and exercise modulated lipid peroxidation and improved steatohepatitis in ovariectomized rats. BMC Complement Med *Ther* 2020; **20**: 162 [PMID: 32482167 DOI: 10.1186/s12906-020-02962-z]



- 152 Wang X, Wang Y, Xu W, Lan L, Li Y, Wang L, Sun X, Yang C, Jiang Y, Feng R. Dietary isoflavones intake is inversely associated with non-alcoholic fatty liver disease, hyperlipidaemia and hypertension. Int J Food Sci Nutr 2022; 73: 60-70 [PMID: 33899670 DOI: 10.1080/09637486.2021.1910630]
- 153 Seidemann L, Krüger A, Kegel-Hübner V, Seehofer D, Damm G. Influence of Genistein on Hepatic Lipid Metabolism in an In Vitro Model of Hepatic Steatosis. Molecules 2021; 26 [PMID: 33671486 DOI: 10.3390/molecules26041156]
- 154 Aoyama Y, Naiki-Ito A, Xiaochen K, Komura M, Kato H, Nagayasu Y, Inaguma S, Tsuda H, Tomita M, Matsuo Y, Takiguchi S, Takahashi S. Lactoferrin Prevents Hepatic Injury and Fibrosis via the Inhibition of NF-KB Signaling in a Rat Non-Alcoholic Steatohepatitis Model. Nutrients 2021; 14 [PMID: 35010924 DOI: 10.3390/nu14010042]
- Guo C, Xue H, Guo T, Zhang W, Xuan WQ, Ren YT, Wang D, Chen YH, Meng YH, Gao HL, Zhao P. Recombinant 155 human lactoferrin attenuates the progression of hepatosteatosis and hepatocellular death by regulating iron and lipid homeostasis in ob/ob mice. Food Funct 2020; 11: 7183-7196 [PMID: 32756704 DOI: 10.1039/d0fo00910e]
- 156 Soulaidopoulos S, Tsiogka A, Chrysohoou C, Lazarou E, Aznaouridis K, Doundoulakis I, Tyrovola D, Tousoulis D, Tsioufis K, Vlachopoulos C, Lazaros G. Overview of Chios Mastic Gum (Pistacia lentiscus) Effects on Human Health. Nutrients 2022; 14 [PMID: 35276949 DOI: 10.3390/nu14030590]
- 157 Kannt A, Papada E, Kammermeier C, D'Auria G, Jiménez-Hernández N, Stephan M, Schwahn U, Madsen AN, Østergaard MV, Dedoussis G, Francino MP; MAST4HEALTH consortium. Mastiha (Pistacia lentiscus) Improves Gut Microbiota Diversity, Hepatic Steatosis, and Disease Activity in a Biopsy-Confirmed Mouse Model of Advanced Non-Alcoholic Steatohepatitis and Fibrosis. Mol Nutr Food Res 2019; 63: e1900927 [PMID: 31599067 DOI: 10.1002/mnfr.201900927
- Kanoni S, Kumar S, Amerikanou C, Kurth MJ, Stathopoulou MG, Bourgeois S, Masson C, Kannt A, Cesarini L, Kontoe 158 MS, Milanović M, Roig FJ, Beribaka M, Campolo J, Jiménez-Hernández N, Milošević N, Llorens C, Smyrnioudis I, Francino MP, Milić N, Kaliora AC, Trivella MG, Ruddock MW, Medić-Stojanoska M, Gastaldelli A, Lamont J, Deloukas P, Dedoussis GV, Visvikis-Siest S. Nutrigenetic Interactions Might Modulate the Antioxidant and Anti-Inflammatory Status in Mastiha-Supplemented Patients With NAFLD. Front Immunol 2021; 12: 683028 [PMID: 34025683 DOI: 10.3389/fimmu.2021.683028
- 159 Amerikanou C, Papada E, Gioxari A, Smyrnioudis I, Kleftaki SA, Valsamidou E, Bruns V, Banerjee R, Trivella MG, Milic N, Medić-Stojanoska M, Gastaldelli A, Kannt A; MAST4HEALTH, Dedoussis GV, Kaliora AC. Mastiha has efficacy in immune-mediated inflammatory diseases through a microRNA-155 Th17 dependent action. Pharmacol Res 2021; 171: 105753 [PMID: 34224858 DOI: 10.1016/j.phrs.2021.105753]
- 160 Zhou DD, Mao QQ, Li BY, Saimaiti A, Huang SY, Xiong RG, Shang A, Luo M, Li HY, Gan RY, Li HB, Li S. Effects of Different Green Teas on Obesity and Non-Alcoholic Fatty Liver Disease Induced by a High-Fat Diet in Mice. Front Nutr 2022; 9: 929210 [PMID: 35811941 DOI: 10.3389/fnut.2022.929210]
- 161 Abdallah AAM, Abdelrahman MM, Attia HMAS, Hafez A, Anwar Rashed S, Amin YA, Hemdan SB. Decreased Serum zinc, selenium, and vitamin E as possible risk factors of hepatic fibrosis in non-alcoholic fatty liver disease. Nutr Health 2022; 2601060221103032 [PMID: 35603860 DOI: 10.1177/02601060221103032]
- Tang C, Li S, Zhang K, Li J, Han Y, Zhan T, Zhao Q, Guo X, Zhang J. Selenium deficiency-induced redox imbalance 162 leads to metabolic reprogramming and inflammation in the liver. Redox Biol 2020; 36: 101519 [PMID: 32531544 DOI: 10.1016/j.redox.2020.101519]
- Pant R, Sharma N, Kabeer SW, Sharma S, Tikoo K. Selenium-Enriched Probiotic Alleviates Western Diet-Induced Non-163 alcoholic Fatty Liver Disease in Rats via Modulation of Autophagy Through AMPK/SIRT-1 Pathway. Biol Trace Elem Res 2022 [PMID: 35499800 DOI: 10.1007/s12011-022-03247-x]
- Zhang Q, Zhou X, Zhang J, Li Q, Qian Z. Selenium and vitamin B(6) cosupplementation improves dyslipidemia and fatty 164 liver syndrome by SIRT1/SREBP-1c pathway in hyperlipidemic Sprague-Dawley rats induced by high-fat diet. Nutr Res 2022; 106: 101-118 [PMID: 36183668 DOI: 10.1016/j.nutres.2022.06.010]
- 165 Cui S, Pan XJ, Ge CL, Guo YT, Zhang PF, Yan TT, Zhou JY, He QX, Cheng LH, Wang GJ, Hao HP, Wang H. Silybin alleviates hepatic lipid accumulation in methionine-choline deficient diet-induced nonalcoholic fatty liver disease in mice via peroxisome proliferator-activated receptor a. Chin J Nat Med 2021; 19: 401-411 [PMID: 34092291 DOI: 10.1016/S1875-5364(21)60039-0
- Stephen Robert JM, Peddha MS, Srivastava AK. Effect of Silymarin and Quercetin in a Miniaturized Scaffold in Wistar 166 Rats against Non-alcoholic Fatty Liver Disease. ACS Omega 2021; 6: 20735-20745 [PMID: 34423182 DOI: 10.1021/acsomega.1c00555
- 167 Mengesha T, Gnanasekaran N, Mehare T. Hepatoprotective effect of silymarin on fructose induced nonalcoholic fatty liver disease in male albino wistar rats. BMC Complement Med Ther 2021; 21: 104 [PMID: 33785007 DOI: 10.1186/s12906-021-03275-5
- Mirhashemi SH, Hakakzadeh A, Yeganeh FE, Oshidari B, Rezaee SP. Effect of 8 Weeks milk thistle powder (silymarin 168 extract) supplementation on fatty liver disease in patients candidates for bariatric surgery. Metabol Open 2022; 14: 100190 [PMID: 35651885 DOI: 10.1016/j.metop.2022.100190]
- Zhang QZ, Liu YL, Wang YR, Fu LN, Zhang J, Wang XR, Wang BM. Effects of telmisartan on improving leptin 169 resistance and inhibiting hepatic fibrosis in rats with non-alcoholic fatty liver disease. Exp Ther Med 2017; 14: 2689-2694 [PMID: 28962213 DOI: 10.3892/etm.2017.4809]
- Wasta Esmail VA, Al-Nimer MSM, Mohammed MO. Effects of Orlistat or Telmisartan on the Serum Free Fatty Acids in 170 Non-alcoholic Fatty Liver Disease Patients: An Open-Labeled Randomized Controlled Study. Turk J Gastroenterol 2022; 33: 421-426 [PMID: 35678800 DOI: 10.5152/tjg.2020.19365]
- 171 Alam S, Abrar M, Islam S, Kamal M, Hasan MJ, Khan MAS, Ahmad N. Effect of telmisartan and vitamin E on liver histopathology with non-alcoholic steatohepatitis: A randomized, open-label, noninferiority trial. JGH Open 2020; 4: 663-669 [PMID: 32782954 DOI: 10.1002/jgh3.12315]
- 172 Devan AR, Nair B, Kumar AR, Nath LR. An insight into the role of telmisartan as PPAR-γ/α dual activator in the management of nonalcoholic fatty liver disease. Biotechnol Appl Biochem 2022; 69: 461-468 [PMID: 33578449 DOI: 10.1002/bab.2123]



- Sen CK, Khanna S, Roy S. Tocotrienols: Vitamin E beyond tocopherols. Life Sci 2006; 78: 2088-2098 [PMID: 16458936 173 DOI: 10.1016/j.lfs.2005.12.001]
- 174 Wong WY, Ward LC, Fong CW, Yap WN, Brown L. Anti-inflammatory y- and δ-tocotrienols improve cardiovascular, liver and metabolic function in diet-induced obese rats. Eur J Nutr 2017; 56: 133-150 [PMID: 26446095 DOI: 10.1007/s00394-015-1064-1]
- Wang H, Yan W, Sun Y, Yang CS. δ-Tocotrienol is the Most Potent Vitamin E Form in Inhibiting Prostate Cancer Cell 175 Growth and Inhibits Prostate Carcinogenesis in Ptenp-/- Mice. Cancer Prev Res (Phila) 2022; 15: 233-245 [PMID: 35144931 DOI: 10.1158/1940-6207.CAPR-21-0508]
- Pervez MA, Khan DA, Ijaz A, Khan S. Effects of Delta-tocotrienol Supplementation on Liver Enzymes, Inflammation, 176 Oxidative stress and Hepatic Steatosis in Patients with Nonalcoholic Fatty Liver Disease. Turk J Gastroenterol 2018; 29: 170-176 [PMID: 29749323 DOI: 10.5152/tjg.2018.17297]
- 177 Suleman F, Khan DA, Pervez MA, Aamir M. Effects of delta-tocotrienol supplementation on glycaemic control in individuals with prediabetes: A randomized controlled study. J Pak Med Assoc 2022; 72: 4-7 [PMID: 35099428 DOI: 10.47391/JPMA.966
- Mahjabeen W, Khan DA, Mirza SA, Pervez MA. Effects of delta-tocotrienol supplementation on Glycemic Control, 178 oxidative stress, inflammatory biomarkers and miRNA expression in type 2 diabetes mellitus: A randomized control trial. Phytother Res 2021; 35: 3968-3976 [PMID: 33899292 DOI: 10.1002/ptr.7113]
- 179 Pervez MA, Khan DA, Slehria AUR, Ijaz A. Delta-tocotrienol supplementation improves biochemical markers of hepatocellular injury and steatosis in patients with nonalcoholic fatty liver disease: A randomized, placebo-controlled trial. Complement Ther Med 2020; 52: 102494 [PMID: 32951743 DOI: 10.1016/j.ctim.2020.102494]
- Nisha R, Kumar P, Gautam AK, Bera H, Bhattacharya B, Parashar P, Saraf SA, Saha S. Assessments of in vitro and 180 in vivo antineoplastic potentials of β-sitosterol-loaded PEGylated niosomes against hepatocellular carcinoma. J Liposome Res 2021; 31: 304-315 [PMID: 32901571 DOI: 10.1080/08982104.2020.1820520]
- 181 Kathiswar Raj R, Ezhilarasan D, Rajeshkumar S. β-Sitosterol-assisted silver nanoparticles activates Nrf2 and triggers mitochondrial apoptosis via oxidative stress in human hepatocellular cancer cell line. J Biomed Mater Res A 2020; 108: 1899-1908 [PMID: 32319188 DOI: 10.1002/jbm.a.36953]
- Bai C, Zhao J, Su J, Chen J, Cui X, Sun M, Zhang X. Curcumin induces mitochondrial apoptosis in human hepatoma cells 182 through BCLAF1-mediated modulation of PI3K/AKT/GSK-3β signaling. Life Sci 2022; 306: 120804 [PMID: 35882275 DOI: 10.1016/j.lfs.2022.120804]
- 183 Abdelhamid AM, Saber S, Youssef ME, Gaafar AGA, Eissa H, Abd-Eldayem MA, Alqarni M, Batiha GE, Obaidullah AJ, Shahien MA, El-Ahwany E, Amin NA, Etman MA, Kaddah MMY, Abd El-Fattah EE. Empagliflozin adjunct with metformin for the inhibition of hepatocellular carcinoma progression: Emerging approach for new application. Biomed Pharmacother 2022; 145: 112455 [PMID: 34844106 DOI: 10.1016/j.biopha.2021.112455]
- 184 Shu G, Yang T, Wang C, Su H, Xiang M. Gastrodin stimulates anticancer immune response and represses transplanted H22 hepatic ascitic tumor cell growth: Involvement of NF-KB signaling activation in CD4+ T cells. Toxicol Appl Pharmacol 2013; 269: 270-279 [PMID: 23578476 DOI: 10.1016/j.taap.2013.02.019]
- 185 El-Far YM, Khodir AE, Emarah ZA, Ebrahim MA, Al-Gayyar MMH. Chemopreventive and hepatoprotective effects of genistein via inhibition of oxidative stress and the versican/PDGF/PKC signaling pathway in experimentally induced hepatocellular carcinoma in rats by thioacetamide. Redox Rep 2022; 27: 9-20 [PMID: 35080474 DOI: 10.1080/13510002.2022.2031515
- 186 Hegazy RR, Mansour DF, Salama AA, Abdel-Rahman RF, Hassan AM. Regulation of PKB/Akt-pathway in the chemopreventive effect of lactoferrin against diethylnitrosamine-induced hepatocarcinogenesis in rats. Pharmacol Rep 2019; 71: 879-891 [PMID: 31442665 DOI: 10.1016/j.pharep.2019.04.019]
- 187 Al-Noshokaty TM, Mesbah NM, Abo-Elmatty DM, Abulsoud AI, Abdel-Hamed AR. Selenium nanoparticles overcomes sorafenib resistance in thioacetamide induced hepatocellular carcinoma in rats by modulation of mTOR, NF-KB pathways and LncRNA-AF085935/GPC3 axis. Life Sci 2022; 303: 120675 [PMID: 35640776 DOI: 10.1016/j.lfs.2022.120675]
- 188 Yassin NYS, AbouZid SF, El-Kalaawy AM, Ali TM, Almehmadi MM, Ahmed OM. Silybum marianum total extract, silymarin and silibinin abate hepatocarcinogenesis and hepatocellular carcinoma growth via modulation of the HGF/c-Met, Wnt/β-catenin, and PI3K/Akt/mTOR signaling pathways. Biomed Pharmacother 2022; 145: 112409 [PMID: 34781148 DOI: 10.1016/j.biopha.2021.112409]
- 189 Ren F, Zhang Y, Qin Y, Shang J, Wang Y, Wei P, Guo J, Jia H, Zhao T. Taraxasterol prompted the anti-tumor effect in mice burden hepatocellular carcinoma by regulating T lymphocytes. Cell Death Discov 2022; 8: 264 [PMID: 35577774 DOI: 10.1038/s41420-022-01059-5]
- 190 Saber S, Khodir AE, Soliman WE, Salama MM, Abdo WS, Elsaeed B, Nader K, Abdelnasser A, Megahed N, Basuony M, Shawky A, Mahmoud M, Medhat R, Eldin AS. Telmisartan attenuates N-nitrosodiethylamine-induced hepatocellular carcinoma in mice by modulating the NF-κB-TAK1-ERK1/2 axis in the context of PPARγ agonistic activity. Naunyn Schmiedebergs Arch Pharmacol 2019; 392: 1591-1604 [PMID: 31367864 DOI: 10.1007/s00210-019-01706-2]
- 191 Lucci A, Vera MC, Comanzo CG, Lorenzetti F, Ferretti AC, Ceballos MP, Quiroga AD, Alvarez ML, Carrillo MC. Deltatocotrienol enhances the anti-tumor effects of interferon alpha through reactive oxygen species and Erk/MAPK signaling pathways in hepatocellular carcinoma cells. Can J Physiol Pharmacol 2022; 100: 453-463 [PMID: 34932399 DOI: 10.1139/cjpp-2021-0606]
- 192 Taheri H, Malek M, Ismail-Beigi F, Zamani F, Sohrabi M, Reza Babaei M, Khamseh ME. Effect of Empagliflozin on Liver Steatosis and Fibrosis in Patients With Non-Alcoholic Fatty Liver Disease Without Diabetes: A Randomized, Double-Blind, Placebo-Controlled Trial. Adv Ther 2020; 37: 4697-4708 [PMID: 32975679 DOI: 10.1007/s12325-020-01498-5]
- 193 Schrieber SJ, Hawke RL, Wen Z, Smith PC, Reddy KR, Wahed AS, Belle SH, Afdhal NH, Navarro VJ, Meyers CM, Doo E. Fried MW. Differences in the disposition of silvmarin between patients with nonalcoholic fatty liver disease and chronic hepatitis C. Drug Metab Dispos 2011; 39: 2182-2190 [PMID: 21865319 DOI: 10.1124/dmd.111.040212]
- 194 Navarro VJ, Belle SH, D'Amato M, Adfhal N, Brunt EM, Fried MW, Reddy KR, Wahed AS, Harrison S; Silymarin in



NASH and C Hepatitis (SyNCH) Study Group. Silymarin in non-cirrhotics with non-alcoholic steatohepatitis: A randomized, double-blind, placebo controlled trial. PLoS One 2019; 14: e0221683 [PMID: 31536511 DOI: 10.1371/journal.pone.0221683]

195 Amanat S, Eftekhari MH, Fararouei M, Bagheri Lankarani K, Massoumi SJ. Genistein supplementation improves insulin resistance and inflammatory state in non-alcoholic fatty liver patients: A randomized, controlled trial. Clin Nutr 2018; 37: 1210-1215 [PMID: 28647291 DOI: 10.1016/j.clnu.2017.05.028]



Saisbideng® WJH | https://www.wjgnet.com



Published by Baishideng Publishing Group Inc 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA Telephone: +1-925-3991568 E-mail: bpgoffice@wjgnet.com Help Desk: https://www.f6publishing.com/helpdesk https://www.wjgnet.com

