

World Journal of *Hepatology*

World J Hepatol 2023 March 27; 15(3): 321-440



REVIEW

- 321 Main factors influencing long-term outcomes of liver transplantation in 2022
Fuochi E, Anastasio L, Lynch EN, Campani C, Dragoni G, Milani S, Galli A, Innocenti T

MINIREVIEWS

- 353 COVID-19 and liver dysfunction in children: Current views and new hypotheses
Yun YF, Feng ZY, Zhang JJ
- 364 May 2022 acute hepatitis outbreak, is there a role for COVID-19 and other viruses?
Elbeltagi R, Al-Beltagi M, Saeed NK, Bediwy AS, Toema O
- 377 Challenges and recommendations when selecting empirical antibiotics in patients with cirrhosis
Dirchwolf M, Gomez Perdiguero G, Grech IM, Marciano S
- 386 Emerging role of engineered exosomes in nonalcoholic fatty liver disease
Ding J, Xu C, Xu M, He XY, Li WN, He F

ORIGINAL ARTICLE

Basic Study

- 393 mRNA transcriptome profiling of human hepatocellular carcinoma cells HepG2 treated with *Catharanthus roseus*-silver nanoparticles
Azhar NA, Abu Bakar SA, Citartan M, Ahmad NH

Retrospective Cohort Study

- 410 Adherence to guideline-directed hepatocellular carcinoma screening: A single-center US experience
King WW, Richhart R, Culpepper T, Mota M, Banerjee D, Ismael M, Chakraborty J, Ladna M, Khan W, Ruiz N, Wilson J, Altshuler E, Clark V, Cabrera R

Retrospective Study

- 419 To scan or not to scan: Use of transient elastography in an integrated health system
Stein L, Mittal R, Song H, Chung J, Sahota A
- 431 Coexistent alcohol-related cirrhosis and chronic pancreatitis have a comparable phenotype to either disease alone: A comparative retrospective analysis
Lu M, Sun Y, Feldman R, Saul M, Althouse A, Arteel G, Yadav D

ABOUT COVER

Editorial Board Member of *World Journal of Hepatology*, Raika Jamali, MD, Gastroenterologist and Hepatologist, Associate Professor, Vice President for Research, Digestive Disease Research Institute, Tehran University of Medical Sciences, Shariati Hospital, Tehran, Iran. jamalira@tums.ac.ir

AIMS AND SCOPE

The primary aim of *World Journal of Hepatology* (*WJH*, *World J Hepatol*) is to provide scholars and readers from various fields of hepatology with a platform to publish high-quality basic and clinical research articles and communicate their research findings online.

WJH mainly publishes articles reporting research results and findings obtained in the field of hepatology and covering a wide range of topics including chronic cholestatic liver diseases, cirrhosis and its complications, clinical alcoholic liver disease, drug induced liver disease autoimmune, fatty liver disease, genetic and pediatric liver diseases, hepatocellular carcinoma, hepatic stellate cells and fibrosis, liver immunology, liver regeneration, hepatic surgery, liver transplantation, biliary tract pathophysiology, non-invasive markers of liver fibrosis, viral hepatitis.

INDEXING/ABSTRACTING

The *WJH* is now abstracted and indexed in PubMed, PubMed Central, Emerging Sources Citation Index (Web of Science), Scopus, Reference Citation Analysis, China National Knowledge Infrastructure, China Science and Technology Journal Database, and Superstar Journals Database. The 2022 edition of Journal Citation Reports® cites the 2021 Journal Citation Indicator (JCI) for *WJH* as 0.52. The *WJH*'s CiteScore for 2021 is 3.6 and Scopus CiteScore rank 2021: Hepatology is 42/70.

RESPONSIBLE EDITORS FOR THIS ISSUE

Production Editor: Yi-Xuan Cai; Production Department Director: Xiang Li; Editorial Office Director: Xiang Li.

NAME OF JOURNAL

World Journal of Hepatology

ISSN

ISSN 1948-5182 (online)

LAUNCH DATE

October 31, 2009

FREQUENCY

Monthly

EDITORS-IN-CHIEF

Nikolaos Pylsopoulos, Ke-Qin Hu, Koo Jeong Kang

EDITORIAL BOARD MEMBERS

<https://www.wjnet.com/1948-5182/editorialboard.htm>

PUBLICATION DATE

March 27, 2023

COPYRIGHT

© 2023 Baishideng Publishing Group Inc

INSTRUCTIONS TO AUTHORS

<https://www.wjnet.com/bpg/gerinfo/204>

GUIDELINES FOR ETHICS DOCUMENTS

<https://www.wjnet.com/bpg/GerInfo/287>

GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH

<https://www.wjnet.com/bpg/gerinfo/240>

PUBLICATION ETHICS

<https://www.wjnet.com/bpg/GerInfo/288>

PUBLICATION MISCONDUCT

<https://www.wjnet.com/bpg/gerinfo/208>

ARTICLE PROCESSING CHARGE

<https://www.wjnet.com/bpg/gerinfo/242>

STEPS FOR SUBMITTING MANUSCRIPTS

<https://www.wjnet.com/bpg/GerInfo/239>

ONLINE SUBMISSION

<https://www.f6publishing.com>



Emerging role of engineered exosomes in nonalcoholic fatty liver disease

Jian Ding, Chen Xu, Ming Xu, Xiao-Yue He, Wei-Na Li, Fei He

Specialty type: Gastroenterology and hepatology

Provenance and peer review: Invited article; Externally peer reviewed.

Peer-review model: Single blind

Peer-review report's scientific quality classification

Grade A (Excellent): 0
Grade B (Very good): B, B
Grade C (Good): 0
Grade D (Fair): D
Grade E (Poor): 0

P-Reviewer: Forlano R, United Kingdom; Thandassery RB, United States

Received: December 26, 2022

Peer-review started: December 26, 2022

First decision: February 1, 2023

Revised: February 20, 2023

Accepted: March 15, 2023

Article in press: March 15, 2023

Published online: March 27, 2023



Jian Ding, Chen Xu, Ming Xu, Department of Hepatobiliary Surgery, Xi-Jing Hospital, The Fourth Military Medical University, Xi'an 710032, Shaanxi Province, China

Xiao-Yue He, The Affiliated Hospital of Jining Medical University, Jining Medical University, Jining 272067, Shandong Province, China

Wei-Na Li, School of Basic Medicine, The Fourth Military Medical University, Xi'an 710032, Shaanxi Province, China

Fei He, Department of Hepatobiliary Surgery, Xi-Jing Hospital, Xi'an 710032, Shaanxi Province, China

Corresponding author: Fei He, PhD, Research Associate, Department of Hepatobiliary Surgery, Xi-Jing Hospital, No. 127 Changle West Road, Xi'an 710032, Shaanxi Province, China. hefei_hefei@163.com

Abstract

Nonalcoholic fatty liver disease (NAFLD) is the most common chronic liver disease worldwide. NAFLD comprises a continuum of liver abnormalities from non-alcoholic fatty liver to nonalcoholic steatohepatitis, and can even lead to cirrhosis and liver cancer. However, a well-established treatment for NAFLD has yet to be identified. Exosomes have become an ideal drug delivery tool because of their high transmissibility, low immunogenicity, easy accessibility and targeting. Exosomes with specific modifications, known as engineered exosomes, have the potential to treat a variety of diseases. Here, we review the treatment of NAFLD with engineered exosomes and the potential use of exosomes as biomarkers and therapeutic targets for NAFLD.

Key Words: Nonalcoholic fatty liver disease; Nonalcoholic steatohepatitis; Exosome; Engineered exosome; Targeted therapy

©The Author(s) 2023. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Nonalcoholic fatty liver disease (NAFLD) is the fastest growing chronic disease in the world. As the disease progresses, NAFLD can lead to liver fibrosis, cirrhosis and even liver cancer. However, a well-established treatment for NAFLD has yet to be identified. Exosomes are small extracellular vesicles secreted by cells. Owing to their high delivery efficiency and biocompatibility, exosomes are expected to become a new means of drug delivery and precise treatment for a variety of diseases, including NAFLD.

Citation: Ding J, Xu C, Xu M, He XY, Li WN, He F. Emerging role of engineered exosomes in nonalcoholic fatty liver disease. *World J Hepatol* 2023; 15(3): 386-392

URL: <https://www.wjgnet.com/1948-5182/full/v15/i3/386.htm>

DOI: <https://dx.doi.org/10.4254/wjh.v15.i3.386>

INTRODUCTION

Nonalcoholic fatty liver disease (NAFLD) is a metabolic disease that is prevalent worldwide affecting at least a quarter of the population[1]. NAFLD is a continuum of liver abnormalities from nonalcoholic fatty liver (NAFL) to nonalcoholic steatohepatitis (NASH) that can even lead to cirrhosis and liver cancer. NAFL is reversible, whereas NASH with cirrhosis is difficult to reverse[2]. Therefore, it is critical to explore the pathogenesis of NAFLD and identify therapeutic targets to treat or prevent its development. Exosomes are extracellular vesicles with a particle size of 30-150 nm that play a crucial role in communication between cells[3]. Some macromolecules such as RNA or proteins in exosomes are associated with the occurrence and development of liver-related diseases and can be used as potential molecular markers in the diagnosis of NAFLD[4]. Processed and modified exosomes (known as engineered exosomes) may also facilitate the study of NAFLD and the development of new therapeutic strategies[5]. In this review, the mechanism and function of engineered exosomes in the development of NAFLD are reviewed (Figure 1).

ENGINEERED EXOSOMES AND LIPID METABOLISM

The liver is the largest metabolic organ and a hub of lipid metabolism. Abnormal changes in lipid metabolism in the liver lead to the development of metabolic diseases[6]. A research team found that the release of exosomes in cultured astrocytes from apolipoprotein E knockout mice was significantly reduced compared to wild-type controls, and a PI3K inhibitor (LY294002) rescued the release of exosomes. They confirmed that the release of exosomes was regulated by cellular cholesterol through stimulation of the PI3K/ Akt signalling pathway[7].

Li *et al*[8] systematically screened for microRNA expression using high-throughput small RNA sequencing and found that miR-199a-5p was significantly upregulated in adipose tissue in a mouse model of high-fat diet (HFD). Further studies confirmed that exosomal miR-199a-5p promoted lipid accumulation in the liver through induction of macrophage stimulating 1 (MST1) expression and fatty acid metabolism. Cheng *et al*[9] found that exosomal miR-627-5p reversed insulin resistance, prevented liver injury, normalized glucose and lipid metabolism and reduced lipid deposition in a rat model of NAFLD.

Brown adipose tissue (BAT) strongly promotes energy expenditure and shows good potential in the treatment of obesity. Zhou *et al*[10] treated HFD-fed mice with engineered exosomes derived from the serum of young healthy mice or from BAT. They found that treatment with BAT exosomes significantly promoted oxygen consumption in recipient cells, thus alleviating metabolic syndrome in HFD-fed mice.

Li *et al*[11] used a low-density lipoprotein receptor-deficient mouse (Ldlr mouse) as a model for hypercholesterolemia. Ldlr mRNA was encapsulated into exosomes by overexpression of Ldlr in donor AML12 mouse hepatocytes. The authors found that engineered exosomes loaded with Ldlr mRNA could restore the expression of Ldlr in the livers of Ldlr-deficient mice and rescue hypercholesterolemia. This study suggests that engineered exosomes may be an effective therapy for patients with hypercholesterolemia.

ENGINEERED EXOSOMES AND INSULIN RESISTANCE

Insulin resistance is now believed to play a key role in the onset and progression of NAFLD[12]. A HFD reduces insulin sensitivity. Kumar *et al*[13] found that feeding a HFD changed the lipid composition of intestinal exosomes. These exosomes were found to be absorbed by macrophages and hepatocytes, resulting in inhibition of the insulin signalling pathway. Castaño *et al*[14] found that obesity can alter the

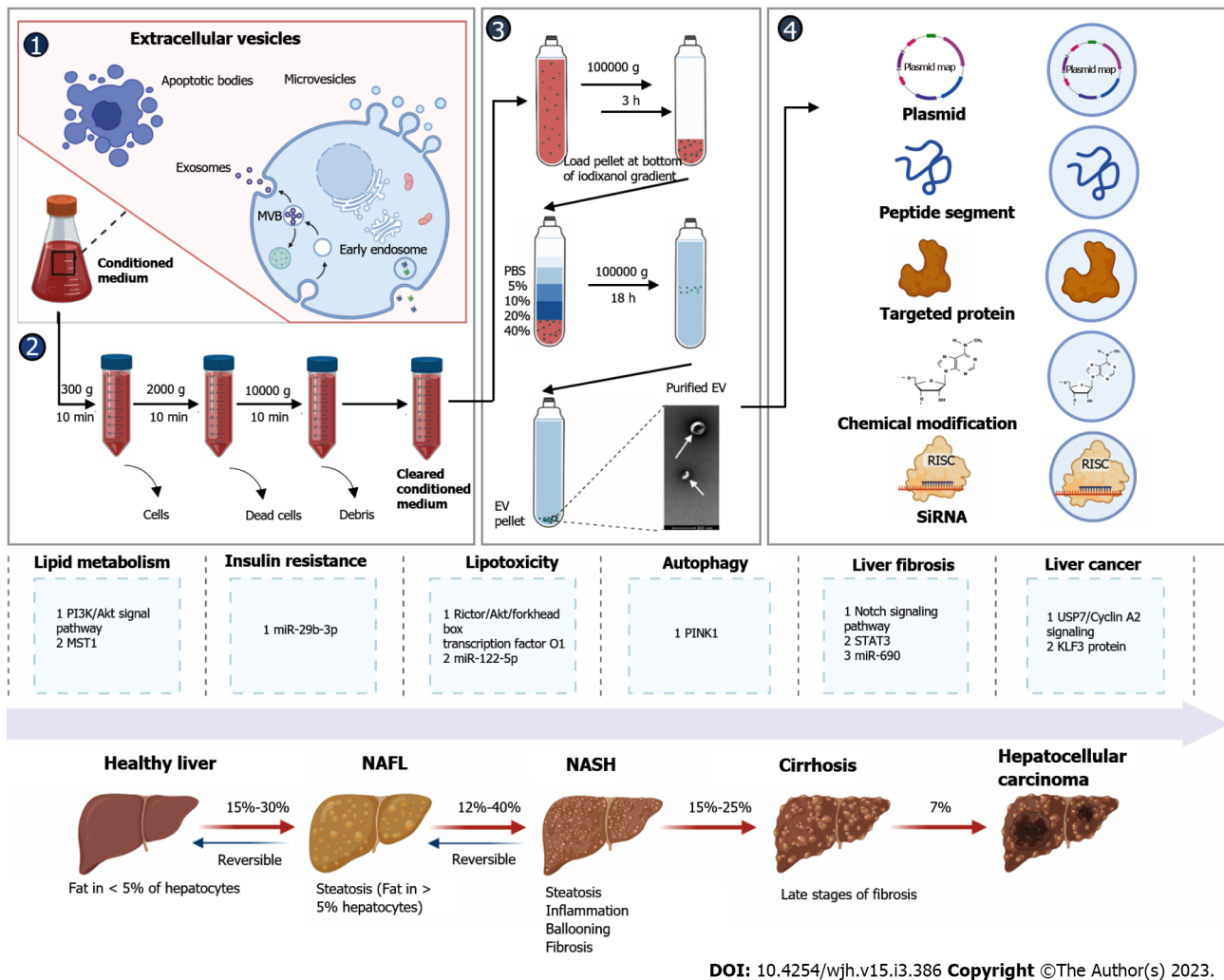


Figure 1 Diagram shows the correlation between nonalcoholic fatty liver disease and engineered exosome. MVB: Multivesicular body; EV: Extracellular vesicles; NASH: Nonalcoholic steatohepatitis; NAFL: Nonalcoholic fatty liver; STAT3: Signal transducer and activator of transcription 3; RISC: RNA-induced silencing complex; MST1: Mammalian STE20-like kinase 1; USP7: Ubiquitin specific peptidase 7; KLF3: Kruppel-like factor 3; PINK: PETN induced kinase 1; PI3K: Phosphatidylinositol-4,5-Bisphosphate 3-Kinase; Akt: Protein kinase B.

expression and composition of miRNAs in mouse plasma exosomes. Ying *et al*[15] found that miR-690, an exosome-derived miRNA from M2-polarized macrophages, improved insulin sensitivity in obese mice. Su *et al*[16] found that exosomes derived from the bone marrow mesenchymal stem cells (BM-MSCs) of aged mice could be ingested by fat, muscle and liver cells, leading to insulin resistance *in vivo* and *in vitro*. The authors found that the amount of miR-29b-3p in exosomes released by BM-MSCs was significantly increased in aged mice. Furthermore, they found that inhibition of miR-29b-3p with an aptamer-mediated nanocomposite delivery system improved insulin resistance in aged mice.

ENGINEERED EXOSOMES AND LIPOTOXICITY

Lipotoxicity promotes proinflammatory M1 polarization of liver macrophages during the development of NAFLD[17,18]. Liu *et al*[19] found that miR-192-5p-rich hepatocyte-exosomes induced by lipotoxic injury promoted macrophage M1 polarization and liver inflammation through Rictor/Akt/forkhead box transcription factor O1 signalling. Zhao *et al*[20] found that cholesterol-induced lysosomal dysfunction increased exosome release from hepatocytes, leading to M1 polarization and macrophage-induced inflammation in a miR-122-5p-dependent manner. Human umbilical cord mesenchymal stem cells (HUC-MSCs) are increasingly being studied in clinical trials of end-stage liver disease due to their excellent tissue repair and anti-inflammatory effects. Shi *et al*[21] found that HUC-MSC-derived exosomes could protect against methionine- and choline-deficient L-amino acid diet (MCD)-induced NASH.

Lipotoxicity can damage mitochondria and induce oxidative stress during the progression of NAFLD [22,23]. Studies have shown that adipocytes respond to mitochondrial stress by rapidly and vigorously

releasing exosomes[24]. Similarly, exosomes derived from chemically induced human hepatic progenitors inhibit cell death induced by oxidative stress[25].

ENGINEERED EXOSOMES AND AUTOPHAGY

Autophagy is a process in which cells degrade and metabolize their own damaged organelles or protein aggregates that plays a key role in maintaining liver homeostasis[26]. Increasing evidence suggests that autophagy plays a very important role in lipid metabolism. Autophagy mainly protects cells and regulates inflammation in NAFLD[26]. Because autophagy and exosomal biogenesis share common elements, some studies have found that plasma exosomal levels are higher in NAFLD patients than in healthy controls[27]. Luo *et al*[28] found that miR-27a inhibited mitochondrial autophagy and promoted NAFLD-associated liver fibrosis by negatively regulating PINK1 expression *via* lipotoxic hepatocyte exosomes. A research team established a model of hepatocyte injury and apoptosis induced by D-galactosamine and lipopolysaccharide (D-GalN/LPS) to study the protective effect of bone marrow mesenchymal stem cell (BMSC)-derived exosomes on liver injury. They found that BMSC-derived exosomes attenuated D-GalN/LPS-induced hepatocyte apoptosis by activating autophagy *in vitro*[29]. Similar studies have shown that upregulation of miR-96-5p in BMSCs and their exosomes ameliorated NASH *via* caspase-2[30].

ENGINEERED EXOSOMES AND LIVER FIBROSIS

It is generally believed that during the development of NAFLD, liver-related cells are replaced by fibrotic scar tissue, giving rise to liver fibrosis or cirrhosis, which are associated with poor prognosis and mortality in patients with NASH[2]. The Notch signalling pathway is a key mediator of cellular differentiation, proliferation and apoptosis[31]. We designed hairpin-type decoy oligodeoxynucleotides (ODNs) for RBP-J to inhibit the activation of Notch signalling. ODNs were loaded into HEK293T-derived exosomes by electroporation. Furthermore, we observed that tail vein-injected exosomes were mainly taken up by hepatic macrophages in mice with hepatic fibrosis. RBP-J decoy ODNs delivered by exosomes efficiently inhibited Notch signalling in macrophages and ameliorated liver fibrosis in mice [32].

Hou *et al*[33] found that myeloid cell-specific IL-6 signalling promoted miR-223-enriched exosome production and attenuated NAFLD-associated fibrosis. Tang *et al*[34] found that exosomes embedded with siRNAs or antisense oligonucleotides targeting signal transducer and activator of transcription 3 (STAT3) could attenuate liver fibrosis. Gao *et al*[35] showed that Kupffer cells produced endogenous miR-690 and shuttled this miRNA to other hepatocytes through exosomal secretion. Treatment with miR-690 inhibitors reduced fibrosis and steatosis in a NASH model. Wang *et al*[36] found that miR-6766-3p-rich 3D human embryonic stem cell (hESC) exosomes could ameliorate liver fibrosis by targeting the TGF β RII-SMADS pathway in hepatic stellate cells. Ji *et al*[37] developed an exosome-liposome hybrid loaded with clodronate-nintedanib that impaired hepatic fibrosis by reducing the activation of Kupffer cells.

CRISPR-Cas9 gene editing has become a powerful therapeutic technology. However, there is a lack of safe and effective *in vivo* delivery systems for CRISPR-Cas9, especially for tissue-specific vectors[38]. Luo *et al*[39] used exosome-mediated CRISPR/dCas9-VP64 delivery to reprogram hepatic stellate cells to construct engineered exosomes for the treatment of liver fibrosis. Similarly, Wan *et al*[40] delivered exosome-mediated Cas9 ribonucleoprotein complexes for tissue-specific gene therapy in liver disease.

ENGINEERED EXOSOMES AND LIVER CANCER

Without timely intervention, NAFLD inevitably results in liver cancer[41]. Liver cancer is the fourth leading cause of cancer-related death worldwide and occurs in patients with various chronic liver diseases[42]. To date, the exact pathogenesis of NAFLD-induced liver cancer is not fully understood, but may involve DNA damage responses, inflammation, autophagy, and disruption of the gut microbiota [41].

Adipose tissue is known to play a role in energy storage and metabolic regulation by secreting adipokines[43]. Studies have demonstrated that exosomal circRNA secreted by adipocytes promotes tumour growth by inhibiting miR-34a and activating the USP7/Cyclin A2 signalling pathway[44].

An acidic microenvironment has been shown to promote the release of exosomes, which are considered to be cell-to-cell communication agents involved in cancer progression and metastasis[45]. Tian *et al*[46] found that exosomal miR-21 and miR-10b induced by the acidic microenvironment in liver cancer could promote cancer cell proliferation and metastasis and be used as prognostic molecular markers and therapeutic targets for liver cancer.

Macrophage-derived exosomes play multiple roles in cancer initiation and progression[47]. Zhang *et al*[48] found that exosomes derived from RBP-J overexpressing macrophages inhibited the progression of liver cancer by miR-499b-5p/JAM3. M2 macrophages can influence tumour development by secreting various cytokines, including exosomes. Some studies suggest that M2 macrophage-derived exosomes modified by miR-660-5p-related oligonucleotides enhanced the development of hepatocellular carcinoma by regulating KLF3[49].

ENGINEERED EXOSOMES INVOLVED IN THE DIAGNOSIS OF NAFLD

Exosomes can be derived from healthy and stressed cells to provide a snapshot of the cell of origin under physiological and pathological conditions. Hepatocyte-derived exosomes released from stressed/injured hepatocytes have been identified as a partial cause of liver disease progression and liver injury, so circulating exosomes may serve as biomarkers of NAFLD. Nanoparticle-enhanced scattering of gold nanoparticles coupled with hepatocyte-specific antibodies was used to identify hepatocyte-derived exosomes[50]. Furthermore, microarray analysis of exosomal miRNAs isolated from the serum of 41 patients with NAFLD (diagnosed using liver biopsy) suggested that serum exosomal miRNAs could be used to assess the severity of NAFLD and identify potential targets for NAFLD treatment[33]. One of the determinants of liver degeneration in the progression of NAFLD is Wnt/frizzled (FZD) signalling; for example, FZD7 delivered by plasma-derived exosomes is a good candidate for a novel and effective biomarker for the diagnosis and prognosis of NAFLD[51].

CONCLUSION

The incidence of NAFLD is rapidly increasing with changes in lifestyle and dietary habits[1]. Exosomes not only mediate communication between cells but can also be engineered to deliver specific substances. Engineered exosomes have shown some effects on NAFLD in animal experiments. Owing to their low immunogenicity and liver targeting[52,53], engineered exosomes have great potential to treat NAFLD.

FOOTNOTES

Author contributions: Ding J contributed to writing the original draft; Xu C contributed to picture making; Xu M, He XY, and Li WN contributed to data collection; He F contributed to designed the review and revised the final version.

Supported by National Natural Science Foundation of China, No. 81970535.

Conflict-of-interest statement: All the authors declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <https://creativecommons.org/licenses/by-nc/4.0/>

Country/Territory of origin: China

ORCID number: Fei He 0000-0001-8368-5030.

S-Editor: Liu JH

L-Editor: A

P-Editor: Liu JH

REFERENCES

- 1 Younossi Z, Anstee QM, Marietti M, Hardy T, Henry L, Eslam M, George J, Bugianesi E. Global burden of NAFLD and NASH: trends, predictions, risk factors and prevention. *Nat Rev Gastroenterol Hepatol* 2018; **15**: 11-20 [PMID: 28930295 DOI: 10.1038/nrgastro.2017.109]
- 2 Powell EE, Wong VW, Rinella M. Non-alcoholic fatty liver disease. *Lancet* 2021; **397**: 2212-2224 [PMID: 33894145 DOI: 10.1016/S0140-6736(20)32511-3]
- 3 Kalluri R, LeBleu VS. The biology, function, and biomedical applications of exosomes. *Science* 2020; **367** [PMID: 32015840 DOI: 10.1126/science.1262130]

- 32029601 DOI: [10.1126/science.aau6977](https://doi.org/10.1126/science.aau6977)]
- 4 **Wang W**, Zhu N, Yan T, Shi YN, Chen J, Zhang CJ, Xie XJ, Liao DF, Qin L. The crosstalk: exosomes and lipid metabolism. *Cell Commun Signal* 2020; **18**: 119 [PMID: [32746850](https://pubmed.ncbi.nlm.nih.gov/32746850/) DOI: [10.1186/s12964-020-00581-2](https://doi.org/10.1186/s12964-020-00581-2)]
 - 5 **Nakao Y**, Amrollahi P, Parthasarathy G, Mauer AS, Schrawat TS, Vanderboom P, Nair KS, Nakao K, Allen AM, Hu TY, Malhi H. Circulating extracellular vesicles are a biomarker for NAFLD resolution and response to weight loss surgery. *Nanomedicine* 2021; **36**: 102430 [PMID: [34174416](https://pubmed.ncbi.nlm.nih.gov/34174416/) DOI: [10.1016/j.nano.2021.102430](https://doi.org/10.1016/j.nano.2021.102430)]
 - 6 **Kojta I**, Chacińska M, Błachnio-Zabielska A. Obesity, Bioactive Lipids, and Adipose Tissue Inflammation in Insulin Resistance. *Nutrients* 2020; **12** [PMID: [32375231](https://pubmed.ncbi.nlm.nih.gov/32375231/) DOI: [10.3390/nu12051305](https://doi.org/10.3390/nu12051305)]
 - 7 **Abdullah M**, Nakamura T, Ferdous T, Gao Y, Chen Y, Zou K, Michikawa M. Cholesterol Regulates Exosome Release in Cultured Astrocytes. *Front Immunol* 2021; **12**: 722581 [PMID: [34721384](https://pubmed.ncbi.nlm.nih.gov/34721384/) DOI: [10.3389/fimmu.2021.722581](https://doi.org/10.3389/fimmu.2021.722581)]
 - 8 **Li Y**, Luan Y, Li J, Song H, Li Y, Qi H, Sun B, Zhang P, Wu X, Liu X, Yang Y, Tao W, Cai L, Yang Z. Exosomal miR-199a-5p promotes hepatic lipid accumulation by modulating MST1 expression and fatty acid metabolism. *Hepatol Int* 2020; **14**: 1057-1074 [PMID: [33037981](https://pubmed.ncbi.nlm.nih.gov/33037981/) DOI: [10.1007/s12072-020-10096-0](https://doi.org/10.1007/s12072-020-10096-0)]
 - 9 **Cheng D**, Chai J, Wang H, Fu L, Peng S, Ni X. Hepatic macrophages: Key players in the development and progression of liver fibrosis. *Liver Int* 2021; **41**: 2279-2294 [PMID: [33966318](https://pubmed.ncbi.nlm.nih.gov/33966318/) DOI: [10.1111/liv.14940](https://doi.org/10.1111/liv.14940)]
 - 10 **Zhou X**, Li Z, Qi M, Zhao P, Duan Y, Yang G, Yuan L. Brown adipose tissue-derived exosomes mitigate the metabolic syndrome in high fat diet mice. *Theranostics* 2020; **10**: 8197-8210 [PMID: [32724466](https://pubmed.ncbi.nlm.nih.gov/32724466/) DOI: [10.7150/thno.43968](https://doi.org/10.7150/thno.43968)]
 - 11 **Li Z**, Zhao P, Zhang Y, Wang J, Wang C, Liu Y, Yang G, Yuan L. Exosome-based Ldlr gene therapy for familial hypercholesterolemia in a mouse model. *Theranostics* 2021; **11**: 2953-2965 [PMID: [33456582](https://pubmed.ncbi.nlm.nih.gov/33456582/) DOI: [10.7150/thno.49874](https://doi.org/10.7150/thno.49874)]
 - 12 **Watt MJ**, Miotto PM, De Nardo W, Montgomery MK. The Liver as an Endocrine Organ-Linking NAFLD and Insulin Resistance. *Endocr Rev* 2019; **40**: 1367-1393 [PMID: [31098621](https://pubmed.ncbi.nlm.nih.gov/31098621/) DOI: [10.1210/er.2019-00034](https://doi.org/10.1210/er.2019-00034)]
 - 13 **Kumar A**, Sundaram K, Mu J, Dryden GW, Sriwastva MK, Lei C, Zhang L, Qiu X, Xu F, Yan J, Zhang X, Park JW, Merchant ML, Bohler HCL, Wang B, Zhang S, Qin C, Xu Z, Han X, McClain CJ, Teng Y, Zhang HG. High-fat diet-induced upregulation of exosomal phosphatidylcholine contributes to insulin resistance. *Nat Commun* 2021; **12**: 213 [PMID: [33431899](https://pubmed.ncbi.nlm.nih.gov/33431899/) DOI: [10.1038/s41467-020-20500-w](https://doi.org/10.1038/s41467-020-20500-w)]
 - 14 **Castaño C**, Kalko S, Novials A, Párrizas M. Obesity-associated exosomal miRNAs modulate glucose and lipid metabolism in mice. *Proc Natl Acad Sci U S A* 2018; **115**: 12158-12163 [PMID: [30429322](https://pubmed.ncbi.nlm.nih.gov/30429322/) DOI: [10.1073/pnas.1808855115](https://doi.org/10.1073/pnas.1808855115)]
 - 15 **Ying W**, Gao H, Dos Reis FCG, Bandyopadhyay G, Ofrecio JM, Luo Z, Ji Y, Jin Z, Ly C, Olefsky JM. MiR-690, an exosomal-derived miRNA from M2-polarized macrophages, improves insulin sensitivity in obese mice. *Cell Metab* 2021; **33**: 781-790.e5 [PMID: [33450179](https://pubmed.ncbi.nlm.nih.gov/33450179/) DOI: [10.1016/j.cmet.2020.12.019](https://doi.org/10.1016/j.cmet.2020.12.019)]
 - 16 **Su T**, Xiao Y, Guo Q, Li C, Huang Y, Deng Q, Wen J, Zhou F, Luo XH. Bone Marrow Mesenchymal Stem Cells-Derived Exosomal MiR-29b-3p Regulates Aging-Associated Insulin Resistance. *ACS Nano* 2019; **13**: 2450-2462 [PMID: [30715852](https://pubmed.ncbi.nlm.nih.gov/30715852/) DOI: [10.1021/acsnano.8b09375](https://doi.org/10.1021/acsnano.8b09375)]
 - 17 **Schuster S**, Cabrera D, Arrese M, Feldstein AE. Triggering and resolution of inflammation in NASH. *Nat Rev Gastroenterol Hepatol* 2018; **15**: 349-364 [PMID: [29740166](https://pubmed.ncbi.nlm.nih.gov/29740166/) DOI: [10.1038/s41575-018-0009-6](https://doi.org/10.1038/s41575-018-0009-6)]
 - 18 **Kazankov K**, Jørgensen SMD, Thomsen KL, Møller HJ, Vilstrup H, George J, Schuppan D, Grønbaek H. The role of macrophages in nonalcoholic fatty liver disease and nonalcoholic steatohepatitis. *Nat Rev Gastroenterol Hepatol* 2019; **16**: 145-159 [PMID: [30482910](https://pubmed.ncbi.nlm.nih.gov/30482910/) DOI: [10.1038/s41575-018-0082-x](https://doi.org/10.1038/s41575-018-0082-x)]
 - 19 **Liu XL**, Pan Q, Cao HX, Xin FZ, Zhao ZH, Yang RX, Zeng J, Zhou H, Fan JG. Lipotoxic Hepatocyte-Derived Exosomal MicroRNA 192-5p Activates Macrophages Through Rictor/Akt/Forkhead Box Transcription Factor O1 Signaling in Nonalcoholic Fatty Liver Disease. *Hepatology* 2020; **72**: 454-469 [PMID: [31782176](https://pubmed.ncbi.nlm.nih.gov/31782176/) DOI: [10.1002/hep.31050](https://doi.org/10.1002/hep.31050)]
 - 20 **Zhao Z**, Zhong L, Li P, He K, Qiu C, Zhao L, Gong J. Cholesterol impairs hepatocyte lysosomal function causing M1 polarization of macrophages via exosomal miR-122-5p. *Exp Cell Res* 2020; **387**: 111738 [PMID: [31759057](https://pubmed.ncbi.nlm.nih.gov/31759057/) DOI: [10.1016/j.yexcr.2019.111738](https://doi.org/10.1016/j.yexcr.2019.111738)]
 - 21 **Shi Y**, Yang X, Wang S, Wu Y, Zheng L, Tang Y, Gao Y, Niu J. Human umbilical cord mesenchymal stromal cell-derived exosomes protect against MCD-induced NASH in a mouse model. *Stem Cell Res Ther* 2022; **13**: 517 [PMID: [36371344](https://pubmed.ncbi.nlm.nih.gov/36371344/) DOI: [10.1186/s13287-022-03201-7](https://doi.org/10.1186/s13287-022-03201-7)]
 - 22 **Paradies G**, Paradies V, Ruggiero FM, Petrosillo G. Oxidative stress, cardiolipin and mitochondrial dysfunction in nonalcoholic fatty liver disease. *World J Gastroenterol* 2014; **20**: 14205-14218 [PMID: [25339807](https://pubmed.ncbi.nlm.nih.gov/25339807/) DOI: [10.3748/wjg.v20.i39.14205](https://doi.org/10.3748/wjg.v20.i39.14205)]
 - 23 **Dabravolski SA**, Bezsonov EE, Orekhov AN. The role of mitochondria dysfunction and hepatic senescence in NAFLD development and progression. *Biomed Pharmacother* 2021; **142**: 112041 [PMID: [34411916](https://pubmed.ncbi.nlm.nih.gov/34411916/) DOI: [10.1016/j.biopha.2021.112041](https://doi.org/10.1016/j.biopha.2021.112041)]
 - 24 **Crewe C**, Funcke JB, Li S, Joffin N, Gliniak CM, Ghaben AL, An YA, Sadek HA, Gordillo R, Akgul Y, Chen S, Samovski D, Fischer-Posovszky P, Kusminski CM, Klein S, Scherer PE. Extracellular vesicle-based interorgan transport of mitochondria from energetically stressed adipocytes. *Cell Metab* 2021; **33**: 1853-1868.e11 [PMID: [34418352](https://pubmed.ncbi.nlm.nih.gov/34418352/) DOI: [10.1016/j.cmet.2021.08.002](https://doi.org/10.1016/j.cmet.2021.08.002)]
 - 25 **Hyung S**, Jeong J, Shin K, Kim JY, Yim JH, Yu CJ, Jung HS, Hwang KG, Choi D, Hong JW. Exosomes derived from chemically induced human hepatic progenitors inhibit oxidative stress induced cell death. *Biotechnol Bioeng* 2020; **117**: 2658-2667 [PMID: [32484909](https://pubmed.ncbi.nlm.nih.gov/32484909/) DOI: [10.1002/bit.27447](https://doi.org/10.1002/bit.27447)]
 - 26 **Filali-Mouncef Y**, Hunter C, Roccio F, Zagkou S, Dupont N, Primard C, Proikas-Cezanne T, Reggiori F. The ménage à trois of autophagy, lipid droplets and liver disease. *Autophagy* 2022; **18**: 50-72 [PMID: [33794741](https://pubmed.ncbi.nlm.nih.gov/33794741/) DOI: [10.1080/15548627.2021.1895658](https://doi.org/10.1080/15548627.2021.1895658)]
 - 27 **Zhang J**, Tan J, Wang M, Wang Y, Dong M, Ma X, Sun B, Liu S, Zhao Z, Chen L, Liu K, Xin Y, Zhuang L. Lipid-induced DRAM recruits STOM to lysosomes and induces LMP to promote exosome release from hepatocytes in NAFLD. *Sci Adv* 2021; **7**: eabh1541 [PMID: [34731006](https://pubmed.ncbi.nlm.nih.gov/34731006/) DOI: [10.1126/sciadv.abh1541](https://doi.org/10.1126/sciadv.abh1541)]
 - 28 **Luo X**, Xu ZX, Wu JC, Luo SZ, Xu MY. Hepatocyte-derived exosomal miR-27a activates hepatic stellate cells through the inhibition of PINK1-mediated mitophagy in MAFLD. *Mol Ther Nucleic Acids* 2021; **26**: 1241-1254 [PMID: [34853724](https://pubmed.ncbi.nlm.nih.gov/34853724/)]

- DOI: [10.1016/j.omtn.2021.10.022](https://doi.org/10.1016/j.omtn.2021.10.022)]
- 29 **Zhao S**, Liu Y, Pu Z. Bone marrow mesenchymal stem cell-derived exosomes attenuate D-GaIN/LPS-induced hepatocyte apoptosis by activating autophagy in vitro. *Drug Des Devel Ther* 2019; **13**: 2887-2897 [PMID: [31695322](https://pubmed.ncbi.nlm.nih.gov/31695322/) DOI: [10.2147/DDDT.S220190](https://doi.org/10.2147/DDDT.S220190)]
- 30 **El-Derany MO**, AbdelHamid SG. Upregulation of miR-96-5p by bone marrow mesenchymal stem cells and their exosomes alleviate non-alcoholic steatohepatitis: Emphasis on caspase-2 signaling inhibition. *Biochem Pharmacol* 2021; **190**: 114624 [PMID: [34052187](https://pubmed.ncbi.nlm.nih.gov/34052187/) DOI: [10.1016/j.bcp.2021.114624](https://doi.org/10.1016/j.bcp.2021.114624)]
- 31 **Xu H**, Wang L. The Role of Notch Signaling Pathway in Non-Alcoholic Fatty Liver Disease. *Front Mol Biosci* 2021; **8**: 792667 [PMID: [34901163](https://pubmed.ncbi.nlm.nih.gov/34901163/) DOI: [10.3389/fmolb.2021.792667](https://doi.org/10.3389/fmolb.2021.792667)]
- 32 **He F**, Li WN, Li XX, Yue KY, Duan JL, Ruan B, Liu JJ, Song P, Yue ZS, Tao KS, Wang L. Exosome-mediated delivery of RBP-J decoy oligodeoxynucleotides ameliorates hepatic fibrosis in mice. *Theranostics* 2022; **12**: 1816-1828 [PMID: [35198075](https://pubmed.ncbi.nlm.nih.gov/35198075/) DOI: [10.7150/thno.69885](https://doi.org/10.7150/thno.69885)]
- 33 **Hou X**, Yin S, Ren R, Liu S, Yong L, Liu Y, Li Y, Zheng MH, Kunos G, Gao B, Wang H. Myeloid-Cell-Specific IL-6 Signaling Promotes MicroRNA-223-Enriched Exosome Production to Attenuate NAFLD-Associated Fibrosis. *Hepatology* 2021; **74**: 116-132 [PMID: [33236445](https://pubmed.ncbi.nlm.nih.gov/33236445/) DOI: [10.1002/hep.31658](https://doi.org/10.1002/hep.31658)]
- 34 **Tang M**, Chen Y, Li B, Sugimoto H, Yang S, Yang C, LeBleu VS, McAndrews KM, Kalluri R. Therapeutic targeting of STAT3 with small interference RNAs and antisense oligonucleotides embedded exosomes in liver fibrosis. *FASEB J* 2021; **35**: e21557 [PMID: [33855751](https://pubmed.ncbi.nlm.nih.gov/33855751/) DOI: [10.1096/fj.202002777RR](https://doi.org/10.1096/fj.202002777RR)]
- 35 **Gao H**, Jin Z, Bandyopadhyay G, Cunha E Rocha K, Liu X, Zhao H, Zhang D, Jouihan H, Pourshahian S, Kisseleva T, Brenner DA, Ying W, Olefsky JM. MiR-690 treatment causes decreased fibrosis and steatosis and restores specific Kupffer cell functions in NASH. *Cell Metab* 2022; **34**: 978-990.e4 [PMID: [35700738](https://pubmed.ncbi.nlm.nih.gov/35700738/) DOI: [10.1016/j.cmet.2022.05.008](https://doi.org/10.1016/j.cmet.2022.05.008)]
- 36 **Wang N**, Li X, Zhong Z, Qiu Y, Liu S, Wu H, Tang X, Chen C, Fu Y, Chen Q, Guo T, Li J, Zhang S, Zern MA, Ma K, Wang B, Ou Y, Gu W, Cao J, Chen H, Duan Y. 3D hESC exosomes enriched with miR-6766-3p ameliorates liver fibrosis by attenuating activated stellate cells through targeting the TGFβRII-SMADS pathway. *J Nanobiotechnology* 2021; **19**: 437 [PMID: [34930304](https://pubmed.ncbi.nlm.nih.gov/34930304/) DOI: [10.1186/s12951-021-01138-2](https://doi.org/10.1186/s12951-021-01138-2)]
- 37 **Ji K**, Fan M, Huang D, Sun L, Li B, Xu R, Zhang J, Shao X, Chen Y. Clodronate-nintedanib-loaded exosome-liposome hybridization enhances the liver fibrosis therapy by inhibiting Kupffer cell activity. *Biomater Sci* 2022; **10**: 702-713 [PMID: [34927632](https://pubmed.ncbi.nlm.nih.gov/34927632/) DOI: [10.1039/d1bm01663f](https://doi.org/10.1039/d1bm01663f)]
- 38 **Sharma G**, Sharma AR, Bhattacharya M, Lee SS, Chakraborty C. CRISPR-Cas9: A Preclinical and Clinical Perspective for the Treatment of Human Diseases. *Mol Ther* 2021; **29**: 571-586 [PMID: [33238136](https://pubmed.ncbi.nlm.nih.gov/33238136/) DOI: [10.1016/j.ymthe.2020.09.028](https://doi.org/10.1016/j.ymthe.2020.09.028)]
- 39 **Luo N**, Li J, Chen Y, Xu Y, Wei Y, Lu J, Dong R. Hepatic stellate cell reprogramming via exosome-mediated CRISPR/dCas9-VP64 delivery. *Drug Deliv* 2021; **28**: 10-18 [PMID: [33336604](https://pubmed.ncbi.nlm.nih.gov/33336604/) DOI: [10.1080/10717544.2020.1850917](https://doi.org/10.1080/10717544.2020.1850917)]
- 40 **Wan T**, Zhong J, Pan Q, Zhou T, Ping Y, Liu X. Exosome-mediated delivery of Cas9 ribonucleoprotein complexes for tissue-specific gene therapy of liver diseases. *Sci Adv* 2022; **8**: eabp9435 [PMID: [36103526](https://pubmed.ncbi.nlm.nih.gov/36103526/) DOI: [10.1126/sciadv.abp9435](https://doi.org/10.1126/sciadv.abp9435)]
- 41 **Ioannou GN**. Epidemiology and risk-stratification of NAFLD-associated HCC. *J Hepatol* 2021; **75**: 1476-1484 [PMID: [34453963](https://pubmed.ncbi.nlm.nih.gov/34453963/) DOI: [10.1016/j.jhep.2021.08.012](https://doi.org/10.1016/j.jhep.2021.08.012)]
- 42 **Anwanwan D**, Singh SK, Singh S, Saikam V, Singh R. Challenges in liver cancer and possible treatment approaches. *Biochim Biophys Acta Rev Cancer* 2020; **1873**: 188314 [PMID: [31682895](https://pubmed.ncbi.nlm.nih.gov/31682895/) DOI: [10.1016/j.bbcan.2019.188314](https://doi.org/10.1016/j.bbcan.2019.188314)]
- 43 **Goossens GH**. The Metabolic Phenotype in Obesity: Fat Mass, Body Fat Distribution, and Adipose Tissue Function. *Obes Facts* 2017; **10**: 207-215 [PMID: [28564650](https://pubmed.ncbi.nlm.nih.gov/28564650/) DOI: [10.1159/000471488](https://doi.org/10.1159/000471488)]
- 44 **Zhang H**, Deng T, Ge S, Liu Y, Bai M, Zhu K, Fan Q, Li J, Ning T, Tian F, Li H, Sun W, Ying G, Ba Y. Exosome circRNA secreted from adipocytes promotes the growth of hepatocellular carcinoma by targeting deubiquitination-related USP7. *Oncogene* 2019; **38**: 2844-2859 [PMID: [30546088](https://pubmed.ncbi.nlm.nih.gov/30546088/) DOI: [10.1038/s41388-018-0619-z](https://doi.org/10.1038/s41388-018-0619-z)]
- 45 **Meng W**, Hao Y, He C, Li L, Zhu G. Exosome-orchestrated hypoxic tumor microenvironment. *Mol Cancer* 2019; **18**: 57 [PMID: [30925935](https://pubmed.ncbi.nlm.nih.gov/30925935/) DOI: [10.1186/s12943-019-0982-6](https://doi.org/10.1186/s12943-019-0982-6)]
- 46 **Tian XP**, Wang CY, Jin XH, Li M, Wang FW, Huang WJ, Yun JP, Xu RH, Cai QQ, Xie D. Acidic Microenvironment Up-Regulates Exosomal miR-21 and miR-10b in Early-Stage Hepatocellular Carcinoma to Promote Cancer Cell Proliferation and Metastasis. *Theranostics* 2019; **9**: 1965-1979 [PMID: [31037150](https://pubmed.ncbi.nlm.nih.gov/31037150/) DOI: [10.7150/thno.30958](https://doi.org/10.7150/thno.30958)]
- 47 **Xia Y**, Rao L, Yao H, Wang Z, Ning P, Chen X. Engineering Macrophages for Cancer Immunotherapy and Drug Delivery. *Adv Mater* 2020; **32**: e2002054 [PMID: [32856350](https://pubmed.ncbi.nlm.nih.gov/32856350/) DOI: [10.1002/adma.202002054](https://doi.org/10.1002/adma.202002054)]
- 48 **Zhang L**, Zhang J, Li P, Li T, Zhou Z, Wu H. Exosomal hsa_circ_0004658 derived from RBPJ overexpressed-macrophages inhibits hepatocellular carcinoma progression via miR-499b-5p/JAM3. *Cell Death Dis* 2022; **13**: 32 [PMID: [35013102](https://pubmed.ncbi.nlm.nih.gov/35013102/) DOI: [10.1038/s41419-021-04345-9](https://doi.org/10.1038/s41419-021-04345-9)]
- 49 **Tian B**, Zhou L, Wang J, Yang P. miR-660-5p-loaded M2 macrophages-derived exosomes augment hepatocellular carcinoma development through regulating KLF3. *Int Immunopharmacol* 2021; **101**: 108157 [PMID: [34673296](https://pubmed.ncbi.nlm.nih.gov/34673296/) DOI: [10.1016/j.intimp.2021.108157](https://doi.org/10.1016/j.intimp.2021.108157)]
- 50 **Nguyen HQ**, Lee D, Kim Y, Bang G, Cho K, Lee YS, Yeon JE, Lubman DM, Kim J. Label-free quantitative proteomic analysis of serum extracellular vesicles differentiating patients of alcoholic and nonalcoholic fatty liver diseases. *J Proteomics* 2021; **245**: 104278 [PMID: [34089894](https://pubmed.ncbi.nlm.nih.gov/34089894/) DOI: [10.1016/j.jprot.2021.104278](https://doi.org/10.1016/j.jprot.2021.104278)]
- 51 **Scavo MP**, Depalo N, Rizzi F, Carrieri L, Serino G, Franco I, Bonfiglio C, Pesole PL, Cozzolongo R, Gianuzzi V, Curri ML, Osella AR, Giannelli G. Exosomal FZD-7 Expression Is Modulated by Different Lifestyle Interventions in Patients with NAFLD. *Nutrients* 2022; **14** [PMID: [35334792](https://pubmed.ncbi.nlm.nih.gov/35334792/) DOI: [10.3390/nu14061133](https://doi.org/10.3390/nu14061133)]
- 52 **Barile L**, Vassalli G. Exosomes: Therapy delivery tools and biomarkers of diseases. *Pharmacol Ther* 2017; **174**: 63-78 [PMID: [28202367](https://pubmed.ncbi.nlm.nih.gov/28202367/) DOI: [10.1016/j.pharmthera.2017.02.020](https://doi.org/10.1016/j.pharmthera.2017.02.020)]
- 53 **Zhang G**, Huang X, Xiu H, Sun Y, Chen J, Cheng G, Song Z, Peng Y, Shen Y, Wang J, Cai Z. Extracellular vesicles: Natural liver-accumulating drug delivery vehicles for the treatment of liver diseases. *J Extracell Vesicles* 2020; **10**: e12030 [PMID: [33335695](https://pubmed.ncbi.nlm.nih.gov/33335695/) DOI: [10.1002/jev2.12030](https://doi.org/10.1002/jev2.12030)]



Published by **Baishideng Publishing Group Inc**
7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA

Telephone: +1-925-3991568

E-mail: bpgoffice@wjgnet.com

Help Desk: <https://www.f6publishing.com/helpdesk>

<https://www.wjgnet.com>

