

Exploring new frontiers in molecular imaging: Emergence of ^{68}Ga PET/CT

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Author contributions: Tan EH is the main author; Goh SW assisted in manuscript review.

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Received: January 26, 2010 Revised: February 4, 2010

Accepted: February 22, 2010

Published online: February 28, 2010

Abstract

Since US Food and Drug Administration approval of 18-fluorodeoxyglucose as a positron tracer, and the development of hybrid positron emission tomography/computed tomography machines, there has been a great increase in clinical application and progress in the field of nuclear molecular imaging. However, not underestimating the value of ^{18}F , there are known limitations in the use of this cyclotron-produced positron tracer. We hence turn our focus to an emerging positron tracer, ^{68}Ga , and examine the advantages, current clinical uses and potential future applications of this radioisotope.

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Key words: Gallium 68; Positron emission tomography; computed tomography; Neuroendocrinology

Peer reviewer: George C Kagadis, Assistant Professor, Department of Medical Physics, School of Medicine University of Patras, GR 265 04, Rion, Greece

Tan EH, Goh SW. Exploring new frontiers in molecular imaging: Emergence of ^{68}Ga PET/CT. *World J Radiol* 2010; 2(2): 55-67 Available from: URL: <http://www.wjgnet.com/1949-8470/full/v2/i2/55.htm> DOI: <http://dx.doi.org/10.4329/wjr.v2.i2.55>

THE DEVELOPMENT OF POSITRON EMISSION TOMOGRAPHY

Positron emission tomography (PET) imaging is essentially a story of a technique in wait of a technology. Since the discovery of positron emission in 1933 by Thibaud and Joliot *et al*, and the subsequent report of the coincident nature of emissions by Klempner and Beringer, it has in effect taken close to half a century for the full realization of PET imaging in mainstream medical practice^[1].

The history of PET development is a fascinating look into the technological advances in molecular medicine, and the account of Terry Jones provides fascinating reading^[1].

The first use of positron tracers was likely performed in the 1940s using ^{11}C in animal models^[2], with its first possible use in humans performed in the 1950s at the Hammersmith Hospital in London, United Kingdom using $^{15}\text{O}_2$ in studies of lung ventilation^[3]. This was followed by increasing use of positron tracers in the physiological assessment of lung function, which resulted in the installation of the world's first hospital-based cyclotron in 1955. Subsequently, development shifted into myocardial perfusion^[4], cerebral perfusion^[5-7], and of course, glucose metabolism^[8-11].

This development in positron tracers mirrored the progress in positron imaging. In the 1970s, Massachusetts General Hospital developed, what was then, the most advanced coincidence positron camera system, with a spatial resolution of approximately $1\text{ cm}^{[12]}$. This was followed by developments predominantly in single photon emission computed tomography (SPECT) with work done by Kuhl, Budinger and Gullberg, and in 1974, there were reports of the development of a dedicated single-plane positron emission transaxial tomograph, which was the precursor to the current PET systems.

These developments explain the slow implementation of PET into clinical practice, as synchronous developments in both tracer and detector technology were re-

quired to reach a certain threshold before adoption by the clinical community.

Molecular imaging has thus seen explosive growth and mainstream adoption since the US Food and Drug Administration (FDA) approval of 18-fluorodeoxyglucose (FDG) as a radiopharmaceutical in 1997, accelerated by the emergence of hybrid imaging typified by PET/computed tomography (CT) scanners. The development of such dual or hybrid modality PET/CT imaging has addressed the fundamental problems of PET imaging; namely, the limited spatial ability and the absence of anatomical landmarks, and this has resulted in widespread clinical adoption and acceptance. However, whole-body PET/CT imaging involves increased patient radiation exposure compared to single modality imaging, with the effective dose per PET/CT scan of approximately 25 mSv^[13]. Hence, patient selection for PET/CT imaging has to be justified, and further dose reduction strategies are needed.

Nevertheless, the success of FDG PET/CT imaging sets the stage for the future development of positron-based functional imaging, buoyed by the realization of the advantages provided by such hybrid diagnostics.

PRINCIPLES OF MOLECULAR IMAGING

The basis of molecular imaging lies with the targeted detection of specific cell targets or receptors. Receptor-specific molecules often bind to their receptors with high affinity and low dissociation rates^[14]. However, for purposes of diagnostic imaging, the concentration of receptor molecules in target tissue may be hard to differentiate from background non-specific binding^[15]. Thus, molecular imaging has often been confined to nuclear-based techniques such as PET or SPECT, which are able to generate images with micromolar to picomolar concentrations of imaging probes^[16].

Theoretically, there are multiple radionuclides that are of potential use in PET imaging. However in practice, most of these radionuclides are unsuitable for a variety of reasons, including availability and cost constraints, production issues, and the intrinsic decay properties of the radionuclides (Table 1). ¹⁸F is the most widely used positron tracer currently, and is produced in a cyclotron, most often utilizing either a neon gas target or an oxygen-enriched water target^[17]. It possesses a half-life of 109.8 min, mass of 18.0009380, with a maximum energy and range of 0.69 MeV and 2.4 mm, respectively.

However, there are distinct limitations in using ¹⁸F as a positron tracer. Firstly, the short half-life of ¹⁸F requires close physical proximity of a cyclotron facility to the imaging center, thus PET imaging centers are in essence “tethered” to cyclotrons.

Secondly, the infrastructure set-up and maintenance of a production cyclotron facility requires substantial financial and manpower resources, and this often limits PET imaging to countries with the necessary resources to support such molecular imaging.

Thirdly, ¹⁸F is available mainly as an ion in aqueous

Table 1 Examples of positron emitting radionuclides

Radionuclide	Half-life	Positron decay (%)	E _{max} (keV)	Production
¹¹ C	20.3 min	100	961	Cyclotron
¹³ N	9.97 min	100	1190	Cyclotron
¹⁵ O	2.1 min	100	1732	Cyclotron
¹⁸ F	110 min	97	634	Cyclotron
⁶⁴ Cu	12.8 h	19	656	Cyclotron
⁶⁸ Ga	67.6 min	89	1899	Generator
⁸² Rb	76 s	95	3150	Generator
¹²⁴ I	4.17 d	23	2100	Cyclotron

solution that must be taken to a dry organic environment for subsequent radiolabeling, and hence requires specially designed radiopharmacy facilities in addition to the cyclotron, which implies yet again significant resource investment.

⁶⁸Ga

⁶⁸Ga is a metallic positron emitter with a physical half-life of approximately 68 min and mass of 67.93. It decays predominantly through positron emission (89%) of 1.92 MeV and partially through electron capture (11%), into stable ⁶⁸Zn^[18].

It is available from a long-lived parent radionuclide ⁶⁸Ge, with a half-life of 288 d, which can be adsorbed to solids from which ⁶⁸Ga can be eluted. Hence, a generator-based source can be developed, and in fact several have been described. Current ⁶⁸Ge/⁶⁸Ga radionuclide generators are most commonly based on a TiO₂ solid phase^[19], from which ionic ⁶⁸Ga³⁺ can be easily eluted.

There are several advantages in using ⁶⁸Ga. Firstly, ⁶⁸Ga is generator-based, and hence there are no requirements for a cyclotron facility, with the isotope continuously available from the generator. Secondly, ⁶⁸Ga is a radiometal and can be more easily complexed using suitable chelating agents to targeting ligands, as compared with the more complex organic chemistry and purification requirements of ¹⁸F and most other cyclotron-based positron emitters. Examples of bifunctional chelators include EDTA and diethylenetriaminepentaacetic acid (DTPA). Thirdly, the long half-life of the parent radionuclide allows the use of the generator for long periods, possibly a year and longer, and the short half-life of ⁶⁸Ga matches the pharmacokinetics of numerous small peptides^[20].

Currently enjoying resurgence, ⁶⁸Ga has in fact been in clinical practice since the 1950s, and increased interest was stirred when a practical gallium parent–daughter generator system was developed in the early 1960s^[21]. Researchers and clinicians then and now understood the advantages of a generator-based positron isotope^[22,23].

However, as mentioned earlier, the development of nuclear-based molecular imaging has only blossomed in recent years due to the concomitant development of synergistic technologies. Despite the recognized potential of ⁶⁸Ga, it was not routinely used in clinical practice until recently. It is probable that advances in the development of radiolabeled peptides for diagnostic and therapeutic

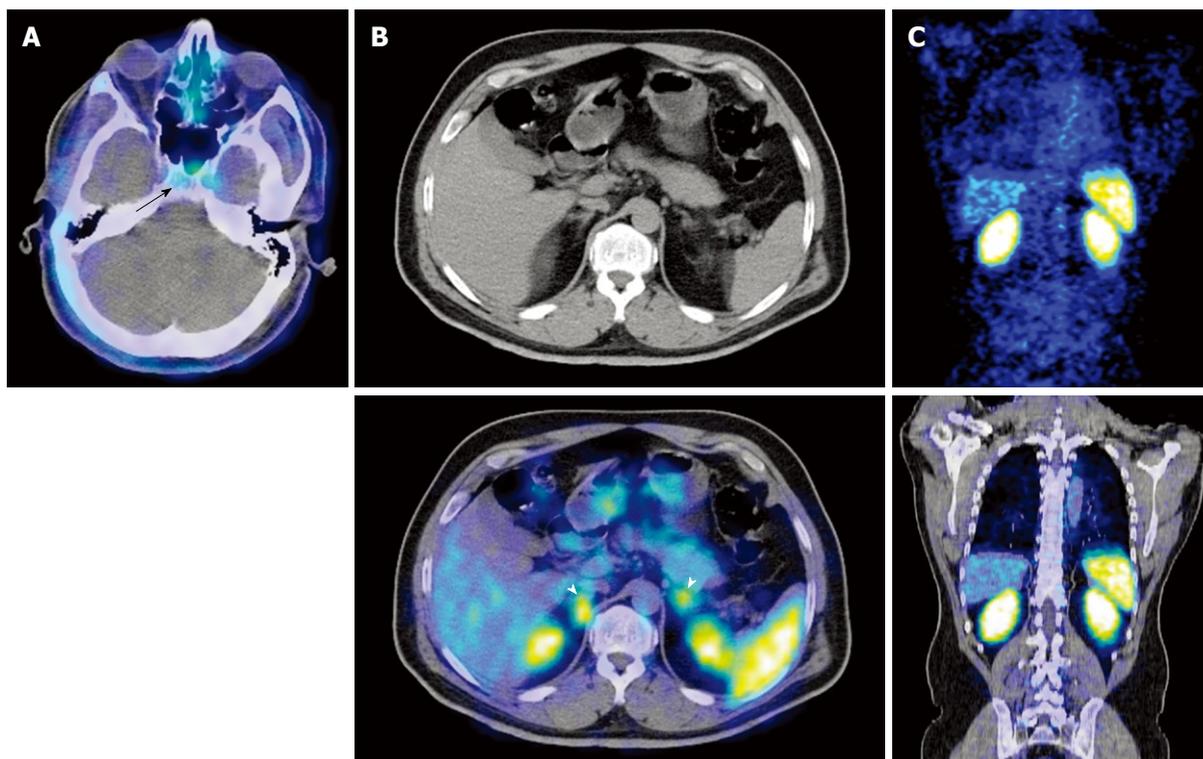


Figure 1 Examples of physiological areas of uptake in somatostatin receptor scintigraphy. A: Axial positron emission tomography (PET)/computed tomography (CT) of the base of skull demonstrates avid uptake in the pituitary fossa (black arrow) corresponding to physiological uptake in the pituitary gland; B: Axial CT and PET/CT of the abdomen demonstrates physiological uptake in the adrenals glands (white arrowheads); C: Coronal PET and PET/CT sections demonstrate avid tracer uptake in the kidneys and spleen.

purposes have contributed substantially to the current emergence of ⁶⁸Ga as a positron tracer, specifically in somatostatin receptor scintigraphy.

SOMATOSTATIN RECEPTOR SCINTIGRAPHY

Somatostatin is a naturally occurring peptide with various biological functions that mediates its action through several membrane-bound somatostatin receptors. Currently, five subtypes of somatostatin receptors have been identified in humans (SSRT1, SSRT2, SSRT3, SSRT4, SSRT5), with SSRT2 further classified into subtypes 2A and 2B^[24].

The distribution of various SSRT subtypes in human organ systems have been studied fairly extensively (Figure 1). Somatostatin receptor expression is found in a large number of tumors. Neuroendocrine tumors are the archetypical class that has been extensively imaged and treated using somatostatin analogues, but other tumors such as neuroblastoma, meningioma, breast cancer, lymphoma, renal cell carcinoma, hepatoma and pheochromocytoma are also known to express somatostatin receptors^[25,26].

The principle of somatostatin receptor imaging is based on linking a stable somatostatin type ligand to a radioisotope (e.g. ¹¹¹In, ^{99m}Tc, ⁶⁸Ga) *via* a chelating agent before administration into the patient, where uptake in the target tissue is dependent on SSRT-mediated inter-

nalization of the radioligand. *In vitro* studies have found that each of the different SSRT subtypes internalizes SSRT-ligands differently, and uptake of octreotide in SSRT-positive organs is determined predominantly by SSRT2^[27].

The half-life of somatostatin itself is short (< 2 min), thus it is unsuitable for use in either diagnosis or therapy. Hence, there has been development of synthetic somatostatin analogues with a sufficiently long half-life for use in diagnostic imaging or therapeutics.

The first radiolabeled somatostatin analogue Octreoscan® [¹¹¹In-DTPA-octreotide, D-Phe-Cys-Phe-D-Trp-Lys-Thr-Cys-Thr(ol)] was approved in the early 1990s for use in patients with neuroendocrine disease^[28]. Octreotide is an eight-amino acid analogue of somatostatin with four identical amino acids, with a longer half-life and greater affinity for SSRT2, SSRT3 and SSRT5, which makes a safe and sensitive imaging modality for the detection of gastroenteropancreatic neuroendocrine tumors^[29,30].

Since then, octreotide derivatives have been developed, allowing stable labeling with radiometals, as well as increased affinity for somatostatin receptor as compared with Octreoscan®. Most of these analogues utilize 1,4,7,10-tetraazacyclododecane-1,4,7,10-tetraacetic acid (DOTA) as the chelating agent, which forms thermodynamically and kinetically stable metal complexes. Common examples of such newer somatostatin analogues include: (1) TOC [D-Phe-Cys-Trp-D-Trp-Lys-Thr-Cys-

Thr(ol)] in which replacement of Phe with Tyr at position 3 results in increased internalization and higher contrast uptake *in vivo* as compared with octreotide; (2) TATE (D-Phe-Cys-Tyr-D-Trp-Lys-Thr-Cys-Thr), an analogue in the hydroxy group at the C terminus is changed to a free carboxylic group. This results in further increased binding affinity, internalization rates and selectivity for SSRT-2^[31]; and (3) NOC [D-Phe-Cys-Nal-D-Trp-Lys-Thr-Cys-Thr(ol)], where replacement of Phe with Nal results in a compound with high affinity to SSRT2, SSRT3 and SSRT5^[32].

⁶⁸Ga SOMATOSTATIN RECEPTOR SCINTIGRAPHY

The team in Zentralklinik Bad Berka, Germany has had extensive experience with receptor PET/CT imaging utilizing ⁶⁸Ga-labeled somatostatin analogues, where more than 2300 cases have been reported as of early 2009^[33]. In general, they found that ⁶⁸Ga somatostatin receptor scintigraphy imaging was a flexible, fast modality, with a low radiation burden and apparently lower costs, as compared with Octreoscan[®]. In addition, semi-quantitative reproducible standardized uptake values were utilized in selecting patients for peptide receptor radionuclide therapy (PRRT) and evaluation of treatment response. Expression of somatostatin analogues has been found in a wide variety of tumors, and therefore, ⁶⁸Ga somatostatin receptor scintigraphy (SRS) has broad clinical applications. Several of these applications are discussed below.

NEUROENDOCRINE TUMORS

Neuroendocrine tumors are a heterogeneous group of tumors that phenotypically are cancers that arise from neural crest tissue, and can hence arise from any part of the body depending on the distribution of the embryological tissue. The term neuroendocrine is derived from the relationship to neural cells in the expression of certain proteins such as synaptophysin, chromogranin, protein gene product 9.5 and neuron specific enolase (NSE).

Oberndorfer first coined the term “carcinoid” in 1907 to describe epithelial cells in the gut with a homogeneous structure with generally less aggressive features as compared with carcinomas^[34]. However, the use of this term is at best heterogeneous among clinicians, and this in turn results in substantial confusion. It is for this reason that the term neuroendocrine tumor is preferred.

Diagnosis and assessment of neuroendocrine tumors are based on morphological, immunohistochemical and functional characteristics. The diagnosis of neuroendocrine tumors relies heavily on the positive detection of markers by immunohistochemistry, such as NSE, protein gene product 9.5, chromogranin A and synaptophysin^[35]. Neuroendocrine tumors associated with hyperfunctional symptoms are termed functional, whereas those not associated with symptoms are termed non-functional.

The World Health Organization (WHO) classification for neuroendocrine tumor for the gastroenteropancreatic

system is divided into several broad categories^[36], with a general categorization based on histomorphology, tumor size, angio-invasion, organ-specific invasion, proliferation index, metastasis and functional/hormonal status^[37].

Neuroendocrine tumors of the gastroenteropancreatic system are by far the most common (70%), with the bronchopulmonary system also accounting for a significant proportion (25%)^[38]. Our discussion will focus predominantly on the gastroenteropancreatic system, but general principles are likely applicable to neuroendocrine tumors in other parts of the body.

These were the first tumors for which SRS was utilized and proven a viable clinical tool, and SRS using Octreoscan[®] was considered the gold standard in the diagnosis, staging and follow-up of patients with neuroendocrine tumors. As such, these were among the first tumors for which ⁶⁸Ga SRS was attempted.

Hofmann *et al.*^[39] have compared the utility of ⁶⁸Ga-DOTA-TOC PET with conventional SRS in patients with histologically proven neuroendocrine tumors, and have found higher tumor to non-tumor contrast ratios, with significantly higher detection ratios for ⁶⁸Ga SRS. This has been followed up by Kowalski *et al.*^[40], who also have compared these two techniques, and have found that ⁶⁸Ga SRS was again able to detect more lesions and was superior in detecting smaller lesions with lower uptake.

These findings were further supported by Buchmann *et al.*^[41], who prospectively evaluated 27 patients with known neuroendocrine tumors with ⁶⁸Ga-DOTA-TOC PET and ¹¹¹In-DTPA-octreotide SPECT for the detection of tumor characteristics. They have concluded that ⁶⁸Ga-DOTA-TOC PET is superior to ¹¹¹In-DTPA-octreotide SPECT for the detection of neuroendocrine tumor manifestations in the lung and skeleton, and similar for their detection in the liver and brain.

Ambrosini *et al.*^[42] have compared the use of ⁶⁸Ga-DOTA-NOC PET/CT with ⁶⁸Ga SRS in the assessment of histologically proven, well-differentiated bronchial carcinoids, and have found that the latter is better than contrast-enhanced CT for the evaluation and determination of disease extent.

Gabriel *et al.*^[43] have compared the diagnostic value of ⁶⁸Ga-DOTA-TOC with conventional scintigraphy and dedicated CT, and have found a sensitivity of 97%, specificity of 92%, and accuracy of 96%, which represents a significantly higher detection rate compared with that of conventional SRS or diagnostic CT.

In comparing the different somatostatin analogues, Antunes *et al.*^[44] have compared ⁶⁸Ga-DOTA-NOC with ⁶⁸Ga-DOTA-TATE and have found that the third-generation somatostatin analogue NOC has better preclinical and pharmacological performance, especially on SSRT2-expressing cells. This has particular clinical implications because SSRT2 receptors are important for ligand internalization.

Overall, ⁶⁸Ga-based SRS has been reported to have higher diagnostic sensitivity, specificity and accuracy compared with conventional diagnostic imaging and first-generation SRS^[45]. PET imaging has intrinsic ad-

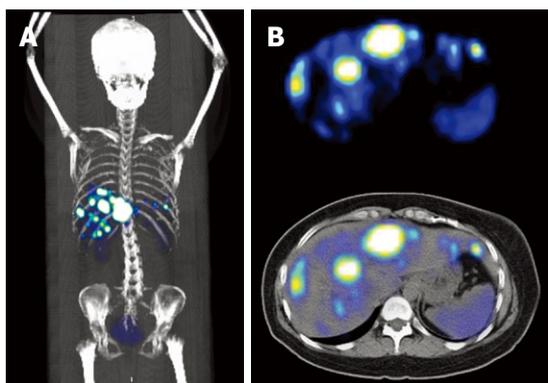


Figure 2 Metastatic pancreatic neuroendocrine carcinoma with multiple liver lesions. A: Coronal 3D reconstruction demonstrates multiple tracer avid lesions sited predominantly in the right hypochondrium; B: Axial PET and PET/CT of the liver shows multiple somatostatin receptor rich lesions in the liver.

vantages compared with conventional SRS: it has faster imaging turnover and increased spatial resolution.

Furthermore, as our understanding of tumor pathogenesis and histopathological characteristics increases, limitations of utilizing a single molecular target have emerged. Neuroendocrine tumors in particular have a wide range of cellular differentiation, and the molecular basis of SRS imaging is influenced by tumor grade (Figures 2 and 3).

Koukouraki *et al.*^[46] have evaluated 15 patients with 63 lesions using ⁶⁸Ga-DOTA-TOC and FDG-PET, and found that the global ⁶⁸Ga-DOTA-TOC uptake was mostly influenced by receptor affinity, whereas FDG uptake was predominantly influenced by fractional blood volume. Of further interest, Kayani *et al.*^[47] have compared ⁶⁸Ga-DOTA-TATE with FDG-PET/CT and have found a significant correlation between tumor uptake and histological grade, with ⁶⁸Ga SRS superior for well-differentiated neuroendocrine tumors and FDG-PET superior for poorly differentiated tumors.

These findings illustrate the heterogeneous nature of tumors and the challenge posed in molecular imaging, and raise the possibility of dual tracer imaging techniques in evaluating neuroendocrine tumors. However, further work in determining patient selection criteria needs to be done to balance the risk of increased radiation exposure against potential benefits.

NEUROGENIC AND NEUROECTODERMAL TUMORS

Although SRS has been used predominantly for neuroendocrine tumors, the expression of SSRTs is also found in a variety of other tumors, making them amenable to SRS.

Somatostatin receptor expression is found in neurogenic tumors (neuroblastoma, ganglioneuroblastoma and ganglioneuroma), which are tumors of the sympathetic nervous system that originate from neural crest tissue^[48], but are histologically differentiated by the stage of neu-

roblast maturation^[49]. The expression of SSRTs in these tumors allows for targeted imaging and therapy, and there is a direct association with the level of differentiation in such tumors, with SSRT expression being higher in benign ganglioneuroma and associated with favorable outcomes in advanced tumors^[50]. The use of ⁶⁸Ga SRS in establishing prognosis and assessing suitability for PRRT are possibilities, and needs further study.

Neuroectodermal tumors (pheochromocytoma and paraganglioma) arise from either the chromaffin tissues of the adrenal medulla or other paraganglionic sites, and often express SSRTs. With regard to pheochromocytoma, immunohistochemical assessments have found positive SSRT3 staining for a large majority of tumors (90%), with a significant portion (25%) demonstrating SSRT2A staining^[51]. Pheochromocytoma is often staged locally using conventional diagnostic imaging modalities (CT or MRI), with metaiodobenzylguanidine (MIBG) scintigraphy utilized for N and M assessment.

Current understanding of the use of functional imaging in pheochromocytoma involves MIBG imaging in the detection of more well-differentiated tumors and FDG-PET in the detection of poorly differentiated types^[52].

The role of SRS in the imaging assessment of pheochromocytoma is still indistinct, with the overall sensitivity of octreotide SRS reported to be about 30%^[53].

However, several studies have demonstrated the complementary nature of SRS and MIBG functional imaging, where conventional SRS is able to detect tumor foci deemed negative by MIBG assessment, possibly because of dedifferentiation of the norepinephrine transporter system^[54]. This result has been corroborated by a study in which ⁶⁸Ga-DOTA-TATE was used to assess malignant pheochromocytoma in comparison with CT and ¹²³I-MIBG in a small group of five patients, in whom SRS findings were positive for several lesions that had negative or low MIBG uptake (Figure 4)^[55].

It is therefore possible to use SRS to determine disease prognosis, as well as suitability for PRRT. With the newer and expanded range of improved somatostatin analogues, and the increased intrinsic resolution of PET over planar imaging, ⁶⁸Ga SRS shows promise for clinical application.

NON-MEDULLARY THYROID TUMORS

The thyroid gland is composed of two major cell types: follicular cells that produce thyroxine and triiodothyronine, and parafollicular cells that produce calcitonin. Thyroid tumors can arise from these epithelial cells, or from the supporting stromal tissue itself, thus exhibiting a wide range of morphological and biological patterns.

Malignant thyroid tumors are classified according to several subtypes: follicular (encapsulated and widely invasive), papillary (classical or variants), anaplastic, and medullary^[56]. Papillary and follicular type thyroid cancer form the majority of thyroid tumors.

The basis of SRS depends on the expression of SSRTs on the target tissue, and such receptors are widely

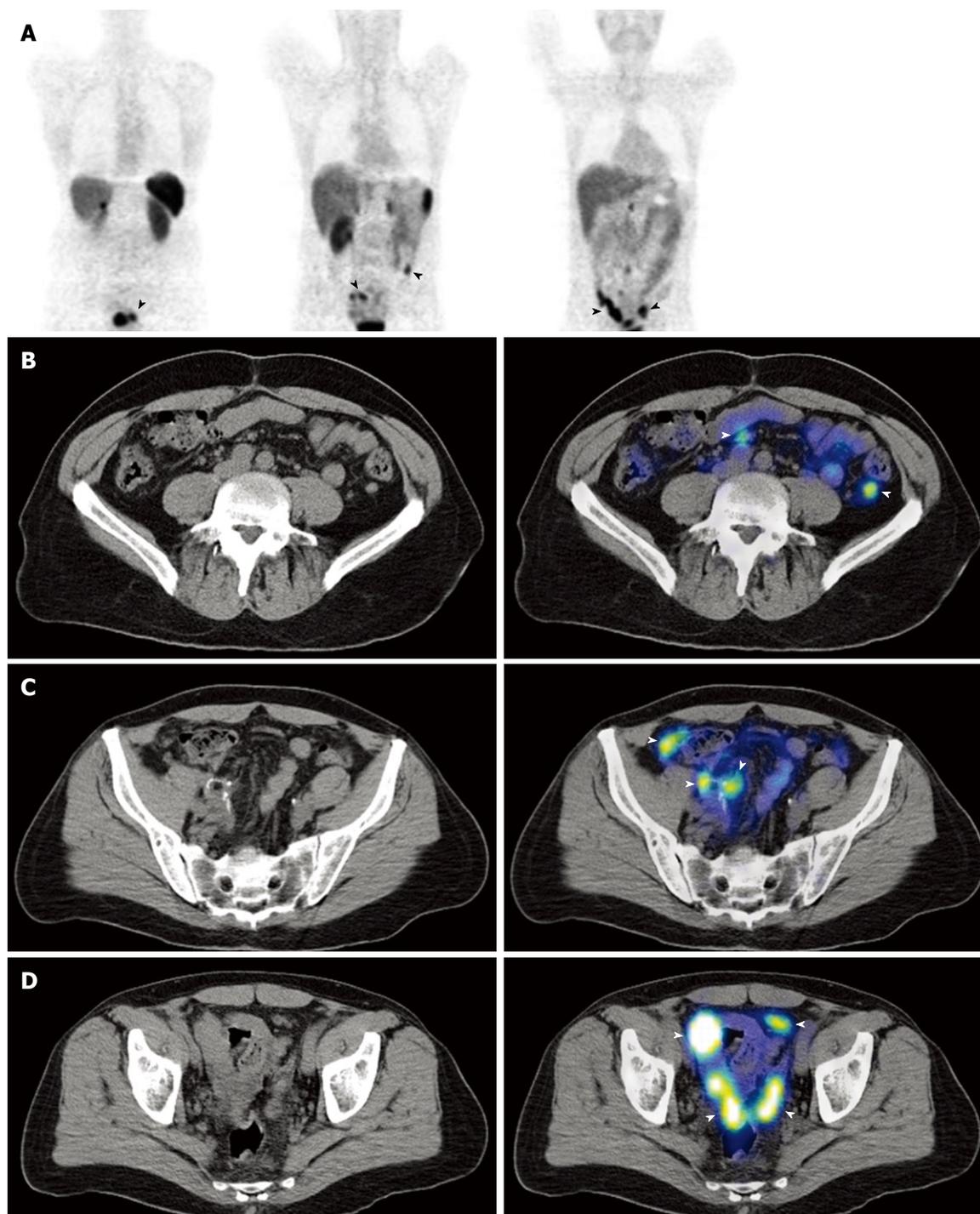


Figure 3 Metastatic appendiceal neuroendocrine carcinoma with multiple peritoneal deposits. A: Coronal PET images demonstrate multiple tracer avid foci projected over the abdomen and pelvis (black arrowheads); B-D: Axial CT and PET/CT sections of the abdomen show multiple somatostatin receptor rich peritoneal deposits (white arrowheads).

distributed in human thyroid diseases. However, there have been conflicting reports on the frequency and intensity of expression of specific receptor subtypes.

Pisarek *et al*^[57] have reported that the SSTR1 protein was expressed in 88.8%, SSTR2A and 2B in 44.4%, SSTR3 in 55.5%, SSTR4 in 11.2%, and SSTR5 in 33.3% of cases, and have concluded that SSTR1 is the dominant form in thyroid gland tumors and hyperplasia.

In contrast, Druckenthaner *et al*^[58] have reported that

SSRT2 is the predominant receptor type in tumor cells, and have found it in > 68% of cases. SSRT3 was found in 31% of cases, SSRT5 in 44%, and SSRT1 and SSRT4 were not detected in any of the samples.

Forsell-Aronsson *et al*^[59] have investigated tissue samples from 68 patients and have reported that all thyroid tumor types regularly express SSRT1, SSRT3, SSRT4 and SSRT5. Although SSRT2 is expressed in most medullary thyroid carcinomas, it is not detected in

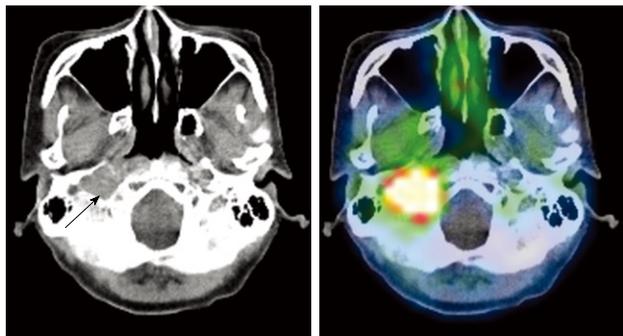


Figure 4 Malignant paraganglioma at the right jugular foramen. Axial CT and PET/CT of the base of skull. Soft tissue mass in the right jugular foramen showing intense tracer avidity, compatible with a somatostatin receptor expressing tumor.

follicular or papillary carcinoma subtypes. Ribonuclease protection assay however has detected SSRT2 expression in several samples deemed negative by northern blotting. In addition, despite the lack of SSRT2 expression detected by northern blotting, most tumors show uptake of radiolabeled octreotide analogues.

Ain *et al*^[60] have reported that most thyroid cancer cell lines express SSRT3 and SSRT5, with varying SSRT1 expression and SSRT2 only faintly detectable. SSRT4 was found to be positive in only one of their samples. It has been suggested that SSRT5-specific analogues are the best suited for targeting, and that SSRT2-specific analogues are of little use.

The literature regarding somatostatin receptor expression on thyroid tumor lines is conflicting, and may be related to the different investigative methods used. SSRT expression in differentiated thyroid carcinoma may also be heterogeneous in nature, possibly accounting for the differences in the findings.

Nevertheless, the value of SRS in thyroid tumor imaging dates back to the mid 1990s, when ¹¹¹In-labeled octreotide was found to visualize non-medullary differentiated thyroid carcinoma^[61]. Subsequently, several studies have followed that predominantly have focused on the clinical value of SRS in non-iodine-avid thyroid carcinoma, with sensitivities ranging from 19% to 100%^[62]. Overall sensitivity by the largest group investigating 43 patients was reported to be 51%. The anatomical location of the tumor has a significant influence on tumor detection; mediastinal tumors had the highest detection rate, while small lesions in the neck and lung were missed^[63]. The limitations of planar imaging and the inherent low-resolution capacity of earlier diagnostic modalities are likely contributors to the overall low sensitivity. With the advent of PET imaging and the development of newer somatostatin analogues, imaging sensitivity and specificity is expected to be significantly improved.

Similarly, ⁶⁸Ga SRS can be used clinically in patients with non-iodine-avid, non-medullary thyroid carcinoma for tumor localization, and assessment of suitability for PRRT and response to treatment. Our own experience of ⁶⁸Ga SRS for thyroid tumors is at an early stage, but

current findings show promise, with imaging showing significant tracer uptake in non-iodine-avid lesions (Figure 5).

FDG PET/CT has demonstrated clinical utility in evaluating non-iodine-avid lesions, with sensitivity approaching 90%^[64,65]. This peculiar phenomenon has been described by Feine *et al*^[66] as the “flip-flop” mechanism, where the dedifferentiated tumors lose iodine trapping capability, but have increased glucose metabolism and *vice versa*.

To the best of our knowledge, there have been no studies comparing FDG with ⁶⁸Ga SRS PET/CT in the evaluation of non-iodine-avid lesions. It is possible that dedifferentiation of thyroid carcinoma results in sequential loss of function, starting from loss of iodine trapping to loss of somatostatin receptors, with an increase in glucose metabolic activity. This suggests the clinical utility of dual tracer PET/CT imaging for non-iodine-avid lesions for evaluating the proportion of somatostatin-receptor-avid lesions, which could have an impact on management decisions. Patients with somatostatin-receptor-avid lesions may benefit from PRRT^[67], while those with solely FDG-avid lesions could benefit from trials of novel chemotherapy agents such as SorafenibTM^[68]. Current studies concerning ⁶⁸Ga SRS for thyroid carcinoma are rare, and further investigations are needed.

MENINGIOMAS

Meningiomas are mesodermal type tumors, which represent a significant percentage (approximately 20%) of intracranial neoplasms, with an overall incidence of 2.3/100 000^[69]. In general, surgical resection continues to be the mainstay of therapy, with complete removal providing the optimal chance for long-term remission. In patients for whom surgical options are not viable, external beam radiotherapy provides a safe palliative option^[70]. Almost all meningiomas express a high SSRT2 density, and kinetic studies using ⁶⁸Ga-DOTA-TOC has demonstrated high uptake in such tumors as expected^[71].

Clinical experiences are promising. Henze *et al*^[72] have shown that ⁶⁸Ga DOTA-TOC PET/CT is able to demonstrate clear visualization of meningioma, even sub-centimeter size lesions, with good sensitivity and specificity. They have concluded that ⁶⁸Ga SRS appears to be a promising modality for imaging meningioma, which offers excellent imaging properties and a high tumor-to-background ratio.

Aside from possible clinical use in surgical or radiotherapy planning, findings from ⁶⁸Ga SRS can be used to determine suitability for PRRT. Bartolomei *et al*^[73] have found a promising response to ⁹⁰Y-DOTA-TOC therapy in 29 patients with histologically proven meningioma, and have alluded to the possible adjuvant role of PRRT.

OTHER ⁶⁸Ga-RADIOLABELED LIGANDS

Somatostatin receptor scintigraphy illustrates the archetypical use of ⁶⁸Ga in PET/CT imaging, but does not

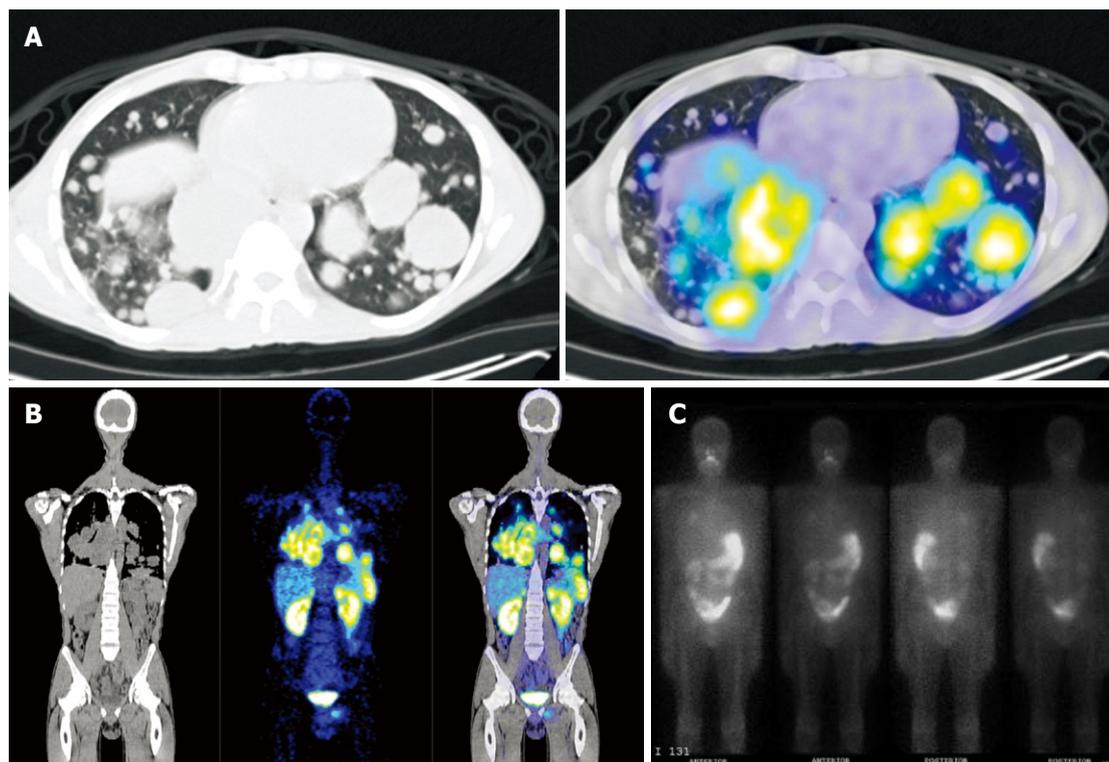


Figure 5 Metastatic non-iodine avid Hurthle cell thyroid carcinoma. A: Axial CT and PET/CT section of the thorax shows multiple pulmonary masses demonstrating avid tracer uptake; B: Coronal CT, PET and PET/CT images show somatostatin receptor rich pulmonary and mediastinal masses, and another discrete lesion projected over the left pelvis; C: I131 Whole body planar scan of the same patient shows only faint iodine uptake projected over the right upper lobe.

encompass its entire clinical spectrum. We thus discuss several promising ⁶⁸Ga-radiolabeled ligands and their potential clinical uses.

BOMBESIN

Bombesin was first described in 1970 by Erspamer *et al.* It was isolated from the skin of the *Bombina bombina* frog, and subsequently described in mammals by McDonald *et al* in 1979 and Minamino *et al* in 1983. Bombesin is a tetradecapeptide with a COOH terminus, which closely resembles gastrin-releasing peptide and neuromedin B^[74].

These peptides elicit a variety of physiological actions in the nervous system and gastrointestinal tract, with current nomenclature describing three classes of receptors: BB1 (384 amino acids, neuromedin receptor), BB2 (390 amino acids, gastrin-releasing peptide receptor) and BB3 (399 amino acids, orphan receptor), each of which is a G-protein-coupled receptor with separate general characteristics^[75].

Activation of the bombesin receptors is proposed to be important in the mediation of several disorders, as well as in the growth/differentiation of cancer. These receptors have been found to be overexpressed in a large number of tumors. For example, BB2 receptors have been found in tumors such as small cell lung cancer (85%-100%), non-small cell lung cancer (74%-78%) breast cancer (38%-72%), pancreatic cancer (10%), prostate cancer (62%-100%) and head/neck squamous cell carcinoma (100%) and neuroblastoma/ glioblastoma (72%-85%)^[76].

In addition, these receptors have been found to have growth-promoting effects in several tumors^[77].

Overexpression of these receptors allows for the targeted imaging and possible therapy of such tumors, with ongoing evaluation on the use of ⁶⁸Ga and other radioisotope-labeled bombesin analogues. Synthetic bombesin analogues can be categorized into two forms: type-A analogues are truncated peptides with only a portion retained, usually the BN (7-14) at the C terminus, and Type-B analogues, which are full-length synthetics. Type-A analogues are usually more accepted, as they are more stable *in vivo* and yet maintain acceptable receptor binding capability^[77]. Examples of bombesin analogues include demobesin-1, AMBA, pesin, DOTABOM and RP527, all with altered amino acid sequences.

Dimitrakopoulou-Strauss *et al*^[78] have investigated 17 patients with proven gastrointestinal stromal tumors (GISTs) using FDG- and ⁶⁸Ga-labeled bombesin analogue PET/CT. They found that, although FDG was superior in sensitivity and general tracer avidity, two false-positive lesions with FDG demonstrated uptake with the bombesin analogue. Thus, there is possibly a sub-group of GISTs that demonstrate enhanced bombesin receptor expression, but the clinical significance of this is still uncertain.

Prostate cancer is one of the most frequently diagnosed cancers in developed countries, and a significant cause of morbidity. Current non-invasive methods for imaging using MRI/CT/ultrasound have limitations and are of insufficient sensitivity in early tumor staging or

tumor recurrence. There have been prior investigations using predominantly ^{99m}Tc-labeled bombesin analogues, but the literature on PET imaging is scarce. Hofmann *et al.*^[79] have evaluated ⁶⁸Ga-labeled DOTABOM in 11 patients with prostate cancer. Peak tumor uptake was found at 15-25 min, with rapid renal excretion (> 75% injected dose recovered in the urine at 60 min). A transient reversible drop in systemic blood pressure was encountered in four patients. All tumor sites identified by the tracer was histologically proven, with the smallest tumor measuring 5 mm delineated, and it was concluded that ⁶⁸Ga-labeled DOTABOM appeared feasible in localizing the primary tumor and locoregional metastasis in patients with prostate carcinoma.

In addition to the above examples, radiolabeled bombesin analogues have shown promise in several other tumors such as breast^[80] and colorectal cancers^[81], using ^{99m}Tc as the radioisotope. Use of the ⁶⁸Ga PET tracer is being studied, and publications can be expected in the near future.

MELANOCYTE STIMULATING HORMONE (MSH) ANALOGUES

Melanomas are aggressive tumors arising from melanocytes, and can occur in any part of the body to which neural crest tissue migrates, with the skin being the most common primary site. There is an age-adjusted incidence rate of 19.6 per 100000 per year^[82], with an estimated 70000 new cases encountered in the United States annually.

Prognosis and management depend on tumor staging, which is influenced by clinical and histological factors, as well as the presence of nodal and distant spread^[83]. Although early-stage disease is curable in a significant proportion of patients, those with distant metastasis are rarely curable with standard therapy, and all such patients are considered candidates for clinical trials for new forms of therapy. Hence, accurate initial staging has a tangible clinical impact on prognosis and therapy stratification.

α -MSH peptides are tridecapeptides that are secreted by the pituitary gland, which target the melanocortin-1 receptor. This has been reported to have a > 80% expression rate in human melanoma^[84], which indicates a potential clinical use of such peptide analogues in patients with melanoma.

In a preclinical study, Wei *et al.*^[85] have utilized DO-TA-ReCCMSH (Arg11) as a melanocortin-1-receptor-targeting ligand, labeled with ⁶⁸Ga, for imaging B16/F1 melanoma tumor-bearing mice. They reported moderate receptor-mediated tumor uptake, fast non-target organ clearance, and high tumor to non-target tissue ratios.

Current strategies already include FDG PET/CT as part of the diagnostic workup for melanoma, with estimated 92% sensitivity and 90% specificity reported in a meta-analysis, and an overall FDG PET directed change in management value of 22%^[86]. Hence, such ⁶⁸Ga-

Table 2 Examples of FDA approved monoclonal antibodies

Generic name	Target	Type	Indication
Rituximab	CD20	Chimeric IgG1	Non-Hodgkins lymphomas
Trastuzumab	HER-2/neu	Humanized IgG1	Breast cancer
Alemtuzumab	CD52	Humanized IgG1	Chronic lymphocytic leukaemia
Bevacizumab	VEGF	Humanized IgG1	Colorectal cancer Non-small cell lung cancer Breast cancer Glioblastoma Renal cancer
Cetuximab	EGFR	Chimeric IgG1	Colorectal cancer Head and neck cancer
Panitumumab	EGFR	Human IgG1	Colorectal cancer
Ofatumumab	CD20	Human IgG1	Chronic lymphocytic leukaemia
Gemtuzumab	CD33	Humanized IgG4 conjugated to calicheamicin	Acute myeloid leukaemia
⁹⁰ Y-Ibritumomab Tiuxetan	CD20	⁹⁰ Y-radiolabeled murine IgG1	Non-Hodgkins lymphoma
¹³¹ I-Tositumomab	CD20	¹³¹ I-radiolabeled murine IgG2a	Non-Hodgkins lymphoma

FDA: Food and Drug Administration; EGFR: Epidermal growth factor receptor; VEGF: Vascular endothelial growth factor.

tagged α -MSH analogue ligands may be used for assessing suitability for PRRT in patients with advanced, non-curative malignant melanoma.

IMMUNOSCINTIGRAPHY USING ⁶⁸Ga

Antibodies have been frequently utilized as targeting ligands for tumors and other lesions because of their ability to bind to specific targets on the intended tissue. Advances in cell biology and molecular techniques have also facilitated the discovery of novel new molecular targets on tumor cells, further spurring development of newer targeting monoclonal antibodies (mAbs). To date, there are multiple mAbs approved by the US FDA for use in the systemic treatment of cancer (Table 2).

The use of positron-emitting radiotracers such as ¹⁸F- and ⁶⁸Ga-tagged mAb ligands is an attractive proposition that allows accurate localization of tumor tissues, with potential use in assessment of therapeutic suitability, prognosis, and response to treatment. We are possibly on the cusp of the “magic bullet” notion proposed by Paul Ehrlich, both in diagnostic and therapeutic applications.

mAbs are produced from a single B-cell clone, and it was the work of Köhler and Milstein in 1975, with the introduction of hybridoma technology, that led to the development of commercially and economically viable mAb production^[87]. This is essentially the production of cell lines formed by the fusion of the specific antibody-

producing B cell with immortal myeloma cell lines, which results in a biological factory that produces a homogeneous and specific line of mAbs.

Immunoglobulins are designated IgG, IgM, IgE, IgA or IgD according to their general structure and can be further subdivided on basis of internal attributes. For example, IgG is a bivalent antibody that comprises two long and two short amino acid chains (heavy and light chains), with a total molecular weight of approximately 150 000 Da. Two antigen-binding Fab fragments contain the variable regions, which are unique for each mAb. The constant region of the antibody molecule is known as the Fc fragment.

An initial drawback in the early years was the immunogenic nature of such antibodies because of their murine origin. Immunogenicity attracted significant attention when such mAbs were utilized, with human anti-mouse antibody immunoreactions being encountered^[88]. However, further developments in DNA recombination technology have led to development of partially and fully humanized antibodies, as well as antibodies with selected deletion of unfavorable domains (domain-deleted antibodies), which further diminishes the problems with immunogenicity^[89].

Generally, whole mAbs have a long residence time *in vivo*, with optimal tumor-to-non-target tissue ratios at 2-4 d post administration. Conversely, mAb fragments reach optimal target tissue concentrations earlier and are cleared rapidly *in vivo*. This is because the smaller molecular size compared with whole antibodies allows for increased blood clearance and soft tissue penetration. However, absolute target concentrations are often lower.

The selection of a suitable mAb depends on several considerations: (1) serum half-life; (2) efficient tumor localization and general tissue penetration; and (3) immunogenicity.

At present, there are several FDA-approved immunoscintigraphic products available for clinical use. For example, OncoScint™, CEA-Scan™ and ProstaScint™, utilizing either ^{99m}Tc or ¹¹¹In as radiotracers^[90].

In considering positron emission immunoscintigraphy, several issues have to be addressed. First, internalization of the selected mAb has implications for those positron tracers that degrade upon internalization (e.g. ¹²⁴I), which results in rapid cellular clearance. Hence, the appropriate positron emitter has to be selected with regard to the mAb ligand. Second, the biodistribution and clearance of the selected mAb has to be considered for diagnostic and therapeutic purposes. In general, mAb fragments are cleared by the kidneys while whole mAbs are cleared by the reticuloendothelial system. This has implications for diagnosis because physiological areas of uptake will be obscured, and it also has dosimetric and clinical implications in therapeutic applications. Third, the choice of delivery technique for the radioisotope is important. For a short-lived positron tracer such as ⁶⁸Ga, whole mAbs may be unsuitable due to the long interval required before optimal target tissue concentration is

reached, therefore, mAb fragments may be the more suitable candidate. However, target tissue concentrations may not be optimal using such mAb fragments.

Several technological methods have been developed to address the problem of optimal radioisotope delivery, such as pre-targeting^[91,92]. Pre-targeting is essentially the technique of giving the non-radioactive antibody first to allow optimal localization in the target tissue (pre-targeting), and subsequently administering the radiolabel that targets the pre-targeting antibody^[93]. There are currently several techniques routinely used in pretargeting; bispecific mAbs constructed with one hapten-binding site and another target binding site; antibodies conjugated to streptavidin or avidin to enable binding of biotin ligands; biotinylated antibodies capable of forming a complex with avidin or streptavidin; antibodies conjugated to DNA to promote binding to complementary nucleotide sequences; and antibodies conjugated to enzymes to activate a pro-drug at the target site^[94]. Specifically, techniques for ⁶⁸Ga PET that utilize bispecific antibody pre-targeting have been developed, possibly opening the door for routine clinical practice and targeted therapies^[95].

Several studies have been carried out using ⁶⁸Ga as the positron tracer. Klivényi *et al.*^[96] in a preclinical study have demonstrated the feasibility of using ⁶⁸Ga chelates in the imaging of human colon carcinoma xenografts pre-targeted with bispecific anti-CD44_{v6}/anti-Ga chelate antibodies. Furthermore, Schuhmacher *et al.*^[97] have conducted a phase I trial of 10 patients with histologically proven breast carcinoma. Using mAb 12H12, a murine IgG₁ mAb that targets carbohydrate side chains of TAG12, which is overexpressed by most epithelial cell adenocarcinomas, and anti-Ga-chelate mAbs (designated 3A10, an IgG₃ mAb), the authors hybridized both mAbs to create a bispecific mAb towards Ga-chelate and TAG12. Next, ⁶⁸Ga was joined to the Ga-chelate ligand. Patients were then infused with the bispecific mAb, followed by a blocker to saturate hapten ligand sites in free floating bispecific mAbs, before the administration of the ⁶⁸Ga-labeled Ga chelate. Fourteen of 17 known lesions were visualized, with three false-negative results, with authors concluding that the technique provided better sensitivity for the detection of breast cancer at low tumor contrasts as compared with conventional immunoscintigraphy.

CONCLUSION

¹⁸F-labeled FDG is currently the most widely used general PET tracer worldwide, but there are several limitations to its use. The emergence of ⁶⁸Ga provides promise of a logistically easy to manage PET tracer with broad applications, and current developments in both diagnostic and therapeutic areas show great clinical potential for this tracer. Further work and investigations are still required, but there is the promise that the long-awaited potential of ⁶⁸Ga might finally be realized.

REFERENCES

- 1 **Jones T.** Historical development of functional in vivo studies using positron emitting tracers. In: Valk PE, Bailey DL, Townsend DW, Maisey MN, editors. Positron emission tomography: basic science and clinical practice. London: Springer-Verlag, 2003: 3-40
- 2 **Tobias C,** Lawrence J, Roughton F, Root W, Gregersen M. The elimination of carbon monoxide from the human body with reference to the possible conversion of CO to CO₂. *Am J Physiol* 1945; **145**: 253-263
- 3 **Dyson NA,** Hugh-Jones P, Newbury GR, West JB. The preparation and use of Oxygen-15 with particular reference to its value in the study of pulmonary malfunction. Conference on Peaceful Uses of Atomic Energy, 1959: 103-115
- 4 **Harper PV,** Lathrop KA, Krizek H, Lembares N, Stark V, Hoffer PB. Clinical feasibility of myocardial imaging with ¹³NH₃. *J Nucl Med* 1972; **13**: 278-280
- 5 **Eichling JO,** Raichle ME, Grubb RL Jr, Larson KB, Ter-Pogossian MM. In vivo determination of cerebral blood volume with radioactive oxygen-15 in the monkey. *Circ Res* 1975; **37**: 707-714
- 6 **Ter-Pogossian MM,** Eichling JO, Davis DO, Welch MJ, Metzger JM. The determination of regional cerebral blood flow by means of water labeled with radioactive oxygen 15. *Radiology* 1969; **93**: 31-40
- 7 **Raichle ME,** Grubb RL Jr, Eichling JO, Ter-Pogossian MM. Measurement of brain oxygen utilization with radioactive oxygen-15: experimental verification. *J Appl Physiol* 1976; **40**: 638-640
- 8 **Lifton JF,** Welch MJ. Preparation of glucose labeled with 20-minute half-lived carbon-11. *Radiat Res* 1971; **45**: 35-40
- 9 **Sokoloff L,** Reivich M, Kennedy C, Des Rosiers MH, Patlak CS, Pettigrew KD, Sakurada O, Shinohara M. The [¹⁴C]deoxyglucose method for the measurement of local cerebral glucose utilization: theory, procedure, and normal values in the conscious and anesthetized albino rat. *J Neurochem* 1977; **28**: 897-916
- 10 **Ido T,** Wan CN, Fowler JS, Wolf AP. Fluorination with molecular fluorine. A convenient synthesis of 2-deoxy-2-fluoro-D-glucose. *J Org Chem* 1977; **42**: 2341-2342
- 11 **Ido T,** Wan CN, Casella V, Fowler JS, Wolf AP, Reivich M, Kuhl ED. Labeled 2-deoxy-D-glucose analogs. 18F-labeled 2-deoxy-2-fluoro-D-glucose, 2-deoxy-2-fluoro-D-mannose and 14C-2-deoxy-2-fluoro-D-glucose. *J Labelled Comp Radiopharm* 1978; **14**: 175-183
- 12 **Burnham CA,** Brownell GL. A multi-crystal positron camera. *IEEE Trans Nucl Sci* 1973; **19**: 201-205
- 13 **Brix G,** Lechel U, Glatting G, Ziegler SI, Münzing W, Müller SP, Beyer T. Radiation exposure of patients undergoing whole-body dual-modality 18F-FDG PET/CT examinations. *J Nucl Med* 2005; **46**: 608-613
- 14 **Krohn KA.** The physical chemistry of ligand-receptor binding identifies some limitations to the analysis of receptor images. *Nucl Med Biol* 2001; **28**: 477-483
- 15 **Katzenellenbogen BS,** Fang H, Ince BA, Pakdel F, Reese JC, Wooge CH, Wrenn CK, William L. McGuire Memorial Symposium. Estrogen receptors: ligand discrimination and antiestrogen action. *Breast Cancer Res Treat* 1993; **27**: 17-26
- 16 **Mankoff DA,** Link JM, Linden HM, Sundararajan L, Krohn KA. Tumor receptor imaging. *J Nucl Med* 2008; **49** Suppl 2: 149S-163S
- 17 **Helus F,** Maier-Borst W, Sahn U, Wiebe LI. F-18 cyclotron production methods. *Radiochem Radioanal Lett* 1979; **38**: 395-410
- 18 **Szelecsényi F,** Boothe TE, Tavano E, Plitnikas ME, Tárkányi F. Compilation of cross sections/thick target yields for ⁶⁶Ga, ⁶⁷Ga and ⁶⁸Ga production using Zn targets up to 30 MeV proton energy. *Appl Radiat Isot* 1994; **45**: 473-500
- 19 **Zhernosekov KP,** Filosofov DV, Baum RP, Aschoff P, Bihl H, Razbash AA, Jahn M, Jennewein M, Rösch F. Processing of generator-produced ⁶⁸Ga for medical application. *J Nucl Med* 2007; **48**: 1741-1748
- 20 **Breeman WA,** Verbruggen AM. The ⁶⁸Ge/ ⁶⁸Ga generator has high potential, but when can we use ⁶⁸Ga-labelled tracers in clinical routine? *Eur J Nucl Med Mol Imaging* 2007; **34**: 978-981
- 21 **Greene MW,** Tucker WD. An improved gallium-68 cow. *Int J Appl Radiat Isot* 1961; **12**: 62-63
- 22 **Hayes RL,** Carlton JE, Byrd BL. Bone scanning with gallium-68: a carrier effect. *J Nucl Med* 1965; **6**: 605-610
- 23 **Welch MJ,** Thakur ML, Coleman RE, Patel M, Siegel BA, Ter-Pogossian M. Gallium-68 labeled red cells and platelets: new agents for positron tomography. *J Nucl Med* 1977; **18**: 558-562
- 24 **Taniyama Y,** Suzuki T, Mikami Y, Moriya T, Satomi S, Sasano H. Systemic distribution of somatostatin receptor subtypes in human: an immunohistochemical study. *Endocr J* 2005; **52**: 605-611
- 25 **Reubi JC,** Waser B, Schaer JC, Laissue JA. Somatostatin receptor sst1-sst5 expression in normal and neoplastic human tissues using receptor autoradiography with subtype-selective ligands. *Eur J Nucl Med* 2001; **28**: 836-846
- 26 **Reubi JC,** Schaer JC, Laissue JA, Waser B. Somatostatin receptors and their subtypes in human tumors and in peritumoral vessels. *Metabolism* 1996; **45**: 39-41
- 27 **Hofland LJ,** Lamberts SW, van Hagen PM, Reubi JC, Schaeffer J, Waaijers M, van Koetsveld PM, Srinivasan A, Krenning EP, Breeman WA. Crucial role for somatostatin receptor subtype 2 in determining the uptake of [¹¹¹In-DTPA-D-Phe¹]octreotide in somatostatin receptor-positive organs. *J Nucl Med* 2003; **44**: 1315-1321
- 28 **Krenning EP,** Kwekkeboom DJ, Bakker WH, Breeman WA, Kooij PP, Oei HY, van Hagen M, Postema PT, de Jong M, Reubi JC. Somatostatin receptor scintigraphy with [¹¹¹In-DTPA-D-Phe¹]- and [¹²³I-Tyr³]-octreotide: the Rotterdam experience with more than 1000 patients. *Eur J Nucl Med* 1993; **20**: 716-731
- 29 **Harris AG.** Somatostatin and somatostatin analogues: pharmacokinetics and pharmacodynamic effects. *Gut* 1994; **35**: S1-S4
- 30 **Jamar F,** Fiasse R, Leners N, Pauwels S. Somatostatin receptor imaging with indium-111-pentetreotide in gastroenteropancreatic neuroendocrine tumors: safety, efficacy and impact on patient management. *J Nucl Med* 1995; **36**: 542-549
- 31 **Virgolini I,** Traub-Weidinger T, Decristoforo C. Nuclear medicine in the detection and management of pancreatic islet-cell tumours. *Best Pract Res Clin Endocrinol Metab* 2005; **19**: 213-227
- 32 **Wild D,** Schmitt JS, Ginj M, Mäcke HR, Bernard BF, Krenning E, De Jong M, Wenger S, Reubi JC. DOTA-NOC, a high-affinity ligand of somatostatin receptor subtypes 2, 3 and 5 for labelling with various radiometals. *Eur J Nucl Med Mol Imaging* 2003; **30**: 1338-1347
- 33 **Baum R.** Ga-68 Labelled Radiopharmaceuticals for Molecular Imaging of Cancer Using PET/CT - Present State and Future Perspectives. 2010 February 25. Available from: URL: http://www.singaporeradiology2010.com/programme_detailed.pdf
- 34 **Oberndorfer S.** Karzinoide Tumoren des Dünndarms. *Frankf Z Pathol* 1907; **1**: 425-432
- 35 **Solcia E,** Rindi G, Paolotti D, La Rosa S, Capella C, Fiocca R. Clinicopathological profile as a basis for classification of the endocrine tumours of the gastroenteropancreatic tract. *Ann Oncol* 1999; **10** Suppl 2: S9-S15
- 36 **Solcia E,** Klöppel G, Sobin LH. Histological typing of endocrine tumours (World Health Organization International Histological Classification of Tumours). 2nd ed. New York: Springer, 2000: 61-68
- 37 **Kaltsas GA,** Besser GM, Grossman AB. The diagnosis and

- medical management of advanced neuroendocrine tumors. *Endocr Rev* 2004; **25**: 458-511
- 38 **Pearse AGE**. Genesis of neuroendocrine system. In: Frisen SR, Thompson NW, editors. *Surgical endocrinology*. Philadelphia: Lippincott, 1990: 25-35
- 39 **Hofmann M**, Maecke H, Börner R, Weckesser E, Schöffski P, Oei L, Schumacher J, Henze M, Heppeler A, Meyer J, Knapp H. Biokinetics and imaging with the somatostatin receptor PET radioligand (⁶⁸Ga)-DOTATOC: preliminary data. *Eur J Nucl Med* 2001; **28**: 1751-1757
- 40 **Kowalski J**, Henze M, Schuhmacher J, Mäcke HR, Hofmann M, Haberkorn U. Evaluation of positron emission tomography imaging using [⁶⁸Ga]-DOTA-D Phe(1)-Tyr(3)-Octreotide in comparison to [¹¹¹In]-DTPAOC SPECT. First results in patients with neuroendocrine tumors. *Mol Imaging Biol* 2003; **5**: 42-48
- 41 **Buchmann I**, Henze M, Engelbrecht S, Eisenhut M, Runz A, Schäfer M, Schilling T, Haufe S, Herrmann T, Haberkorn U. Comparison of ⁶⁸Ga-DOTATOC PET and ¹¹¹In-DTPAOC (Octreoscan) SPECT in patients with neuroendocrine tumours. *Eur J Nucl Med Mol Imaging* 2007; **34**: 1617-1626
- 42 **Ambrosini V**, Castellucci P, Rubello D, Nanni C, Musto A, Allegrì V, Montini GC, Mattioli S, Grassetto G, Al-Nahhas A, Franchi R, Fanti S. ⁶⁸Ga-DOTA-NOC: a new PET tracer for evaluating patients with bronchial carcinoid. *Nucl Med Commun* 2009; **30**: 281-286
- 43 **Gabriel M**, Decristoforo C, Kendler D, Dobrozemsky G, Heute D, Uprimny C, Kovacs P, Von Guggenberg E, Bale R, Virgolini IJ. ⁶⁸Ga-DOTA-Tyr3-octreotide PET in neuroendocrine tumors: comparison with somatostatin receptor scintigraphy and CT. *J Nucl Med* 2007; **48**: 508-518
- 44 **Antunes P**, Ginj M, Zhang H, Waser B, Baum RP, Reubi JC, Maecke H. Are radiogallium-labelled DOTA-conjugated somatostatin analogues superior to those labelled with other radiometals? *Eur J Nucl Med Mol Imaging* 2007; **34**: 982-993
- 45 **Modlin IM**, Oberg K, Chung DC, Jensen RT, de Herder WW, Thakker RV, Caplin M, Delle Fave G, Kaltsas GA, Krenning EP, Moss SF, Nilsson O, Rindi G, Salazar R, Ruzniewski P, Sundin A. Gastroenteropancreatic neuroendocrine tumours. *Lancet Oncol* 2008; **9**: 61-72
- 46 **Koukouraki S**, Strauss LG, Georgoulas V, Eisenhut M, Haberkorn U, Dimitrakopoulou-Strauss A. Comparison of the pharmacokinetics of ⁶⁸Ga-DOTATOC and [¹⁸F]FDG in patients with metastatic neuroendocrine tumours scheduled for ⁹⁰Y-DOTATOC therapy. *Eur J Nucl Med Mol Imaging* 2006; **33**: 1115-1122
- 47 **Kayani I**, Bomanji JB, Groves A, Conway G, Gacinovic S, Win T, Dickson J, Caplin M, Ell PJ. Functional imaging of neuroendocrine tumors with combined PET/CT using ⁶⁸Ga-DOTATATE (DOTA-DPhe1,Tyr3-octreotate) and ¹⁸F-FDG. *Cancer* 2008; **112**: 2447-2455
- 48 **Rha SE**, Byun JY, Jung SE, Chun HJ, Lee HG, Lee JM. Neurogenic tumors in the abdomen: tumor types and imaging characteristics. *Radiographics* 2003; **23**: 29-43
- 49 **Shimada H**, Ambros IM, Dehner LP, Hata J, Joshi VV, Roald B. Terminology and morphologic criteria of neuroblastic tumors: recommendations by the International Neuroblastoma Pathology Committee. *Cancer* 1999; **86**: 349-363
- 50 **Kogner P**, Borgström P, Bjellerup P, Schilling FH, Refai E, Jonsson C, Dominici C, Wassberg E, Bihl H, Jacobsson H, Theodorsson E, Hassan M. Somatostatin in neuroblastoma and ganglioneuroma. *Eur J Cancer* 1997; **33**: 2084-2089
- 51 **Mundschenk J**, Unger N, Schulz S, Höllt V, Schulz S, Steinke R, Lehnert H. Somatostatin receptor subtypes in human pheochromocytoma: subcellular expression pattern and functional relevance for octreotide scintigraphy. *J Clin Endocrinol Metab* 2003; **88**: 5150-5157
- 52 **Shulkin BL**, Thompson NW, Shapiro B, Francis IR, Sisson JC. Pheochromocytomas: imaging with 2-[fluorine-¹⁸]fluoro-2-deoxy-D-glucose PET. *Radiology* 1999; **212**: 35-41
- 53 **Pacak K**, Eisenhofer G, Carrasquillo JA, Chen CC, Li ST, Goldstein DS. 6-[¹⁸F]fluorodopamine positron emission tomographic (PET) scanning for diagnostic localization of pheochromocytoma. *Hypertension* 2001; **38**: 6-8
- 54 **Tenenbaum F**, Lumbroso J, Schlumberger M, Mure A, Plouin PF, Caillou B, Parmentier C. Comparison of radio-labeled octreotide and meta-iodobenzylguanidine (MIBG) scintigraphy in malignant pheochromocytoma. *J Nucl Med* 1995; **36**: 1-6
- 55 **Win Z**, Al-Nahhas A, Towey D, Todd JF, Rubello D, Lewington V, Gishen P. ⁶⁸Ga-DOTATATE PET in neuroectodermal tumours: first experience. *Nucl Med Commun* 2007; **28**: 359-363
- 56 **Scopa CD**. Histopathology of thyroid tumors. An overview. *Hormones (Athens)* 2004; **3**: 100-110
- 57 **Pisarek H**, Stepień T, Kubiak R, Borkowska E, Pawlikowski M. Expression of somatostatin receptor subtypes in human thyroid tumors: the immunohistochemical and molecular biology (RT-PCR) investigation. *Thyroid Res* 2009; **2**: 1
- 58 **Druckenthaner M**, Schwarzer C, Ensinger C, Gabriel M, Prommegger R, Riccabona G, Decristoforo C. Evidence for Somatostatin receptor 2 in thyroid tissue. *Regul Pept* 2007; **138**: 32-39
- 59 **Forsell-Aronsson EB**, Nilsson O, Bejegård SA, Kölbj L, Bernhardt P, Mölne J, Hashemi SH, Wängberg B, Tisell LE, Ahlman H. ¹¹¹In-DTPA-D-Phe1-octreotide binding and somatostatin receptor subtypes in thyroid tumors. *J Nucl Med* 2000; **41**: 636-642
- 60 **Ain KB**, Taylor KD, Tofiq S, Venkataraman G. Somatostatin receptor subtype expression in human thyroid and thyroid carcinoma cell lines. *J Clin Endocrinol Metab* 1997; **82**: 1857-1862
- 61 **Postema PT**, De Herder WW, Reubi JC, Oei HY, Kwekkeboom DJ, Bruining HJ, Bonjer J, van Toor H, Hennemann G, Krenning EP. Somatostatin receptor scintigraphy in non-medullary thyroid cancer. *Digestion* 1996; **57** Suppl 1: 36-37
- 62 **Teunissen JJM**, Kwekkeboom DJ, Krenning EP. Staging and treatment of differentiated thyroid carcinoma with radiolabeled somatostatin analogs. Trends in endocrinology and metabolism. *Trends Endocrinol Metab* 2006; **17**: 19-25
- 63 **Giammarile F**, Houzard C, Bournaud C, Hafdi Z, Sassolas G, Borson-Chazot F. Diagnostic management of suspected metastatic thyroid carcinoma: clinical value of octreotide scintigraphy in patients with negative high-dose radioiodine scans. *Eur J Endocrinol* 2004; **150**: 277-283
- 64 **Helal BO**, Merlet P, Toubert ME, Franc B, Schwartz C, Gauthier-Koelesnikov H, Prigent A, Syrota A. Clinical impact of (¹⁸F)-FDG PET in thyroid carcinoma patients with elevated thyroglobulin levels and negative (¹³¹I) scanning results after therapy. *J Nucl Med* 2001; **42**: 1464-1469
- 65 **Ong SC**, Ng DC, Sundram FX. Initial experience in use of fluorine-¹⁸-fluorodeoxyglucose positron emission tomography/computed tomography in thyroid carcinoma patients with elevated serum thyroglobulin but negative iodine-¹³¹ whole body scans. *Singapore Med J* 2005; **46**: 297-301
- 66 **Feine U**, Lietzenmayer R, Hanke JP, Held J, Wöhrle H, Müller-Schauenburg W. Fluorine-¹⁸-FDG and iodine-¹³¹-iodide uptake in thyroid cancer. *J Nucl Med* 1996; **37**: 1468-1472
- 67 **Teunissen JJ**, Kwekkeboom DJ, Kooij PP, Bakker WH, Krenning EP. Peptide receptor radionuclide therapy for non-radioiodine-avid differentiated thyroid carcinoma. *J Nucl Med* 2005; **46** Suppl 1: 107S-114S
- 68 **Kloos RT**, Ringel MD, Knopp MV, Hall NC, King M, Stevens R, Liang J, Wakely PE Jr, Vasko VV, Saji M, Rittenberry J, Wei L, Arbogast D, Collamore M, Wright JJ, Grever M, Shah MH. Phase II trial of sorafenib in metastatic thyroid cancer. *J Clin Oncol* 2009; **27**: 1675-1684
- 69 **Bondy M**, Ligon BL. Epidemiology and etiology of intracranial meningiomas: a review. *J Neurooncol* 1996; **29**: 197-205
- 70 **D'Ambrosio AL**, Bruce JN. Treatment of meningioma: an

- update. *Curr Neurol Neurosci Rep* 2003; **3**: 206-214
- 71 **Henze M**, Dimitrakopoulou-Strauss A, Milker-Zabel S, Schuhmacher J, Strauss LG, Doll J, Mäcke HR, Eisenhut M, Debus J, Haberkorn U. Characterization of ⁶⁸Ga-DOTA-D-Phe1-Tyr3-octreotide kinetics in patients with meningiomas. *J Nucl Med* 2005; **46**: 763-769
 - 72 **Henze M**, Schuhmacher J, Hipp P, Kowalski J, Becker DW, Doll J, Mäcke HR, Hofmann M, Debus J, Haberkorn U. PET imaging of somatostatin receptors using [⁶⁸Ga]DOTA-D-Phe1-Tyr3-octreotide: first results in patients with meningiomas. *J Nucl Med* 2001; **42**: 1053-1056
 - 73 **Bartolomei M**, Bodei L, De Cicco C, Grana CM, Cremonesi M, Botteri E, Baio SM, Aricò D, Sansovini M, Paganelli G. Peptide receptor radionuclide therapy with (90)Y-DOTA-TOC in recurrent meningioma. *Eur J Nucl Med Mol Imaging* 2009; **36**: 1407-1416
 - 74 **Gonzalez N**, Moody TW, Igarashi H, Ito T, Jensen RT. Bombesin-related peptides and their receptors: recent advances in their role in physiology and disease states. *Curr Opin Endocrinol Diabetes Obes* 2008; **15**: 58-64
 - 75 **Jensen RT**, Batten JF, Spindel ER, Benya RV. International Union of Pharmacology. LXVIII. Mammalian bombesin receptors: nomenclature, distribution, pharmacology, signaling, and functions in normal and disease states. *Pharmacol Rev* 2008; **60**: 1-42
 - 76 **Jensen RT**, Moody TW. Bombesin-related peptides and neurotensin: effects on cancer growth/proliferation and cellular signaling in cancer. In: Kastin AJ, editor. *Handbook of Biologically Active Peptides*. Amsterdam: Elsevier, 2006: 429-434
 - 77 **Schroeder RP**, van Weerden WM, Bangma C, Krenning EP, de Jong M. Peptide receptor imaging of prostate cancer with radiolabelled bombesin analogues. *Methods* 2009; **48**: 200-204
 - 78 **Dimitrakopoulou-Strauss A**, Hohenberger P, Haberkorn U, Mäcke HR, Eisenhut M, Strauss LG. ⁶⁸Ga-labeled bombesin studies in patients with gastrointestinal stromal tumors: comparison with ¹⁸F-FDG. *J Nucl Med* 2007; **48**: 1245-1250
 - 79 **Hofmann M**, Machtens S, Stief C, Maecke H, Boerner AR, Knapp WH. Feasibility of Ga-68-DOTABOM PET in prostate carcinoma patients [abstract]. *J Nucl Med* 2004; **45**: 449
 - 80 **Scopinaro F**, Varvarigou AD, Ussof W, De Vincentis G, Sourlingas TG, Evangelatos GP, Datsteris J, Archimandritis SC. Technetium labeled bombesin-like peptide: preliminary report on breast cancer uptake in patients. *Cancer Biother Radiopharm* 2002; **17**: 327-335
 - 81 **Scopinaro F**, De Vincentis G, Corazziari E, Massa R, Osti M, Pallotta N, Covotta A, Remediani S, Paolo MD, Monteleone F, Varvarigou A. Detection of colon cancer with ^{99m}Tc-labeled bombesin derivative (^{99m}Tc-leu13-BN1). *Cancer Biother Radiopharm* 2004; **19**: 245-252
 - 82 SEER Cancer Statistics Review 1975-2006. Available from: URL: http://seer.cancer.gov/csr/1975_2006/index.html
 - 83 **Manola J**, Atkins M, Ibrahim J, Kirkwood J. Prognostic factors in metastatic melanoma: a pooled analysis of Eastern Cooperative Oncology Group trials. *J Clin Oncol* 2000; **18**: 3782-3793
 - 84 **Tatro JB**, Atkins M, Mier JW, Hardarson S, Wolfe H, Smith T, Entwistle ML, Reichlin S. Melanotropin receptors demonstrated in situ in human melanoma. *J Clin Invest* 1990; **85**: 1825-1832
 - 85 **Wei L**, Miao Y, Gallazzi F, Quinn TP, Welch MJ, Vävere AL, Lewis JS. Gallium-68-labeled DOTA-rhenium-cyclized alpha-melanocyte-stimulating hormone analog for imaging of malignant melanoma. *Nucl Med Biol* 2007; **34**: 945-953
 - 86 **Schwimmer J**, Essner R, Patel A, Jahan SA, Shepherd JE, Park K, Phelps ME, Czernin J, Gambhir SS. A review of the literature for whole-body FDG PET in the management of patients with melanoma. *Q J Nucl Med* 2000; **44**: 153-167
 - 87 **Köhler G**, Milstein C. Continuous cultures of fused cells secreting antibody of predefined specificity. *Nature* 1975; **256**: 495-497
 - 88 **Reilly RM**, Sandhu J, Alvarez-Diez TM, Gallinger S, Kirsh J, Stern H. Problems of delivery of monoclonal antibodies. Pharmaceutical and pharmacokinetic solutions. *Clin Pharmacokinet* 1995; **28**: 126-142
 - 89 **Keenan AM**, Harbert JC, Larson SM. Monoclonal antibodies in nuclear medicine. *J Nucl Med* 1985; **26**: 531-537
 - 90 **van Dongen GA**, Visser GW, Lub-de Hooge MN, de Vries EG, Perk LR. Immuno-PET: a navigator in monoclonal antibody development and applications. *Oncologist* 2007; **12**: 1379-1389
 - 91 **Boerman OC**, van Schaijk FG, Oyen WJ, Corstens FH. Pre-targeted radioimmunotherapy of cancer: progress step by step. *J Nucl Med* 2003; **44**: 400-411
 - 92 **Meredith RF**, Buchsbaum DJ. Pretargeted radioimmunotherapy. *Int J Radiat Oncol Biol Phys* 2006; **66**: S57-S59
 - 93 **Goodwin DA**, Meares CF, McCall MJ, McTigue M, Chaovapong W. Pre-targeted immunoscintigraphy of murine tumors with indium-111-labeled bifunctional haptens. *J Nucl Med* 1988; **29**: 226-234
 - 94 **Goldenberg DM**, Sharkey RM, Paganelli G, Barbet J, Chatal JF. Antibody pretargeting advances cancer radioimmunodetection and radioimmunotherapy. *J Clin Oncol* 2006; **24**: 823-834
 - 95 **Griffiths GL**, Chang CH, McBride WJ, Rossi EA, Sheerin A, Tejada GR, Karacay H, Sharkey RM, Horak ID, Hansen HJ, Goldenberg DM. Reagents and methods for PET using bispecific antibody pretargeting and ⁶⁸Ga-radiolabeled bivalent hapten-peptide-chelate conjugates. *J Nucl Med* 2004; **45**: 30-39
 - 96 **Klivényi G**, Schuhmacher J, Patzelt E, Hauser H, Matys R, Moock M, Regiert T, Maier-Borst W. Gallium-68 chelate imaging of human colon carcinoma xenografts pretargeted with bispecific anti-CD44V6/anti-gallium chelate antibodies. *J Nucl Med* 1998; **39**: 1769-1776
 - 97 **Schuhmacher J**, Kaul S, Klivényi G, Junkermann H, Magener A, Henze M, Doll J, Haberkorn U, Amelung F, Bastert G. Immunoscintigraphy with positron emission tomography: gallium-68 chelate imaging of breast cancer pretargeted with bispecific anti-MUC1/anti-Ga chelate antibodies. *Cancer Res* 2001; **61**: 3712-3717

S- Editor Cheng JX L- Editor Kerr C E- Editor Zheng XM