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## Britanin – a beacon of hope against gastrointestinal tumors?

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### Abstract

Britanin is a bioactive sesquiterpene lactone known for its potent anti-inflammatory and anti-oxidant properties. It also exhibits significant anti-tumor activity, suppressing tumor growth *in vitro* and *in vivo*. The current body of research on Britanin includes thirty papers predominantly related to neoplasms, the majority of which are gastrointestinal tumors that have not been summarized before. To drive academic debate, the present paper reviews the available research on Britanin in gastrointestinal tumors. It also outlines novel research directions using data not directly concerned with the digestive system, but which could be adopted in future gastrointestinal research. Britanin was found to counteract liver, colorectal, pancreatic, and gastric tumors, by regulating proliferation, apoptosis, autophagy, immune response, migration, and angiogenesis. As confirmed in pancreatic, gastric, and liver cancer, its most commonly noted molecular effects include nuclear factor kappa B and B-cell lymphoma 2 downregulation, as well as Bcl-2-associated X protein upregulation. Moreover, it has been found to induce the Akt kinase and Forkhead box O1 axis, activate the AMP-activated protein kinase pathway, elevate interleukin-2 and peroxisome proliferator-activated receptor- $\gamma$  levels, reduce interleukin-10, as well as downregulate matrix metalloproteinase-9, Twist family bHLH transcription factor 1, and cyclooxygenase-2. It

also inhibits Myc-HIF1 $\alpha$  interaction and programmed death ligand 1 transcription by interrupting the Ras/RAF/MEK/ERK pathway and mTOR/P70S6K/4EBP1 signaling. Future research should aim to unravel the link between Britanin and acetylcholinesterase, mast cells, osteolysis, and ischemia, as compelling data have been provided by studies outside the gastrointestinal context. Since the cytotoxicity of Britanin on noncancerous cells is significantly lower than that on tumor cells, while still being effective against the latter, further in-depth studies with the use of animal models are merited. The compound exhibits pleiotropic biological activity and offers considerable promise as an anti-cancer agent, which may address the current paucity of treatment options and high mortality rate among patients with gastrointestinal tumors.

**Key Words:** Britanin; Sesquiterpene lactones; Chemotherapeutics; Gastrointestinal tumors; *In vitro*; *In vivo*

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**Core Tip:** Natural compounds have settled in the development of novel drugs. Britanin is a sesquiterpene lactone whose effect on gastrointestinal tumors has not been summarized before. Our paper reviews the current state of knowledge and proposes novel research directions. Britanin was found to counteract liver, colorectal, pancreatic, and gastric tumors *via* the regulation of proliferation, apoptosis, autophagy, immune response, migration, and angiogenesis. Future research should examine the link between Britanin and acetylcholinesterase, mast cells, osteolysis, and ischemia. The compound holds promise as an anti-cancer agent and may overcome the paucity of treatment options or high mortality rate in gastrointestinal tumors.

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## INTRODUCTION

Natural compounds have long been established in the development of novel drugs. One such group, the sesquiterpene lactones, are organic terpenoids that exhibit a broad spectrum of biological activities, with their anti-cancer, anti-parasitic, and anti-inflammatory properties being the most prominent[1-3]. One of the representatives of this group is a compound termed Britanin (C<sub>19</sub>H<sub>26</sub>O<sub>7</sub>), a pseudoguaianolide-type sesquiterpene lactone present in various *Inula* species. It has been found to demonstrate anti-cancer agent activities by affecting tumor cell survival[3,4]. Although Britanin has been present in the PubChem database since 2005, the current body of research is limited to about thirty papers in total, mostly related to cancer. The vast majority of the literature concerns gastrointestinal tumors that have not been summarized before. Britanin has also been evaluated in leukemia[5-8] and tumors of the breast[9-12], head and neck[13], kidney[14], prostate[15], or lung[14]; however, insufficient data exists on each disease type to draw firm conclusions. Given its promising implications in oncology, Britanin is likely to be the subject of considerable research in the upcoming years. To drive academic debate, the present paper reviews and discusses available research on Britanin in gastrointestinal tumors. A literature search was performed *via* PubMed using the “britanin” and “britannin” terms, focusing on gastrointestinal tumors. Moreover, the present paper outlines novel research directions using data outside the scope of the digestive system, which could be adopted in future gastrointestinal research.

## RESEARCH ON BRITANIN IS FOCUSED ON LIVER, COLORECTAL, PANCREATIC, AND GASTRIC TUMORS

The first report on the anti-proliferative properties of Britanin was published in 2012 by Moghadam *et al*[14] who extracted a compound from *Inula aucheriana*. A strong cytotoxic effect was noted on the liver cancer cell line HepG2 based on MTT assay, *i.e.*, utilizing 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide, with the half-maximal inhibitory concentration (IC<sub>50</sub>) of 2.2  $\mu$ g/mL[14]. In the following year, Fishedick *et al*[16] found that 10  $\mu$ mol/L of *Inula britannica*-derived Britanin inhibited cell growth by ~80%, as estimated on colorectal cancer cell line DLD1 and its multi-drug resistant counterpart with P-glycoprotein overexpression.

In 2016, Piao *et al*[17] evaluated the activity of fourteen *Inula japonica*-derived compounds that inhibit DNA topoisomerases. Among them, Britanin exhibited better inhibitory activity against topoisomerase II (IC<sub>50</sub> = 6.9  $\mu$ mol/L) than against topoisomerase I (IC<sub>50</sub> > 80  $\mu$ mol/L). Interestingly, the inhibitory capabilities of Britanin directed at topoisomerase II were found to surpass those of Etoposide (IC<sub>50</sub> = 26.9  $\mu$ mol/L), a commonly used inhibitor. Moreover, Britanin showed low toxicity against liver hepatoblastoma (HepG2 cell line) and colon adenocarcinoma (HT-29 cell line), with IC<sub>50</sub> values



of 35.5  $\mu\text{mol/L}$  and 3.9  $\mu\text{mol/L}$ , respectively[17].

In 2017, Moenifard *et al*[18] assessed the chemotherapeutic potential of Britanin derived from *Inula aucheriana* in pancreatic cancer therapy. The results indicated that the compound induces apoptosis in human pancreatic cancer cell lines AsPC-1 and PANC-1 by simultaneously decreasing B-cell lymphoma 2 (BCL-2) expression and increasing that of Bcl-2-associated X protein (BAX). Additionally, Britanin increased the generation of reactive oxygen species (ROS) and activated the axis of Akt kinase and Forkhead box O1 (AKT-FOXO1), inducing the mitochondrial apoptotic pathway in both cell lines[18].  $\text{IC}_{50}$  values for AsPC-1 and PANC-1 cell lines equaled  $30 \pm 4.61 \mu\text{mol/L}$  and  $40 \pm 5.63 \mu\text{mol/L}$ , respectively.

In the following year, Cui *et al*[19] reported that Britanin extracted from *Inula aucheriana* could induce apoptosis and autophagy *via* ROS-driven activation of the AMP-activated protein kinase (AMPK) pathway in the liver cancer cell lines HuH-7, SMMC-7721, and HepG2. Britanin reduced the survival rate of the cells in a dose- and time-dependent manner, with respective  $\text{IC}_{50}$  values of  $27.86 \pm 1.35 \mu\text{mol/L}$ ,  $28.92 \pm 1.09 \mu\text{mol/L}$  and  $15.69 \pm 1.58 \mu\text{mol/L}$  after 24-h treatment ( $8.81 \pm 0.95 \mu\text{mol/L}$ ,  $8.12 \pm 1.15 \mu\text{mol/L}$ , and  $6.86 \pm 1.05 \mu\text{mol/L}$  after 48 h). Furthermore, the compound exhibited no cytotoxicity against normal human liver cells. Further *in vivo* tests on the most susceptible cell line (HepG2) found Britanin to suppress liver cancer proliferation in a dose-dependent manner[19].

In 2020, Shi *et al*[20] found *Inula japonica*-derived Britanin to inhibit the growth and progression of gastric cancer cells using *in vitro* and *in vivo* models. The *in vitro* study examined the influence of Britanin on the proliferation and migration of BGC-823 and SGC-7901 gastric cell lines, while the mouse xenograft model involving the BGC-823 allowed for real-time tracking of tumor growth through bioluminescent imaging. Cytotoxicity testing indicated  $\text{IC}_{50}$  values of 4.999  $\mu\text{mol/L}$  for BGC-823 and 2.243  $\mu\text{mol/L}$  for SGC-7901. Treatment with Britanin was associated with alterations in the nuclear factor kappa B (NF- $\kappa$ B) pathway which reduced the proliferation of gastric cancer cells. It also resulted in elevated interleukin-2 levels (activator of Natural Killer cells, B-cells, CD4+ and CD8+ T-cells) and decreased interleukin-10 levels (CD4+ T-cell inactivator), thus promoting the immune response and inhibiting cancer cell development[20].

A study by Li *et al*[21] found Britanin to have similar effects on hepatocellular carcinoma. The cytotoxicity and anti-tumor effects were studied on HepG2 and BEL-7402 cell lines *in vitro* and a subcutaneous BEL-7402 tumor model in mice *in vivo*. The  $\text{IC}_{50}$  values were found to be 2.702  $\mu\text{mol/L}$  in the BEL-7402 and 6.006  $\mu\text{mol/L}$  in the HepG2 cells. Colony formation assay, transwell migration, and tumor size measurements showed that Britanin possesses a reliable anti-tumor effect. Additionally, Western Blotting indicated that Britanin inhibited p65 protein and modulated the BCL-2/BAX ratio [21].

The effect of Britanin from *Inula linearifolia* on pancreatic cancer was examined by Li *et al*[22]. The anti-tumor effects were determined *in vitro* on three pancreatic cancer cell lines: PANC-1, MIA CaPa-2, and BxPC-3. Respective  $\text{IC}_{50}$  values equaled 1.348, 3.104, and 3.367  $\mu\text{mol/L}$ . PANC-1 was utilized to establish a murine xenograft model. Britanin exhibited very low toxicity *in vivo* and excellent inhibitory effects against pancreatic cancer *in vivo* and *in vitro*. The compound diminished cell proliferation and migration by inhibiting the p50-p65/NF- $\kappa$ B pathway. The authors suggest that, due to its very low toxicity, Britanin could be safer for use than small molecule inhibitors[22].

In 2021, Zhang *et al*[23] investigated the potential of Britanin in cancer immunotherapy, specifically its impact on the Programmed death receptor 1 and ligand 1 (PD-1/PD-L1) immune pathway. The study used Hep3B liver cancer cells and HCT116 colorectal cancer cells, with the latter utilized to establish a mouse xenograft model. It was found that Britanin maintains the activity of T-cells and reduces proliferation and angiogenesis by inhibiting PD-L1 transcription; this was achieved by interrupting the Ras/RAF/MEK/ERK pathway and mTOR/P70S6K/4EBP1 signaling, ultimately affecting communication between myelocytomatosis oncogene (Myc) and hypoxia-inducible factor 1 $\alpha$  (HIF-1 $\alpha$ ). Moreover, molecular docking data revealed that Britanin interacts with PD-L1, HIF-1 $\alpha$ , and Myc[23]. A later docking analysis of Britanin, and fifteen of its analogues, to the PD-L1 protein was used in the design of novel molecules based on the structure of pseudoguaianolide-type sesquiterpene lactones[4].

The most recent gastrointestinal study was conducted by Abdolmohammadi *et al*[24] who evaluated the mode of action of Britanin from *Inula aucheriana* in gastric cancer. Growth inhibition and apoptosis induction were noticed in AGS and MKN45 cell lines, where Britanin suppressed the NF- $\kappa$ B pathway by increasing the mRNA and protein levels of peroxisome proliferator-activated receptor- $\gamma$  (PPAR $\gamma$ ). Upregulation of BAX and downregulation of BCL-2, matrix metalloproteinase-9 (MMP-9), Twist family bHLH transcription factor 1 (TWIST-1), and cyclooxygenase-2 (COX-2) were also noted. The authors concluded that Britanin is an encouraging anti-cancer agent that still requires further examination [24].

The main biological and molecular findings from the above studies are briefly summarized in Figure 1, whereas available  $\text{IC}_{50}$  values are collected in Table 1. It is worth recapitulating a few aspects that make Britanin a promising anti-cancer agent. Above data certify that the compound exhibits pleiotropic biological activity, providing a multimodal approach against gastrointestinal tumors. Combining these properties with the impact of Britanin on the PD-1/PD-L1 pathway[4,23], it seems that the compound might be valuable for both chemotherapeutic and immunotherapeutic settings. Moreover, available studies report that the cytotoxic effect of Britanin on noncancerous cells is significantly lower than that on tumor cells, while still being effective against the latter[18,19,22]. Ultimately, it has been suggested that Britanin could be safer than small molecule inhibitors[22], which are currently used for targeting gastrointestinal tumors [25,26].

## FUTURE PROSPECTS

A wealth of data on the effect of Britanin has been obtained from studies other than those associated with liver, colorectal, pancreatic, and gastric cancer. Such information may suggest the direction of further research on gastrointestinal tumors.

Table 1 Efficacy of Britanin from various plant sources in inhibiting gastrointestinal cancer cell lines				
Gastrointestinal tumor	Cell line	Source of Britanin	IC <sub>50</sub> (μmol/L)	Ref.
Liver cancer	HuH-7	<i>Inula aucheriana</i>	27.86 ± 1.35 <sup>3</sup>	Cui <i>et al</i> [19]
Liver cancer	SMMC-7721	<i>Inula aucheriana</i>	28.92 ± 1.09 <sup>3</sup>	Cui <i>et al</i> [19]
Liver cancer	HepG2	<i>Inula aucheriana</i>	15.69 ± 1.58 <sup>3</sup>	Cui <i>et al</i> [19]
Liver cancer	HepG2	<i>Inula aucheriana</i>	6.004 <sup>1,4</sup>	Moghadam <i>et al</i> [14]
Liver cancer	HepG2	<i>Inula japonica</i>	35.5 <sup>5</sup>	Piao <i>et al</i> [17]
Liver cancer	HepG2	Unspecified <sup>2</sup>	6.006 <sup>5</sup>	Li <i>et al</i> [21]
Liver cancer	BEL-7402	Unspecified <sup>2</sup>	2.702 <sup>5</sup>	Li <i>et al</i> [21]
Colorectal cancer	HT-29	<i>Inula japonica</i>	3.9 <sup>5</sup>	Piao <i>et al</i> [17]
Pancreatic cancer	MIA CaPa-2	<i>Inula linearifolia</i>	3.104 <sup>4</sup>	Li <i>et al</i> [22]
Pancreatic cancer	BxPC-3	<i>Inula linearifolia</i>	3.367 <sup>4</sup>	Li <i>et al</i> [22]
Pancreatic cancer	PANC-1	<i>Inula linearifolia</i>	1.348 <sup>4</sup>	Li <i>et al</i> [22]
Pancreatic cancer	PANC-1	<i>Inula aucheriana</i>	40 ± 5.63 <sup>3</sup>	Moeinifard <i>et al</i> [18]
Pancreatic cancer	AsPC-1	<i>Inula aucheriana</i>	30 ± 4.61 <sup>3</sup>	Moeinifard <i>et al</i> [18]
Gastric cancer	BGC-832	<i>Inula japonica</i>	4.999 <sup>4</sup>	Shi <i>et al</i> [20]
Gastric cancer	SGC-7901	<i>Inula japonica</i>	2.243 <sup>4</sup>	Shi <i>et al</i> [20]

<sup>1</sup>Recalculated from μg/mL to μmol/L to standardize the unit (molecular weight of Britanin, i.e., 366.4 g/mol, was acquired from PubChem 2.1).

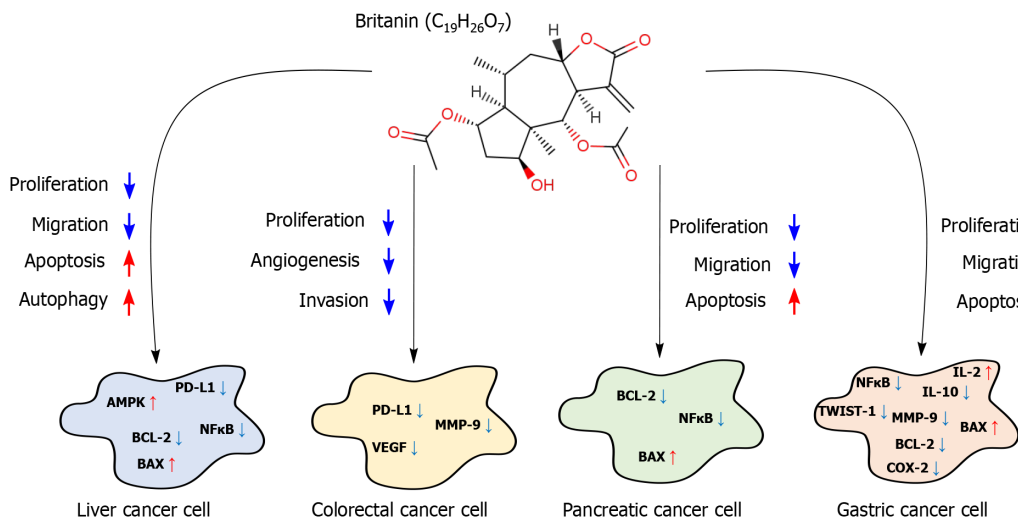
<sup>2</sup>Unspecified Britanin source (non-open access paper with no data in abstract).

<sup>3</sup>24-h incubation time with Britanin.

<sup>4</sup>72-h incubation time with Britanin.

<sup>5</sup>Unspecified incubation time with Britanin (non-open access paper or no data).

IC<sub>50</sub>: Half-maximal inhibitory concentration.



**Figure 1 Influence of Britanin on biological processes and related proteins in gastrointestinal tumors.** A red upward pointing arrow (“↑”) indicates biological process activation by Britanin, whereas a blue downward pointing arrow (“↓”) signifies biological process inhibition by the same compound. Similar applies to the level of proteins, the symbols of which are located in four multicolored areas representing liver, colorectal, pancreatic, and gastric cancer cells.

Firstly, Hajimehdipoor *et al*[27] discovered that three sesquiterpene lactones extracted from *Inula aucheriana* hold promise as inhibitors of acetylcholinesterase (AChE). While the research was primarily focused on Alzheimer’s disease, Britanin emerged as the second most potent inhibitor of AChE, exhibiting 25.2% inhibitory activity at a concentration of 300 μg/mL. The researchers suggest that altering the structure of Britanin could enhance its AChE inhibitory potential and reduce its cytotoxicity[27]. This could be of value in cancer treatment, as the cholinergic system and AChE activity are known to play important roles in tumor development and microenvironmental alterations[28]. Modifying the structure of Britanin to reduce cytotoxicity is noteworthy since gastrointestinal toxicity remains a common complication



of cytotoxic anti-cancer chemotherapy[29].

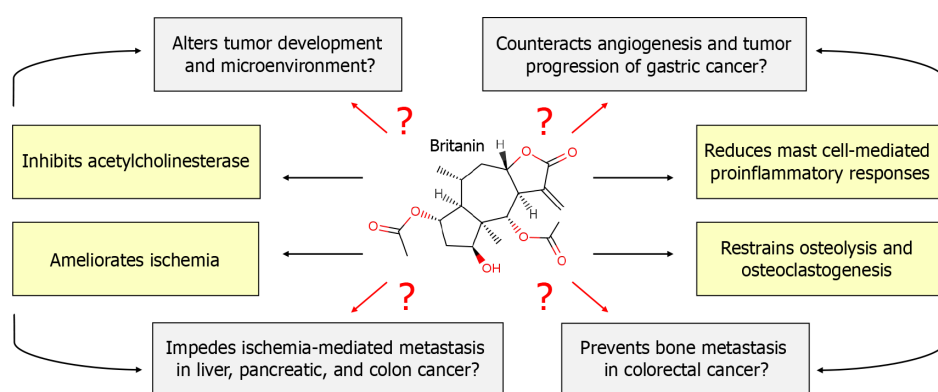
Secondly, gastrointestinal research on Britanin should be directed at mast cells, which appear to play pro-tumorigenic and anti-tumorigenic roles[30]. Lu *et al*[31] assessed the anti-allergic activity of an *Inula japonica* extract *in vivo* and investigated its mode of action on mast cells *in vitro*. Britanin was found to be one of the most abundant sesquiterpenes. The extract attenuated the mast cell-mediated passive cutaneous anaphylaxis reaction and exhibited an anti-allergic effect by modulating eicosanoid generation and degranulation *in vitro*[31]. Park *et al*[32] found *Inula japonica*-derived Britanin to ameliorate mast cell-mediated pro-inflammatory responses, which they attributed to NF- $\kappa$ B activation. Similarly, Lu *et al*[33] found the mast cell-suppressing ability of Britanin to be associated with the inhibition of the spleen tyrosine kinase (Syk) pathway *via* Syk protein dephosphorylation, as well as deactivation of NF- $\kappa$ B and mitogen-activated protein kinases.

It has been observed that mast cell density appears to correlate with angiogenesis and progression in patients with gastric carcinoma[34]. Moreover, mast cells were found to be abundant in gastric cancer, which shorten patient survival [35]. The latter study also revealed that cancer-derived tumor necrosis factor alpha induces PD-L1 overexpression in mast cells *via* activation of the NF- $\kappa$ B signaling pathway. PD-L1+ mast cells suppressed T-cell growth and function in a PD-L1-dependent manner. Given that Britanin is associated with NF- $\kappa$ B, PD-L1, and T-cells, future gastrointestinal research should include Britanin and mast cells.

Thirdly, Britanin has been found to inhibit osteoclastogenesis and osteolysis. The compound inhibited osteoclast differentiation by downregulation of B lymphocyte-induced maturation protein 1 and nuclear factor of activated T cells 1 *in vitro*, as well as protected bone from titanium-induced calvarial osteolysis *in vivo*[36]. Although osteolysis is a complication among patients carrying titanium-based implants after long-term usage[37], it also occurs as an outcome of bone metastasis in colorectal cancer. The mechanism by which colorectal cancer cells influence the differentiation of bone marrow-derived monocytes into osteoclasts has been described previously[38]. However, further studies are needed to confirm whether Britanin can prevent metastasis of colorectal cancer while also counteracting the tumor itself.

Lastly, Britanin was found to relieve ischemic injury, a phenomenon characterized by tissue damage due to the lack of perfusion and oxygenation. Although a higher risk of hypoxia is typically associated with organ transplantation, the tumor microenvironment is similar to ischemic tissue in this regard[39]. Outside the gastrointestinal context, Britanin was found to ameliorate cerebral and myocardial ischemia *via* pathways incorporating the nuclear factor erythroid 2-related factor 2, which is one of the most important defenders against oxidative stress[40,41]. Thus, Britanin might be an important protector against negative outcomes of oxidative stress, to which rapidly dividing cells of colonic mucosa are steadily exposed[42]. Moreover, subsequent research on gastrointestinal tumors is necessary, since ischemia mediates metastasis in liver, pancreatic, and colon cancer[43-45].

The novel research directions which could be adopted in future gastrointestinal research on Britanin are recapitulated in Figure 2. Regardless of the topic, any studies of the relationship between Britanin and its influence on signaling pathways or the proteome should be supported by molecular docking. Existing data indicates that Britanin interacts with such essential proteins as NF- $\kappa$ B, PD-L1, Myc, and HIF-1 $\alpha$ [4,12,23,46], and it may also influence other important proteins and pathways, such as BCL-2, BAX, AMPK, MMP-9, TWIST-1, COX-2, or PPAR $\gamma$ .



**Figure 2** Novel research directions which could be adopted in future gastrointestinal research on Britanin. The light-yellow rectangles represent data on Britanin obtained from studies other than those associated with liver, colorectal, pancreatic, and gastric cancer. Processes included therein are linked to various tumor-related phenomena, which are depicted in gray rectangles. Britanin was not yet investigated in these tumor-related phenomena, which was marked with solid red arrows and question marks ("?"). Such information may suggest the direction of further research on gastrointestinal tumors.

## CONCLUSION

Britanin is a natural compound that counteracts liver, colorectal, pancreatic, and gastric tumors by regulating proliferation, apoptosis, autophagy, immune response, migration, and angiogenesis. Its cytotoxicity on noncancerous cells is significantly lower than that on tumor cells, while still being effective against the latter, warranting further in-depth studies based on animal models. The ability to reduce the cytotoxicity of Britanin *via* structural modification may be useful in limiting gastrointestinal toxicity after cytotoxic anti-cancer chemotherapy. Outside the chemotherapeutic context, Britanin might also be valuable in an immunotherapeutic setting since it affects the PD-1/PD-L1 pathway. The

compound acts against negative outcomes of oxidative stress, to which rapidly dividing cells of colonic mucosa are steadily exposed. Given the pleiotropic biological activity, Britanin ensures a multimodal approach against gastrointestinal tumors, which may provide additional treatment options or reduce the high mortality rate. However, it has yet to be included in clinical trials as no data on its use exists in the National Institutes of Health. Future research should incorporate molecular docking simulations and focus on the link between Britanin and acetylcholinesterase, mast cells, osteolysis, and ischemia, as considerable data on its potential already exists outside the gastrointestinal context.

## FOOTNOTES

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## REFERENCES

- Matos MS, Anastácio JD, Nunes Dos Santos C. Sesquiterpene Lactones: Promising Natural Compounds to Fight Inflammation. *Pharmaceutics* 2021; **13** [PMID: 34208907 DOI: 10.3390/pharmaceutics13070991]
- Ranasinghe S, Armson A, Lymbery AJ, Zahedi A, Ash A. Medicinal plants as a source of antiparasitics: an overview of experimental studies. *Pathog Glob Health* 2023; **117**: 535-553 [PMID: 36805662 DOI: 10.1080/20477724.2023.2179454]
- Paço A, Brás T, Santos JO, Sampaio P, Gomes AC, Duarte MF. Anti-Inflammatory and Immunoregulatory Action of Sesquiterpene Lactones. *Molecules* 2022; **27** [PMID: 35164406 DOI: 10.3390/molecules27031142]
- Vergoten G, Bailly C. Molecular docking study of britanin binding to PD-L1 and related anticancer pseudoguaianolide sesquiterpene lactones. *J Recept Signal Transduct Res* 2022; **42**: 454-461 [PMID: 34789056 DOI: 10.1080/10799893.2021.2003816]
- Park HH, Kim MJ, Li Y, Park YN, Lee J, Lee YJ, Kim SG, Park HJ, Son JK, Chang HW, Lee E. Britanin suppresses LPS-induced nitric oxide, PGE2 and cytokine production via NF-kappaB and MAPK inactivation in RAW 264.7 cells. *Int Immunopharmacol* 2013; **15**: 296-302 [DOI: 10.1016/j.intimp.2012.12.005]
- Mohammadlou H, Hamzeloo-Moghadam M, Yami A, Feizi F, Moeinifard M, Gharehbaghian A. Britanin a Sesquiterpene Lactone from *Inula aucheriana* Exerted an Anti-leukemic Effect in Acute Lymphoblastic Leukemia (ALL) Cells and Enhanced the Sensitivity of the Cells to Vincristine. *Nutr Cancer* 2022; **74**: 965-977 [PMID: 34060394 DOI: 10.1080/01635581.2021.1931700]
- Mohammadlou H, Hamzeloo-Moghadam M, Mohammadi MH, Yami A, Gharehbaghian A. Britanin, a sesquiterpene lactone induces ROS-dependent apoptosis in NALM-6, REH, and JURKAT cell lines and produces a synergistic effect with vincristine. *Mol Biol Rep* 2021; **48**: 6249-6258 [PMID: 34478011 DOI: 10.1007/s11033-021-06572-x]
- Mohammadlou H, Hamzeloo-Moghadam M, Moeinifard M, Gharehbaghian A. Cytotoxic Effects of Britanin on Acute and Chronic Myeloid Leukemia Cells Through Inducing p21-Mediated Apoptotic Cell Death. *Turk J Pharm Sci* 2022; **19**: 314-321 [PMID: 35775388 DOI: 10.4274/tjps.galenos.2021.88655]
- Hamzeloo-Moghadam M, Aghaei M, Fallahian F, Jafari SM, Dolati M, Abdolmohammadi MH, Hajiahmadi S, Esmacili S. Britanin, a sesquiterpene lactone, inhibits proliferation and induces apoptosis through the mitochondrial signaling pathway in human breast cancer cells. *Tumour Biol* 2015; **36**: 1191-1198 [PMID: 25342596 DOI: 10.1007/s13277-014-2744-9]
- Hamzeloo-Moghadam M, Aghaei M, Abdolmoham Madi MH, Fallahian F. Anticancer activity of britanin through the downregulation of cyclin D1 and CDK4 in human breast cancer cells. *J Cancer Res Ther* 2019; **15**: 1105-1108 [PMID: 31603118 DOI: 10.4103/jcrt.jcrt\_517\_17]
- Lu H, Wu Z, Wang Y, Zhao D, Zhang B, Hong M. Study on inhibition of Britanin on triple-negative breast carcinoma through degrading ZEB1 proteins. *Phytomedicine* 2022; **104**: 154291 [PMID: 35839735 DOI: 10.1016/j.phymed.2022.154291]
- Xu X, Guo Y, Du G, Liu H, Wang L, Chen D. Bioluminescence Imaging-Based Assessment of the Anti-Triple-Negative Breast Cancer and NF-Kappa B Pathway Inhibition Activity of Britanin. *Front Pharmacol* 2020; **11**: 575 [PMID: 32431613 DOI: 10.3389/fphar.2020.00575]
- Kumar S, Das A. A Cocktail of Natural Compounds Holds Promise for New Immunotherapeutic Potential in Head and Neck Cancer. *Chin J Integr Med* 2024; **30**: 42-51 [PMID: 37118529 DOI: 10.1007/s11655-023-3694-0]
- Moghadam MH, Hajimehdipoor H, Saeidnia S, Atoofi A, Shahrestani R, Read RW, Mosaddegh M. Anti-proliferative activity and apoptotic potential of britanin, a sesquiterpene lactone from *Inula aucheriana*. *Nat Prod Commun* 2012; **7**: 979-980 [PMID: 22978209 DOI: 10.1002/npc.10000]

- 10.1177/1934578X1200700804]
- 15 **Zeng Q**, Zeng Y, Nie X, Guo Y, Zhan Y. Britanin Exhibits Potential Inhibitory Activity on Human Prostate Cancer Cell Lines Through PI3K/Akt/NF- $\kappa$ B Signaling Pathways. *Planta Med* 2020; **86**: 1401-1410 [PMID: 32781474 DOI: 10.1055/a-1211-4656]
  - 16 **Fischedick JT**, Pesic M, Podolski-Renic A, Bankovic J, de Vos RCH, Perić M, Todorović S, Tanic N. Cytotoxic activity of sesquiterpene lactones from *Inula britannica* on human cancer cell lines. *Phytochem Lett* 2013; **6**: 246-252 [DOI: 10.1016/j.phytol.2013.02.006]
  - 17 **Piao D**, Kim T, Zhang HY, Choi HG, Lee CS, Choi HJ, Chang HW, Woo MH, Son JK. DNA Topoisomerase Inhibitory Activity of Constituents from the Flowers of *Inula japonica*. *Chem Pharm Bull (Tokyo)* 2016; **64**: 276-281 [PMID: 26936053 DOI: 10.1248/cpb.c15-00780]
  - 18 **Moeinifard M**, Hassan ZM, Fallahian F, Hamzeloo-Moghadam M, Taghikhani M. Britannin induces apoptosis through AKT-FOXO1 pathway in human pancreatic cancer cells. *Biomed Pharmacother* 2017; **94**: 1101-1110 [PMID: 28821161 DOI: 10.1016/j.biopha.2017.08.025]
  - 19 **Cui YQ**, Liu YJ, Zhang F. The suppressive effects of Britannin (Bri) on human liver cancer through inducing apoptosis and autophagy via AMPK activation regulated by ROS. *Biochem Biophys Res Commun* 2018; **497**: 916-923 [PMID: 29288670 DOI: 10.1016/j.bbrc.2017.12.144]
  - 20 **Shi K**, Liu X, Du G, Cai X, Zhan Y. In vivo antitumour activity of Britanin against gastric cancer through nuclear factor- $\kappa$ B-mediated immune response. *J Pharm Pharmacol* 2020; **72**: 607-618 [PMID: 31943207 DOI: 10.1111/jphp.13230]
  - 21 **Li H**, Du G, Yang L, Pang L, Zhan Y. The Antitumor Effects of Britanin on Hepatocellular Carcinoma Cells and its Real-Time Evaluation by In Vivo Bioluminescence Imaging. *Anticancer Agents Med Chem* 2020; **20**: 1147-1156 [PMID: 32106805 DOI: 10.2174/1871520620666200227092623]
  - 22 **Li K**, Zhou Y, Chen Y, Zhou L, Liang J. A novel natural product, britanin, inhibits tumor growth of pancreatic cancer by suppressing nuclear factor- $\kappa$ B activation. *Cancer Chemother Pharmacol* 2020; **85**: 699-709 [PMID: 32185482 DOI: 10.1007/s00280-020-04052-w]
  - 23 **Zhang YF**, Zhang ZH, Li MY, Wang JY, Xing Y, Ri M, Jin CH, Xu GH, Piao LX, Zuo HX, Jin HL, Ma J, Jin X. Britannin stabilizes T cell activity and inhibits proliferation and angiogenesis by targeting PD-L1 via abrogation of the crosstalk between Myc and HIF-1 $\alpha$  in cancer. *Phytomedicine* 2021; **81**: 153425 [PMID: 33310309 DOI: 10.1016/j.phymed.2020.153425]
  - 24 **Abdolmohammadi MH**, Roozbehani M, Hamzeloo-Moghadam M, Heidari F, Fallahian F. Targeting PPAR $\gamma$ /NF- $\kappa$ B Signaling Pathway by Britannin, a Sesquiterpene Lactone from *Inula aucheriana* DC., in Gastric Cancer. *Anticancer Agents Med Chem* 2023; **23**: 2102-2110 [PMID: 37723632 DOI: 10.2174/1871520623666230918140559]
  - 25 **Liu GH**, Chen T, Zhang X, Ma XL, Shi HS. Small molecule inhibitors targeting the cancers. *MedComm (2020)* 2022; **3**: e181 [PMID: 36254250 DOI: 10.1002/mco2.181]
  - 26 **Godesi S**, Lee J, Nada H, Quan G, Elkamhawry A, Choi Y, Lee K. Small Molecule c-KIT Inhibitors for the Treatment of Gastrointestinal Stromal Tumors: A Review on Synthesis, Design Strategies, and Structure-Activity Relationship (SAR). *Int J Mol Sci* 2023; **24** [PMID: 37298401 DOI: 10.3390/ijms24119450]
  - 27 **Hajimehdipoor H**, Mosaddegh M, Naghibi F, Haeri A, Hamzeloo-Moghadam M. Natural sesquiterpene lactones as acetylcholinesterase inhibitors. *An Acad Bras Cienc* 2014; **86**: 801-806 [PMID: 24838542 DOI: 10.1590/0001-3765201420130005]
  - 28 **Pérez-Aguilar B**, Marquardt JU, Muñoz-Delgado E, López-Durán RM, Gutiérrez-Ruiz MC, Gomez-Quiroz LE, Gómez-Olivares JL. Changes in the Acetylcholinesterase Enzymatic Activity in Tumor Development and Progression. *Cancers (Basel)* 2023; **15** [PMID: 37760598 DOI: 10.3390/cancers15184629]
  - 29 **Akbarali HI**, Muchhala KH, Jessup DK, Cheatham S. Chemotherapy induced gastrointestinal toxicities. *Adv Cancer Res* 2022; **155**: 131-166 [PMID: 35779873 DOI: 10.1016/bs.acr.2022.02.007]
  - 30 **Molfetta R**, Paolini R. The Controversial Role of Intestinal Mast Cells in Colon Cancer. *Cells* 2023; **12** [PMID: 36766801 DOI: 10.3390/cells12030459]
  - 31 **Lu Y**, Li Y, Jin M, Yang JH, Li X, Chao GH, Park HH, Park YN, Son JK, Lee E, Chang HW. *Inula japonica* extract inhibits mast cell-mediated allergic reaction and mast cell activation. *J Ethnopharmacol* 2012; **143**: 151-157 [PMID: 22728246 DOI: 10.1016/j.jep.2012.06.015]
  - 32 **Park HH**, Kim SG, Park YN, Lee J, Lee YJ, Park NY, Jeong KT, Lee E. Suppressive effects of britanin, a sesquiterpene compound isolated from *Inulae flos*, on mast cell-mediated inflammatory responses. *Am J Chin Med* 2014; **42**: 935-947 [PMID: 25004884 DOI: 10.1142/S0192415X14500591]
  - 33 **Lu Y**, Li X, Park YN, Kwon O, Piao D, Chang YC, Kim CH, Lee E, Son JK, Chang HW. Britanin Suppresses IgE/Ag-Induced Mast Cell Activation by Inhibiting the Syk Pathway. *Biomol Ther (Seoul)* 2014; **22**: 193-199 [PMID: 25009699 DOI: 10.4062/biomolther.2014.038]
  - 34 **Ribatti D**, Guidolin D, Marzullo A, Nico B, Annese T, Benagiano V, Crivellato E. Mast cells and angiogenesis in gastric carcinoma. *Int J Exp Pathol* 2010; **91**: 350-356 [PMID: 20412338 DOI: 10.1111/j.1365-2613.2010.00714.x]
  - 35 **Lv Y**, Zhao Y, Wang X, Chen N, Mao F, Teng Y, Wang T, Peng L, Zhang J, Cheng P, Liu Y, Kong H, Chen W, Hao C, Han B, Ma Q, Zou Q, Chen J, Zhuang Y. Increased intratumoral mast cells foster immune suppression and gastric cancer progression through TNF- $\alpha$ -PD-L1 pathway. *J Immunother Cancer* 2019; **7**: 54 [PMID: 30808413 DOI: 10.1186/s40425-019-0530-3]
  - 36 **Kim JA**, Lim S, Ihn HJ, Kim JE, Yea K, Moon J, Choi H, Park EK. Britanin inhibits titanium wear particle-induced osteolysis and osteoclastogenesis. *Mol Med Rep* 2023; **28** [PMID: 37732549 DOI: 10.3892/mmr.2023.13092]
  - 37 **Saadi SB**, Ranjbarzadeh R, Ozeir Kazemi, Amirabadi A, Ghouschi SJ, Kazemi O, Azadikhah S, Bendeche M. Osteolysis: A Literature Review of Basic Science and Potential Computer-Based Image Processing Detection Methods. *Comput Intell Neurosci* 2021; **2021**: 4196241 [PMID: 34646317 DOI: 10.1155/2021/4196241]
  - 38 **Zi-Chen G**, Jin Q, Yi-Na Z, Wei W, Xia K, Wei X, Juan W, Wei Z. Colorectal cancer cells promote osteoclastogenesis and bone destruction through regulating EGF/ERK/CCL3 pathway. *Biosci Rep* 2020; **40** [PMID: 32478376 DOI: 10.1042/BSR20201175]
  - 39 **Nemeth DV**, Baldini E, Sorrenti S, D'Andrea V, Bellini MI. Cancer Metabolism and Ischemia-Reperfusion Injury: Two Sides of the Same Coin. *J Clin Med* 2022; **11** [PMID: 36079025 DOI: 10.3390/jcm11175096]
  - 40 **Wu G**, Zhu L, Yuan X, Chen H, Xiong R, Zhang S, Cheng H, Shen Y, An H, Li T, Li H, Zhang W. Britanin Ameliorates Cerebral Ischemia-Reperfusion Injury by Inducing the Nrf2 Protective Pathway. *Antioxid Redox Signal* 2017; **27**: 754-768 [PMID: 28186440 DOI: 10.1089/ars.2016.6885]
  - 41 **Lu H**, Xiao H, Dai M, Xue Y, Zhao R. Britanin relieves ferroptosis-mediated myocardial ischaemia/reperfusion damage by upregulating GPX4 through activation of AMPK/GSK3 $\beta$ /Nrf2 signalling. *Pharm Biol* 2022; **60**: 38-45 [PMID: 34860639 DOI: 10.1080/13880209.2021.2007269]
  - 42 **Atay AE**, Esen B, Gokmen ES. Oxidative Stress and Gastrointestinal System Cancers. *Gastrointestinal Tissue* 2017; 29-51 [DOI: 10.1016/b978-0-12-805377-5.00003-5]
  - 43 **van der Blit JD**, Kranenburg O, Nijkamp MW, Smakman N, Veenendaal LM, Te Velde EA, Voest EE, van Diest PJ, Borel Rinkes IH. Ischemia/reperfusion accelerates the outgrowth of hepatic micrometastases in a highly standardized murine model. *Hepatology* 2005; **42**: 165-

- 175 [PMID: [15962318](#) DOI: [10.1002/hep.20739](#)]
- 44 **Tashiro Y**, Nishino H, Higuchi T, Sugisawa N, Fukuda Y, Yamamoto J, Inubushi S, Aoki T, Murakami M, Singh SR, Bouvet M, Hoffman RM. Ischemia reperfusion-induced metastasis is resistant to PPAR $\gamma$  agonist pioglitazone in a murine model of colon cancer. *Sci Rep* 2020; **10**: 18565 [PMID: [33122687](#) DOI: [10.1038/s41598-020-75210-6](#)]
- 45 **Yoshimoto K**, Tajima H, Ohta T, Okamoto K, Sakai S, Kinoshita J, Furukawa H, Makino I, Hayashi H, Nakamura K, Oyama K, Inokuchi M, Nakagawara H, Itoh H, Fujita H, Takamura H, Ninomiya I, Kitagawa H, Fushida S, Fujimura T, Wakayama T, Iseki S, Shimizu K. Increased E-selectin in hepatic ischemia-reperfusion injury mediates liver metastasis of pancreatic cancer. *Oncol Rep* 2012; **28**: 791-796 [PMID: [22766603](#) DOI: [10.3892/or.2012.1896](#)]
- 46 **Bailly C**. Anticancer Targets and Signaling Pathways Activated by Britannin and Related Pseudoguaianolide Sesquiterpene Lactones. *Biomedicines* 2021; **9** [PMID: [34680439](#) DOI: [10.3390/biomedicines9101325](#)]



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