

## Interplay of adipokines and myokines in cancer pathophysiology: Emerging therapeutic implications

Maria Dalamaga

Maria Dalamaga, Department of Clinical Biochemistry, Medical School, University of Athens, "Attikon" General University Hospital, 12462 Athens, Greece

Author contributions: Dalamaga M solely contributed to this paper.

Correspondence to: Maria Dalamaga, MD, PhD, MS, MPH, Assistant Professor, Department of Clinical Biochemistry, Medical School, University of Athens, "Attikon" General University Hospital, 1 Rimini Street, 12462 Athens, Greece. [madalamaga@med.uoa.gr](mailto:madalamaga@med.uoa.gr)

Telephone: +30-210-5831915 Fax: +30-210-6082467

Received: June 23, 2013 Revised: July 19, 2013

Accepted: August 16, 2013

Published online: August 20, 2013

### Abstract

Excess body weight constitutes a worldwide health problem with epidemic proportions impacting on the risk and prognosis of several disease states including malignancies. It is believed that the metabolic changes associated with weight gain, particularly visceral obesity, and physical inactivity could lead to dysfunctional adipose and muscle tissues causing insulin resistance, low-grade chronic inflammation and abnormal secretion of adipokines and myokines. The complex paracrine and endocrine interconnection between adipokines and myokines reflects a yin-yang balance with important implications in processes such as lipolysis control, insulin sensitivity and prevention from obesity-driven chronic low-grade inflammation and cancer promotion through anti-inflammatory adipokines and myokines. Furthermore, the complex pathophysiology of cancer cachexia is based on the interplay between muscle and adipose tissue mediated by free fatty acids, various adipokines and myokines. The purpose of this editorial is to explore the role of the adipose and muscle tissue interplay in carcinogenesis, cancer progression and cachexia, and to examine the mechanisms underpinning their association with malignancy. Understanding of

the mechanisms connecting the interplay of adipokines and myokines with cancer pathophysiology is expected to be of importance in the development of therapeutic strategies against cancer cachexia. Advances in the field of translational investigation may lead to tangible benefits to obese and inactive persons who are at increased risk of cancer as well as to cancer patients with cachexia.

© 2013 Baishideng. All rights reserved.

**Key words:** Adipokine; Myokine; Cancer; Cachexia; Interleukin-15; Interleukin-6; Obesity; Myostatin

**Core tip:** The complex paracrine and endocrine interconnection between adipokines and myokines reflects a yin-yang balance with important implications in processes such as lipolysis control, insulin sensitivity and prevention from obesity-driven chronic low-grade inflammation and cancer promotion through anti-inflammatory adipokines and myokines. In addition, the complex pathophysiology of cancer cachexia is based on the interplay between muscle and adipose tissue mediated by free fatty acids, various adipokines and myokines. Advances in the field of translational investigation may lead to tangible benefits to obese and inactive persons who are at increased risk of cancer as well as to cancer patients with cachexia.

Dalamaga M. Interplay of adipokines and myokines in cancer pathophysiology: Emerging therapeutic implications. *World J Exp Med* 2013; 3(3): 26-33 Available from: URL: <http://www.wjgnet.com/2220-315X/full/v3/i3/26.htm> DOI: <http://dx.doi.org/10.5493/wjem.v3.i3.26>

### INTRODUCTION

Excess body weight constitutes a worldwide health prob-

lem with epidemic proportions impacting on the risk and prognosis of several disease states including cardiovascular disease (CVD), type 2 diabetes mellitus (t2DM) and common forms of cancer, such as colon cancer, postmenopausal breast cancer, endometrial cancer, renal cell cancer and esophageal adenocarcinoma<sup>[1-10]</sup>. Globally, about 25% of cancer cases are due to overweight/obesity and sedentary lifestyle<sup>[11]</sup>.

Obesity prevents muscle gain and the combination of obesity and loss of muscle mass could lead to elevated health risks including obesity-associated malignancies. It is believed that the metabolic changes associated with weight gain, particularly visceral obesity, and physical inactivity could lead to dysfunctional adipose and muscle tissues causing insulin resistance, low-grade chronic inflammation and abnormal secretion of adipokines and myokines<sup>[6,12,13]</sup>. Therefore, the adipose-muscle cross-talk plays a critical role in cancer promotion. On the other hand, in the context of cancer cachexia which characterizes cancer patients with advanced stage, the interplay between adipose tissue and skeletal muscle that occurs through adipokines and myokines is an exciting field of research with emerging novel therapeutic implications<sup>[14-16]</sup>.

The purpose of this editorial is to explore the role of the adipose and muscle tissue interplay in carcinogenesis, cancer progression and cachexia, and to examine the mechanisms underpinning their association with malignancy. Understanding of the mechanisms connecting the interplay of adipokines and myokines with cancer pathophysiology is expected to be of importance in the development of preventive and therapeutic strategies against cancer.

## INTERPLAY OF ADIPOKINES AND MYOKINES IN CANCER ETIOPATHOGENESIS

### **Adipose tissue, main adipokines and cancer**

In addition to its inert lipid-storing capacity, adipose tissue represents the largest endocrine organ modulating energy homeostasis, metabolism, inflammation, immunity and endocrine balance<sup>[6]</sup>. Adipose tissue synthesizes and secretes more than fifty hormones and cytokines, known as adipokines<sup>[6]</sup>. As adipose tissue expands in obesity, the amount of anti-inflammatory adipokines, particularly adiponectin, decreases and the amount of pro-inflammatory adipokines with an oncogenic potential, such as leptin, resistin, visfatin and chemerin, and cytokines such as tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), interleukin (IL)-1 and IL-6 increases<sup>[6,17]</sup>. Obesity-driven chronic low-grade inflammation is also involved in insulin resistance (IR), which is characterized by hyperinsulinemia, increased levels of growth factors such as insulin-like growth factor-I (IGF-I) and activation of transcription factors participating in pro-inflammatory response and cell-cycle regulation, like nuclear factor kappa-B (NF- $\kappa$ B), which can promote carcinogenesis<sup>[6,17,18]</sup>. Important cancer-related adipokine effects are summarized below.

Adiponectin is a 30-kDa, 244-amino-acid adipokine exerting insulin-sensitizing, anti-inflammatory and anti-neoplastic effects<sup>[6]</sup>. The majority of epidemiologic evidence has connected *in vivo* hypoadiponectinemia with an increased risk for IR, metabolic syndrome (Mets), t2DM, CVD and obesity-associated malignancies<sup>[6,19]</sup> as well as with a more aggressive cancer phenotype characterized by higher histologic grade, large size of tumor, lymph node invasion, distal metastases or estrogen receptor negativity for breast cancer<sup>[6,20-25]</sup>. In summary, adiponectin presents anti-tumorigenic effects *via* two mechanisms: (1) it can act directly on cancer cells by modulating receptor-mediated signaling pathways, including mitogen-activated protein kinase (MAPK), AMP-activated protein kinase (AMPK), Wnt/ $\beta$ -catenin and estrogen receptor (ER) signaling; and (2) it can act indirectly by regulating insulin sensitivity, influencing tumor angiogenesis and modulating inflammatory responses by inhibiting NF- $\kappa$ B signaling<sup>[6,24,25]</sup>. On the contrary, leptin, a 167-amino acid pleiotropic adipokine that regulates food intake, energy expenditure, immunity, and inflammation<sup>[26,27]</sup>, has been shown *in vitro* to promote growth and proliferation of neoplastic cells *via* activation of various growth and survival signaling pathways including canonical: Janus kinase 2/signal transducer and activator of transcription 3 (JAK2/STAT3), phosphatidylinositol 3-kinase/v-Akt murine thymoma viral oncogene homolog/mammalian target of rapamycin (PI3K/Akt/mTOR), MAPK/Extracellular signal-related kinase 1/2 (ERK1/2) and non-canonical signaling pathways such as protein kinase C, c-Jun N-terminal kinase (JNK) and p38 MAPK<sup>[25-29]</sup>. Additionally, leptin may act indirectly by diminishing insulin tissue sensitivity causing hyperinsulinemia, by shifting inflammatory responses towards a T-helper 1 phenotype with oversecretion of pro-inflammatory cytokines and by influencing tumor angiogenesis; though such leptin effects were not seen *in vivo*<sup>[26,27]</sup>. Resistin, another pro-inflammatory adipokine synthesized predominantly in visceral macrophages in humans, is a 12 kDa cysteine-rich polypeptide<sup>[30-32]</sup>. Visfatin or nicotinamide phosphoribosyl-transferase (Nampt), a novel pleiotropic adipokine found in the visceral fat, acts as a pro-inflammatory cytokine, a growth factor and an enzyme in the cellular energy metabolism, particularly nicotinamide adenine dinucleotide (NAD) biosynthesis, which is required in a plethora of intracellular processes such as redox reactions, DNA repair, transcriptional regulation and activity of poly-ADP ribosyltransferases (PARPs) and deacetylases (sirtuins) modulating cell survival and cytokine responses<sup>[33-35]</sup>. The majority of epidemiologic studies has indicated that *in vivo* hyperresistinemia and hypervisfatinemia are associated with some obesity-related malignancies such as colon cancer, postmenopausal breast cancer and prostate cancer<sup>[7,31-34,36-42]</sup>; though their ontological role in the association between obesity and cancer needs to be clarified. Resistin and visfatin may: (1) upregulate pro-inflammatory cytokines *via* the NF- $\kappa$ B pathway<sup>[32,33]</sup>; (2) stimulate signaling pathways which are

important components of cancer-promoting machinery<sup>[32,33,41-43]</sup>; and (3) induct pro-angiogenic proteins such as the vascular endothelial growth (VEGF) and the expression of metalloproteases (MMPs) participating in tumor invasiveness and metastasis<sup>[32,33]</sup>. Much less is known about a novel pro-inflammatory adipokine, chemerin, which is found elevated in obese individuals<sup>[44]</sup>. Chemerin may cause IR in human skeletal muscle at the level of glycogen synthase kinase 3 (GSK3) and Akt phosphorylation, and glucose uptake. Finally, chemerin may activate signaling pathways pertinent to inflammation and cancer promotion, such as NF- $\kappa$ B, p38 MAPK and ERK 1/2<sup>[45]</sup>.

### **Skeletal muscle, main myokines and cancer prevention**

Skeletal muscle accounts approximately for 40% of body weight in non-obese individuals, constituting therefore the largest human organ<sup>[46]</sup>. There has been accumulating evidence that skeletal muscle is an important secretory organ producing several proteins and low molecular weight molecules<sup>[45,46]</sup>. Myokines are muscle-derived cytokines that exert autocrine/paracrine and endocrine effects. Myokines play a pivotal role in metabolism as mediators of muscle-to-adipose tissue cross-talk and regulators of muscular glucose and fat homeostasis, and in cancer prevention as mediators of the beneficial effects of physical activity counteracting the harmful effects of pro-inflammatory adipokines<sup>[45,46]</sup>. It seems that the complex paracrine and endocrine interconnection between adipokines and myokines reflects a yin-yang balance with important implications in processes such as lipolysis control, insulin sensitivity and prevention from obesity-driven chronic low-grade inflammation and cancer promotion through anti-inflammatory adipokines and myokines. At the same time, skeletal muscle cells may secrete adipokines such as adiponectin, which can exert beneficial local metabolic effects enhancing insulin sensitivity and inhibiting inflammatory processes<sup>[47]</sup>. It is important to underscore that adipose tissue is not the exclusive source of adipokines. Although adipose tissue constitutes the primary site of adipokines production, several adipokines are synthesized by both fat and muscle, playing a critical role for autocrine/paracrine loops<sup>[45]</sup>. For example, IL-6 and IL-8 are considered adipokines but also myokines with different roles in inflammation, exercise, skeletal muscle development and insulin sensitivity.

It is well known that physical activity offers protection against a variety of chronic diseases including obesity, t2DM, CVD, osteoporosis, depression and cancer<sup>[45]</sup>. Recent meta-analyses and epidemiological studies have underscored the protective effect of physical activity on reducing colorectal, prostate and breast cancer risk by 20%-40%<sup>[45]</sup>. Interestingly, moderate-intensity physical activity after breast and colorectal cancer diagnosis may improve prognosis and reduce the risk of cancer-specific and overall mortality<sup>[48-51]</sup>. Below is discussed the role of

major beneficial myokines.

IL-6 was the first described myokine produced in an exponential manner in response to muscle contraction after exercise in a strictly TNF-independent fashion<sup>[12,52]</sup>. IL-6 release from muscle is associated with exercise intensity and duration as well as muscle mass involved in the mechanical load<sup>[52]</sup>. Muscular IL-6 is involved in AMPK-mediated fat oxidation, skeletal muscle lipolysis and insulin-stimulated glucose uptake enhancing insulin sensitivity<sup>[12]</sup>. IL-6 also mediates some of the immunoregulatory and anti-inflammatory properties of regular exercise as it modulates TNF- $\alpha$  levels<sup>[52]</sup> and stimulates the secretion of classic anti-inflammatory cytokines such as IL-10 and IL-1ra<sup>[12]</sup>. In contrast to the beneficial effects of muscular IL-6, chronic elevated serum IL-6 levels synthesized by adipocytes and immune cells in the visceral adipose tissue are closely associated with *in vivo* IR, Mets, obesity and physical inactivity<sup>[12,45]</sup>. Interestingly, oncostatin M (OSM), a member belonging to the IL-6 superfamily, represents a pleiotropic myokine released by contracting myotubes<sup>[12]</sup>. OSM has been shown to exert *in vitro* important apoptotic effects on tumor cell lines by inhibiting proliferation in a variety of tissues comprising mammary epithelial cells, melanoma, ovarian and lung cells<sup>[12]</sup>.

IL-15 is a 15 kDa myokine that is highly expressed in skeletal muscle especially after aerobic exercise and resistance, and acts as a myokine that inhibits adiposity<sup>[13]</sup>. Apart from its hypertrophic and anabolic effects on muscle tissue as an authentic myokine, IL-15 exerts many metabolic actions by enhancing glucose uptake and fat oxidation in muscle tissue, stimulating lipolysis and inhibiting preadipocyte differentiation and lipogenesis as part of the muscle-adipose cross-talk<sup>[13]</sup>. Obese individuals exhibit low plasma IL-15 levels<sup>[46]</sup>. Interestingly, IL-15 may stimulate the production of anti-inflammatory and anti-neoplastic adiponectin downregulating visceral obesity while it reduces white adipocyte size and serum leptin levels in male mice<sup>[12]</sup>.

A new myokine, irisin, was recently discovered and named after the Greek messenger goddess Iris<sup>[46,53-55]</sup>. Physical activity increases the muscular expression levels of the transcriptional co-activator PGC-1 $\alpha$  upregulating the expression of the type I membrane protein FNDC5, which is C-terminally cleaved and secreted into the circulation as irisin<sup>[53]</sup>. In turn, irisin increases the expression of uncoupled protein-1 (UCP-1) contributing to the "browning" of white adipose tissue characterized by enhanced mitochondrial density, oxygen consumption and non-shivering thermogenesis<sup>[55]</sup>. Therefore, the muscle-derived irisin exhibits beneficial metabolic actions by increasing energy expenditure, causing small weight loss and improving metabolic parameters such as insulin signaling and sensitivity<sup>[55]</sup>. Basal plasma irisin levels may increase in response to 10 wk of regular exercise in humans and correlate with physical activity levels both in

mice and humans<sup>[46,54,55]</sup>.

## INTERPLAY OF ADIPOKINES AND MYOKINES IN CANCER CACHEXIA

Almost 50% of patients suffering from advanced cancer stage present cachexia which is responsible for 25% of deaths due to cancer<sup>[14-16]</sup>. Cachexia is a complex metabolic state characterized by loss of skeletal muscle mass and adipose tissue leading to progressive functional impairment. Cachexia is usually associated with asthenia, anorexia, anemia, weight loss, hypoalbuminemia, IR and abnormal metabolism of carbohydrates, lipids and proteins<sup>[56]</sup>. Cancer cachexia may be caused by anorexia, dysphagia related to advanced esophageal cancer, an imbalance between protein synthesis and catabolism with an increase in energy expenditure, or a combination of the two<sup>[14-16]</sup>. However, the complex pathophysiology of cancer cachexia is based on the interplay between muscle and adipose tissue mediated by free fatty acids, various adipokines and myokines<sup>[14,15]</sup>.

As cancer progresses, a variety of cytokines (IL-6 and TNF- $\alpha$ ) and tumor-derived mediators such as proteolysis-inducing factor (PIF) and parathyroid hormone-related protein (PTHrp), activate the pro-inflammatory catabolic cytokine cascade and deactivate the anti-inflammatory anabolic network (IL-4, IL-10, IL-12 and IL-15) leading to a systemic, chronic inflammation in cancer patients<sup>[16]</sup>. Pro-inflammatory and pro-cachectic cytokines, mainly TNF- $\alpha$ , IL-6 and interferon- $\alpha$ , and a lipid mobilizing factor (LIF), which is homologous to the soluble plasma protein Zinc- $\alpha$ 2-glycoprotein (ZAG), activate adipose triglyceride lipase (Atgl) triggering lipolysis which results in net mobilization of white adipose tissue and an augmentation of plasma free fatty acids levels<sup>[14,15]</sup>. Interestingly, ZAG, a recently identified 43-kDa adipokine, acts as a lipid-mobilizing factor stimulating lipolysis in adipocytes, and is enhanced in mice and humans with cancer cachexia<sup>[57,58]</sup>. Based on its lipid-mobilizing role, ZAG could also contribute to adipose tissue atrophy associated with cancer cachexia<sup>[58]</sup>. At the same time, the process of protein catabolism in cachexia starts and may be regulated by the cross-talk between adipose and muscle tissue mediated by free fatty acids, adipokines, cytokines and myokines. Interestingly, in cancer-bearing mice in which the *Atgl* gene is ablated, lipolysis is not activated and both adipose tissue mass and skeletal muscle mass are preserved<sup>[59]</sup>. TNF- $\alpha$ , named originally cachectin, presents a critical mediatory role in cancer cachexia. IL-6 and leptin may also inhibit synthesis and enhance lipid and protein catabolism in adipocytes and myocytes respectively<sup>[60]</sup>. Nevertheless, hypoleptinemia and hyperadiponectinemia characterize the cancer cachectic state in human studies<sup>[61,62]</sup>. Our group has shown that low leptin and elevated adiponectin levels were seen in pancreatic cancer cases compared to controls<sup>[62]</sup>. Hyperadiponectinemia may be a compensatory response to inflammation, IR and/or the disease-induced weight

loss possibly through altering the size of adipocytes<sup>[62]</sup>. Besides, cachectic patients exhibit frequently a relative glucose intolerance and IR due to alterations in fat metabolism, hypoleptinemia, a pro-inflammatory state and an increased activity of the Cori cycle<sup>[16]</sup>. Muscle wasting in cancer cachexia mediated by free fatty acids, adipokines, cytokines and myokines results in: (1) an activation of the ATP-dependent ubiquitin-proteasome pathway which targets not only structural and sarcomeric proteins such as myosin, troponin and titin but also important myogenic transcription factors such as calcineurin and Myo D<sup>[63,64]</sup>; (2) a defective muscle regeneration capacity due to an abnormal regulation of satellite cells in skeletal muscle<sup>[14]</sup>; and (3) a hyperexpression of myokines that play an important role in muscle atrophy such as myostatin<sup>[65]</sup>. Muscle regeneration may be further compromised in cachexia due to the reprogramming of protein metabolism toward an increased production of acute phase response proteins sustained by the aminoacids secreted by skeletal muscle catabolism. In agreement with this concept, there is also evidence that TNF- $\alpha$  inhibits skeletal muscle regeneration *in vivo* via a caspase-dependent stem cell response<sup>[66]</sup>. Besides its role as a potent cachexia inducer, TNF- $\alpha$  may be a potent inhibitor of *in vivo* myogenesis<sup>[67]</sup>.

Myostatin is a protein belonging to the transforming growth factor- $\beta$  (TGF- $\beta$ ) superfamily, playing a pivotal role in the negative regulation of muscle growth and determining the size and mass of skeletal muscle<sup>[68]</sup>. Myostatin is an authentic myokine as it is exclusively produced by skeletal muscle and to a lesser extent by adipose tissue<sup>[65]</sup>. Deletion of myostatin in mice results in an increased number of satellite cells that are involved in muscle growth<sup>[65]</sup> leading to an enhanced muscle regeneration and skeletal mass hypertrophy and a reduction in total adipose tissue<sup>[46]</sup>. Physical activity attenuates myostatin expression, whereas myostatin deactivation may stimulate the beneficial effects of exercise on metabolism<sup>[46]</sup>. High myostatin gene expression and signaling enhancement have been associated with cancer cachexia<sup>[68]</sup>. In blood, myostatin is inhibited by its propeptide or other binding proteins such as follistatin, a hepatokine which belongs to the TGF- $\beta$  superfamily<sup>[68]</sup>.

Other myokines that could play a role in cachexia are Leukemia inhibitory factor (LIF), IL-7 and IL-8<sup>[46]</sup>. LIF, an IL-6 cytokine superfamily member that affects cell growth by inhibiting differentiation, represents a contraction-induced myokine acting in an autocrine/paracrine manner to promote satellite cell proliferation for muscle regeneration. IL-7 and IL-8 are novel myokines participating in the regulation of skeletal muscle development<sup>[46]</sup>; however, their exact biologic functions remain unknown.

## EMERGING PREVENTIVE AND THERAPEUTIC IMPLICATIONS

High-fat diet, weight gain and physical inactivity may

lead to visceral obesity and muscle loss, and consequently to the enhancement of a network of inflammatory pathways promoting the development of IR, Mets and malignancy growth. Physical activity offers protection against metabolic disorders and obesity-associated malignancies<sup>[45]</sup>.

The capacity of adiponectin to stimulate insulin sensitivity synergistically with its apoptotic properties has rendered this adipokine a promising diagnostic and prognostic biomarker as well as a novel therapeutic tool in the pharmacologic armamentarium for treating cancer<sup>[3]</sup>. However, since adiponectin is extremely difficult to synthesize, research should be conducted in identifying pathways to augment endogenous circulating adiponectin levels in order to attenuate the obesity/physical inactivity-cancer connection<sup>[6]</sup>.

Modulating adipokines and myokines could be a particularly attractive goal for cancer prevention, specifically in overweight/obese and physical inactive individuals. Regular moderate exercise, adoption of a balanced diet, weight reduction and bariatric surgery for morbidly obese persons may increase plasma adiponectin, irisin, IL-15 and the hepatokine follistatin<sup>[65]</sup>, and decrease plasma leptin, resistin, visfatin, chemerin and myostatin concentrations, reducing thus the risk of developing cancer. Very recently, L-4F, an apolipoprotein peptide mimetic used for the pharmacologic upregulation of adiponectin, decreased multiple myeloma (MM) tumor burden through induction of apoptosis, increased survival of myeloma-bearing mice and provided protection against myeloma destructive osteolytic bone disease, an important clinical feature of MM<sup>[69]</sup>. Interestingly, MM as well as monoclonal gammopathy of undetermined significance which may subsequently progress to MM are characterized by hypoadiponectinemia<sup>[69,70]</sup>. ADP 355, a new adiponectin-based short peptide mimicking adiponectin action, decreased proliferation in several adiponectin receptor-positive cancer cell lines, modulated several key adiponectin signaling pathways and suppressed the growth of orthotopic human breast cancer xenografts by 31% *in vivo*<sup>[71]</sup>. Additionally, anti-Nampt (anti-visfatin) agents such as FK866, CHS-828 and APO866 inhibited tumor growth in a broad range of tumor cell lines by diminishing NAD levels, enhanced apoptosis or autophagy, and abrogated tumor growth in animal models of hematological malignancies without significant toxicity<sup>[53,72,73]</sup>.

The pathway of IL-15 and irisin could be explored as a potential therapeutic avenue to combat disease states such as obesity and muscle loss, Mets and obesity-associated malignancies. Increased formation of brown fat instead of white fat has been shown to exhibit beneficial metabolic effects by improving glucose homeostasis and insulin sensitivity in multiple murine models<sup>[53-55]</sup>. Through regular physical activity, irisin and other myokines could ameliorate insulin sensitivity and attenuate the link between IR and cancer<sup>[46,52]</sup>. Exercise-induced myokines have been found to inhibit mammary tumor

cell growth<sup>[12]</sup>. There is accumulating evidence that hyperinsulinemia, the hallmark of IR, and the increase of bioavailable IGF-I may promote cancer. Insulin exerts its oncogenic potential through enhancing growth factor-dependent cell proliferation and through abnormal stimulation of multiple cellular signaling cascades<sup>[74]</sup>. Recent data have consistently underscored the strong link between anti-diabetic treatment, which improves insulin sensitivity and adiponectin production, and decrease in cancer incidence and mortality<sup>[6,75]</sup>.

Finally, the long-term beneficial effects of physical exercise on cancer prevention may be ascribed to the anti-inflammatory actions of myokines and adipokines<sup>[11,52]</sup>. The upregulation of pro-inflammatory cytokines *via* the NF- $\kappa$ B pathway in a chronic low-grade inflammatory disease state such as obesity is a significant component of the cancer-promoting machinery<sup>[6]</sup>.

Regarding cancer cachexia, the metabolic dysfunction precludes the accretion of skeletal muscle mass, even if additional proteins and calories are provided. Furthermore, the use of anti-TNF (and anti-IL-6) antibodies against the main cachectic factor (TNF- $\alpha$ ) in reversing cachexia has led to moderate results<sup>[16]</sup>. Due to the complex pathophysiology of cachexia, combined approaches to deactivate various pathways implicated in cachexia may open up a new era of significant therapeutic progress. In particular, targeting myostatin may represent a novel therapeutic strategy by using potential myostatin inhibitors such as soluble myostatin receptors, follistatin-related proteins, myostatin propeptide, anti-myostatin antibodies and small interfering RNAs<sup>[68]</sup>. Anabolic factors such as insulin-like growth factor I enhancing muscle precursor cell proliferation and regeneration are at the forefront of future therapeutic modalities for cachexia<sup>[16]</sup>.

Nevertheless, more intensive basic research studies, *in vivo* animal studies, observational human studies, and larger prospective and longitudinal studies are needed in order to fully clarify the mechanisms underlying the effects of adipokines and myokines on cancer pathophysiology. Further studies are required for the development of reliable laboratory techniques (*e.g.*, enzyme-linked immunosorbent assays) to assess adipokines and myokines as well as their physiologic relevance. Which levels of adipokines and myokines should be considered abnormal needs also to be determined along with standardization of levels and assay procedures. Proteomics will identify new adipokines, myokines and the extent of the “adipo-myokinome”.

Whilst understanding the interplay of adipokines and myokines with cancer might provide potential therapeutic targets, lifestyle amelioration remains the most important component in preventing obesity-related malignancies. Reduction of body weight, daily physical exercise and a balanced diet with fruit and vegetables consumption may improve energy balance and reduce the risk of developing IR, Mets, t2DM, CVD and obesity-associated malignancies. Advances in the field of translational in-

vestigation may lead to tangible benefits to obese and inactive persons who are at increased risk of cancer as well as to cancer patients with cachexia.

## REFERENCES

- 1 **Juonala M**, Magnussen CG, Berenson GS, Venn A, Burns TL, Sabin MA, Srinivasan SR, Daniels SR, Davis PH, Chen W, Sun C, Cheung M, Viikari JS, Dwyer T, Raitakari OT. Childhood adiposity, adult adiposity, and cardiovascular risk factors. *N Engl J Med* 2011; **365**: 1876-1885 [PMID: 22087679 DOI: 10.1056/NEJMoa1010112]
- 2 **Wilson PW**, D'Agostino RB, Sullivan L, Parise H, Kannel WB. Overweight and obesity as determinants of cardiovascular risk: the Framingham experience. *Arch Intern Med* 2002; **162**: 1867-1872 [PMID: 12196085]
- 3 **Langenberg C**, Sharp SJ, Schulze MB, Rolandsson O, Overvad K, Forouhi NG, Spranger J, Drogan D, Huerta JM, Arriola L, de Lauzon-Guillan B, Tormo MJ, Ardanaz E, Balkau B, Beulens JW, Boeing H, Bueno-de-Mesquita HB, Clavel-Chapelon F, Crowe FL, Franks PW, Gonzalez CA, Griioni S, Halkjaer J, Hallmans G, Kaaks R, Kerrison ND, Key TJ, Khaw KT, Mattiello A, Nilsson P, Norat T, Palla L, Palli D, Panico S, Quirós JR, Romaguera D, Romieu I, Sacerdote C, Sanchez MJ, Slimani N, Sluijs I, Spijkerman AM, Teucher B, Tjønneland A, Tumino R, van der A DL, van der Schouw YT, Feskens EJ, Riboli E, Wareham NJ. Long-term risk of incident type 2 diabetes and measures of overall and regional obesity: the EPIC-InterAct case-cohort study. *PLoS Med* 2012; **9**: e1001230 [PMID: 22679397 DOI: 10.1371/journal.pmed.1001230]
- 4 **Renahan AG**, Tyson M, Egger M, Heller RF, Zwahlen M. Body-mass index and incidence of cancer: a systematic review and meta-analysis of prospective observational studies. *Lancet* 2008; **371**: 569-578 [PMID: 18280327 DOI: 10.1016/S0140-6736(08)60269-X]
- 5 **Doyle SL**, Donohoe CL, Lysaght J, Reynolds JV. Visceral obesity, metabolic syndrome, insulin resistance and cancer. *Proc Nutr Soc* 2012; **71**: 181-189 [PMID: 22051112 DOI: 10.1017/S002966511100320X]
- 6 **Dalamaga M**, Diakopoulos KN, Mantzoros CS. The role of adiponectin in cancer: a review of current evidence. *Endocr Rev* 2012; **33**: 547-594 [PMID: 22547160 DOI: 10.1210/er.2011-1015]
- 7 **Dalamaga M**, Karmaniolas K, Papadavid E, Pelekanos N, Sotiropoulos G, Lekka A. Hyperresistinemia is associated with postmenopausal breast cancer. *Menopause* 2013; **20**: 845-851 [PMID: 23481121]
- 8 **Dalamaga M**. Nicotinamide phosphoribosyl-transferase/visfatin: a missing link between overweight/obesity and postmenopausal breast cancer? Potential preventive and therapeutic perspectives and challenges. *Med Hypotheses* 2012; **79**: 617-621 [PMID: 22922056 DOI: 10.1016/j.mehy.2012.07.036]
- 9 **Wysocki PJ**, Wierusz-Wysocka B. Obesity, hyperinsulinemia and breast cancer: novel targets and a novel role for metformin. *Expert Rev Mol Diagn* 2010; **10**: 509-519 [PMID: 20465505 DOI: 10.1586/erm.10.22]
- 10 **Lorincz AM**, Sukumar S. Molecular links between obesity and breast cancer. *Endocr Relat Cancer* 2006; **13**: 279-292 [PMID: 16728564]
- 11 **McTiernan A**. Mechanisms linking physical activity with cancer. *Nat Rev Cancer* 2008; **8**: 205-211 [PMID: 18235448 DOI: 10.1038/nrc2325]
- 12 **Hojman P**, Dethlefsen C, Brandt C, Hansen J, Pedersen L, Pedersen BK. Exercise-induced muscle-derived cytokines inhibit mammary cancer cell growth. *Am J Physiol Endocrinol Metab* 2011; **301**: E504-E510 [PMID: 21653222 DOI: 10.1152/ajpendo.00520.2010]
- 13 **Quinn LS**, Anderson BG, Strait-Bodey L, Stroud AM, Argilés JM. Oversecretion of interleukin-15 from skeletal muscle reduces adiposity. *Am J Physiol Endocrinol Metab* 2009; **296**: E191-E202 [PMID: 19001550 DOI: 10.1152/ajpendo.90506.2008]
- 14 **Johns N**, Stephens NA, Fearon KC. Muscle wasting in cancer. *Int J Biochem Cell Biol* 2013; **301**: E504-E510 [PMID: 23770121]
- 15 **Fearon KC**. Cancer cachexia and fat-muscle physiology. *N Engl J Med* 2011; **365**: 565-567 [PMID: 21830971 DOI: 10.1056/NEJMcibr1106880]
- 16 **Lelbach A**, Muzes G, Feher J. Current perspectives of catabolic mediators of cancer cachexia. *Med Sci Monit* 2007; **13**: RA168-RA173 [PMID: 17767131]
- 17 **Duan XF**, Tang P, Li Q, Yu ZT. Obesity, adipokines and hepatocellular carcinoma. *Int J Cancer* 2013; **133**: 1776-1783 [PMID: 23404222 DOI: 10.1002/ijc.28105]
- 18 **Pichard C**, Plu-Bureau G, Neves-E Castro M, Gompel A. Insulin resistance, obesity and breast cancer risk. *Maturitas* 2008; **60**: 19-30 [PMID: 18485631 DOI: 10.1016/j.maturitas.2008.03.002]
- 19 **Ziemke F**, Mantzoros CS. Adiponectin in insulin resistance: lessons from translational research. *Am J Clin Nutr* 2010; **91**: 258S-261S [PMID: 19906806 DOI: 10.3945/ajcn.2009.28449C]
- 20 **Mantzoros C**, Petridou E, Dessypris N, Chavelas C, Dalamaga M, Alexe DM, Papadiamantis Y, Markopoulos C, Spanos E, Chrousos G, Trichopoulos D. Adiponectin and breast cancer risk. *J Clin Endocrinol Metab* 2004; **89**: 1102-1107 [PMID: 15001594]
- 21 **Miyoshi Y**, Funahashi T, Kihara S, Taguchi T, Tamaki Y, Matsuzawa Y, Noguchi S. Association of serum adiponectin levels with breast cancer risk. *Clin Cancer Res* 2003; **9**: 5699-5704 [PMID: 14654554]
- 22 **Macis D**, Gandini S, Guerrieri-Gonzaga A, Johansson H, Magni P, Ruscica M, Lazzeroni M, Serrano D, Cazzaniga M, Mora S, Feroce I, Pizzamiglio M, Sandri MT, Gulisano M, Bonanni B, Decensi A. Prognostic effect of circulating adiponectin in a randomized 2 x 2 trial of low-dose tamoxifen and fenretinide in premenopausal women at risk for breast cancer. *J Clin Oncol* 2012; **30**: 151-157 [PMID: 22162577 DOI: 10.1200/JCO.2011.35.2237]
- 23 **Körner A**, Pazaitou-Panayiotou K, Kelesidis T, Kelesidis I, Williams CJ, Kaprara A, Bullen J, Neuwirth A, Tseleni S, Mitsiades N, Kiess W, Mantzoros CS. Total and high-molecular-weight adiponectin in breast cancer: *in vitro* and *in vivo* studies. *J Clin Endocrinol Metab* 2007; **92**: 1041-1048 [PMID: 17192291]
- 24 **Delort L**, Jardé T, Dubois V, Vasson MP, Caldefie-Chézet F. New insights into anticarcinogenic properties of adiponectin: a potential therapeutic approach in breast cancer? *Vitam Horm* 2012; **90**: 397-417 [PMID: 23017724 DOI: 10.1016/B978-0-12-398313-8.00015-4]
- 25 **Grossmann ME**, Cleary MP. The balance between leptin and adiponectin in the control of carcinogenesis - focus on mammary tumorigenesis. *Biochimie* 2012; **94**: 2164-2171 [PMID: 22728769 DOI: 10.1016/j.biochi.2012.06.013]
- 26 **Moon HS**, Dalamaga M, Kim SY, Polyzos SA, Hamnvik OP, Magkos F, Paruthi J, Mantzoros CS. Leptin's role in lipodystrophic and nonlipodystrophic insulin-resistant and diabetic individuals. *Endocr Rev* 2013; **34**: 377-412 [PMID: 23475416 DOI: 10.1210/er.2012-1053]
- 27 **Dalamaga M**, Chou SH, Shields K, Papageorgiou P, Polyzos SA, Mantzoros CS. Leptin at the intersection of neuroendocrinology and metabolism: current evidence and therapeutic perspectives. *Cell Metab* 2013; **18**: 29-42 [PMID: 23770129 DOI: 10.1016/j.cmet.2013.05.010]
- 28 **Catalano S**, Mauro L, Bonofiglio D, Pellegrino M, Qi H, Rizza P, Vizza D, Bossi G, Andò S. *In vivo* and *in vitro* evidence that PPAR $\gamma$  ligands are antagonists of leptin signaling in breast cancer. *Am J Pathol* 2011; **179**: 1030-1040 [PMID: 21704006 DOI: 10.1016/j.ajpath.2011.04.026]
- 29 **Ray A**. Adipokine leptin in obesity-related pathology of breast cancer. *J Biosci* 2012; **37**: 289-294 [PMID: 22581334]
- 30 **Steppan CM**, Bailey ST, Bhat S, Brown EJ, Banerjee RR,

- Wright CM, Patel HR, Ahima RS, Lazar MA. The hormone resistin links obesity to diabetes. *Nature* 2001; **409**: 307-312 [PMID: 11201732]
- 31 **Schwartz DR**, Lazar MA. Human resistin: found in translation from mouse to man. *Trends Endocrinol Metab* 2011; **22**: 259-265 [PMID: 21497511 DOI: 10.1016/j.tem.2011.03.005]
- 32 **Filkovα M**, Haluzik M, Gay S, Senolt L. The role of resistin as a regulator of inflammation: Implications for various human pathologies. *Clin Immunol* 2009; **133**: 157-170 [PMID: 19740705 DOI: 10.1016/j.clim.2009.07.013]
- 33 **Garten A**, Petzold S, Körner A, Imai S, Kiess W. Nampt: linking NAD biology, metabolism and cancer. *Trends Endocrinol Metab* 2009; **20**: 130-138 [PMID: 19109034 DOI: 10.1016/j.tem.2008.10.004]
- 34 **Zhang LQ**, Heruth DP, Ye SQ. Nicotinamide Phosphoribosyltransferase in Human Diseases. *J Bioanal Biomed* 2011; **3**: 13-25 [PMID: 22140607]
- 35 **Fukuhara A**, Matsuda M, Nishizawa M, Segawa K, Tanaka M, Kishimoto K, Matsuki Y, Murakami M, Ichisaka T, Murakami H, Watanabe E, Takagi T, Akiyoshi M, Ohtsubo T, Kihara S, Yamashita S, Makishima M, Funahashi T, Yamanaoka S, Hiramatsu R, Matsuzawa Y, Shimomura I. Visfatin: a protein secreted by visceral fat that mimics the effects of insulin. *Science* 2005; **307**: 426-430 [PMID: 15604363]
- 36 **Dalamaga M**, Sotiropoulos G, Karmaniolas K, Pelekanos N, Papadavid E, Lekka A. Serum resistin: a biomarker of breast cancer in postmenopausal women? Association with clinicopathological characteristics, tumor markers, inflammatory and metabolic parameters. *Clin Biochem* 2013; **46**: 584-590 [PMID: 23321342 DOI: 10.1016/j.clinbiochem.2013.01.001]
- 37 **Lee YC**, Chen YJ, Wu CC, Lo S, Hou MF, Yuan SS. Resistin expression in breast cancer tissue as a marker of prognosis and hormone therapy stratification. *Gynecol Oncol* 2012; **125**: 742-750 [PMID: 22370603 DOI: 10.1016/j.ygyno.2012.02.032]
- 38 **Dalamaga M**, Karmaniolas K, Papadavid E, Pelekanos N, Sotiropoulos G, Lekka A. Elevated serum visfatin/nicotinamide phosphoribosyl-transferase levels are associated with risk of postmenopausal breast cancer independently from adiponectin, leptin, and anthropometric and metabolic parameters. *Menopause* 2011; **18**: 1198-1204 [PMID: 21712732 DOI: 10.1097/gme.0b013e31821e21f5]
- 39 **Dalamaga M**, Archondakis S, Sotiropoulos G, Karmaniolas K, Pelekanos N, Papadavid E, Lekka A. Could serum visfatin be a potential biomarker for postmenopausal breast cancer? *Maturitas* 2012; **71**: 301-308 [PMID: 22261365 DOI: 10.1016/j.maturitas.2011.12.013]
- 40 **Lee YC**, Yang YH, Su JH, Chang HL, Hou MF, Yuan SS. High visfatin expression in breast cancer tissue is associated with poor survival. *Cancer Epidemiol Biomarkers Prev* 2011; **20**: 1892-1901 [PMID: 21784959 DOI: 10.1158/1055-9965.EPI-11-0399]
- 41 **Kim HJ**, Lee YS, Won EH, Chang IH, Kim TH, Park ES, Kim MK, Kim W, Myung SC. Expression of resistin in the prostate and its stimulatory effect on prostate cancer cell proliferation. *BJU Int* 2011; **108**: E77-E83 [PMID: 21050358 DOI: 10.1111/j.1464-410X.2010.09813.x]
- 42 **Patel ST**, Mistry T, Brown JE, Digby JE, Adya R, Desai KM, Randeve HS. A novel role for the adipokine visfatin/pre-B cell colony-enhancing factor 1 in prostate carcinogenesis. *Peptides* 2010; **31**: 51-57 [PMID: 19819277 DOI: 10.1016/j.peptides.2009.10.001]
- 43 **Calabro P**, Samudio I, Willerson JT, Yeh ET. Resistin promotes smooth muscle cell proliferation through activation of extracellular signal-regulated kinase 1/2 and phosphatidylinositol 3-kinase pathways. *Circulation* 2004; **110**: 3335-3340 [PMID: 15545519]
- 44 **Sell H**, Divoux A, Poutou C, Basdevant A, Bouillot JL, Bedossa P, Tordjman J, Eckel J, Clément K. Chemerin correlates with markers for fatty liver in morbidly obese patients and strongly decreases after weight loss induced by bariatric surgery. *J Clin Endocrinol Metab* 2010; **95**: 2892-2896 [PMID: 20375212 DOI: 10.1210/jc.2009-2374]
- 45 **Trayhurn P**, Drevon CA, Eckel J. Secreted proteins from adipose tissue and skeletal muscle - adipokines, myokines and adipose/muscle cross-talk. *Arch Physiol Biochem* 2011; **117**: 47-56 [PMID: 21158485 DOI: 10.3109/13813455.2010.535835]
- 46 **Pedersen BK**, Febbraio MA. Muscles, exercise and obesity: skeletal muscle as a secretory organ. *Nat Rev Endocrinol* 2012; **8**: 457-465 [PMID: 22473333 DOI: 10.1038/nrendo.2012.49]
- 47 **Liu Y**, Chewchuk S, Lavigne C, Brulé S, Pilon G, Houde V, Xu A, Marette A, Sweeney G. Functional significance of skeletal muscle adiponectin production, changes in animal models of obesity and diabetes, and regulation by rosiglitazone treatment. *Am J Physiol Endocrinol Metab* 2009; **297**: E657-E664 [PMID: 19531641 DOI: 10.1152/ajpendo.00186.2009]
- 48 **Holmes MD**, Chen WY, Feskanich D, Kroenke CH, Colditz GA. Physical activity and survival after breast cancer diagnosis. *JAMA* 2005; **293**: 2479-2486 [PMID: 15914748]
- 49 **Meyerhardt JA**, Giovannucci EL, Holmes MD, Chan AT, Chan JA, Colditz GA, Fuchs CS. Physical activity and survival after colorectal cancer diagnosis. *J Clin Oncol* 2006; **24**: 3527-3534 [PMID: 16822844]
- 50 **Irwin ML**, Smith AW, McTiernan A, Ballard-Barbash R, Cronin K, Gilliland FD, Baumgartner RN, Baumgartner KB, Bernstein L. Influence of pre- and postdiagnosis physical activity on mortality in breast cancer survivors: the health, eating, activity, and lifestyle study. *J Clin Oncol* 2008; **26**: 3958-3964 [PMID: 18711185 DOI: 10.1200/JCO.2007.15.9822]
- 51 **Holick CN**, Newcomb PA, Trentham-Dietz A, Titus-Ernstoff L, Bersch AJ, Stampfer MJ, Baron JA, Egan KM, Willett WC. Physical activity and survival after diagnosis of invasive breast cancer. *Cancer Epidemiol Biomarkers Prev* 2008; **17**: 379-386 [PMID: 18250341 DOI: 10.1158/1055-9965.EPI-07-0771]
- 52 **Mathur N**, Pedersen BK. Exercise as a mean to control low-grade systemic inflammation. *Mediators Inflamm* 2008; **2008**: 109502 [PMID: 19148295 DOI: 10.1155/2008/109502]
- 53 **Castillo-Quan JI**. From white to brown fat through the PGC-1α-dependent myokine irisin: implications for diabetes and obesity. *Dis Model Mech* 2012; **5**: 293-295 [PMID: 22566556 DOI: 10.1242/dmm.009894]
- 54 **Enerbäck S**. Adipose tissue metabolism in 2012: Adipose tissue plasticity and new therapeutic targets. *Nat Rev Endocrinol* 2013; **9**: 69-70 [PMID: 23296169 DOI: 10.1038/nrendo.2012.242]
- 55 **Boström P**, Wu J, Jedrychowski MP, Korde A, Ye L, Lo JC, Rasbach KA, Boström EA, Choi JH, Long JZ, Kajimura S, Zingaretti MC, Vind BF, Tu H, Cinti S, Højlund K, Gygi SP, Spiegelman BM. A PGC1-α-dependent myokine that drives brown-fat-like development of white fat and thermogenesis. *Nature* 2012; **481**: 463-468 [PMID: 22237023 DOI: 10.1038/nature10777]
- 56 **Fearon K**, Strasser F, Anker SD, Bosaeus I, Bruera E, Fainsinger RL, Jatoi A, Loprinzi C, MacDonald N, Mantovani G, Davis M, Muscaritoli M, Ottery F, Radbruch L, Ravasco P, Walsh D, Wilcock A, Kaasa S, Baracos VE. Definition and classification of cancer cachexia: an international consensus. *Lancet Oncol* 2011; **12**: 489-495 [PMID: 21296615 DOI: 10.1016/S1470-2045(10)70218-7]
- 57 **Bing C**, Bao Y, Jenkins J, Sanders P, Manieri M, Cinti S, Tisdale MJ, Trayhurn P. Zinc-alpha2-glycoprotein, a lipid mobilizing factor, is expressed in adipocytes and is up-regulated in mice with cancer cachexia. *Proc Natl Acad Sci USA* 2004; **101**: 2500-2505 [PMID: 14983038]
- 58 **Mracek T**, Stephens NA, Gao D, Bao Y, Ross JA, Rydén M, Arner P, Trayhurn P, Fearon KC, Bing C. Enhanced ZAG production by subcutaneous adipose tissue is linked to weight loss in gastrointestinal cancer patients. *Br J Cancer* 2011; **104**: 441-447 [PMID: 21245862 DOI: 10.1038/sj.bjc.6606083]
- 59 **Das SK**, Eder S, Schauer S, Diwoky C, Temmel H, Guertl B, Gorkiewicz G, Tamilarasan KP, Kumari P, Trauner M, Zim-

- mermann R, Vesely P, Haemmerle G, Zechner R, Hoefler G. Adipose triglyceride lipase contributes to cancer-associated cachexia. *Science* 2011; **333**: 233-238 [PMID: 21680814 DOI: 10.1126/science.1198973]
- 60 **Barton BE**. IL-6-like cytokines and cancer cachexia: consequences of chronic inflammation. *Immunol Res* 2001; **23**: 41-58 [PMID: 11417859]
- 61 **Simons JP**, Schols AM, Campfield LA, Wouters EF, Saris WH. Plasma concentration of total leptin and human lung-cancer-associated cachexia. *Clin Sci (Lond)* 1997; **93**: 273-277 [PMID: 9337643]
- 62 **Dalamaga M**, Migdalis I, Fargnoli JL, Papadavid E, Bloom E, Mitsiades N, Karmaniolas K, Pelecanos N, Tseleni-Balafouta S, Dionyssiou-Asteriou A, Mantzoros CS. Pancreatic cancer expresses adiponectin receptors and is associated with hypoleptinemia and hyperadiponectinemia: a case-control study. *Cancer Causes Control* 2009; **20**: 625-633 [PMID: 19051043 DOI: 10.1007/s10552-008-9273-z]
- 63 **Witt SH**, Granzier H, Witt CC, Labeit S. MURF-1 and MURF-2 target a specific subset of myofibrillar proteins redundantly: towards understanding MURF-dependent muscle ubiquitination. *J Mol Biol* 2005; **350**: 713-722 [PMID: 15967462]
- 64 **Tintignac LA**, Lagirand J, Battonnet S, Sirri V, Leibovitch MP, Leibovitch SA. Degradation of MyoD mediated by the SCF (MAFbx) ubiquitin ligase. *J Biol Chem* 2005; **280**: 2847-2856 [PMID: 15531760]
- 65 **Tsuchida K**. Targeting myostatin for therapies against muscle-wasting disorders. *Curr Opin Drug Discov Devel* 2008; **11**: 487-494 [PMID: 18600566]
- 66 **Moresi V**, Pristerà A, Scicchitano BM, Molinaro M, Teodori L, Sassoon D, Adamo S, Coletti D. Tumor necrosis factor-alpha inhibition of skeletal muscle regeneration is mediated by a caspase-dependent stem cell response. *Stem Cells* 2008; **26**: 997-1008 [PMID: 18258721 DOI: 10.1634/stemcells.2007-0493]
- 67 **Coletti D**, Moresi V, Adamo S, Molinaro M, Sassoon D. Tumor necrosis factor-alpha gene transfer induces cachexia and inhibits muscle regeneration. *Genesis* 2005; **43**: 120-128 [PMID: 16158413]
- 68 **Costelli P**, Muscaritoli M, Bonetto A, Penna F, Reffo P, Bosola M, Bonelli G, Doglietto GB, Baccino FM, Rossi Fanelli F. Muscle myostatin signalling is enhanced in experimental cancer cachexia. *Eur J Clin Invest* 2008; **38**: 531-538 [PMID: 18578694 DOI: 10.1111/j.1365-2362.2008.01970.x]
- 69 **Fowler JA**, Lwin ST, Drake MT, Edwards JR, Kyle RA, Mundy GR, Edwards CM. Host-derived adiponectin is tumor-suppressive and a novel therapeutic target for multiple myeloma and the associated bone disease. *Blood* 2011; **118**: 5872-5882 [PMID: 21908434 DOI: 10.1182/blood-2011-01-330407]
- 70 **Dalamaga M**, Karmaniolas K, Panagiotou A, Hsi A, Chamberland J, Dimas C, Lekka A, Mantzoros CS. Low circulating adiponectin and resistin, but not leptin, levels are associated with multiple myeloma risk: a case-control study. *Cancer Causes Control* 2009; **20**: 193-199 [PMID: 18814045 DOI: 10.1007/s10552-008-9233-7]
- 71 **Otvos L**, Haspinger E, La Russa F, Maspero F, Graziano P, Kovalszky I, Lovas S, Nama K, Hoffmann R, Knappe D, Cassone M, Wade J, Surmacz E. Design and development of a peptide-based adiponectin receptor agonist for cancer treatment. *BMC Biotechnol* 2011; **11**: 90 [PMID: 21974986 DOI: 10.1186/1472-6750-11-90]
- 72 **Olesen UH**, Christensen MK, Björkling F, Jäätelä M, Jensen PB, Sehested M, Nielsen SJ. Anticancer agent CHS-828 inhibits cellular synthesis of NAD. *Biochem Biophys Res Commun* 2008; **367**: 799-804 [PMID: 18201551 DOI: 10.1016/j.bbrc.2008.01.019]
- 73 **Cea M**, Zoppoli G, Bruzzone S, Fruscione F, Moran E, Garuti A, Rocco I, Cirmena G, Casciaro S, Olcese F, Pierri I, Cagnetta A, Ferrando F, Ghio R, Gobbi M, Ballestrero A, Patrone F, Nencioni A. APO866 activity in hematologic malignancies: a preclinical *in vitro* study. *Blood* 2009; **113**: 6035-6037; author reply 6035-6037; [PMID: 19498032 DOI: 10.1182/blood-2009-03-209213]
- 74 **Djiogue S**, Nwabo Kamdje AH, Vecchio L, Kipanyula MJ, Farahna M, Aldebasi Y, Seke Etet PF. Insulin resistance and cancer: the role of insulin and IGFs. *Endocr Relat Cancer* 2013; **20**: R1-R17 [PMID: 23207292 DOI: 10.1530/ERC-12-0324]
- 75 **Pierotti MA**, Berrino F, Gariboldi M, Melani C, Mogavero A, Negri T, Pasanisi P, Pilotti S. Targeting metabolism for cancer treatment and prevention: metformin, an old drug with multi-faceted effects. *Oncogene* 2013; **32**: 1475-1487 [PMID: 22665053 DOI: 10.1038/onc.2012.181]

P- Reviewers Bing C, Coletti D, Vlachostergios PJ  
S- Editor Wen LL L- Editor A E- Editor Liu XM





百世登

**Baishideng**®

Published by **Baishideng Publishing Group Co., Limited**

Flat C, 23/F., Lucky Plaza, 315-321 Lockhart Road,

Wan Chai, Hong Kong, China

Fax: +852-65557188

Telephone: +852-31779906

E-mail: [bpgoffice@wjgnet.com](mailto:bpgoffice@wjgnet.com)

<http://www.wjgnet.com>

