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Nicotinic acid: Do we know how it works after 55 years of clinical experience?

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Author contributions: Kei A and Elisaf MS contributed to the conception, design, acquisition and interpretation of data, revised the paper critically and gave their final approval of the version to be published.

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Received: August 26, 2011 Revised: December 27, 2011

Accepted: April 10, 2012

Published online: June 9, 2012

Abstract

Nicotinic acid (NA) comprises the oldest hypolipidemic drug, in use since 1955. Despite its established broad spectrum effect on lipid profile and the associated reduction in cardiovascular morbidity and mortality, the mechanisms by which NA achieves its beneficial effects remain elusive. Regarding the NA-associated reduction in triglyceride and low density lipoprotein cholesterol levels, data are controversial. The prevailing view which suggested that NA inhibits lipolysis and decreases free fatty acid (FFA) release both *via* activation of adipose tissue G-protein receptor-109A (GPR109A) and *via* inhibition of hepatic triglyceride synthesis is currently debated by the observation that the initially decreased FFA levels rebound during long-term NA treatment even though the beneficial NA effects on lipid metabolism are preserved, while other mechanisms involving modulation of transcription and translation pathways are emerging. In addition, NA has been demonstrated to affect high density lipoprotein (HDL) particles remodeling in a number of ways, including reducing cholesterol ester transfer protein levels and activity, increasing apolipoprotein A-I levels, eliminating HDL hepatic uptake, increasing cholesterol efflux *via* ATP-binding

cassette A1, inhibiting hepatic lipase, thereby overall increasing the plasma residence time of HDL and apoA-I with retention of cholesterol esters in HDL. Focus of this article is to present the mechanisms by which NA exerts its broad spectrum hypolipidemic actions.

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Key words: Dyslipidemia; Nicotinic acid; Niacin; Mechanisms; GPR109A; Free fatty acids

Peer reviewer: Josh Burk, Associate Professor, Department of Psychology, College of William and Mary, 540 Landrum Drive, Williamsburg, VA 23187, United States

Kei A, Elisaf MS. Nicotinic acid: Do we know how it works after 55 years of clinical experience? *World J Pharmacol* 2012; 1(3): 50-54 Available from: URL: <http://www.wjgnet.com/2220-3192/full/v1/i3/50.htm> DOI: <http://dx.doi.org/10.5497/wjp.v1.i3.50>

INTRODUCTION

Nicotinic acid (NA), also known as the water-soluble vitamin B3 (niacin), comprises the oldest hypolipidemic agent, being in use since 1955^[1]. It decreases by 5%-25% and 20%-50% the levels of low density lipoprotein (LDL) cholesterol and triglycerides (TG), respectively, while NA remains the most effective currently available agent for raising high density lipoprotein (HDL) cholesterol levels (by 20%-25%) and decreasing the levels of lipoprotein (a) [Lp(a)] (by 28%-40%)^[2,3]. Moreover, a number of clinical trials have demonstrated that NA may decrease cardiovascular events^[4] and total mortality^[5] in patients with coronary heart disease. Overall, NA comprises a hypolipidemic agent with unique broad-spectrum lipid-modifying properties and possible clinical benefits^[6,7]. However, the mechanisms by which NA exerts its lipid-modifying

effects remain elusive. Focus of this paper is the presentation of these mechanisms. We searched PubMed up to 10 August 2011 using combinations of the following keywords: niacin, NA, mechanism, action, GPR109A, free fatty acids (FFA), dyslipidemia, hypolipidemic, liver, adipose tissue, macrophages and receptor. The references of these articles were scrutinized for relevant articles.

TG AND LDL CHOLESTEROL LEVEL REDUCTION

The mechanisms which contribute to the NA-associated TG and LDL cholesterol level reduction are not fully understood. However, the prevailing view holds that NA achieves TG and LDL cholesterol level reduction, primarily by affecting both adipose tissue's and liver's TG metabolism. (Figure 1).

NA's effect on adipose tissue

G-protein receptor-109A (GPR109A), also known as HM74A receptor in humans^[8] is expressed in adipose tissue, spleen and immune cells^[9-11]. NA binds and activates GPR109A in adipose tissue inducing a Gi-mediated inhibition of adenylyl cyclase activity, thereby resulting in a decrease of cyclic adenosine monophosphate (cAMP) intracellular levels^[12]. This leads to decreased lipolysis, as cAMP is the main intracellular mediator of prolipolytic stimuli. cAMP normally activates protein kinase A (PKA) to phosphorylate various proteins, including perilipin and hormone-sensitive lipase (HSL), thereby promoting lipolysis^[12]. The decrease in circulating FFA results in a substrate shortage for hepatic very low density lipoprotein (VLDL) production, consequently reducing plasma levels of LDL cholesterol and TG^[13,14]. Paradoxically, it has been established that the initially decreased FFA levels rebound during long-term NA treatment even though the beneficial NA effects on lipid metabolism are preserved^[15,16]. But how that happens? Phosphoenolpyruvate carboxykinase (PEPCK1) is a key enzyme in adipose tissue gluconeogenesis and its deficit leads to increased FFA release. Recently, NA was associated with decreased expression of PEPCK1 in adipose tissue and thus increased FFA release, partly explaining the rebound phenomenon^[17]. Another contributing mechanism to FFA rebound could also be the NA-induced up-regulation of tumor necrosis factor- α (TNF- α) transcription and the consequent increase of interleukin-6 (IL-6), as both of them comprise cytokines with lipolytic properties, thereby increasing FFA release^[17]. Of note, MK-0354, a partial agonist of GPR109A which resulted in decreased plasma FFA concentrations, paradoxically failed to affect LDL cholesterol and TG levels^[18]. Overall, it seems that the contribution of GPR109A activation to the long-term hypolipidemic effects of NA remains debatable.

NA's effect on liver

NA has been demonstrated to inhibit diacylglycerol acyl transferase 2 (DGAT2) which comprises a key enzyme

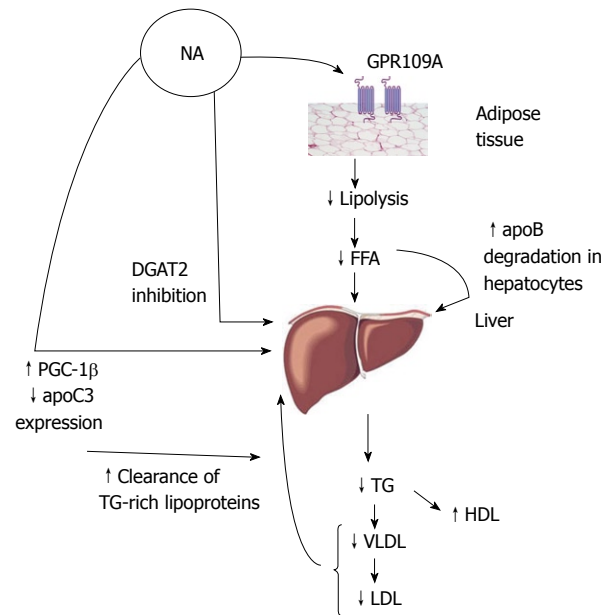


Figure 1 How nicotinic acid decreases triglycerides and low density lipoprotein cholesterol levels. NA: Nicotinic acid; TG: Triglycerides; LDL: Low density lipoprotein; FFA: Free fatty acids; VLDL: Very low density lipoprotein; DGAT2: Diacylglycerol acyl transferase 2; PGC-1 β : Peroxisome proliferator-activated receptor γ coactivator-1 β ; apoC3: Apolipoprotein C3; apoB: Apolipoprotein B.

for the hepatic TG synthesis^[19]. However, DGAT2 inhibition has been observed at NA concentrations 100-fold higher than those associated with maximal pharmacological effects of NA on FFA and TG levels^[20].

Current evidence indicates that the post-translational apolipoprotein B (apoB) degradative processes regulate the hepatic assembly and secretion of VLDL and the subsequent generation of LDL particles. The availability of TG for the addition to apoB during intracellular processing appears to play a central role in targeting apoB for either intracellular degradation or assembly and secretion as VLDL particles. NA-induced TG synthesis inhibition has been demonstrated to create a favorable environment for protease-mediated intracellular apoB degradation in hepatocytes, thereby resulting in decreased apoB-containing VLDL and thus LDL particle formation^[21,22]. In addition, stable isotope methodologies in dyslipidemic patients demonstrated NA-enhanced plasma clearance of TG-rich lipoproteins containing either apoB100 or apoB48, thereby implying that NA may affect both hepatic and intestinal TG-rich lipoproteins' metabolism^[22]. Of note, NA failed to interact with hepatic LDL-receptors^[23].

Further actions on TGs have been associated with the NA-induced inhibition of peroxisome proliferator-activated receptor γ (PPAR γ) coactivator-1 β (PGC-1 β). PGC-1 β is a transcriptional co-activator that is regulated by FFA. Specifically, PGC-1 β induces hypertriglyceridemia in response to dietary fats through activation of hepatic lipogenesis and lipoprotein secretion. Moreover, PGC-1 β regulates plasma TG levels by stimulating apolipoprotein C3 (apoC3) expression, thereby inhibiting

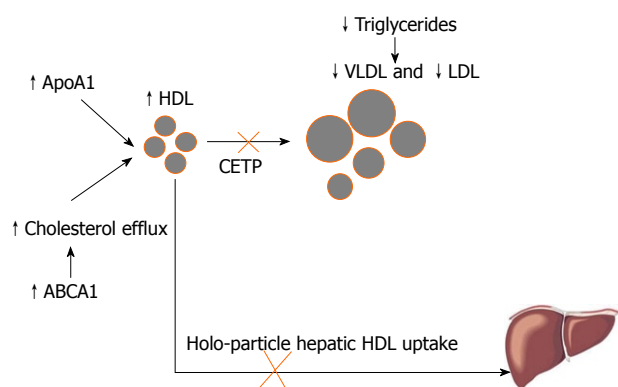


Figure 2 How nicotinic acid can increase high density lipoprotein cholesterol level. NA: Nicotinic acid; HDL: High density lipoprotein; VLDL: Very low density lipoprotein; LDL: Low density lipoprotein; CETP: Cholesterol ester transfer protein; ApoA1: Apolipoprotein A1; ABCA1: ATP-binding cassette protein A1.

apolipoprotein E (apoE) driven clearance of TG-rich lipoproteins^[24]. Of note, both acute and chronic treatment with NA were associated with reduced hepatic expression of PGC-1 β and apoC3, while knockdown of PGC-1 β or APOC3 in mice liver recapitulated NA's hypolipidemic effect^[24].

HDL CHOLESTEROL LEVEL ELEVATION

The mechanisms by which NA elevates HDL cholesterol level have not been fully elucidated. However, a number of mechanisms have been shown to contribute to the observed NA-induced HDL cholesterol level elevation. (Figure 2).

NA's effect on cholesterol ester transfer protein

The NA-induced decrease in TG levels in apoB-containing lipoproteins (LDL and VLDL) eliminates the exchange of TG for cholesteryl-esters from HDL particles mediated by cholesterol ester transfer protein (CETP), resulting in increased HDL concentration^[25,26]. In fact, NA-associated HDL cholesterol elevation depended on the presence of CETP in mice^[27]. However, the partial GPR109A agonist MK-0354 failed to raise HDL levels despite the reduction in plasma FFA^[18]. NA has been also associated with reduced CETP activity per se as a result of reduced hepatic CETP gene expression and reduced release of CETP in plasma^[28]. Of note, the reduction in CETP activity can also explain how NA promotes the maturation of HDL into large particles^[22].

NA's effect on the holo-particle uptake pathway

NA has been associated with reduced hepatic uptake of HDL, potentially by the holo-particle uptake pathway. In fact, NA has been shown to inhibit the surface-expressed ATP-synthase β -chain which acts as a HDL holoparticle receptor leading to slower HDL catabolism^[29].

NA's effect on apolipoprotein A-I metabolism

Data regarding the effect of NA on apolipoprotein A-I

(apoA-I) metabolism are controversial. NA has been shown to increase production rate of apoA-I both in liver and intestinal cells^[22,30]. In fact, NA activates both mitogen activated protein (MAP) kinase and the PPAR transcription factors pathways, which both affect hepatic apoA-I production^[31-33]. On the contrary, other studies with hepatic cells and mice reported no effect of NA on apoA-I production rate, while NA administration was associated with decreased apoA-I hepatic removal^[28,34].

NA's effect on ATP-binding cassette protein A1

NA, potentially *via* GPR109A activation, enhances transcription of cholesterol efflux transporters ATP-binding cassette protein A1 and G1 (ABCA1 and ABCG1, respectively). Thus, NA-induced cholesterol efflux from macrophages could also contribute to the reported increase in HDL cholesterol levels^[23,35]. Moreover, NA dose-dependently stimulated PPAR γ and ABCA1 expression and promoted ApoA-I-induced cholesterol efflux in adipocytes. In fact, treatment of PPAR γ -selective antagonist GW9662 significantly abolished the NA-induced increase in ABCA1 mRNA expression and cholesterol efflux to ApoA-I^[36]. Of note, NA had no effect in HDL cholesterol levels in GPR109A knock-out mice^[36]. On the other hand, overexpression of GPR109A reduced hepatocyte ABCA1 expression and activity, thereby decreasing cholesterol efflux to nascent apoA-I and reducing HDL cholesterol levels in mice^[37]. Overall, it seems that NA effect on ABCA1 is mediated *via* GPR109A and we can speculate that a phenomenon of tachyphylaxis may occur in case of GPR109A overstimulation.

NA's effect on hepatic lipase

In mice NA has been shown to inhibit hepatic lipase activity. This results in decreased remodeling of plasma HDL, thereby limiting HDL clearance^[28].

Lp(a) REDUCTION

No particular mechanisms regarding the NA-induced reduction of Lp(a) have been reported. However, the NA-associated reduction in the circulating FFA by both GPR109A-mediated lipolysis and DGAT2 inhibition results in reduced VLDL and subsequently LDL levels^[13,19]. As LDL particles comprise the substrate for Lp(a) it comes as no surprise that NA also reduces Lp(a) levels.

CONCLUSION

Overall, NA exerts broad spectrum effects on lipids through a number of elusive and even controversial mechanisms. Regarding the NA-associated reduction in TG and LDL cholesterol levels, the prevailing view which suggested that NA inhibits lipolysis *via* GPR109A activation is currently debated by both the rebound phenomenon and the failure of partial GPR109A agonist, MK-0354 to reduce TG and LDL cholesterol levels despite the decrease in plasma FFA. On the other hand,

the NA-associated DGAT2 inhibition was reported at NA levels much higher than those used in clinical setting. However, it can be argued that plasma levels of NA may not reflect its bioavailability at the liver. In addition, PGC-1 β mediated increased clearance of TG-rich lipoproteins may also contribute to hypolipidemic effects of NA.

Similarly, data regarding the mechanisms by which NA increases HDL cholesterol level are scant. However, it seems that NA affects HDL particles remodeling in a number of ways, including reducing CETP levels and activity, increasing apoA-I levels, eliminating HDL hepatic uptake, increasing cholesterol efflux *via* ABCA1, inhibiting hepatic lipase, thereby overall increasing the plasma residence time of HDL and apoA-I with retention of cholesterol esters in HDL.

Conclusively, the mechanisms by which the oldest hypolipidemic drug exerts its lipid-modifying effects remain elusive even after 55 years of clinical experience. However, it is undebatable that NA targets a number of different receptors expressed in a variety of cells including hepatic, intestinal and adipose tissue cells in order to achieve its broad spectrum effect on lipid profile. More research effort especially with genetically modified animals which do not express or overexpress a number of receptors or transporters including GPR109A, ABCA1, PGC-1 β is needed in order to decode how NA really works.

REFERENCES

- 1 Altschul R, Hoffer A, Stephen JD. Influence of nicotinic acid on serum cholesterol in man. *Arch Biochem Biophys* 1955; **54**: 558-559
- 2 Capuzzi DM, Guyton JR, Morgan JM, Goldberg AC, Kreisberg RA, Brusco OA, Brody J. Efficacy and safety of an extended-release niacin (Niaspan): a long-term study. *Am J Cardiol* 1998; **82**: 74U-81U; discussion 85U-86U
- 3 Vogt A, Kassner U, Hostalek U, Steinhagen-Thiessen E. Evaluation of the safety and tolerability of prolonged-release nicotinic acid in a usual care setting: the NAUTILUS study. *Curr Med Res Opin* 2006; **22**: 417-425
- 4 Brown BG, Zhao XQ, Chait A, Fisher LD, Cheung MC, Morse JS, Dowdy AA, Marino EK, Bolson EL, Alaupovic P, Frohlich J, Albers JJ. Simvastatin and niacin, antioxidant vitamins, or the combination for the prevention of coronary disease. *N Engl J Med* 2001; **345**: 1583-1592
- 5 Canner PL, Berge KG, Wenger NK, Stamler J, Friedman L, Prineas RJ, Friedewald W. Fifteen year mortality in Coronary Drug Project patients: long-term benefit with niacin. *J Am Coll Cardiol* 1986; **8**: 1245-1255
- 6 Florentin M, Liberopoulos EN, Kei A, Mikhailidis DP, Elisaf MS. Pleiotropic effects of nicotinic acid: beyond high density lipoprotein cholesterol elevation. *Curr Vasc Pharmacol* 2011; **9**: 385-400
- 7 Kei A, Elisaf M, Moutzouri E, Tsiara S, Liberopoulos E. Add-on-Statins Extended Release Nicotinic Acid/Laropiprant but Not the Switch to High-Dose Rosuvastatin Lowers Blood Pressure: An Open-Label Randomized Study. *Int J Hypertens* 2011; **2011**: 830434
- 8 Benyó Z, Gille A, Kero J, Csiky M, Suchánková MC, Nüsing RM, Moers A, Pfeffer K, Offermanns S. GPR109A (PUMA-G/HM74A) mediates nicotinic acid-induced flushing. *J Clin Invest* 2005; **115**: 3634-3640
- 9 Soga T, Kamohara M, Takasaki J, Matsumoto S, Saito T, Ohishi T, Hiyama H, Matsuo A, Matsushima H, Furuichi K. Molecular identification of nicotinic acid receptor. *Biochem Biophys Res Commun* 2003; **303**: 364-369
- 10 Lorenzen A, Stanek C, Lang H, Andrianov V, Kalvinsh I, Schwabe U. Characterization of a G protein-coupled receptor for nicotinic acid. *Mol Pharmacol* 2001; **59**: 349-357
- 11 Maciejewski-Lenoir D, Richman JG, Hakak Y, Gaidarov I, Behan DP, Connolly DT. Langerhans cells release prostaglandin D2 in response to nicotinic acid. *J Invest Dermatol* 2006; **126**: 2637-2646
- 12 Zechner R, Strauss JG, Haemmerle G, Lass A, Zimmermann R. Lipolysis: pathway under construction. *Curr Opin Lipidol* 2005; **16**: 333-340
- 13 Carlson LA, Oro L. The effect of nicotinic acid on the plasma free fatty acid; demonstration of a metabolic type of sympathicolysis. *Acta Med Scand* 1962; **172**: 641-645
- 14 Kamanna VS, Ganji SH, Kashyap ML. The mechanism and mitigation of niacin-induced flushing. *Int J Clin Pract* 2009; **63**: 1369-1377
- 15 Poynten AM, Gan SK, Kriketos AD, O'Sullivan A, Kelly JJ, Ellis BA, Chisholm DJ, Campbell LV. Nicotinic acid-induced insulin resistance is related to increased circulating fatty acids and fat oxidation but not muscle lipid content. *Metabolism* 2003; **52**: 699-704
- 16 Vega GL, Cater NB, Meguro S, Grundy SM. Influence of extended-release nicotinic acid on nonesterified fatty acid flux in the metabolic syndrome with atherogenic dyslipidemia. *Am J Cardiol* 2005; **95**: 1309-1313
- 17 Choi S, Yoon H, Oh KS, Oh YT, Kim YI, Kang I, Youn JH. Widespread effects of nicotinic acid on gene expression in insulin-sensitive tissues: implications for unwanted effects of nicotinic acid treatment. *Metabolism* 2011; **60**: 134-144
- 18 Lai E, Waters MG, Tata JR, Radziszewski W, Perevozskaya I, Zheng W, Wenning L, Connolly DT, Semple G, Johnson-Levonas AO, Wagner JA, Mitchel Y, Paolini JF. Effects of a niacin receptor partial agonist, MK-0354, on plasma free fatty acids, lipids, and cutaneous flushing in humans. *J Clin Lipidol* 2008; **2**: 375-383
- 19 Ganji SH, Tavintharan S, Zhu D, Xing Y, Kamanna VS, Kashyap ML. Niacin noncompetitively inhibits DGAT2 but not DGAT1 activity in HepG2 cells. *J Lipid Res* 2004; **45**: 1835-1845
- 20 Svedmyr N, Harthorn L, Lundholm L. The relationship between the plasma concentration of free nicotinic acid and some of its pharmacologic effects in man. *Clin Pharmacol Ther* 1969; **10**: 559-570
- 21 Jin FY, Kamanna VS, Kashyap ML. Niacin accelerates intracellular ApoB degradation by inhibiting triacylglycerol synthesis in human hepatoblastoma (HepG2) cells. *Arterioscler Thromb Vasc Biol* 1999; **19**: 1051-1059
- 22 Lamon-Fava S, Diffenderfer MR, Barrett PH, Buchsbaum A, Nyaku M, Horvath KV, Asztalos BF, Otokozaawa S, Ai M, Matthan NR, Lichtenstein AH, Dolnikowski GG, Schaefer EJ. Extended-release niacin alters the metabolism of plasma apolipoprotein (Apo) A-I and ApoB-containing lipoproteins. *Arterioscler Thromb Vasc Biol* 2008; **28**: 1672-1678
- 23 Rubic T, Trottmann M, Lorenz RL. Stimulation of CD36 and the key effector of reverse cholesterol transport ATP-binding cassette A1 in monocytoic cells by niacin. *Biochem Pharmacol* 2004; **67**: 411-419
- 24 Hernandez C, Molusky M, Li Y, Li S, Lin JD. Regulation of hepatic ApoC3 expression by PGC-1 β mediates hypolipidemic effect of nicotinic acid. *Cell Metab* 2010; **12**: 411-419
- 25 Kontush A, Chapman MJ. Functionally defective high-density lipoprotein: a new therapeutic target at the crossroads of dyslipidemia, inflammation, and atherosclerosis. *Pharmacol Rev* 2006; **58**: 342-374
- 26 Joy T, Hegele RA. Is raising HDL a futile strategy for atheroprotection? *Nat Rev Drug Discov* 2008; **7**: 143-155
- 27 Hernandez M, Wright SD, Cai TQ. Critical role of chole-

- terol ester transfer protein in nicotinic acid-mediated HDL elevation in mice. *Biochem Biophys Res Commun* 2007; **355**: 1075-1080
- 28 **van der Hoorn JW**, de Haan W, Berbée JF, Havekes LM, Jukema JW, Rensen PC, Princen HM. Niacin increases HDL by reducing hepatic expression and plasma levels of cholesteryl ester transfer protein in APOE*3Leiden.CETP mice. *Arterioscler Thromb Vasc Biol* 2008; **28**: 2016-2022
- 29 **Zhang LH**, Kamanna VS, Zhang MC, Kashyap ML. Niacin inhibits surface expression of ATP synthase beta chain in HepG2 cells: implications for raising HDL. *J Lipid Res* 2008; **49**: 1195-1201
- 30 **Haas MJ**, Alamir AR, Sultan S, Chehade JM, Wong NC, Mooradian AD. Nicotinic acid induces apolipoprotein A-I gene expression in HepG2 and Caco-2 cell lines. *Metabolism* 2011; **60**: 1790-1796
- 31 **Lamon-Fava S**, Micherone D. Regulation of apoA-I gene expression: mechanism of action of estrogen and genistein. *J Lipid Res* 2004; **45**: 106-112
- 32 **Pandey NR**, Renwick J, Misquith A, Sokoll K, Sparks DL. Linoleic acid-enriched phospholipids act through peroxisome proliferator-activated receptors alpha to stimulate hepatic apolipoprotein A-I secretion. *Biochemistry* 2008; **47**: 1579-1587
- 33 **Watt MJ**, Southgate RJ, Holmes AG, Febbraio MA. Suppression of plasma free fatty acids upregulates peroxisome proliferator-activated receptor (PPAR) alpha and delta and PPAR coactivator 1alpha in human skeletal muscle, but not lipid regulatory genes. *J Mol Endocrinol* 2004; **33**: 533-544
- 34 **Jin FY**, Kamanna VS, Kashyap ML. Niacin decreases removal of high-density lipoprotein apolipoprotein A-I but not cholesterol ester by Hep G2 cells. Implication for reverse cholesterol transport. *Arterioscler Thromb Vasc Biol* 1997; **17**: 2020-2028
- 35 **Lukasova M**, Malaval C, Gille A, Kero J, Offermanns S. Nicotinic acid inhibits progression of atherosclerosis in mice through its receptor GPR109A expressed by immune cells. *J Clin Invest* 2011; **121**: 1163-1173
- 36 **Wu ZH**, Zhao SP. Niacin promotes cholesterol efflux through stimulation of the PPARgamma-LXRalpha-ABCA1 pathway in 3T3-L1 adipocytes. *Pharmacology* 2009; **84**: 282-287
- 37 **Li X**, Millar JS, Brownell N, Briand F, Rader DJ. Modulation of HDL metabolism by the niacin receptor GPR109A in mouse hepatocytes. *Biochem Pharmacol* 2010; **80**: 1450-1457

S- Editor Yan JL L- Editor A E- Editor Zheng XM