

66006_Auto_Edited-check.docx

Name of Journal: *World Journal of Clinical Cases*

Manuscript NO: 66006

Manuscript Type: REVIEW

Encouraging specific biomarkers-based therapeutic strategies for hepatocellular carcinoma

Yao M *et al.* Biomarkers in HCC therapy

Min Yao, Jun-Ling Yang, De-Feng Wang, Li Wang, Ying Chen, Deng-Fu Yao

Abstract

The prevention, early diagnosis, effectively treatment, and poor prognosis of patients with hepatocellular carcinoma (HCC) remain a global medical challenge. Surgery operation is still mainly treated for HCC with auxiliary vascular embolization, radio frequency, radiotherapy, chemotherapy, and biological therapy. Applications of multikinase inhibitor sorafenib, chimeric antigen receptor T cells, or PD-1/PD-L1 inhibitors can prolong the median survival of HCC patients. However, the treatment efficacy is still unsatisfactory because of HCC metastasis and postoperative recurrence. Liver tissues in hepatocarcinogenesis or hepatocyte malignant transformation can express and secrete a variety of molecules with specific biomarkers or oncogenic antigens into blood, for example, alpha-fetoprotein, glypican-3, Wnt3a (one of the key signaling molecules in the Wnt/ β -catenin pathway), insulin-like growth factor (IGF)-II or IGF-I receptor, vascular endothelial growth factor, secretory clusterin, and so on. In addition, combinatorial immunotherapy with non-coding RNAs might improve anticancer efficacy. These biomarkers not only contribute to the diagnosis and prognosis of HCC, they but also might become the molecular-targets for HCC treatment under developing or clinical trials. This article reviews the recent progress of some emerging biomarkers in basic researches or clinical trials for HCC immunotherapy.

Key Words: Hepatocellular carcinoma; Immunotherapy; Carcinoembryonic proteins; Specific biomarkers; Wnt/ β -catenin pathway; Signal molecules

Yao M, Yang JL, Wang DF, Wang L, Chen Y, Yao DF. Encouraging specific biomarkers-based therapeutic strategies for hepatocellular carcinoma. *World J Clin Cases* 2021; In press

Core Tip: Tissues in hepatocellular carcinoma (HCC) or hepatocyte malignant transformation can express and secrete a variety of molecules with specific biomarkers or oncogenic antigens into blood. These biomarkers not only contribute to the diagnosis and prognosis of HCC, they but also might become the molecular-targets for HCC treatment under developing or clinical trials. This article reviews the recent progress of some emerging biomarkers in basic researches or clinical trials for HCC immunotherapy.

INTRODUCTION

The prevention, early monitoring or diagnosis and accurate or effectively treatment of hepatocellular carcinoma (HCC) are still urgent medical problems^[1,2]. Occurrence of HCC is mainly associated with chronic persistent infection of hepatitis B virus (HBV) or hepatitis C virus (HCV), chemical carcinogens intake, and nonalcoholic fatty liver disease (NAFLD)^[3]. In the past decade, NAFLD has become a leading cause of chronic hepatitis and liver cirrhosis, as well as an important risk factor for HCC^[4]. **Innate and adaptive immunity play a pivotal role in determining tumor control vs progression. Genomic instability and abnormal signaling in the setting of chronic liver inflammation** that promotes fibrogenesis and angiogenesis lead to tumorigenesis, along with how they may be exploited in the development of novel therapeutics^[5]. The activation of oncogenes or HCC-related genes, inactivation of anti-oncogenes or reactivations of some oncogenes during the embryonic period can induce malignant transformation of

hepatocytes^[5], many kinds of specific markers can be expressed, and then secreted into blood in the process of initiation, promotion and evolution^[1]. Notably, HCC oncoimmunology ⁵ depends on diverse genetic and environmental factors that together shape cancer-promoting inflammation and immune dysfunction- critical processes that control HCC malignant progression and response to therapy^[6,7].

Nowdays, surgery operation is still mainly treated for HCC with auxiliary vascular embolization, radio frequency, radiotherapy, chemotherapy, and biological therapy^[8,9]. Application of multikinase inhibitor sorafenib can prolong the median survival of HCC patients. However, the treatment efficacy is still unsatisfactory because of HCC metastasis and postoperative occurrence^[10,11]. Undoubtedly, the ¹³ integration of data obtained from both preclinical models and human studies can help to accelerate the identification of robust predictive biomarkers of response to targeted or immune-therapy^[12,13]. HCC tissues express the specific antigens such as the key molecules of HCC-related signal pathways, growth factors and receptors, vascular endothelial growth factor (VEGF), and the products of oncogenes that some mediated tumor progression and could be potential molecular-targeted for anti-cancer therapy with highly specificity and application prospects^[14,15]. This review presents new advances of few promising carcino-embryonic biomarkers for HCC immunotherapy on basic studies or clinical trials.

ALPHA-FETOPROTEIN

A glycoprotein of alpha-fetoprotein (AFP) synthesized from human fetal liver or HCC tissues^[16], consisting of 609 single-chain amino acid polypeptides and containing 24 Leading signal points (9-10 amino acid) residues located in three N-terminal domains, the major histocompatibility complex (MHC) class I or II molecules recognize these precursor signals and present them to CD4⁺ T cells and CD8⁺ T cells, the activated T cells recognize the body's immunodominant or sub-immunodominant epitopes^[17]. Amino acid peptide sequences and immunogenicity of human AFP epitopes are shown in Table 1. These immunogenic or sub-immunogenic AFP peptide chains could play an

immune-modulatory role in human, having the function and ability of polypeptide vaccine, and could induce and stimulate human anti-AFP specific immune responses.

AFP peptide chains have several fragments showing immunodominant or sub-immunodominant epitopes, which could be recognized by the MHC-I molecules, specifically induce T cells to activate and recognize AFP antigen. AFP positive peripheral blood mononuclear cells (PBMC) containing five human leukocyte antigen (HLA)-A*24:02 restricted T cell epitopes, AFP-derived peptide induces cytotoxic T lymphocyte (CTL) to produce interferon- γ (INF- γ), which can kill AFP positive HCC cells. Although it has been in clinical trials, the function of dendritic cells (DC), specific CTL, CD8⁺ T cells response and targeting therapy for AFP positive HCC cells remains to be studied. Now, T cell receptor (TCR) has been prepared by induction and screening *in vitro*, which can specifically recognize and bind AFP/HLA-A*02 antigen that is restricted to AFP158-166 peptide (FMNKFIYEI) to lay a foundation for HCC immunotherapy^[18]. A novel HLA-A*24:02 antigen was found to be more common than the HLA-A*02:01 in Asian HCC patients. Its restrictive peptide (KWVESIFLIF, AFP2-11signal) was found to be soluble in healthy human monocyte AFP 2-11-HLA-A*24:02-specific TCR (KWV3.1). T cells could be activated specifically and kill AFP positive T2-A24 HCC cells that contained AFP 2-11 and HLA-A*24:02⁺ antigen, indicated that AFP⁺HLA-A*24:02⁺ antigen might be a new immunotherapeutic target for HCC^[19].

Combination of anti-CTL-A-4 therapy (Tremelimumab) together with ablation in advanced HCC cases has ¹⁸ shown that the killing tumors by direct methods can result in immune system being activated or switched on. There are new drugs available known as immune checkpoint inhibitors (ICIs) which could enhance anti-HCC effect. After the patients with Tremelimumab treatment, the blood ¹⁴ CD4⁺- HLA-DR⁺, CD4⁺PD-1⁺, CD8⁺HLA-DR⁺, CD8⁺PD-1⁺, CD4⁺ICOS⁺, and CD8⁺ICOS⁺ T cells increased, the patients with higher CD⁺PD-1⁺ cells responded well to the treatment, with increasing specific CD8⁺PD-1 T cells for AFP & survivin, and higher CD3⁺T cells for tumor infiltrating, suggesting that Tremelimumab with ablation is a novel potential method with

increasing CD8⁺ T cells and decreasing circulating HCV amount for advanced HCC effective therapy^[20].

ANGIOGENIC FACTORS

Most patients with HCC are diagnosed at an advanced stage of disease. Until recently, systemic treatment options that showed survival benefits in HCC have been limited to tyrosine kinase inhibitors, antibodies targeting oncogenic signaling pathways or VEGF receptors^[21]. Angiogenesis plays an important role in HCC progression, and VEGF and angiopoietin (Ang) are key drivers of HCC angiogenesis. A better understanding of the relation between VEGF and HCC angiogenesis or progression may reveal their potential as biomarker for HCC diagnosis and therapy. VEGF- targeting strategies already represent an important component of today's systemic treatment landscape of HCC, whereas targeting the Ang/Tie2 signaling pathway may harbor future potential in this context due to reported beneficial anticancer effects when targeting this pathway^[22,23]. Following a decade of negative Phase III trials since the approval of sorafenib, more recently several drugs have proven efficacy both in first line *vs* sorafenib (lenvatinib) or in second line *vs* placebo (regorafenib, cabozantinib, ramucirumab/Cyramza®). A fully human anti-VEGFR-2 recombinant IgG1 monoclonal antibody (Ramucirumab) has been approved as monotherapy for HCC patients with AFP levels ≥ 400 ng/mL who have been treated with sorafenib, with significantly prolonged overall survival (OS) and progression-free survival. Its safety profile was consistent with that expected for agents targeting the VEGF/ VEGFR axis. The potential clinical development of systemic treatments in HCC, focusing on combination therapies with immunotherapy and treatment sequences as a way to maximize survival benefit^[24,25].

HCC microenvironment is characterized by a dysfunction of the immune system through multiple mechanisms, including accumulation of various immune- suppressive factors, recruitment of regulatory T cells and myeloid-derived suppressor cells, and induction of T cell exhaustion accompanied with the interaction between immune

checkpoint ligands and receptors. ICIs have been interfered this interaction and have altered therapeutic landscape of multiple cancer types including HCC. Intermediate-stage HCC¹⁶ with different levels of liver function, tumor size, and number of lesions may all have intermediate-stage disease according to the BCLC staging system. Their treatments includes conventional or drug-eluting bead transarterial chemoembolization, yttrium-90 radioembolization, thermal ablation, bland embolization, and combination therapy with VEGF inhibitors or ICIs. The clinical evidence supported available locoregional treatment options for intermediate-stage HCC^[26]. Although optimal sequencing is an area of ongoing investigation, multiple targeted therapies have improved OS in intermediate or advanced HCC^[27]. Several⁴ targeted agents including multi-tyrosine kinase inhibitors and immunotherapy agents have been approved for use beyond the frontline setting in patients with advanced HCC, combinatorial therapeutic strategies is an evolving approach showing early promising signal^[23,28]. Success of PD-1 monotherapy, combinatorial regimens with PD-1/PD-L1 inhibitors plus VEGF targeted agents shown positive results in various malignancies including HCC.⁴ These innovative approaches enhance the intensity of cancer-directed immune responses and will potentially impact the outlook of this aggressive disease^[29].

GLYPICAN-3

Regarding HCC, one promising antigen appears to be glypican-3 (GPC3) that is over-expressed by HCC tissues¹ and has been associated with worse disease-free survival and overall survival. GPC3 is involved in many signaling cascades that promote cell growth and invasion, including the Wnt pathway that is well-known for its role in embryogenesis.³ GPC3 as an oncofetal proteoglycan anchored to the cell membrane of HCC, is normally detected in the fetal liver but not in the healthy adult liver^[30,31]. However, the abnormal levels of GPC3 expression in tissues or sera of HCC patients are expressed at GPC3 mRNA gene transcription or protein levels, and predicts a poor prognosis of HCC.³ Mechanistic studies have revealed that GPC3 functions by binding

to molecules such as the Wnt/ β -catenin signaling or growth factors during HCC occurrence and progression. Moreover, specific serological GPC3 has been used as a diagnostic or prognostic serological marker, and a molecular-targeted for molecular imaging or therapeutic intervention in HCC^[32-34]. GPC3 as a molecular target for HCC immunotherapy is shown in Table 2. Up to date, GPC3-targeted magnetic resonance imaging, positron emission tomography, and near-infrared imaging have been investigated at early stage of HCC, and immunotherapeutic protocols targeting GPC3 have been developed, including the use of humanized anti-GPC3 cytotoxic antibodies, peptide/DNA vaccines, immuno- toxin therapies, and genetic therapies.

Different synergisms have been postulated based on the potential interplay between anti-angiogenic drugs and immunotherapy, with several clinical trials currently testing. Since the most extensively tested combination regimens for advanced HCC comprise anti-PD-1/anti-PD-L1 agents plus anti-angiogenic agents, oncogenic GPC3 is becoming an ideal promising candidate for HCC immunotherapy because of highly expressed in cancerous tissues but limited in normal liver tissues. Recently, the adoptive transfer of hGPC3-specific chimeric antigen receptor T (CAR-T) cells for HCC treatment has been conducted in clinical trials. Due to the rigid construction, conventional CAR-T cells have some intrinsic limitations, like uncontrollable over- activation and inducing severe cytokine release syndrome. By using co-culturing assays and a xenograft mouse model, the *in vitro* and *in vivo* cytotoxicity and cytokine release of the split anti-hGPC3 CAR-T cells were evaluated against various HCC cell lines and compared with conventional CAR-T cells. *In vitro* data demonstrated that split anti-hGPC3 CAR-T cells could recognize and lyse hGPC3 positive HepG2 and Huh7 cells in a dose-dependent manner. Impressively, the split anti-hGPC3 CAR-T cells produced and released a significantly lower amount of pro-inflammatory cytokines, including IFN- γ , TNF- α , IL-6, and GM-CSF, than conventional CAR-T cells. When injected into immune-deficient mice inoculated subcutaneously with HepG2 cells, our split anti-hGPC3 CAR-T cells could suppress HCC tumor growth, but released significantly lower levels of cytokines than conventional CAR-T cells. The split anti-hGPC3 CAR-T cells could suppress tumor

growth and reduce cytokine release, and represent a more versatile and safer alternative to conventional CAR-T cells for HCC treatment^[35,36]. The most recent data on novel combination strategies and targets, as well as looking ahead to the future role of molecular therapies in the treatment of advanced HCC. Current barriers of CAR-T therapy include its high production cost and need to identify validated extracellular HCC-specific antigens^[33,37].

WNT3a

Several signaling pathways involved in HCC have been studied, including STAT3-NF κ B, JAK-STAT, RAS MAPK, PI3K-AKT-mTOR, and Wnt- β -catenin. Of these, cascades involving mitogen-activated protein kinase (MAPK) emerge as key regulators of HCC. Bath of HBV and HCV infection can induce the activation of Wnt/ β -catenin signal pathway and participate in HCC progression^[38,39]. Oncogenic HBx of HBV can activate Src kinase to inhibit GSK3 β activity and make intracellular β -catenin accumulation, promote the expression of DNA methyl-transferase I and Wnt3a to bind and silence secreted frizzled related protein 1 and 5^[40]. HBx can reduce the inhibiting role for deacetylase 1 to β -catenin, and activating Wnt pathway promotes HCC development^[41]. Also, the core protein of HCV can promote Wnt3a expression, induce TCF dependent transcription, inhibit GSK3 β , increase and stabilize intracellular β -catenin to nucleus transport, and up-regulate the expressions of cyclinD1, c-myc, WISP2, Wnt3a, Wnt1 and CTGF to promote the HCC growth, DNA synthesis for HCC progression^[42]. Wnt3a is a critical signal molecule among the 19 mammalian Wnt proteins. The higher level of Wnt3a expression was only found in sera or tissues of HCC patients from a cohort cases with chronic liver diseases^[43,44], and it is the first time to report as a novel specific marker for HCC diagnosis and prognosis^[45,46].

Abnormal Wnt3a expression is involved in the development and metastasis of HCC^[47], and might be a novel strategy for HBV or HCV-related HCC therapy. Hepatic higher LINC00662 correlated with poor survival of HCC patients^[48,49], and might up-regulate Wnt3a expression by competitively binding miR-15a, miR-16 and miR-107,

with tumor-associated macrophages as a major component of tumor microenvironment in HCC progression, and they have been revealed ¹ the associations between Wnt3a signaling and cancer initiation, tumor growth, metastasis, dormancy, immunity and tumor stem cell maintenance^[40]. Wnt3a is one of HCC-related Wnt signals exhibited numerous genetic abnormalities as well as epigenetic alterations including modulation of DNA methylation. Targeted *Wnt3a* gene transcription might be an effective molecule-targeted therapy for HCC. Novel Crispr/Cas9-gsRNA lentiviral vector system with the advantages of higher targeting accuracy has been successfully used to affect the Wnt3a gene transcription in human HCC cell lines at mRNA level *in vitro* and confirmed at protein level *in vivo* with transplanted tumor studies^[44,50].

The inhibitory effect of Wnt3a on the proliferation of HCC cells or the growth of HCC xenograft models has been demonstrated that interfering with *Wnt3a* gene transcription could significantly inhibit the expressions of down-stream β -catenin and related-signal molecules^[51]. The xenograft model of knockout of Wnt3a gene in HepG2 cells resulted in a slower growth, significant reduction of tumor size or loss of weight. The molecular mechanism is Wnt3a cascade reaction involving multiple targets, can block upstream GPC-3 signal and downstream β -catenin to nucleus transport^[52,53], and inhibiting or delaying cancer progression by using specific antibodies (OMP-54F28, OTSA101)^[54] and small size peptide SAH-BCL-9^[55]. As the abnormality of liver and circulating Wnt3a expression in HCC has provided initial evidence, the tumor volume after intervening Wnt3a mRNA transcription with specific shRNA was $355.0 \pm 99.9 \text{ mm}^3$ in the intervention group with significantly lower than that ($869.4 \pm 222.5 \text{ mm}^3$) in the negative group, and the tumor formation days of the intervention group were longer than that of the negative group; the tumor weight ($0.35 \pm 0.11 \text{ g}$) of the intervention group was markedly lower than that ($0.88 \pm 0.20 \text{ g}$) of the negative group. Immunohistochemistry ¹ showed that Wnt3a was strongly inhibited in the intervention group^[56], and indicated that targeted-Wnt3a signaling could be a promising target or an effective target for HCC therapy.

CLUSTERIN

Secretory clusterin (sCLU) is a stress-induced heterodimer sulfated glycoprotein, located on chromosome 8q21-q12, which is highly conserved between species and has cytoprotective effect. Its biological function as a small molecule partner is almost similar to that of heat shock protein^[57]. Basic and clinical studies showed that sCLU expression was low in normal liver tissues and its activation during the malignant transformation of hepatocytes was progressive over-expression^[58,59], which was closely associated with HCC progression by contributing to angiogenesis, chemo- resistance, cell survival, and metastasis^[60]. The positive rate of hepatic sCLU expression was up to 73.3% at stage I of HCC by immunohistochemical analysis. Its expression at mRNA or protein level were increased with the clinical staging of HCC, indicated that sCLU could be a biomarker for differentiating benign from malignant liver diseases^[61].

Recurrence and metastasis after hepatectomy are the main causes of poor prognosis of HCC^[62]. Hepatic sCLU plays an important role in the proliferation, multidrug resistance, invasion and metastasis of HCC cells^[63,64]. The sCLU mediated the expression of MMP-2, p-AKT and E-cadherin in human HCC BEL-7402 and SMMC-7721 cell lines, and down-regulating sCLU expression can significantly reduce the invasive ability of HCC cells by selective COX-2 inhibitor meloxicam combined with specific sCLU-shRNA plasmids^[65,66]. These data indicated that sCLU should be a new effective target for the occurrence, invasion and metastasis of HCC, and should have a bright future in HCC immunotherapy.

INSULIN-LIKE GROWTH FACTOR AXIS

Hepatic insulin-like growth factor (IGF) axis contains ligands, receptors, substrates, and ligand binding proteins. Accumulating data demonstrated that aberrant IGFs signaling molecules may lead to malignant transformation of hepatocytes and HCC progression, especially in IGF-II or IGF-I receptor (IGF-IR) as key molecules in hepatocarcinogenesis^[67] or rat xenograft models^[68], affects the molecular pathogenesis of HCC, thus providing the rationale for targeting IGF axis in HCC^[69]. The biological

activities of IGF-II or IGF-IR not only promote HCC cell proliferation or xenograft growth, but also confer resistance to standard treatments^[70]. Several strategies targeting this system including monoclonal antibodies against IGF-1R and small molecule inhibitors of the tyrosine kinase function of IGF-1R are under active investigation. For example, DX-2647, a recombinant human antibody, potentially neutralizes the action of IGF-II, which is overexpressed in primary HCC^[71] and impairs the xenograft growth of Hep3B but not HepG2 cell line with high p-STAT3 Levels, suggesting that STAT3 activation as one pathway that might mediate resistance to IGF-II-targeted therapy in HCC^[72].

The over-expression of hepatic IGF-IR in human HCC promotes HCC cell proliferation, and attaching importance to IGF-IR might improve the prognostic or therapy of HCC^[73]. Enhancer of zeste 2 polycomb repressive complex 2 subunit (EZH2) as a regulator of promoting IGF-IR induced sorafenib resistance of HCC *in vitro* by directly transcriptionally repressing a set of microRNAs including miR-101, miR-122, miR-125b, and miR-139^[74-76]. A model for an EZH2-miRNAs-IGF-IR regulatory axis might provide insights into how to reversal sorafenib resistance in HCC. Silencing IGF-IR gene by specific shRNA on the inhibition of cell proliferation *in vitro* or rat xenograft growth *in vivo* to elucidate it as a novel molecular-targeted therapy for HCC. Several strategies targeting this system including monoclonal antibodies against IGF-IR and inhibitors of the tyrosine kinase function of IGF-IR are under active investigation. Gene-specific shRNA against IGF-signaling molecules as well as IGF-IR selective receptor tyrosine kinase (RTK)-inhibitors (tyrphostins) may therefore offer new therapeutic options^[77,78]. However, since specific shRNA is currently not applicable in HCC therapy, selective RTK-inhibitors represent the most promising approach for future therapeutic strategies.

SYNERGY OF NON-CODING RNAS

While immunotherapy holds great promise for combating cancer, the limited efficacy due to an immunosuppressive tumor microenvironment and systemic toxicity hinder

the broader application of immunotherapy^[79,80]. Combinatorial immunotherapy approach that uses a highly efficient and tumor-selective gene carrier to improve anticancer efficacy and circumvent the systemic toxicity. HCC is one of the multi-genetic diseases, and multiple studies have highlighted the key roles of noncoding RNAs (ncRNAs) in chemoresistance of HCC such as biomarkers and functional modulation of cellular response to sorafenib^[81-83]. Targeted chemotherapeutic agent, sorafenib, is known to show a statistically significant but limited overall survival advantage in advanced HCC, linked with the modulation of several intracellular signaling pathways through diverse operating biomolecules including ncRNAs^[84-86]. Accumulated evidences have demonstrated that ncRNAs (miRNAs, long ncRNAs or lncRNAs, and circular RNA or circRNA) could serve as biomarkers in diagnosis, prognosis, and treatment of HCC^[87,88] that have been well-documented to participate in HCC progression with promoting or inhibiting roles^[89,90].

Interestingly, miRNAs varied responses have been linked with the modulation of several intracellular signaling pathways^[91]. The abnormality of miR-218 expression was investigated in human HCC tissues or HCC cell lines for evaluating its function and underlying mechanisms of HCC. Gain-of-function and loss-of-function assays indicated that forced expression of miR-218 by inhibited HCC cell migration/invasion and reversed epithelial-mesenchymal transition to mesenchymal-epithelial transition. Serpine mRNA binding protein 1 (SERBP1) was a target gene of miR-218, and targeting the miR-218/SERBP1 signal pathway that inhibit the malignant phenotype formation might be a potential novel way for HCC therapeutics, because of miR-218 functions as a HCC suppressor involves in many biological processes such as tumor initiation, development, and metastasis^[92]. Nanotechnology-enabled dual delivery of siRNA and plasmid DNA that selectively targets and reprograms the immune-suppressive tumor microenvironment to improve HCC immunotherapy^[93-95].

HCC-associated circRNAs are abundant, and their over/Low expression might promote/inhibit HCC cell proliferation or tumor growth^[96-98]. Abnormality of circ-homer1 in HCC cells or tissues was related to tumor size, lymph node metastasis, high

clinical staging and poor prognosis. The molecular mechanism of circ-homer1 over-expression promoted the growth and invasiveness of HCC cell lines *via* mir-1322/cxc16 axis^[99]; conversely, interfering the circ-homer1 activation inhibited the proliferation, migration and invasion of HCC cell lines by increasing cell apoptosis. The circ-0051443 from circulating exosomes or HCC tissues could regulated BAK1 expression by combination of mir-331-3p to promote the cell apoptosis or cell cycle arrest of HCC, inhibit the biological behavior of HCC cells *in vivo* or nude mice HCC xenografts^[100]. Another interesting study also showed that has_circ_0008450 expression in HCC tissues or cells might inhibit HCC progression by regulating mir-214-3p/ezh2 axis^[101,102]. These data suggested that specific ncRNAs were useful molecular targets for HCC therapy.

CONCLUSION

In conclusion, HCC is a multi-gene variant malignant tumor with DNA methylation, microRNA, lncRNA expression and immune response^[103]. Immunotherapy for HCC has begun to produce better results, and HCC-specific molecules may be combined with comprehensive intervention such as surgery, interventional therapy, radiotherapy, and chemotherapy to improve the efficacy and prolong the survival time of HCC patients^[104]. In spite of the rapid development of genomics and proteomics, advances in molecular pathology, pharmacology and genetic engineering, DNA splicing, gene silencing, interfering transcription, and monoclonal antibody for more specific and less side effects immune therapy techniques^[105] that can directly block the signaling molecules involved in HCC growth related signaling pathways (Figure 1) or serve as molecular targets such as radionuclide, drug carriers, and immunotherapy play a unique role in specific or comprehensive treatment of HCC.

REFERENCES

- 1 **Craig AJ**, von Felden J, Garcia-Lezana T, Sarcognato S, Villanueva A. Tumour evolution in hepatocellular carcinoma. *Nat Rev Gastroenterol Hepatol* 2020; **17**: 139-152 [PMID: 31792430 DOI: 10.1038/s41575-019-0229-4]

- 2 **Forner A**, Reig M, Bruix J. Hepatocellular carcinoma. *Lancet* 2018; **391**: 1301-1314 [PMID: 29307467 DOI: 10.1016/S0140-6736(18)30010-2]
- 3 **Chen JG**, Zhu J, Zhang YH, Zhang YX, Yao DF, Chen YS, Lu JH, Ding LL, Chen HZ, Zhu CY, Yang LP, Zhu YR, Qiang FL. Cancer survival in Qidong between 1972 and 2011: A population-based analysis. *Mol Clin Oncol* 2017; **6**: 944-954 [PMID: 28588795 DOI: 10.3892/mco.2017.1234]
- 4 **Negro F**. Natural history of NASH and HCC. *Liver Int* 2020; **40 Suppl 1**: 72-76 [PMID: 32077608 DOI: 10.1111/liv.14362]
- 5 **Wang SZ**, Lee SD, Sarkar D, Lee HM, Khan A, Bhati C, Sharma A, Kumaran V, Bruno D, Cotterell A, Levy MF. Immunological characterization of hepatocellular carcinoma. *Hepatoma Res* 2021; **7**: 6 [DOI: 10.20517/2394-5079.2020.113]
- 6 **Hou J**, Zhang H, Sun B, Karin M. The immunobiology of hepatocellular carcinoma in humans and mice: Basic concepts and therapeutic implications. *J Hepatol* 2020; **72**: 167-182 [PMID: 31449859 DOI: 10.1016/j.jhep.2019.08.014]
- 7 **Rebouissou S**, Nault JC. Advances in molecular classification and precision oncology in hepatocellular carcinoma. *J Hepatol* 2020; **72**: 215-229 [PMID: 31954487 DOI: 10.1016/j.jhep.2019.08.017]
- 8 **Finn RS**, Zhu AX, Farah W, Almasri J, Zaiem F, Prokop LJ, Murad MH, Mohammed K. Therapies for advanced stage hepatocellular carcinoma with macrovascular invasion or metastatic disease: A systematic review and meta-analysis. *Hepatology* 2018; **67**: 422-435 [PMID: 28881497 DOI: 10.1002/hep.29486]
- 9 **Doycheva I**, Thuluvath PJ. Systemic Therapy for Advanced Hepatocellular Carcinoma: An Update of a Rapidly Evolving Field. *J Clin Exp Hepatol* 2019; **9**: 588-596 [PMID: 31695249 DOI: 10.1016/j.jceh.2019.07.012]
- 10 **D'Agnano I**, Berardi AC. Extracellular Vesicles, A Possible Theranostic Platform Strategy for Hepatocellular Carcinoma-An Overview. *Cancers (Basel)* 2020; **12** [PMID: 31973229 DOI: 10.3390/cancers12020261]

- 11 **Xia S**, Pan Y, Liang Y, Xu J, Cai X. The microenvironmental and metabolic aspects of sorafenib resistance in hepatocellular carcinoma. *EBioMedicine* 2020; **51**: 102610 [PMID: 31918403 DOI: 10.1016/j.ebiom.2019.102610]
- 12 **Reig M**, da Fonseca LG, Faivre S. New trials and results in systemic treatment of HCC. *J Hepatol* 2018; **69**: 525-533 [PMID: 29653122 DOI: 10.1016/j.jhep.2018.03.028]
- 13 **Casadei-Gardini A**, Orsi G, Caputo F, Ercolani G. Developments in predictive biomarkers for hepatocellular carcinoma therapy. *Expert Rev Anticancer Ther* 2020; **20**: 63-74 [PMID: 31910040 DOI: 10.1080/14737140.2020.1712198]
- 14 **Luo P**, Wu S, Yu Y, Ming X, Li S, Zuo X, Tu J. Current Status and Perspective Biomarkers in AFP Negative HCC: Towards Screening for and Diagnosing Hepatocellular Carcinoma at an Earlier Stage. *Pathol Oncol Res* 2020; **26**: 599-603 [PMID: 30661224 DOI: 10.1007/s12253-019-00585-5]
- 15 **Cheng AL**, Hsu C, Chan SL, Choo SP, Kudo M. Challenges of combination therapy with immune checkpoint inhibitors for hepatocellular carcinoma. *J Hepatol* 2020; **72**: 307-319 [PMID: 31954494 DOI: 10.1016/j.jhep.2019.09.025]
- 16 **Li XJ**, Shao DH, He ML, Liang GW. Association of Common Variants in HNF1A Gene with Serum AFP Level in Healthy Chinese Individuals and HCC Patients. *Dis Markers* 2019; **2019**: 6273497 [PMID: 31915469 DOI: 10.1155/2019/6273497]
- 17 **Docta RY**, Ferronha T, Sanderson JP, Weissensteiner T, Pope GR, Bennett AD, Pumphrey NJ, Ferjentsik Z, Quinn LL, Wiedermann GE, Anderson VE, Saini M, Maroto M, Norry E, Gerry AB. Tuning T-Cell Receptor Affinity to Optimize Clinical Risk-Benefit When Targeting Alpha-Fetoprotein-Positive Liver Cancer. *Hepatology* 2019; **69**: 2061-2075 [PMID: 30561769 DOI: 10.1002/hep.30477]
- 18 **Li Z**, Gong H, Liu Q, Wu W, Cheng J, Mei Y, Chen Y, Zheng H, Yu X, Zhong S, Li Y. Identification of an HLA-A*24:02-restricted α -fetoprotein signal peptide-derived antigen and its specific T-cell receptor for T-cell immunotherapy. *Immunology* 2020; **159**: 384-392 [PMID: 31849039 DOI: 10.1111/imm.13168]
- 19 **Agdashian D**, ElGindi M, Xie C, Sandhu M, Pratt D, Kleiner DE, Figg WD, Rytlewski JA, Sanders C, Yusko EC, Wood B, Venzon D, Brar G, Duffy AG, Greten TF, Korangy F.

The effect of anti-CTLA4 treatment on peripheral and intra-tumoral T cells in patients with hepatocellular carcinoma. *Cancer Immunol Immunother* 2019; **68**: 599-608 [PMID: 30688989 DOI: 10.1007/s00262-019-02299-8]

20 **Duffy AG**, Ulahannan SV, Makorova-Rusher O, Rahma O, Wedemeyer H, Pratt D, Davis JL, Hughes MS, Heller T, ElGindi M, Uppala A, Korangy F, Kleiner DE, Figg WD, Venzon D, Steinberg SM, Venkatesan AM, Krishnasamy V, Abi-Jaoudeh N, Levy E, Wood BJ, Greten TF. Tremelimumab in combination with ablation in patients with advanced hepatocellular carcinoma. *J Hepatol* 2017; **66**: 545-551 [PMID: 27816492 DOI: 10.1016/j.jhep.2016.10.029]

21 **Lee HW**, Cho KJ, Park JY. Current Status and Future Direction of Immunotherapy in Hepatocellular Carcinoma: What Do the Data Suggest? *Immune Netw* 2020; **20**: e11 [PMID: 32158599 DOI: 10.4110/in.2020.20.e11]

22 **Vanderborght B**, Lefere S, Vlierberghe HV, Devisscher L. The Angiopoietin/Tie2 Pathway in Hepatocellular Carcinoma. *Cells* 2020; **9** [PMID: 33143149 DOI: 10.3390/cells9112382]

23 **Saeed A**, Hildebrand H, Park R, Al-Jumayli M, Abbasi S, Melancon T, Saeed A, Al-Rajabi R, Kasi A, Baranda J, Williamson S, Sun W. Immune Checkpoint Inhibitors versus VEGF Targeted Therapy as Second Line Regimen in Advanced Hepatocellular Carcinoma (HCC): A Retrospective Study. *J Clin Med* 2020; **9** [PMID: 32824968 DOI: 10.3390/jcm9092682]

24 **Syed YY**. Ramucirumab: A Review in Hepatocellular Carcinoma. *Drugs* 2020; **80**: 315-322 [PMID: 32034692 DOI: 10.1007/s40265-020-01263-6]

25 **De Luca E**, Marino D, Di Maio M. Ramucirumab, A Second-Line Option For Patients With Hepatocellular Carcinoma: A Review Of The Evidence. *Cancer Manag Res* 2020; **12**: 3721-3729 [PMID: 32547208 DOI: 10.2147/CMAR.S216220]

26 **Chai NX**, Chapiro J. Therapy of Intermediate-Stage Hepatocellular Carcinoma: Current Evidence and Clinical Practice. *Semin Intervent Radiol* 2020; **37**: 456-465 [PMID: 33328701 DOI: 10.1055/s-0040-1719186]

- 27 **Lim H**, Ramjeesingh R, Liu D, Tam VC, Knox JJ, Card PB, Meyers BM. Optimizing Survival and the Changing Landscape of Targeted Therapy for Intermediate and Advanced Hepatocellular Carcinoma: A Systematic Review. *J Natl Cancer Inst* 2021; **113**: 123-136 [PMID: 32898239 DOI: 10.1093/jnci/djaa119]
- 28 **Mao CS**, Yin H, Ning HB, Peng Z, Li K, Ding GQ. Levels of HBx, VEGF, and CEACAM1 in HBV-related hepatocellular carcinoma and their correlation with cancer prognosis. *Eur Rev Med Pharmacol Sci* 2017; **21**: 3827-3833 [PMID: 28975984]
- 29 **Park R**, Eshrat F, Al-Jumayli M, Saeed A, Saeed A. Immuno-Oncotherapeutic Approaches in Advanced Hepatocellular Carcinoma. *Vaccines (Basel)* 2020; **8** [PMID: 32784389 DOI: 10.3390/vaccines8030447]
- 30 **Yao M**, Wang L, Fang M, Zheng W, Dong Z, Yao D. Advances in the study of oncofetal antigen glypican-3 expression in HBV-related hepatocellular carcinoma. *Biosci Trends* 2016; **10**: 337-343 [PMID: 27795482 DOI: 10.5582/bst.2016.01176]
- 31 **Zhou F**, Shang W, Yu X, Tian J. Glypican-3: A promising biomarker for hepatocellular carcinoma diagnosis and treatment. *Med Res Rev* 2018; **38**: 741-767 [PMID: 28621802 DOI: 10.1002/med.21455]
- 32 **Cao W**, Sharma M, Imam R, Yu J. Study on Diagnostic Values of Astrocyte Elevated Gene 1 (AEG-1) and Glypican 3 (GPC-3) in Hepatocellular Carcinoma. *Am J Clin Pathol* 2019; **152**: 647-655 [PMID: 31305883 DOI: 10.1093/ajcp/aqz086]
- 33 . Erratum for the Research Article: "PI4KIII β is a therapeutic target in chromosome 1q-amplified lung adenocarcinoma" by X. Tan, P. Banerjee, E. A. Pham, F. U. N. Rutaganira, K. Basu, N. Bota-Rabassedas, H.-F. Guo, C. L. Grzeskowiak, X. Liu, J. Yu, L. Shi, D. H. Peng, B. L. Rodriguez, J. Zhang, V. Zheng, D. Y. Duose, L. M. Solis, B. Mino, M. G. Raso, C. Behrens, I. I. Wistuba, K. L. Scott, M. Smith, K. Nguyen, G. Lam, I. Choong, A. Mazumdar, J. L. Hill, D. L. Gibbons, P. H. Brown, W. K. Russell, K. Shokat, C. J. Creighton, J. S. Glenn, J. M. Kurie. *Sci Transl Med* 2020; **12** [PMID: 32188723 DOI: 10.1126/scitranslmed.abb5995]
- 34 **Nishida T**, Kataoka H. Glypican 3-Targeted Therapy in Hepatocellular Carcinoma. *Cancers (Basel)* 2019; **11** [PMID: 31510063 DOI: 10.3390/cancers11091339]

- 35 **Liu X**, Wen J, Yi H, Hou X, Yin Y, Ye G, Wu X, Jiang X. Split chimeric antigen receptor-modified T cells targeting glypican-3 suppress hepatocellular carcinoma growth with reduced cytokine release. *Ther Adv Med Oncol* 2020; **12**: 1758835920910347 [PMID: 32215059 DOI: 10.1177/1758835920910347]
- 36 **Faivre S**, Rimassa L, Finn RS. Molecular therapies for HCC: Looking outside the box. *J Hepatol* 2020; **72**: 342-352 [PMID: 31954496 DOI: 10.1016/j.jhep.2019.09.010]
- 37 **Vormittag P**, Gunn R, Ghorashian S, Veraitch FS. A guide to manufacturing CAR T cell therapies. *Curr Opin Biotechnol* 2018; **53**: 164-181 [PMID: 29462761 DOI: 10.1016/j.copbio.2018.01.025]
- 38 **Piconese S**, Cammarata I, Barnaba V. Viral hepatitis, inflammation, and cancer: A lesson for autoimmunity. *J Autoimmun* 2018; **95**: 58-68 [PMID: 30509387 DOI: 10.1016/j.jaut.2018.10.021]
- 39 **Jiang XH**, Xie YT, Cai YP, Ren J, Ma T. Effects of hepatitis C virus core protein and nonstructural protein 4B on the Wnt/ β -catenin pathway. *BMC Microbiol* 2017; **17**: 124 [PMID: 28545480 DOI: 10.1186/s12866-017-1032-4]
- 40 **Janda CY**, Dang LT, You C, Chang J, de Lau W, Zhong ZA, Yan KS, Marecic O, Siepe D, Li X, Moody JD, Williams BO, Clevers H, Piehler J, Baker D, Kuo CJ, Garcia KC. Surrogate Wnt agonists that phenocopy canonical Wnt and β -catenin signalling. *Nature* 2017; **545**: 234-237 [PMID: 28467818 DOI: 10.1038/nature22306]
- 41 **Timperi E**, Barnaba V. Viral Hepatitides, Inflammation and Tumour Microenvironment. *Adv Exp Med Biol* 2020; **1263**: 25-43 [PMID: 32588321 DOI: 10.1007/978-3-030-44518-8_3]
- 42 **Wang L**, Yao M, Fang M, Zheng WJ, Dong ZZ, Pan LH, Zhang HJ, Yao DF. Expression of hepatic Wnt5a and its clinicopathological features in patients with hepatocellular carcinoma. *Hepatobiliary Pancreat Dis Int* 2018; **17**: 227-232 [PMID: 29709351 DOI: 10.1016/j.hbpd.2018.03.005]
- 43 **Sai WL**, Yao M, Zheng WJ, Wu MN, Sun JY, Pan LH, Dong ZZ, Yao DF. [Abnormal expression of Wnt3a and inhibiting role of its molecular-targeted intervening in

hepatocellular carcinoma]. *Zhonghua Gan Zang Bing Za Zhi* 2019; **27**: 866-871 [PMID: 31941241 DOI: 10.3760/cma.j.issn.1007-3418.2019.11.009]

44 **Zheng W**, Yao M, Fang M, Pan L, Wang L, Yang J, Dong Z, Yao D. Oncogenic Wnt3a: A Candidate Specific Marker and Novel Molecular Target for Hepatocellular Carcinoma. *J Cancer* 2019; **10**: 5862-5873 [PMID: 31737122 DOI: 10.7150/jca.31599]

45 **Pan L**, Yao M, Zheng W, Gu J, Yang X, Qiu L, Cai Y, Wu W, Yao D. Abnormality of Wnt3a expression as novel specific biomarker for diagnosis and differentiation of hepatocellular carcinoma. *Tumour Biol* 2016; **37**: 5561-5568 [PMID: 26577850 DOI: 10.1007/s13277-015-4413-z]

46 **Jingjing H**, Hongna H, Wenfu Z, Jianlin L, Guochu H, Yuanjia L, Songlin C, Yueqiang H. Bie Jia Jian Pill Combined with Bone Mesenchymal Stem Cells Regulates microRNA-140 to Suppress Hepatocellular Carcinoma Stem Cells. *Int J Stem Cells* 2021 [PMID: 33632990 DOI: 10.15283/ijsc20157]

47 **Lu C**, He Y, Duan J, Yang Y, Zhong C, Zhang J, Liao W, Huang X, Zhu R, Li M. Expression of Wnt3a in hepatocellular carcinoma and its effects on cell cycle and metastasis. *Int J Oncol* 2017; **51**: 1135-1145 [PMID: 28902357 DOI: 10.3892/ijo.2017.4112]

48 **Tian X**, Wu Y, Yang Y, Wang J, Niu M, Gao S, Qin T, Bao D. Long noncoding RNA LINC00662 promotes M2 macrophage polarization and hepatocellular carcinoma progression *via* activating Wnt/ β -catenin signaling. *Mol Oncol* 2020; **14**: 462-483 [PMID: 31785055 DOI: 10.1002/1878-0261.12606]

49 **You Y**, Que K, Zhou Y, Zhang Z, Zhao X, Gong J, Liu Z. MicroRNA-766-3p Inhibits Tumour Progression by Targeting Wnt3a in Hepatocellular Carcinoma. *Mol Cells* 2018; **41**: 830-841 [PMID: 30145863 DOI: 10.14348/molcells.2018.0181]

50 **Abd Elhameed AG**, Helal MG, Said E, Salem HA. Saxagliptin Defers Thioacetamide-Induced Hepatocarcinogenesis in Rats; A Novel Suppressive Impact on Wnt/Hedgehog /Notch1 Signaling. *Environ Toxicol Pharmacol* 2021: 103668 [PMID: 33945853 DOI: 10.1016/j.etap.2021.103668]

51 **Li N**, Wei L, Liu X, Bai H, Ye Y, Li D, Li N, Baxa U, Wang Q, Lv L, Chen Y, Feng M, Lee B, Gao W, Ho M. A Frizzled-Like Cysteine-Rich Domain in Glypican-3 Mediates

Wnt Binding and Regulates Hepatocellular Carcinoma Tumor Growth in Mice. *Hepatology* 2019; **70**: 1231-1245 [PMID: 30963603 DOI: 10.1002/hep.30646]

52 **Chen J**, Rajasekaran M, Xia H, Zhang X, Kong SN, Sekar K, Seshachalam VP, Deivasigamani A, Goh BK, Ooi LL, Hong W, Hui KM. The microtubule-associated protein PRC1 promotes early recurrence of hepatocellular carcinoma in association with the Wnt/ β -catenin signalling pathway. *Gut* 2016; **65**: 1522-1534 [PMID: 26941395 DOI: 10.1136/gutjnl-2015-310625]

53 **Yang Y**, Ye YC, Chen Y, Zhao JL, Gao CC, Han H, Liu WC, Qin HY. Crosstalk between hepatic tumor cells and macrophages *via* Wnt/ β -catenin signaling promotes M2-like macrophage polarization and reinforces tumor malignant behaviors. *Cell Death Dis* 2018; **9**: 793 [PMID: 30022048 DOI: 10.1038/s41419-018-0818-0]

54 **Sebio A**, Kahn M, Lenz HJ. The potential of targeting Wnt/ β -catenin in colon cancer. *Expert Opin Ther Targets* 2014; **18**: 611-615 [PMID: 24702624 DOI: 10.1517/14728222.2014.906580]

55 **Takada K**, Zhu D, Bird GH, Sukhdeo K, Zhao JJ, Mani M, Lemieux M, Carrasco DE, Ryan J, Horst D, Fulciniti M, Munshi NC, Xu W, Kung AL, Shivdasani RA, Walensky LD, Carrasco DR. Targeted disruption of the BCL9/ β -catenin complex inhibits oncogenic Wnt signaling. *Sci Transl Med* 2012; **4**: 148ra117 [PMID: 22914623 DOI: 10.1126/scitranslmed.3003808]

56 **Chikhaliwala P**, Rai R, Chandra S. Simultaneous voltammetric immunodetection of alpha-fetoprotein and glypican-3 using a glassy carbon electrode modified with magnetite-conjugated dendrimers. *Mikrochim Acta* 2019; **186**: 255 [PMID: 30904972 DOI: 10.1007/s00604-019-3354-4]

57 **Li Y**, Liu F, Zhou W, Zhang S, Chu P, Lin F, Wang HL. Diagnostic value of clusterin immunostaining in hepatocellular carcinoma. *Diagn Pathol* 2020; **15**: 127 [PMID: 33054843 DOI: 10.1186/s13000-020-01041-8]

58 **Zheng W**, Yao M, Qian Q, Sai W, Qiu L, Yang J, Wu W, Dong Z, Yao D. Oncogenic secretory clusterin in hepatocellular carcinoma: Expression at early staging and

emerging molecular target. *Oncotarget* 2017; **8**: 52321-52332 [PMID: 28881732 DOI: 10.18632/oncotarget.13674]

59 **Udomsinprasert W**, Poovorawan Y, Chongsrisawat V, Vejchapipat P, Honsawek S. Decreased circulating clusterin reflects severe liver complications after hepatopertoenterostomy of biliary atresia. *Sci Rep* 2020; **10**: 19736 [PMID: 33184463 DOI: 10.1038/s41598-020-76875-9]

60 **Wang X**, Liu Y, Qin Q, Zheng T. Clusterin role in hepatocellular carcinoma patients treated with oxaliplatin. *Biosci Rep* 2020; **40** [PMID: 32039450 DOI: 10.1042/BSR20200071]

61 **Park JS**, Lee WK, Kim HS, Seo JA, Kim DH, Han HC, Min BH. Clusterin overexpression protects against western diet-induced obesity and NAFLD. *Sci Rep* 2020; **10**: 17484 [PMID: 33060605 DOI: 10.1038/s41598-020-73927-y]

62 **Yao M**, Sai W, Zheng W, Wang L, Dong Z, Yao D. Secretory Clusterin as a Novel Molecular-targeted Therapy for Inhibiting Hepatocellular Carcinoma Growth. *Curr Med Chem* 2020; **27**: 3290-3301 [PMID: 31232234 DOI: 10.2174/0929867326666190624161158]

63 **Fu N**, Du H, Li D, Lu Y, Li W, Wang Y, Kong L, Du J, Zhao S, Ren W, Han F, Wang R, Zhang Y, Nan Y. Clusterin contributes to hepatitis C virus-related hepatocellular carcinoma by regulating autophagy. *Life Sci* 2020; **256**: 117911 [PMID: 32504756 DOI: 10.1016/j.lfs.2020.117911]

64 **Wang X**, Zou F, Zhong J, Yue L, Wang F, Wei H, Yang G, Jin T, Dong X, Li J, Xiu P. Secretory Clusterin Mediates Oxaliplatin Resistance *via* the Gadd45a/PI3K/Akt Signaling Pathway in Hepatocellular Carcinoma. *J Cancer* 2018; **9**: 1403-1413 [PMID: 29721050 DOI: 10.7150/jca.23849]

65 **Wu B**, Shang H, Liang X, Sun Y, Jing H, Han X, Cheng W. Preparation of novel targeting nanobubbles conjugated with small interfering RNA for concurrent molecular imaging and gene therapy *in vivo*. *FASEB J* 2019; **33**: 14129-14136 [PMID: 31657628 DOI: 10.1096/fj.201900716RR]

66 **Kuo PC**, Chau IY, Li AF, Chau YP, Hsia CY, Chau GY. Clusterin expression in nontumor tissue in patients with resectable hepatocellular carcinoma related with

postresectional survival. *J Chin Med Assoc* 2019; **82**: 929-934 [PMID: 31800534 DOI: 10.1097/JCMA.0000000000000195]

67 **Tai BJ**, Yao M, Zheng WJ, Shen YC, Wang L, Sun JY, Wu MN, Dong ZZ, Yao DF. Alteration of oncogenic IGF-II gene methylation status associates with hepatocyte malignant transformation. *Hepatobiliary Pancreat Dis Int* 2019; **18**: 158-163 [PMID: 30692043 DOI: 10.1016/j.hbpd.2019.01.003]

68 **Ngo MT**, Jeng HY, Kuo YC, Diony Nanda J, Brahmadhi A, Ling TY, Chang TS, Huang YH. The Role of IGF/IGF-1R Signaling in Hepatocellular Carcinomas: Stemness-Related Properties and Drug Resistance. *Int J Mol Sci* 2021; **22** [PMID: 33669204 DOI: 10.3390/ijms22041931]

69 **Wang L**, Yao M, Zheng W, Fang M, Wu M, Sun J, Dong Z, Yao D. Insulin-like Growth Factor I Receptor: A Novel Target for Hepatocellular Carcinoma Gene Therapy. *Mini Rev Med Chem* 2019; **19**: 272-280 [PMID: 30360707 DOI: 10.2174/1389557518666181025151608]

70 **Hua H**, Kong Q, Yin J, Zhang J, Jiang Y. Insulin-like growth factor receptor signaling in tumorigenesis and drug resistance: a challenge for cancer therapy. *J Hematol Oncol* 2020; **13**: 64 [PMID: 32493414 DOI: 10.1186/s13045-020-00904-3]

71 **Qian J**, Yao D, Dong Z, Wu W, Qiu L, Yao N, Li S, Bian Y, Wang Z, Shi G. Characteristics of hepatic igf-ii expression and monitored levels of circulating igf-ii mRNA in metastasis of hepatocellular carcinoma. *Am J Clin Pathol* 2010; **134**: 799-806 [PMID: 20959664 DOI: 10.1309/AJCPTFDSE2V3LCZP]

72 **Greenall SA**, Donoghue J, Johns TG, Adams TE. Differential Sensitivity of Human Hepatocellular Carcinoma Xenografts to an IGF-II Neutralizing Antibody May Involve Activated STAT3. *Transl Oncol* 2018; **11**: 971-978 [PMID: 29933129 DOI: 10.1016/j.tranon.2018.05.011]

73 **Bie C**, Chen Y, Tang H, Li Q, Zhong L, Peng X, Shi Y, Lin J, Lai J, Wu S, Tang S. Insulin-Like Growth Factor 1 Receptor Drives Hepatocellular Carcinoma Growth and Invasion by Activating Stat3-Midkine-Stat3 Loop. *Dig Dis Sci* 2021 [PMID: 33559791 DOI: 10.1007/s10620-021-06862-1]

74 **Cheng Z**, Wei-Qi J, Jin D. New insights on sorafenib resistance in liver cancer with correlation of individualized therapy. *Biochim Biophys Acta Rev Cancer* 2020; **1874**: 188382 [PMID: 32522600 DOI: 10.1016/j.bbcan.2020.188382]

75 **Ghosh MK**, Patra F, Ghosh S, Hossain CM, Mukherjee B. Antisense oligonucleotides directed against insulin-like growth factor-II messenger ribonucleic acids delay the progress of rat hepatocarcinogenesis. *J Carcinog* 2014; **13**: 2 [PMID: 24737950 DOI: 10.4103/1477-3163.126761]

76 **Wei L**, Wang X, Lv L, Liu J, Xing H, Song Y, Xie M, Lei T, Zhang N, Yang M. The emerging role of microRNAs and long noncoding RNAs in drug resistance of hepatocellular carcinoma. *Mol Cancer* 2019; **18**: 147 [PMID: 31651347 DOI: 10.1186/s12943-019-1086-z]

77 **Yao N**, Yao D, Wang L, Dong Z, Wu W, Qiu L, Yan X, Yu D, Chen J, Sai W, Zhang H, Yang J. Inhibition of autocrine IGF-II on effect of human HepG2 cell proliferation and angiogenesis factor expression. *Tumour Biol* 2012; **33**: 1767-1776 [PMID: 22684773 DOI: 10.1007/s13277-012-0436-x]

78 **Mohamed YI**, Lee S, Xiao L, Hassan MM, Qayyum A, Hiatia R, Pestana RC, Haque A, George B, Rashid A, Duda DG, Elghazaly H, Wolff RA, Morris JS, Yao J, Amin HM, Kaseb AO. Insulin-like growth factor 1/Child-Turcotte-Pugh composite score as a predictor of treatment outcomes in patients with advanced hepatocellular carcinoma treated with sorafenib. *Oncotarget* 2021; **12**: 756-766 [PMID: 33889299 DOI: 10.18632/oncotarget.27924]

79 **Kanthaje S**, Makol A, Chakraborti A. Sorafenib response in hepatocellular carcinoma: MicroRNAs as tuning forks. *Hepatol Res* 2018; **48**: 5-14 [PMID: 29055114 DOI: 10.1111/hepr.12991]

80 **Huang KW**, Hsu FF, Qiu JT, Chern GJ, Lee YA, Chang CC, Huang YT, Sung YC, Chiang CC, Huang RL, Lin CC, Dinh TK, Huang HC, Shih YC, Alson D, Lin CY, Lin YC, Chang PC, Lin SY, Chen Y. Highly efficient and tumor-selective nanoparticles for dual-targeted immunogene therapy against cancer. *Sci Adv* 2020; **6**: eaax5032 [PMID: 31998834 DOI: 10.1126/sciadv.aax5032]

- 81 **Niu L**, Liu L, Yang S, Ren J, Lai PBS, Chen GG. New insights into sorafenib resistance in hepatocellular carcinoma: Responsible mechanisms and promising strategies. *Biochim Biophys Acta Rev Cancer* 2017; **1868**: 564-570 [PMID: 29054475 DOI: 10.1016/j.bbcan.2017.10.002]
- 82 **Zhou B**, Yang H, Yang C, Bao YL, Yang SM, Liu J, Xiao YF. Translation of noncoding RNAs and cancer. *Cancer Lett* 2021; **497**: 89-99 [PMID: 33038492 DOI: 10.1016/j.canlet.2020.10.002]
- 83 **Gramantieri L**, Pollutri D, Gagliardi M, Giovannini C, Quarta S, Ferracin M, Casadei-Gardini A, Callegari E, De Carolis S, Marinelli S, Benevento F, Vasuri F, Ravaioli M, Cescon M, Piscaglia F, Negrini M, Bolondi L, Fornari F. MiR-30e-3p Influences Tumor Phenotype through *MDM2/TP53* Axis and Predicts Sorafenib Resistance in Hepatocellular Carcinoma. *Cancer Res* 2020; **80**: 1720-1734 [PMID: 32015093 DOI: 10.1158/0008-5472.CAN-19-0472]
- 84 **Li Y**, He X, Zhang X, Xu Y, Wu Y, Xu X. Immune-related microRNA signature for predicting prognosis and the immune microenvironment in hepatocellular carcinoma. *Life Sci* 2021; **265**: 118799 [PMID: 33220285 DOI: 10.1016/j.lfs.2020.118799]
- 85 **Yugawa K**, Yoshizumi T, Mano Y, Itoh S, Harada N, Ikegami T, Kohashi K, Oda Y, Mori M. Cancer-associated fibroblasts promote hepatocellular carcinoma progression through downregulation of exosomal miR-150-3p. *Eur J Surg Oncol* 2021; **47**: 384-393 [PMID: 32883551 DOI: 10.1016/j.ejso.2020.08.002]
- 86 **Lim LJ**, Wong SYS, Huang F, Lim S, Chong SS, Ooi LL, Kon OL, Lee CG. Roles and Regulation of Long Noncoding RNAs in Hepatocellular Carcinoma. *Cancer Res* 2019; **79**: 5131-5139 [PMID: 31337653 DOI: 10.1158/0008-5472.CAN-19-0255]
- 87 **Zhou Y**, Ren H, Dai B, Li J, Shang L, Huang J, Shi X. Hepatocellular carcinoma-derived exosomal miRNA-21 contributes to tumor progression by converting hepatocyte stellate cells to cancer-associated fibroblasts. *J Exp Clin Cancer Res* 2018; **37**: 324 [PMID: 30591064 DOI: 10.1186/s13046-018-0965-2]

- 88 **Sun C**, Xiao T, Xiao Y, Li Y. Silencing of long non-coding RNA NEAT1 inhibits hepatocellular carcinoma progression by downregulating SMO by sponging microRNA-503. *Mol Med Rep* 2021; **23** [PMID: 33398379 DOI: 10.3892/mmr.2020.11807]
- 89 **Sukowati CHC**, Cabral LKD, Tiribelli C, Pascut D. Circulating Long and Circular Noncoding RNA as Non-Invasive Diagnostic Tools of Hepatocellular Carcinoma. *Biomedicines* 2021; **9** [PMID: 33477833 DOI: 10.3390/biomedicines9010090]
- 90 **Salerno D**, Chiodo L, Alfano V, Floriot O, Cottone G, Paturel A, Pallocca M, Plissonnier ML, Jeddari S, Belloni L, Zeisel M, Levrero M, Guerrieri F. Hepatitis B protein HBx binds the DLEU2 lncRNA to sustain cccDNA and host cancer-related gene transcription. *Gut* 2020; **69**: 2016-2024 [PMID: 32114505 DOI: 10.1136/gutjnl-2019-319637]
- 91 **Onuma AE**, Zhang H, Huang H, Williams TM, Noonan A, Tsung A. Immune Checkpoint Inhibitors in Hepatocellular Cancer: Current Understanding on Mechanisms of Resistance and Biomarkers of Response to Treatment. *Gene Expr* 2020; **20**: 53-65 [PMID: 32340652 DOI: 10.3727/105221620X15880179864121]
- 92 **Wang T**, Xu L, Jia R, Wei J. MiR-218 suppresses the metastasis and EMT of HCC cells via targeting SERBP1. *Acta Biochim Biophys Sin (Shanghai)* 2017; **49**: 383-391 [PMID: 28369267 DOI: 10.1093/abbs/gmx017]
- 93 **Aspichueta P**. Lipid-rich environment: a key role promoting carcinogenesis in obesity-related non-alcoholic fatty liver disease. *Gut* 2018; **67**: 1376-1377 [PMID: 29540438 DOI: 10.1136/gutjnl-2018-316047]
- 94 **Zayac A**, Almhanna K. Hepatobiliary cancers and immunotherapy: where are we now and where are we heading? *Transl Gastroenterol Hepatol* 2020; **5**: 8 [PMID: 32190776 DOI: 10.21037/tgh.2019.09.07]
- 95 **Gan L**, Liu Z, Sun C. Obesity linking to hepatocellular carcinoma: A global view. *Biochim Biophys Acta Rev Cancer* 2018; **1869**: 97-102 [PMID: 29366974 DOI: 10.1016/j.bbcan.2017.12.006]
- 96 **Zhang H**, Deng T, Ge S, Liu Y, Bai M, Zhu K, Fan Q, Li J, Ning T, Tian F, Li H, Sun W, Ying G, Ba Y. Exosome circRNA secreted from adipocytes promotes the growth of

hepatocellular carcinoma by targeting deubiquitination-related USP7. *Oncogene* 2019; **38**: 2844-2859 [PMID: 30546088 DOI: 10.1038/s41388-018-0619-z]

97 **Zhu C**, Su Y, Liu L, Wang S, Liu Y, Wu J. Circular RNA hsa_circ_0004277 Stimulates Malignant Phenotype of Hepatocellular Carcinoma and Epithelial-Mesenchymal Transition of Peripheral Cells. *Front Cell Dev Biol* 2020; **8**: 585565 [PMID: 33511111 DOI: 10.3389/fcell.2020.585565]

98 **Wang S**, Zhang K, Tan S, Xin J, Yuan Q, Xu H, Xu X, Liang Q, Christiani DC, Wang M, Liu L, Du M. Circular RNAs in body fluids as cancer biomarkers: the new frontier of liquid biopsies. *Mol Cancer* 2021; **20**: 13 [PMID: 33430880 DOI: 10.1186/s12943-020-01298-z]

99 **Zhao M**, Dong G, Meng Q, Lin S, Li X. Circ-HOMER1 enhances the inhibition of miR-1322 on CXCL6 to regulate the growth and aggressiveness of hepatocellular carcinoma cells. *J Cell Biochem* 2020; **121**: 4440-4449 [PMID: 32037619 DOI: 10.1002/jcb.29672]

100 **Chen W**, Quan Y, Fan S, Wang H, Liang J, Huang L, Chen L, Liu Q, He P, Ye Y. Exosome-transmitted circular RNA hsa_circ_0051443 suppresses hepatocellular carcinoma progression. *Cancer Lett* 2020; **475**: 119-128 [PMID: 32014458 DOI: 10.1016/j.canlet.2020.01.022]

101 **Du Q**, Han J, Gao S, Zhang S, Pan Y. Hypoxia-induced circular RNA hsa_circ_0008450 accelerates hepatocellular cancer progression *via* the miR-431/AKAP1 axis. *Oncol Lett* 2020; **20**: 388 [PMID: 33193848 DOI: 10.3892/ol.2020.12251]

102 **Lin T**, Dai Y, Guo X, Chen W, Zhao J, Cao L, Wu Z. Silencing Of hsa_circ_0008450 Represses Hepatocellular Carcinoma Progression Through Regulation Of microRNA-214-3p/EZH2 Axis. *Cancer Manag Res* 2019; **11**: 9133-9143 [PMID: 31695501 DOI: 10.2147/CMAR.S222716]

103 **Sutti S**, Albano E. Adaptive immunity: an emerging player in the progression of NAFLD. *Nat Rev Gastroenterol Hepatol* 2020; **17**: 81-92 [PMID: 31605031 DOI: 10.1038/s41575-019-0210-2]

- 104 **Shen S**, Peng H, Wang Y, Xu M, Lin M, Xie X, Peng B, Kuang M. Screening for immune-potentiating antigens from hepatocellular carcinoma patients after radiofrequency ablation by serum proteomic analysis. *BMC Cancer* 2018; **18**: 117 [PMID: 29386009 DOI: 10.1186/s12885-018-4011-8]
- 105 **Nishida N**, Kudo M. Immune Phenotype and Immune Checkpoint Inhibitors for the Treatment of Human Hepatocellular Carcinoma. *Cancers (Basel)* 2020; **12** [PMID: 32443599 DOI: 10.3390/cancers12051274]

28%

SIMILARITY INDEX

PRIMARY SOURCES

1	hrjournal.net Internet	185 words — 4%
2	doaj.org Internet	155 words — 4%
3	onlinelibrary.wiley.com Internet	154 words — 4%
4	www.mdpi.com Internet	128 words — 3%
5	www.ncbi.nlm.nih.gov Internet	102 words — 2%
6	pesquisa.bvsalud.org Internet	99 words — 2%
7	www.scilit.net Internet	68 words — 2%
8	academic.oup.com Internet	46 words — 1%
9	Yahiya Y. Syed. "Ramucirumab: A Review in Hepatocellular Carcinoma", Drugs, 2020 Crossref	36 words — 1%

10	Jing Hu, Jing Zhang, Feifei Sun, Mei Qi, Peng Su, Hui Liu, Lin Gao, Meng Jiao, Zhen Wu, Lei Xiang, Bo Han. "Enhancer of zeste 2 polycomb repressive complex 2 subunit promotes sorafenib resistance of hepatocellular carcinoma though insulin-like growth factor 1 receptor", Anti-Cancer Drugs, 2019 Crossref	35 words — 1%
11	www.dovepress.com Internet	34 words — 1%
12	beta.eurekaselect.com Internet	31 words — 1%
13	explora.unex.es Internet	25 words — 1%
14	Padmaraju Vasudevaraju, Malla Rama Rao. "Chapter 6 Immune Checkpoint Inhibitors in Gastrointestinal Malignancies", Springer Science and Business Media LLC, 2020 Crossref	21 words — < 1%
15	lib.bioinfo.pl Internet	21 words — < 1%
16	www.journaltocs.ac.uk Internet	21 words — < 1%
17	research.kindai.ac.jp Internet	20 words — < 1%
18	pubmed.ncbi.nlm.nih.gov Internet	18 words — < 1%

EXCLUDE QUOTES ON
EXCLUDE BIBLIOGRAPHY ON

EXCLUDE MATCHES < 16 WORDS