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Micelles as potential drug delivery systems for colorectal cancer treatment

Zaynab Fatfat, Maamoun Fatfat, Hala Gali-Muhtasib

Abstract

Despite the significant progress in cancer therapy, colorectal cancer remains one of the most fatal malignancies worldwide. Chemotherapy is currently the mainstay therapeutic modality adopted for colorectal cancer treatment. However, the long-term effectiveness of chemotherapeutic drugs has been hampered by their low bioavailability, non-selective tumor targeting mechanisms, non-specific biodistribution associated with low drug concentrations at the tumor site and undesirable side effects. Over the last decade, there has been increasing interest in using nanotechnology-based drug delivery systems to circumvent these limitations. Various nanoparticles have been developed for delivering chemotherapeutic drugs among which polymeric micelles are attractive candidates. Polymeric micelles are biocompatible nanocarriers that can bypass the biological barriers and preferentially accumulate in tumors via the enhanced permeability and retention effect. They can be easily engineered with stimuliresponsive and tumor targeting moieties to further ensure their selective uptake by cancer cells and controlled drug release at the desirable tumor site. They have been shown to effectively improve the pharmacokinetic properties of chemotherapeutic drugs, enhance their safety profile and anticancer efficacy in different types of cancers. Given that combination therapy is the new strategy implemented in cancer therapy, polymeric micelles are suitable for multi-drug delivery and allow drugs to act concurrently at the action site to achieve synergistic therapeutic outcomes. They also allow the delivery of anticancer genetic material along with chemotherapy drugs

offering a novel approach for colorectal cancer therapy. Here, we highlight the properties of polymeric micelles that make them promising drug delivery systems for colorectal cancer treatment. We also review their application in colorectal cancer chemotherapy and gene therapy, as well as in combination cancer chemotherapy.

Key Words: Polymeric micelles; Drug delivery; Colorectal cancer; Chemotherapy; Gene therapy; Combination cancer therapy

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Core Tip: Polymeric micelles-based drug delivery has demonstrated promising therapeutic outcomes against colorectal cancer. These safe nanocarriers exhibit high encapsulation efficiency of chemotherapeutic drugs, improve their water solubility, and enhance the stability of nucleic acid-based therapeutics. They also accumulate preferentially at colorectal cancer sites, increase the anticancer effect of the delivered therapeutics, and reduce their side effects. Incorporating stimuli-responsive and tumor targeting moieties to their structure further enhances their targeting and therapeutic efficacy. This platform also provides an opportunity to simultaneously deliver different chemotherapeutic drugs or nucleic acid-based therapeutics with chemotherapeutic drugs to the colorectal tumor to achieve an enhanced anticancer response.

INTRODUCTION

Colorectal cancer is the third most prevalent cancer and the second leading cause of cancer death in men and women globally with estimated 1.9 million new cases and 0.9 million deaths in 2020^[1]. These numbers are predicted to reach 3.1 and 1.6 million respectively within the next two decades^[2]. The majority of colorectal cancer patients present with localized tumors and nearly 20–30% of colorectal cancer patients have unresectable metastasis^[3]. While the 5-year survival rate for localized colorectal cancers is nearly 90%, it is reduced to about 12% for distant metastatic colorectal cancers^[4].

Surgery is the primary treatment for patients in early stages of colorectal cancer, whereas preoperative or postoperative radiotherapy and chemotherapy are added for patients in advanced stages^[5]. Numerous chemotherapeutic drugs, approved by the Food and Drug Administration, have been used for colorectal cancer treatment including 5-fluorouracil (5-FU), irinotecan (IRI), oxaliplatin (OXA), and capecitabine (CPT)[3]. However, their clinical efficacy has been constrained by the adverse side effects and drug resistance which causes cancer relapse and treatment failure^[6-8]. To address these drawbacks, combination therapy has been used as an alternative therapeutic approach to cancer monotherapy. Combining therapeutic agents with different modes of action demonstrated great potential in achieving synergistic therapeutic outcomes with reduced side effects in addition to overcoming drug resistance^[9]. Several combination chemotherapeutic regimens such as FOLFIRI (folinic acid, 5-FU and IRI), XELOX/CAPOX (OXA and CPT), and FOLFOX (folinic acid, 5-FU and OXA) are widely used in clinical treatment for colorectal cancer^[10]. However, and despite the continued efforts devoted to advance colorectal cancer chemotherapy, the prognosis of this disease is still unsatisfactory, particularly for metastatic colorectal cancer patients^[11].

The main concerns limiting the success of single agent or combination chemotherapy are poor solubility, short half-life, rapid metabolism of the anticancer drugs, in addition to lack of selectivity for the tumor sites, non-specific biodistribution and subsequent failure to achieve effective therapeutic concentrations at the action sites^[12,13]. Over the past decades, nanotechnology-based drug delivery systems have emerged as a modern approach to overcome these limitations and improve drugs performance. They enhance the pharmacokinetics proprieties of the drugs, ensure their delivery to the tumor, prevent toxicity to healthy tissues, increase drug concentration at the tumor site and therefore potentiate their anticancer activity while mitigating their associated side effects^[14]. In combination therapy, they can effectively carry several drugs with dissimilar pharmacokinetics profiles and maintain the optimized synergistic drug ratio until they reach the target cancer cells^[15]. Additionally, nanosized drug delivery systems opened up new opportunities for the delivery of novel therapeutics along with chemotherapeutic agents, such as genetic anticancer agents which are considered to be innovative approaches in colorectal cancer management^[15].

Among the broad range of nanocarriers, polymeric micelles have received particular attention for therapeutic applications in cancer. Many anticancer agents are polycyclic compounds with poor aqueous solubility^[16], and polymeric micelles have proven to be powerful vehicles for loading and delivering hydrophobic drugs^[17]. Besides being biocompatible and biodegradable, polymeric micelles provide distinct advantages including small size, low-cost, ease of preparation, high drug encapsulation capacity and structural stability^[18-20]. They can be easily functionalized with stimuli-responsive and tumor targeting moieties to achieve site-specific delivery and controlled drug release^[21]. They also allow the integration of multiple functions other than drug delivery including gene delivery and imaging^[21]. To date, two polymeric micelles Genexol-PM loading paclitaxel (PTX) and Nanoxel loading docetaxel (DTX) have been approved in Korea and India respectively for the treatment of different types of cancer including breast and non-small-cell lung cancers^[22]. As these nanocarriers have shown therapeutic potential for these solid tumors, they could serve as drug delivery platforms to alleviate the current limitations of colorectal cancer chemotherapy and promote the introduction of gene therapy in the management of this disease. Here, we review the composition and characteristics of polymeric micelles in addition to the achievements in micelles-based drug delivery for colorectal cancer chemotherapy and gene therapy. Published studies included in this minireview were identified through searching PubMed and Google scholar using different permutations of these keywords "colorectal cancer" or "colon cancer", "chemotherapy" or "gene therapy", "combination", and "micelle". Clinical trials were identified through searching https://clinicaltrials.gov using two keywords "micelle" and "cancer".

COMPOSITION AND CHARACTERISTICS OF POLYMERIC MICELLES

Polymeric micelles are formed by self-assembly of amphiphilic di-block, tri-block or grafted copolymers in aqueous solutions^[23]. Micelles have a two-phase structure including an inner core composed of hydrophobic blocks surrounded by a corona of hydrophilic blocks^[24,25]. The widely used core-forming polymers include polyesters, polyethers, or polyamino acids, while the most used hydrophilic corona-forming polymer is polyethylene glycol (PEG)[26]. Alternatively, other hydrophilic polymers could be used including chitosan, dextran (DEX), and hyaluronic acids (HA)[27]. The hydrophilic corona confers steric stability to the micelle, decreases its interactions with serum components, prevents its recognition and early elimination by the reticuloendothelial system, thus enhancing its retention in the systemic circulation^[28,29]. The hydrophobic core forms a loading site for lipophilic drugs. It was reported that encapsulating drugs in micelles enhanced their water solubility by 10- to 500-fold^[30]. Non-polar drugs can be loaded in micelles by physical entrapment or chemical conjugation[30]. Polymeric micelles used in gene delivery are known as micelleplexes and have cationic properties which allow the association of nucleic acids via electrostatic interactions. The hydrophilic blocks of these micelles are usually composed of polycations including polyethyleneimine (PEI), poly(2-dimethylaminoethyl methacrylate) (PDMAEMA) and polyaminoacids[31].

The concentration of copolymers that is needed to form micelles is known as the critical micelle concentration (CMC). The CMC should be low enough to prevent the

dissociation of the micelles upon dilution in the bloodstream^[32]. The size of polymeric micelles typically ranges from 10 to 100 nm^[33]. Given that nanoparticles that are less than 10 nm can easily undergo renal clearance, and those larger than 100 nm can be rapidly eliminated by the liver and spleen, the suitable size of micelles limits their rapid clearance from the circulation^[34]. In addition, the nano-size of micelles allows them to passively accumulate in tumor tissues by the enhanced permeability and retention effects^[27]. This passive targeting is due to the aberrant and leaky vascular architecture coupled with defective lymphatic drainage that uniquely characterize solid tumors^[35]. Being unable to pass through the normal blood vessel walls, the micelles extravasate through the leaky tumor blood vessels, accumulate preferentially at the tumor site and consequently promote the retention of the delivered drug at the tumor tissue^[36].

The polymeric micelles deliver the drug payload to the cancer cells by two mechanisms. The therapeutic agents can enter the cancer cell as free drugs after their release from the micelles or be internalized as drugs encapsulated within the micelles to be released within the cell^[37]. Chemically conjugated drugs are released by bulk degradation or surface erosion of the micelles while the physically entrapped drugs are mainly released by diffusion^[38]. Intact drug-loaded micelles are internalized *via* cell- dependent pinocytosis which include macropinocytosis, clathrin-mediated endocytosis, caveolae-mediated endocytosis and clathrin- and caveolae-independent endocytosis^[39-42]. Internalized micelles have been shown to localize in acidic organelles, namely endosomes or lysosomes^[40,43]. The loaded drugs were suggested to be released from the micelles in these organelles^[40,43] and to subsequently exert their therapeutic effect at the action site^[40] (Figure 1).

Active targeting can be used to complement passive targeting for enhanced tumor selectivity, improved cellular uptake, and reduced off-target effects^[21,23]. The surface of the micelles can be modified with ligands that can specifically recognize and bind to receptors or proteins overexpressed on the tumor cells but which are less expressed on normal cells^[44]. Different types of ligands have been conjugated to micelles to ensure their active targeting to cancer cells including antibodies and antibody fragments,

proteins including transferrin, peptides such as iRGD, nucleic acid-based ligands such as the AS1411 aptamer, and small molecules including folic acid (FA) and HA[45]. Another interesting approach for active targeting is to achieve adequate drug release at the tumor site by designing stimuli-responsive micelles taking advantage of the unique features of the tumor microenvironment. The tumor tissues are characterized by a low pH, upregulation of specific enzymes and elevated levels of glutathione (GSH) compared with the normal tissues^[34]. Stimuli-sensitive polymeric micelles are developed by introducing pH-sensitive linkers in their structures, such as hydrazone bonds or pH-sensitive ionizable functional groups, enzyme-sensitive moieties, or redoxsensitive linkers including disulfide bonds[34]. When the micelles reach the tumor, the linker degrades in response to the internal stimulus leading to the disassembly of micelles and subsequent drug release^[34]. In addition to internal stimuli, polymeric micelles responsive to external stimuli such as magnetic fields, ultrasound, and light are designed to achieve a temporal and spatial controlled release of the drug in the desired tumor tissues[34]. After releasing the drug, micelles can dissociate into monomers that are excreted by the kidneys, therefore avoiding any long-term adverse effects^[46]. All these properties make polymeric micelles an attractive platform for drug delivery against colorectal cancer.

APPLICATION OF POLYMERIC MICELLES IN COLORECTAL CANCER TREATMENT

The properties and the cellular and molecular mechanisms of action of polymeric micelles used for drug delivery in colorectal cancer treatment are presented in table 1.

Chemotherapy

Numerous studies have demonstrated the potential of polymeric micelles to ensure the targeted delivery of chemotherapeutic drugs to colorectal tumor sites and enhance their therapeutic effectiveness.

Irinotecan is a topoisomerase I inhibitor that is converted by enzymatic activation in the liver and tumors to 7-ethyl-10-hydroxy-camptothecin (SN-38). The latter is about 100-1000 times more potent than IRI against cancer cells[47]. However, only 2-5% of the administered dose of IRI is converted to SN-38 in the clinic^[48]. Although SN-38 demonstrated effective anticancer activity, its direct use as a free drug in the clinic was limited by its hydrophobicity^[49]. To circumvent this problem, Koizumi et al^[50] conjugated SN-38 to PEG-poly(glutamic acid) copolymer with a phenyl ester bond to develop SN-38-loaded polymeric micelles (NK012). The micelle formulation of SN-38 demonstrated extended blood circulation compared with free IRI in mice bearing colorectal cancer xenografts. It also prolonged and improved the distribution of SN-38 in the tumor and produced stronger antitumor activities compared to free IRI. To test the correlation between the efficacy of micellar drugs and tumor hypervascularity and hyperpermeability, the effect of NK012 and free IRI was tested in a vascular endothelial growth factor (VEGF)-secreting tumor model namely SBC-3/VEGF. Interestingly, NK012 was found to improve the accumulation and the antitumor activity of SN-38 in SBC-3/VEGF tumors compared to SBC-3/Neo tumors. This was due to the high vascular density and permeability induced by VEGF. However, treatment with free IRI neither significantly increased SN-38 concentration in SBC-3/VEGF tumors nor effectively suppressed tumor growth^[50].

Doxorubicin (Dox) is an anthracycline antibiotic that is commonly used to treat several types of solid cancers. However, its clinical use is hampered by dose-dependent cardiotoxicity due to its non-specific biodistribution^[51]. Recently, Brunato *et al*^[52] developed methoxyPEG (mPEG)-polyaminoacid based micelles for the delivery and controlled release of Dox in colorectal cancer. The hydrophobic block of the co-polymer included 6 glutamic acid-γ-hydrazide and 10 Leucine. Dox was conjugated to glutamic acid through pH-sensitive hydrazone bond to trigger its release in the acidic lysosomal microenvironment. The release of Dox was faster at pH 5.5 which mimics the lysosomal environment. In addition, the intracellular delivery of Dox by the micelles and its trafficking through the lysosomal compartment of colorectal cancer cells, where the

acidic environment can induce the cleavage of the hydrazone bond, were confirmed by confocal microscopy. Therefore, encapsulating Dox in the formulated micelles may prevent its undesired release in the bloodstream. Interestingly, Dox loaded micelles had good tolerability, produced a higher antitumor effect compared to the free drug and had limited toxicity in colorectal cancer bearing mice^[52].

Docetaxel is a *microtubule inhibitor* that is approved to treat numerous types of cancers. Due to its low water solubility, a mixture of Tween 80 and ethanol is required to solubilize DTX. Yet, Tween 80 used in the available marketed formulation of DTX Taxotere® was associated with adverse effects^[53]. Recently, FA-targeted PEG-polyester micelles were formulated for the targeted delivery of DTX against colon cancer. The targeted formulation resulted in an accelerated internalization of the micelles by FA receptor-positive colon cancer cells *in vitro* and in a higher and preferential accumulation of the micelles in colon tumors *in vivo* compared to the non-targeted formulation. Besides being biocompatible and safe, the FA-targeted DTX-loaded micelles exhibited a remarkable 97% tumor-inhibiting efficiency which was higher than that of non-targeted DTX-loaded micelles (85%) and free DTX (31%) in mice bearing colon tumor xenografts^[54].

Paclitaxel is a microtubule stabilizing agent commonly used against a variety of cancers including colorectal cancer^[55]. Because of its low aqueous solubility, Taxol®, the commercial formulation of PTX uses a nonionic surfactant namely Cremophor® EL^[56]. However, Cremophor® EL has been associated with serious side effects such as hypersensitivity, nephrotoxicity, and neurotoxicity^[57]. Zhu *et al*^[58] developed smart micelles for the targeted delivery and controlled release of PTX in colon cancer. Micelles are made from the self-assembly of D-α-tocopherol succinate (TOS) conjugated to HA *via* a redox responsive disulfide bond (HA-SS-TOS). Their findings demonstrated that these micelles could specifically accumulate in orthotopic colon tumors and metastatic tumor cells rather than in normal intestinal tissue through binding of HA to CD44 receptors overexpressed on primary and metastatic colon tumor cells. They could also selectively release PTX in orthotopic colon tumors and metastatic tumor cells where the

high levels of GSH break disulfide bonds thus causing drug release. PTX-encapsulated micelles induced the highest survival rate (100%) in mice bearing orthotopic colon tumors compared to untreated group (60%) and free PTX (Taxol) treated group (40%). They also suppressed primary colon tumors and metastatic sites in the intestine and decreased metastasis to other organs and to the peritoneum^[58].

Gene therapy

Polymeric micelles were also reported to successfully deliver anticancer genetic material to colorectal cancer. Cationic hybrid micelles composed of amphiphilic cationic lipid N-[1-(2,3-dioleoyloxy) propyl]-N, N, N-trimethylammonium methyl sulfate (DOTAP) and mPEG-poly (ε -caprolactone) (PCL) have been used to deliver genes or small interfering RNA (siRNA) for the treatment of colon cancer in preclinical studies. The cationic hydrophilic head of DOTAP ensures the adsorption of the anionic nucleic acid onto the surface of the micelles via electrostatic interaction. These hybrid micelles demonstrated an effective and safe delivery of siRNA targeting colon cancer cells. The micelle-based delivery of siRNA caused inhibition of the two anti-apoptotic genes Bcl-xl and Mcl1 which consequently induced apoptosis in colon cancer cells in vitro and suppressed the growth of colon tumors in vivo[59]. In another report, these hybrid micelles were used for the delivery of interleukin-22 binding protein (IL-22BP) gene which is known to play a role in blocking IL-22/IL-22R1 signal axis involved in tumor growth and metastasis. The micelles-delivered IL-22BP gene was efficiently expressed in colon tumors in vivo leading to the inhibition of the growth of abdominal cavity metastases. Its anticancer mechanism involved blockage of IL-22, induction of apoptosis and anticancer immune response in addition to inhibition of angiogenesis^[60]. In a study conducted by Duan et al^[61], these hybrid micelles were found to efficiently deliver the classic suicide gene Survivin-T34A that acts as a competitor in the survivin pathway involved in apoptosis inhibition^[62]. The Survivin-T34A gene loaded micelles inhibited the proliferation of colon cancer cells in vitro and the growth of abdominal metastatic colon cancer and malignant ascites in vivo by inducing apoptosis[61].

Importantly, in all these studies, this micellar carrier did not induce any pathological changes in the vital organs of the mice with colon cancer.

Combination of chemotherapy with gene therapy

The micellar dual delivery of chemotherapeutic drugs with agents targeted against specific genes have been shown to produce synergistic therapeutic responses against colorectal cancer. Cationic micelles based on PEI-deoxycholic acid conjugates have been used for the co-delivery of PTX and siRNA targeting the X-linked inhibitor of apoptosis (XIAP) gene to colorectal cancer. The micellar co-delivery of XIAP siRNA and PTX effectively decreased the expression of XIAP at the mRNA and protein levels in vitro and in vivo. In addition, it was more potent than PTX loaded micelles as it caused a greater reduction in cell viability of colorectal cancer cells and inhibited the growth of colorectal tumors in vivo [63]. In another study, Sanati et al[64] synthesized cationic micelles for the targeted dual delivery of Camptothecin (CPN), a topoisomerase inhibitor, and short hairpin RNA targeting survivin (surshRNA) to colon cancer. These micelles containing PEI-poly (DL-lactic acid) copolymers were coated with poly carboxylic acid dextran and were subsequently modified with AS1411 aptamer that has high affinity for the nucleolin receptors expressed on the surface of colon cancer cells. Cell surface nucleolin expression is known to be more elevated in cancer cells than in normal cells^[65]. The conjugation of AS1411 aptamer on the micelles improved their cellular internalization through nucleolin-mediated endocytosis in vitro and enhanced their accumulation in colon tumors in vivo in comparison with their non-targeted counterparts. While the non-targeted micellar formulation of each of CPN and surshRNA exhibited 3% and 5% tumor inhibitory effect in vivo, the targeted and nontargeted formulations of the combination of CPN and surshRNA exerted 93% and 87% tumor inhibitory effects, respectively. It is important to note that the free CPN did not have any anticancer effects in vivo, and its micellar formulation prevented the development of its associated systemic toxicity^[64]. Moreover, Lee et al^[66] designed theranostic micelles from cationic PDMA-b-PCL and mPEG-PCL polymers for the

combined delivery of SN-38, siRNA targeting human VEGF in addition to a magnetic resonance imaging (MRI) contrast agent namely ultra-small superparamagnetic iron oxide nanoparticles (USPIO). The SN-38/USPIO/siRNA loaded mixed micelles were targeted passively to colorectal tumor regions *in vivo*. They did not only induce a synergistic tumor growth suppression but also allowed the tracking of anticancer effects by MRI during the treatment^[66]. Additionally, hybrid micelles co-self-assembled from PEI-poly (D,L lactide) and 1,2-distearoyl-sn-glycero-3-phosphoethanolamine-N-mPEG were used to deliver IRI and a tumor suppressive microRNA-34a (miR-34a) to colon cancer. The micellar co-delivery of the two therapeutic agents showed higher efficacy in inducing apoptosis and inhibiting migration of colon cancer cells *in vitro* compared with free IRI and the single agent formulation. The micellar IRI and miR-34a combination also accumulated in colon tumors *in vivo* and showed superior tumor growth inhibition with low acute systemic toxicity compared with free IRI and the micellar formulation of individual agents^[67].

Combination cancer chemotherapy

For dual combination therapy, the encapsulation of drugs in a micellar system could be performed in two ways (Figure 2). In the first method, both drugs are physically loaded in the micelles by hydrophobic interactions. Alternatively, one hydrophobic drug is conjugated to the amphiphilic co-polymer which self-assembles and forms prodrug micelles to subsequently encapsulate the second hydrophobic drug in its core^[68].

The chemotherapeutic drug Dox and the tyrosine kinase inhibitor dasatinib were effectively co- encapsulated in the inner hydrophobic core of micelles formed by PEG-peptidic conjugate namely PEG-lysyl-(α-fluorenylmethyloxycarbonyl-ε-Cbz-lysine)2. The dual drugs loaded micelles exhibited greater inhibition of colon cancer cell proliferation compared with the micellar formulation or the free form of individual drugs^[69]. In another study, Dox was co-delivered with the tumor necrosis factor-related apoptosis-inducing ligand (TRAIL) in cationic micelles to colorectal cancer. The micelles were prepared with poly {(*N*-methyldietheneamine sebacate)-*co*-[(cholesteryl

oxocarbonylamido ethyl) methyl bis(ethylene) ammonium bromide] sebacate. Dox was encapsulated in the micelles whereas TRAIL was complexed onto their surface. The internalization of these micelles by colorectal cancer cells was mediated by the death receptors expressed on the cell surface. Compared to the formulation of single agents, Dox and TRAIL loaded micelles enhanced the inhibition of colorectal cancer cell viability, the increase of sub-G1 population and the reduction of long-term survival and clonogenicity of cancer cells after the treatment. Interestingly, these remarkable anticancer effects were reported in both TRAIL sensitive and resistant colorectal cancer cells. In addition, the cytotoxic effect of the co-loaded micelles was selectively higher against cancer cells than normal cells. This was suggested to be due to the relative disparities in expression levels of decoy and death receptors between normal and cancer cells[70]. A third study reported the combined delivery of Dox and the proapoptotic cationic peptide KLA using DNA micelles. These micelles were made using cholesteryl-modified single strand DNA and its complementary sequence to which the KLA peptide was conjugated. The cationic peptides enhanced the stability of the micelles by protecting them from digestion by nucleases. Since oligonucleotides cannot cross cell membranes, DNA micelles were conjugated to a mucine1 (MUC1) aptamer to target MUC1 receptors overexpressed on the surface of cancer cells. These micelles were found to specifically bind to the MUC1 overexpressing cells and to accumulate within them. The co-delivery of KLA and DOX using the targeted micelles potentiated the inhibition of colon tumor growth in vivo compared to free Dox in addition to attenuating its toxic effects^[71]. A recent study reported the formulation of a folate receptor-targeted and redox-responsive micellar system for the co-delivery of PTX and adjudin (ADD), a mitochondrial inhibitor, to reverse multidrug resistance in colon cancer. In this drug delivery system, PTX was conjugated to DEX via a disulfide bond to form the PTX prodrug and FA was conjugated to the side chain of DEX-PTX. The resulting PTX prodrug micelles were further used to encapsulate ADD in its hydrophobic core. In vitro studies showed that conjugation of FA to the surface of these micelles enhanced their internalization by PTX resistant colorectal cancer cells

overexpressing the FA-receptor. They also demonstrated that ADD inhibited PTX efflux which consequently improved intracellular accumulation of PTX and therefore its cytotoxic activity. In addition, PTX and ADD loaded micelles exhibited a prolonged circulation in the blood compared with free PTX in vivo. They also increased the concentrations of drugs in resistant colon tumors compared to their non-targeted counterparts and free PTX. Importantly, they demonstrated a strong tumor growth inhibition owing to the active tumor-targeting, the co-delivery of two therapeutic drugs, and GSH-responsive drug release^[72]. In another study, micelles engineered with polylactic-co-glycolic acid grafted branched PEI were used to co-deliver 5-FU and methotrexate (MTX) which is an antagonist of FA and is known to have anticancer effects against different types of neoplasms. MTX was conjugated to the copolymer forming the prodrug micelles which was further used to encapsulate 5-FU. In addition to its therapeutic potential, MTX has structural similarity with FA and can therefore act as a FA receptor targeting agent. The conjugation of MTX to the micelles improved their internalization by colon cancer cells compared to the non-conjugated micelles. The combined drugs loaded micelles were more effective in attenuating the viability of colon cancer cells compared to free 5-FU or the micellar MTX^[73]. Doxifluridine, a prodrug of 5-FU, has been commonly used in cancer treatment to circumvent the toxicity of 5-FU. Doxifluridine is converted into toxic 5-FU by endogenous thymidine phosphorylase in cancer cells^[74]. In a study conducted by Wang et $al^{[75]}$, doxifluridine was conjugated to PCL to form a hydrophobic segment which was grafted with hydrophilic chitosan. The produced polymeric prodrugs self-assembled into micelles used for SN-38 encapsulation. The dually loaded micelles were found to reduce the viability of colon cancer cells in vitro to a greater extent than doxifluridine loaded micelles. In another study, doxifluridine-PCL hydrophobic segment was grafted with mPEG to form prodrug micelles serving as carriers to further encapsulate Dox or SN-38. The micellar co-delivery of doxifluridine with Dox or SN-38 exhibited superior inhibition of colorectal cancer cell viability with respect to doxifluridine loaded micelles^[74].

Polymeric micelles in clinical trials

Despite their promising potential in preclinical studies against colon cancer, few polymeric micelles have reached the clinical setting. A phase I clinical trial of NK012 was conducted in Japan with a total of 24 patients with solid tumors including 12 patients with colorectal cancer. NK012 was found to be well tolerated at doses up to 28 mg/m² when administered every 3 wk. None of the colorectal cancer patients had a partial response. However, five colorectal cancer patients with a history of treatment with OXA and IRI had stable disease; four of them received the NK012 regimen for six cycles of treatment or more^[76]. Subsequently, the efficacy and safety of NK012 was evaluated in a phase II clinical trial at an initial dose of 28 mg/m² in 58 Japanese patients with unresectable metastatic colorectal cancer who had been treated with OXAbased chemotherapy. Administration of NK012 resulted in a response rate similar to that of free IRI. In addition, febrile neutropenia and grade ≥ 3 neutropenia were reported in these patients suggesting that an initial dose of 28 mg/m² of NK012 may be too high. Further studies should be conducted to determine the optimal dose of NK012 and improve its efficacy and safety[77]. In another study, a phase I clinical trial of NK105, PTX loaded micelles, was conducted with 19 patients with advanced solid tumors including one patient with colon cancer. NK105 was well tolerated, and the recommended phase II dose was determined to be 150 mg/m² every 3 wk. The colon cancer patient was reported to have a stable disease for longer than 4 wk at the time of the completion of the study^[78]. Two additional clinical trials on polymeric micelles in colorectal cancer were launched but their results have not been reported yet. The first is a phase I clinical trial that aims to determine the maximum tolerated dose/recommended phase II dose of the combination of NK012 with 5-FU and leucovorin in patients with colorectal cancer (NCT01238939)[79]. The second is a phase II clinical trial that aims to investigate the safety and the diagnostic performance of ONM-100, a polymer micelle covalently conjugated to an intraoperative fluorescence imaging agent indocyanine green, for the detection of cancer in patients with solid

tumors including colorectal cancer patients undergoing routine surgery (NCT03735680)[80].

CONCLUSION

Polymeric micelles are potential candidates for delivery of chemotherapeutic drugs and therapeutic genetic material in single or combination treatment against colorectal cancer. Polymeric micelles exhibit a safe profile, help in solubilizing hydrophobic chemotherapeutic drugs, and achieve prolonged, and stabilized drug circulation in the blood. They also promote the passive accumulation of drugs at colorectal cancer sites, potentiate their anticancer activity and reduce their unwanted side effects. Conjugating polymeric micelles with stimuli-responsive and tumor targeting moieties further increases their internalization by cancer cells, thus achieving selective drug release at colorectal tumor sites and enhancing drug anticancer effects. In addition, polymeric micelles ensure an effective and safe delivery of genetic material to modulate the expression of apoptotic players in colorectal cancer. Co-delivering genetic anticancer agents with chemotherapeutic drugs using micellar systems is known to augment the therapeutic outcome of the latter and therefore represent a promising combination strategy for combating colorectal cancer. This nanocarrier system also allows the incorporation of imaging agents with therapeutics and offers great potential to achieve tumor diagnosis, in addition to tracking the delivery and release of the therapeutic agent and evaluating its therapeutic performance. Although most of the studies evaluating polymeric micelles-mediated combination chemotherapy in colorectal cancer were performed in vitro, they yielded interesting results. The delivery of drug combinations using polymeric micelles could further intensify their inhibitory effects against colorectal cancer and help overcome drug resistance. However, further pharmacokinetics and pharmacodynamics studies are needed prior to the clinical translation of drug loaded polymeric micelles. Few polymeric micelles have been used in the clinical setting for colorectal cancer treatment and diagnosis with promising outcomes, thus their application merits further investigation.

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