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Adiponectin as a therapeutic target for diabetic foot ulcer

Abdalla MMI *et al.* Adiponectin and DFUs

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INTRODUCTION

Diabetic foot ulcers (DFUs), a serious complication of diabetes mellitus, pose a significant economic burden on patients and healthcare systems worldwide. The development of these ulcers is often due to poor foot care, inadequate glycaemic control, underlying neuropathy, and peripheral vascular disease. Left untreated, these ulcers can result in amputations. The global prevalence of DFUs ranges from 3% in Oceania to 13% in North America, with a global average of 6.4%^[1].

Healing time for these ulcers can take up to 12 mo, with a recurrence rate estimated to be 65% within 5 years^[2]. Studies have shown that the impact of DFUs on individuals is profound, with loss of ambulatory function, financial strain, and emotional suffering being common outcomes^[3]. The economic impact on patients and their families due to medical bills, loss of income, and emotional distress can be significant. Participants in a recent study reported experiencing depression, isolation, and hurtful comments from others^[3].

DFUs continue to pose a significant public health challenge, and they are a major cause of morbidity and mortality worldwide^[4].

Adiponectin, a fat-derived hormone, has been shown to protect against insulin resistance, type 2 diabetes (T2DM), and atherosclerosis. Reduced circulating levels of adiponectin are thought to play a role in the development of T2DM. In cases of obesity, the production of endogenous adiponectin is impaired. It is, therefore, suggested that pharmacological or dietary interventions be considered to restore the capacity of adipose tissue to secrete adiponectin^[5].

DIABETIC FOOT SYNDROME

Diabetes mellitus (DM), is a main cause of death and poor quality of life worldwide, affecting 463 million individuals in 2019 and is estimated to reach 700 million by 2045^[2]. People with diabetes often have foot problems that impose economic burden on the individual, and about half of all foot amputations are observed to be among diabetic. The lifetime chance of a diabetic having a foot ulcer is as high as 25%^[3], and it is estimated that every 30 s a lower limb is lost due to diabetes somewhere in the globe^[4]. DFUs can be prevented by ensuring that diabetics get regular foot exams and treating any neuropathy that may be present^[5]. The International Diabetes Foundation has called for greater awareness of diabetes foot concerns due to the psychological, social, medical, and economic effects of what should be one of the most preventable long-term complications of diabetes^[6,7]. In most Western nations, the yearly incidence of DFUs is roughly 2%, however greater rates have been observed in select populations, including medicare recipients (6%) and United States veterans (5%)^[8]. The projected annual cost to the NHS in the United Kingdom is around 580 million, with 307 million spent on ulceration in primary care^[9].

Diabetic foot syndrome is defined, according to the World health Organization, as “ulceration of the foot (distally from the ankle and including the ankle) associated with neuropathy and different grades of ischemia and infection”^[10]. It is a significant, long-term consequence of diabetes that can result in amputations, disability, and diminished quality of life.

Since peripheral neuropathy and vascular disease are present in more than 10% of individuals at the time of diagnosis of T2DM^[11] and because the first year following the diagnosis of diabetes is a risky time for foot ulcers and amputations, the burden of diabetic foot disease is expected to rise in the future^[12]. Furthermore, emerging nations in Africa, Asia, and South America, where foot ulcers are more likely to have neuropathic origins^[13] and are thus very avoidable, are anticipated to have the biggest growth in the prevalence of T2DM^[6]. Deploying screening, education, and treatment

programmes most effectively around the globe is still the dilemma facing the worldwide diabetes community^[14].

The simplest definition of diabetic foot infection is “any infra-malleolar infection in a diabetic patient”. These include paronychia, cellulitis, myositis, abscesses, necrotizing fasciitis, septic arthritis, tendonitis, and osteomyelitis. DFUs are complex and rarely caused by a single condition. Several risk factors cause DFUs^[15,16]. Understanding the pathobiology helps diagnose and treat DFUs, which is one of the leading indicators for amputations.

Neuropathy is the primary contributing factor leading to ulceration, in diabetics. Diabetic peripheral neuropathy (DPN) is a disruption of normal nerve function that can change autonomic, motor, and sensory functioning throughout the body^[17]. Due to the absence of protective sense in patients with sensory neuropathy, the foot is more likely to sustain untreated minor injuries as a result of excessive pressure as well as mechanical or thermal damage. Individuals with diabetes who also have sensory neuropathy were found to be at the highest risk for developing ulcers, as revealed by a significant prospective multicentred investigation^[18].

There are various types of neuropathies, and some of them may cause foot ulcers. Motor neuropathy may lead to foot deformities, decreased joint mobility, and abnormal foot loading. These modifications may cause a shift in the distribution of loads that are experienced when walking, with a subsequent reactive thickening of the skin known as callus at unusual load areas. Ischaemic necrosis of the tissues underneath the callus also contributes to the development of a neuropathic ulcer. Autonomic neuropathy often results in changes to the skin's texture and turgor, such as dryness and fissuring, which makes the skin more susceptible to infection since it provides an entry site to the bacteria^[19].

Another condition that contributes to the development of foot ulcers is peripheral vascular disease, which affects both small and major blood vessels. It is possible for both macrovascular and microvascular diseases to contribute to the symptoms of peripheral vascular disease, which ultimately results in the delay in wound healing. In

both diabetics and non-diabetics, there is an increase in the incidence and prevalence of peripheral arterial disease with age, while the condition is worse with diabetes. Individuals who have diabetes are at an increased risk for vascular disease because of the prevalence of risk factors such as hypertension, smoking, and hyperlipidaemia^[20,21].

The ulcerated diabetic foot is the result of a complex interaction between several factors, including neuropathy, peripheral vascular disease, trauma, and infections. Neuropathy and ischaemia, also called neuro ischaemia, are the initial mechanisms, while infection is typically a result of this condition. Studies have indicated that diabetics acquire peripheral vascular disease at a younger age more frequently than others in the same age group^[22]. In 35% of cases, proximal arterial disease-related peripheral ischaemia was cited as an important cause of ulceration among diabetics in a two-center study of causal pathways^[22]. In another study that compared diabetic patients with peripheral artery disease to non-diabetic patients with the same condition, it was found that diabetic patients had more distal disease and a worse prognosis in terms of amputation and mortality^[23].

Hence the pathogenesis of DFUs, a complication of longstanding uncontrolled diabetes, involves multifactorial influences such as neuropathy, peripheral vascular disease, foot deformity, trauma, infection, and inadequate glycaemic control. The loss of sensation brought on by neuropathy can result in repeated damage to the foot, while peripheral vascular disease can reduce blood flow and slow healing. Injuries and infections can exacerbate already-existing ulcers, while foot abnormalities can create pressure points and raise the risk of skin deterioration. Moreover, poor glycaemic management might hinder wound healing and raise the danger of infection. Thus, for the prevention and management of DFUs, a multidisciplinary strategy that takes these aspects into account is essential.

RISK FACTORS

Multiple variables contribute to the emergence of DFUs. Peripheral neuropathy and ischaemia that result from peripheral vascular disease that reduces the protective

components of the tissues are the primary underlying causes. In addition, the skin can be subjected to stress, such as pressure, shear, or trauma, which also contributes to the condition. Antonio et.al. in their study identified general and local factors predisposing to the development of DFUs^[24]. The general factors include duration and severity of diabetes, associated comorbidities such as hypertension, dyslipidaemia, chronic renal disease, peripheral vascular disease and age while the local factors included foot deformity, trauma, callus presence, previous amputation, impaired joint mobility and shoe defects^[25].

DIAGNOSIS

Individuals who have diabetes are required to have their neurological, vascular, dermatological, and musculoskeletal conditions evaluated on a yearly basis, at the very least. The American Diabetes Association (ADA) developed a comprehensive foot examination and risk assessment tool that is fast and require very little specialised medical equipment^[26]. Patients who come in exhibiting tissue loss are placed in a higher risk category than those who do not. In situations like these, an evaluation of the overall level of limb threat should be performed.

Many measures exist to assess the severity of a diabetic ulcer by analysing the ulcer's features, ischaemia, and infection. Wagner, University of Texas, and PEDIS are the most widely used and globally recognised scales^[27,28]. These scales have demonstrated their utility correlating the degree of severity of the ulcers with the risk of amputation^[29]. The wound scales are a valuable tool for classifying the severity of DFUs, but they should not be used to determine the need for amputation. The microbiology of wounds should be examined in each region to further determine the appropriate empiric therapy in the management of DFU.

COMPLICATIONS

DFUs are the major cause of hospitalisation and amputation in diabetes patients^[5,25]. Foot ulcer complications include excruciating pain, infection, gangrene, osteomyelitis,

amputation, and death^[30]. Coexisting diabetes-related problems, such as diminished peripheral sensations and absence of pain along with this sustained ambulation further incites additional damage^[31].

Studies demonstrated a higher death rate in diabetic patients with DFUs, with a death rate almost double that of diabetic patients without foot ulcers^[22,32]. DFUs have been also reported to be associated with a greater frequency of major cardiovascular risk factors, subclinical signs of past and new-onset cardiovascular and cerebrovascular events^[33].

TREATMENT

Current treatment emphasises patient education, regular foot self-examinations, and annual diabetic foot evaluations. These annual examinations comprise patient history, peripheral vascular exam, and sensory nerve function evaluation to detect DPN early. Pressure analysis studies on lowering foot pressure or changing gait offer promising technology for early detection and prevention of DFU^[34,35]. Depending on DFU categorization, DFU patients need unloading, infection or ischaemia treatment, wound debridement, and wound dressings^[36]. Tissue volume and type are often used to classify DFUs^[37]. Granulation tissue is red/pink and symbolises healing tissue, whereas slough tissue is more yellow and represents infected tissue, and necrotic tissue is dark/black and shows tissue death. Many studies show that DFU diagnosis and treatment can greatly reduce or prevent serious consequences^[37,38]. Despite national and international guidelines, DFU administration varies. Under this ambit, patients suffering from DFUs need reliable and quick therapy, which can only be facilitated with deeper understanding of the metabolic marker of DFU such as advanced glycated end-products (AGE's), inflammatory markers, lipid profile, while newer markers such as adiponectin as a prospective diagnostic tool needs to be further explored. Emerging technologies such as bioprinting and electrospinning^[39], stem and somatic cell monotherapy^[40] and grafting techniques^[41] offer promising alternatives by overcoming the limitation in conventional approaches.

ADIPONECTIN

Adipose tissue produces adipokines, which are peptides that communicate with other tissues such as the brain, liver, pancreas, immune system, vasculature, and muscle about their functional state. Thus, adipose tissue dysfunction is often related with alterations in the secretion of adipokines such as leptin, adiponectin, fibroblast growth factor 21 (FGF21), retinol-binding protein 4, dipeptidyl peptidase 4, bone morphogenetic protein (BMP)-4, BMP-7, vaspin, apelin, and progranulin. Although the complete repertoire of human adipokines has not yet been described, it has been established that adipose tissue is a reservoir for more than 600 secretory proteins^[42].

Adipokines control many physiological processes, including appetite and fullness, fat distribution, insulin secretion and sensitivity, energy expenditure, endothelial function, inflammation, blood pressure, and blood clotting^[43,44]. As the mRNA transcript for adipokines was most robustly expressed in adipocytes, adiponectin was first discovered in mice shortly after leptin's discovery in 1995^[45]. Two different adiponectin receptors, ADIPOR1 and ADIPOR2, are responsible for relaying signals from the 30-kilodalton, 244-amino-acid protein, adiponectin, to its target cells^[45].

Adiponectin undergoes post-translational modifications that lead to the secretion of oligomers of 90-kDa trimers, which are subsequently detected in the bloodstream as 180-kDa hexamers (low molecular weight)^[45,46]. Adiponectin structure consists of trimers, hexamers, and higher order complexes that can be formed in the collagen domain of adiponectin before secretion^[47,48].

ADIPONECTIN RECEPTORS

Many different receptors, including adiponectin receptors 1 and 2, play roles in mediating adiponectin's effects^[49]. These receptors are functionally dissimilar from G-protein-coupled receptors, primarily due to the fact that their polarity is in the opposite direction. It is projected that they include seven transmembrane sections. large level of functional redundancy appears to exist between the adiponectin receptors, as suggested

by both ² single- and double-knockout mice for the receptors^[50]. Although the ²⁹ relative ratios of ADIPOR1 and ADIPOR2 expression in different tissues may differ, in general, both are expressed in a very high proportion of tissues. T-cadherin is the name given to a newly discovered molecule that may be found on the cell surface and possesses a considerable affinity for the protein adiponectin^[51]. It is not technically a signalling receptor since it does not have an intracellular signalling domain, even though it is capable of binding adiponectin. T-cadherin is necessary, however, in order for adiponectin to reach its full potential in terms of its cardioprotective effects^[52].

SECRETION AND RELEASE

Adiponectin is a secretory protein that is only produced by adipocytes. Constitutively synthesized, it accounts for 0.01%-0.05% of plasma protein, which places it in the range of 2-20 g/mL and makes it a component of plasma that is reasonably abundant. Adiponectin is a protein that is fairly stable in circulation, despite the fact that its plasma half-life is only 45-75 min^[53]. Other cell types, such as beta cells in the pancreas and certain cell types in the heart and kidneys, also have a strong affinity for adiponectin and can bind to it. Adiponectin is primarily removed from the bloodstream in the liver, making it an important organ in this process^[53]. In spite of the fact that ² adiponectin is secreted by adipose tissue, circulating levels mysteriously decrease when there is an increase in the amount of central adiposity^[54]. Despite this, greater degrees of adiposity in the lower extremities and the truncal region are associated with greater concentrations of adiponectin.

Adiponectin's insulin-sensitizing, anti-inflammatory, and antiapoptotic effects have generated considerable interest^[45,55]. In addition, numerous cohort studies in various groups have shown that adiponectin levels are inversely related to either ²² the presence of glucose intolerance or the risk of developing T2DM^[56].

²² EFFECTS OF ADIPONECTIN

On Beta-cell function

The beta cells of pancreatic islets express both ADIPOR1 and ADIPOR2, the two receptors for adiponectin^[57,58]. Recombinant adiponectin given to adiponectin-deficient mice shows that it targets beta cells. Adiponectin may enhance glucose-mediated insulin production and promote insulin and related gene transcription^[59], however the effect of adiponectin on insulin release in individuals with normal insulin sensitivity is not well-established^[57,60].

On cardiac and renal function

The strong and long-standing correlation between adiponectin levels and the development of cardiovascular disease has been well-documented. Pischon *et al*^[61] found in a large cohort that men with high plasma adiponectin levels had a lower risk of myocardial infarction. In preclinical ischaemia/reperfusion trials, the Walsh group showed that recombinant adiponectin strongly improves cardiomyocyte survival^[62]. However, why end-stage cardiovascular disease has a significant positive correlation between mortality and high adiponectin levels, unlike early stages, is unknown^[63].

A similar scenario exists in the kidney, where low adiponectin levels correlate with albuminuria in both animals and humans^[64]. In animal models with adiponectin gene knockout, the lack of adiponectin has been linked to increased podocyte damage and albuminuria, and adiponectin therapy has demonstrated the ability to reverse certain renal dysfunction^[65]. In patients with chronic kidney disease, adiponectin levels are positively correlated with proteinuria^[31]. This upregulation is similar to that seen in cardiovascular disease, particularly end-stage cardiovascular disease. The mechanisms are unknown. This is especially challenging given that adiponectin is not cleared through the kidney except in cases of severe proteinuria. This is especially challenging given that adiponectin is not cleared through the kidney except in cases of severe proteinuria^[66], making it difficult to determine which mechanisms are responsible.

On insulin sensitivity

Skeletal muscle is an important factor in insulin sensitivity because it is the primary source of glucose for the body as a whole. It should not come as a surprise, consequently, that a substantial amount of attention has been paid to the potential metabolic effects that adiponectin has on this tissue. High-molecular-weight adiponectin correlated better with systemic insulin sensitivity than low-molecular weight in rodents and humans^[45,46]. Skeletal muscle has a high concentration of ADIPOR1, through which adiponectin regulates energy metabolism^[67]. Most investigations into the effects of adiponectin have focused on its binding to globular adiponectin, which exhibits greater binding strength and biological activity in skeletal muscle compared to most other tissues^[68,69]. Adiponectin binding leads to increased glucose uptake and nonoxidative glycolysis, while simultaneously reducing intramyocellular triacylglycerol content and enhancing fatty acid oxidation^[68,69]. Additionally, adiponectin influences the number of mitochondria and the types of oxidative fibers present in skeletal muscle^[70]. However, in diseased states, the effects of adiponectin on skeletal muscle are attenuated.

The liver is affected in a number of different ways by adiponectin. One of the most notable effects is hepatic glucose production inhibition, which lowers body glucose levels. Hepatocytes are insulin-sensitive at physiological adiponectin levels. As a result, glucose production is significantly inhibited in response to any given dose of insulin^[50]. Adiponectin inhibits both the expression^[68,71] and activity of important regulators in the process of gluconeogenesis^[71,72]. Studies using murine euglycemic clamps have shown that the rates of glucose disposal, glycolysis, and glycogen synthesis are not affected by the presence of intravenous adiponectin infusion^[72]. This suggests that the primary mechanism by which adiponectin lowers blood sugar levels is through the suppression of hepatic glucose output, rather than through enhancing glucose disposal.

On adipose tissue

The adiponectin receptors (ADIPOR1 > R2) are also reported to be expressed by adipocytes. This data further implies that adiponectin may alter adipose tissue function locally, either with modifying autocrine or paracrine function.

As anti-inflammatory effector

Adiponectin's impact on inflammation is not limited to adipose tissue, and its ⁶⁵anti-inflammatory effects have been ⁴⁰observed in other contexts. This is significant because systemic inflammation is thought to play a role in the development of insulin resistance^[73]. These researchers have shown that adiponectin can inhibit the development ²and proliferation of bone marrow-derived granulocyte and macrophage progenitors, but it does ²⁹not have this effect on other haematopoietic cell lines. In addition, it is also reported that inflammatory processes in macrophages can be disrupted, by suppressing the phagocytic activity in human macrophages that have been ²treated with adiponectin^[73], as is the production of pro-inflammatory cytokines^[73]. In the setting of the development of atherosclerosis, adiponectin is shown to limit the transition of macrophages into lipid-laden foam cells^[74].

On other tissues

Adiponectin works in the brain to increase the amount of energy that is expended, which might lead to weight reduction^[46]. In clinical research, circulating adiponectin has been ³shown to have an independent and unfavourable relationship with components of the metabolic syndrome. These components include insulin resistance, body weight, blood pressure, and serum lipids^[43,55].

Adiponectin's molecular functions imply ³that the molecule or agonists of its receptors might cure obesity and related comorbidities^[45]. Studies showed that adiponectin ²⁸improved insulin sensitivity, glucose metabolism, insulin secretion, and body weight in rodent models^[75]. Recently, it was shown that a synthetic small molecule adiponectin receptor agonist, known as "AdipoRon", greatly increased insulin sensitivity and decreased glucose intolerance in rats^[76]. AdipoRon treatment prolonged the lives of

high-fat-fed db/db mice, adding support to the idea that higher blood adiponectin levels are associated with a later average age of mortality in obese people^[76]. Levels of adiponectin have been shown to have a negative correlation with obesity, visceral fat, T2DM, and other complications that are associated with obesity^[45,55]. Not only adiponectin has a promising and readily detectable stable marker for a variety of illnesses, but it also has the potential to play a big role in the future of clinical importance as a therapeutic agent. This is because adiponectin has the ability to regulate fat storage^[44].

The potential therapeutic role of adiponectin in DFUs

Many studies have shown that diabetic patients have lower adiponectin levels than healthy controls^[77-79]. Adiponectin deficiency has been correlated with an increased susceptibility to diabetes and its associated complications, such as DFUs^[80,81]. Some investigations have also indicated that low adiponectin levels may be a potential biomarker of poor wound healing and increased amputation risk in diabetic foot ulcer patients^[78,81,82]. Figure 2 depicts the probable mechanisms *via* which reduced adiponectin levels may contribute to the development of DFUs. Nonetheless, more research is needed to completely understand the role of adiponectin in the pathophysiology and therapy of DFUs.

ADIPONECTIN AND KERATINOCYTES

Diabetic patients often experience impaired wound healing due to continuous hyperglycaemia, leading to alterations in various stages of the healing process such as haemostasis, inflammation, proliferation, and remodelling. Factors that contribute to this include hypercoagulability, impairment of skin function^[83], imbalanced inflammatory and growth factors^[84], reduced neutrophil function^[85], and insufficient wound re-epithelialization^[86]. Adipose tissue has recently been recognized as a key endocrine organ with a role in wound healing. This role is due to the secretion of bioactive substances known as adipokines, which regulate paracrine signaling^[87]. There

is evidence from numerous research that adipose tissue contributes to the healing of wounds^[88-90]. Despite the current understanding, the exact role of adipocytes in the wound healing process remains unknown. However, it has been demonstrated that applying ⁵⁸ adipose tissue extracts over skin wounds can improve wound repair^[91]. The healing process is believed to take place *via* paracrine signaling, highlighting the significance of the adipokines released by adipose tissue in wound healing^[92].

Adiponectin, that is secreted from adipocytes has been found to aid in wound healing through its effects on keratinocytes, the most abundant cellular component of the epidermis^[93]. Adiponectin promotes keratinocyte proliferation and migration, which is crucial for proper re-epithelialization and wound closure. This is mediated *via* the AdipR1/AdipR2 and ERK signaling pathways^[94]. Adiponectin also elevates the ³⁸ intracellular and reconstructed epidermal lipid content of keratinocytes, and regulates ³⁸ the expression of lipid biosynthesis enzymes and nuclear hormone receptors, which helps maintain skin barrier integrity, an action that is mediated through SIRT1 signaling molecule (SIRT1)^[95].

Furthermore, adiponectin possesses reactive oxygen species (ROS)-scavenging abilities and can mitigate oxidative stress-induced DNA damage while regulating antimicrobial peptide production in senescent keratinocytes^[96-98]. Studies have shown that adiponectin can reverse premature cellular aging in keratinocytes and restore normal antimicrobial peptide levels by activating AMP-activated protein kinase (AMPK), increasing SIRT1 deacetylation, recovering FoxO1 and FoxO3 transcription activity, and suppressing NF- κ B p65, thereby preventing abnormal expression of human β -defensin 2 induced by hydrogen peroxide^[99]. Additionally, it restores filaggrin expression and normalizes keratinocyte activity, which is crucial for maintaining skin integrity as an immune barrier^[100,101]. Therefore, one way in which adiponectin may promote DFU healing is through its impact on skin integrity, keratinocyte proliferation, and migration. However, further ¹ research is necessary to fully understand the potential mechanisms of adiponectin in DFU healing.

ADIPONECTIN AND EXTRACELLULAR MATRIX REMODELLING

The extracellular matrix (ECM) that is produced, assembled, and remodelled by fibroblasts is crucial for maintaining skin integrity, but when it is damaged, as in skin ulcers, it undergoes repair and remodelling. Matrix metalloproteinases (MMPs), a family of proteins that includes collagenases and gelatinases, are ECM enzymes that break down damaged fibrils during ECM remodelling^[102]. Normal ECM remodelling includes a delicate balance of ECM breakdown, generation, and maturation. In poor wound healing, such as DFU, the process of ECM remodelling tends to yield more degraded, non-soluble fibrils, resulting in a disorderly ECM network and callus formation^[103,104].

Abnormal expression of MMPs and differential expression of ECM contribute to poor healing in DFUs^[105-107]. Elevated MMP activity and imbalanced ⁶ tissue inhibitors of metalloproteinases (TIMPs) have been reported ⁵ in the skin of diabetic ulcer patients. A study ⁵ reported that enhanced expression of MMP-1 is necessary for wound healing in DFU, while enhanced ⁵ MMP-8 and MMP-9 contribute to delayed wound healing. Furthermore, a higher MMP-1/TIMP-1 ratio may indicate a proteolytic environment in the wound^[106,107]. Adiponectin has been found to suppress fibroblast proliferation, migration, and ECM formation^[108], as well as increase the expression of fibroblasts and type 1 collagen components of the ECM^[109,110]. The endogenous expression of adiponectin and its malfunctioning may play a fundamental role in skin fibrosis and exert a substantial negative regulatory impact on fibrosis^[111].

In summary, adiponectin has been shown to influence ECM composition by regulating the activity of ECM-associated molecules, such as collagen, elastin, and glycosaminoglycans, implicating that as a potential mechanism through which adiponectin may help promote DFU healing.

ADIPONECTIN' ANTI-INFLAMMATORY PROPERTIES AND WOUND HEALING

Another contributing factor to the poor wound healing in diabetic patients is the presence of excessive inflammatory reaction^[112,113]. DFUs which are characterized by

chronic inflammation and infection, can lead to tissue necrosis and amputation^[114-116]. In individuals with diabetes, the wound healing process is often hindered as the wounded tissues remain in the late inflammatory phase. In such cases, macrophages are unable to transition into the repair phenotype and release the necessary factors that promote tissue repair. As a result, the wound fails to progress from the inflammatory to the proliferative phase of healing, leading to persistent inflammation^[117]. The persistent inflammation in DFUs is contributed by the activation of pro-inflammatory cytokines such as TNF- α , IL-1 β , and IL-6, as well as the activation of the nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B) signaling pathway^[112,113,118]. In addition, downregulation of neuropeptides that are essential for wound healing and biofilm development also contributes to the persistent inflammatory state in DFUs. Biofilms, which disrupt wound healing by creating a prolonged inflammatory response, limit macrophage phagocytosis and keratinocyte growth migration, and transmit antimicrobial resistance genes^[119-121].

Adiponectin has an anti-inflammatory effect and is a potential therapeutic option for preventing and treating DFUs. It has been demonstrated that adiponectin reduces the expression of pro-inflammatory markers and inhibits the activation of the NF- κ B signaling pathway in human aortic endothelial cells and monocytes^[122-124]. Activation of AdipoR1 and AdipoR2 receptors increase the activity of AMPK and the inhibit the NF- κ B signaling in various cells including macrophages, liver, and skeletal muscle. Both contribute to adiponectin's anti-inflammatory properties^[50,125]. The crucial role of AMPK signaling activity in wound healing is highlighted by the successful improvement of DFU healing achieved through the reactivation of AMPK signaling^[126]. Adiponectin may also inhibit the activation of the inflammasome, a complex of proteins which plays a key role in the inflammatory response^[127,128].

Adiponectin stimulates the nuclear receptor peroxisome proliferator-activated receptor-gamma (PPAR- γ), which affects glucose and lipid metabolism^[129,130]. When PPAR- is activated, pro-inflammatory cytokines are suppressed, and anti-inflammatory genes are activated. Adiponectin also suppresses the creation of reactive oxygen species

and the activation of NADPH oxidase^[131], which contribute to inflammation and oxidative stress. Adiponectin may also limit the migration and proliferation of vascular smooth muscle cells, as well as the development of new blood vessels^[132-134], while promoting the regression of existing blood vessels, which can also contribute to its anti-inflammatory effects^[135-137]. Hence, adiponectin's anti-inflammatory properties may aid wound healing by minimising prolonged inflammation and accelerating the wound's transition into the proliferative phase of recovery.

ADIPONECTIN AS AN ANTIBACTERIAL IN DFUS

A meta-analysis by Macdonald *et al*^[138] found that DFUs are caused by several genera of bacteria, mainly gram-positive. Another study by Smith *et al*^[139] revealed populations of gram-positive bacteria and both aerobes and anaerobes. These bacteria can form biofilms, making it more difficult for antimicrobials to access and thus slowing down the healing process^[140].

Diabetics are also susceptible to periodontitis, which is associated with dysbiotic plaque biofilms and eventually leads to destruction of the tooth-supporting structures. DFUs are similar in that they comprise bacteria that form biofilms and eventually lead to destruction of the underlying bone structures. A study by Wang *et al* (2021) suggested that the level of adiponectin has an inverse association with periodontitis. Treatment with adiponectin in animal experiments better improved tissue destruction and suppressed inflammation, which improved bone regeneration^[140]. There is little literature on the use of adiponectin as an antibacterial for DFUs. However, a study by Wang *et al* 2021 suggests that adiponectin may inhibit inflammation stimulated by obesity or by periodontal pathogens and somehow influence antibacterial outcomes.

Given these findings, further research is needed to explore the antibacterial effects of adiponectin in DFUs and its use as a candidate for the treatment of this chronic condition.

ADIPONECTIN AND IMMUNE RESPONSE

One contributing factor to the delayed healing and susceptibility to bacterial infection in DFUs is the low immune response. Research has shown that adiponectin has the ability to modulate immune cell activity by inhibiting the activation and differentiation of T-helper 1 (Th1) cells, which leads to the emergence of inflammatory and autoimmune diseases, while promoting the activation and differentiation of Th2 cells, which regulate immune responses^[141]. Adiponectin achieves its anti-inflammatory effects by regulating multiple signaling pathways and modulating cellular processes involved in inflammation, making it a promising therapeutic target for various inflammatory and metabolic disorders. Additionally, studies suggest that adiponectin can modulate bacterial infection by regulating the activity of molecules responsible for bacterial uptake and killing^[142,143].

ADIPONECTIN AND FIBROBLAST GROWTH FACTORS IN DFU HEALING

Fibroblast growth factors (FGFs) are proteins that are expressed in various tissues and play a crucial role in wound repair^[144]. There are two types of FGFs: Paracrine and endocrine. Endocrine FGF regulates various metabolic processes and cell survival, while paracrine FGFs regulate neural development, angiogenesis, and wound healing. Studies have shown that specific types of FGFs for instance aFGF, bFGF, and the FGF 15/19 subfamily may have a positive effect on diabetic wound healing. aFGF aids in diabetic ulcer healing by stimulating capillaries, fibroblasts, and proliferative proteins in ulcer tissue^[145]. Regulating the release of bFGF has also been shown to enhance skin wound healing and epithelium development in diabetic mice, while also minimizing scar formation by promoting fibroblast and myofibroblast apoptosis^[146]. Additionally, FGF-19 and FGF-21 have been found to be excessively expressed in the serum of diabetes patients^[147,148]. FGFs are more potent angiogenesis factors than platelet-derived growth factor and vascular endothelial growth factor (VEGF). FGFs enhance the development of granulation tissue by increasing fibroblast proliferation and angiogenesis^[149].

ADIPONECTIN AND ANGIOGENESIS

Studies have shown that impaired angiogenesis contributes to the poor healing of DFUs^[150,151]. This is due to various factors such as the failure of macrophages to change into a repair phenotype^[150], elevated levels of plasma pigment epithelium-derived factor (PEDF), and dysregulation of angiopoietin-1 (Ang 1) and Ang 2. Macrophages are a key source of VEGF and other pro-angiogenic substances in wounds. On the other hand, PEDF has been found to delay wound healing and decrease angiogenesis in diabetic wounds^[151].

Adiponectin has both pro-angiogenic and anti-angiogenic effects, depending on the signaling pathways involved. Adiponectin can promote the formation of new blood vessels through various mechanisms. For example, it increases the production of pro-angiogenic factors like VEGF and FGF-2, and stimulates the migration, proliferation, and differentiation of endothelial cells. This is thought to happen because adiponectin activates signaling pathways like Akt and AMPK^[152,153]. However, adiponectin can also inhibit angiogenesis in some contexts. It decreases the production of pro-angiogenic factors and inhibits the expression of angiogenic factors like FGF-2. Additionally, it can inhibit the migration and invasion of certain cancer cells through the modulation of signaling pathways like Akt and AMPK^[154].

Hence, the effects of adiponectin on angiogenesis could help promote wound healing. However, those effects are context-dependent and complex. Further research is needed to understand the molecular mechanisms behind these effects and to determine its potential therapeutic applications in different contexts.

ADIPONECTIN AND APOPTOSIS

Impaired apoptosis is another factor that contributes to poor healing of DFUs^[155]. During the wound healing process, different cell groups go through various stages of clearance, culminating in apoptosis. DFU trauma causes mitochondrial damage, which increases the expression of pro-apoptotic proteins while decreasing the expression of anti-apoptotic proteins such as B-cell lymphoma-2 (Bcl-2). This results in apoptosis in

cells such as fibroblasts and vascular smooth muscle cells. Low expression of FGF-2, a factor related to fibroblast mitosis and cell survival, has been observed in diabetic wound cells. Reduced expression of other factors related to fibroblast regeneration, such as adiponectin, also contributes to this process^[156].

Furthermore, delayed apoptosis has been reported during the inflammatory phase of wound healing in diabetic mice, which may contribute to the persistent inflammatory state in DFUs^[157]. Excessive cell death due to hyperglycaemia can lead to poor structural recombination and difficulty in generating granulation tissue, making the wound more susceptible to infection^[155]. In addition, chronic hyperglycaemia associated with altered lipid and glucose metabolism promotes a condition of oxidative stress, which results in long-term chronic inflammation of wounds across all stages of wound healing.

Adiponectin has been shown to have anti-apoptotic effects, which may prevent cell death in the wound area and promote wound healing. Studies have shown that adiponectin can inhibit the activation of the intrinsic apoptotic pathway, leading to the prevention of cell death and promotion of wound healing. For example, a study showed that treatment with recombinant human adiponectin promoted wound healing in diabetic mice by inhibiting the activation of the intrinsic apoptotic pathway. However, further research is needed to understand the molecular mechanisms behind adiponectin's effects on apoptosis and its potential therapeutic applications in different contexts^[158].

ADIPONECTIN AND WOUND CONTRACTION

Adiponectin has been identified as a mediator of wound contraction, a process that involves the reduction of wound size through the convergence of wound edges. This action is considered to occur *via* a variety of molecular mechanisms, including collagen synthesis stimulation, MMP inhibition, and myofibroblast migration and proliferation boosting.

Adiponectin has been found to increase the production of collagen; a critical component of the ECM necessary for wound healing. This is achieved through the

activation of the Transforming Growth Factor- β (TGF- β) signalling pathway, which stimulates collagen synthesis *via* the phosphorylation of Smad2 and Smad3. Adiponectin also enhances the activity of procollagen type I and III mRNA, which are necessary for collagen synthesis^[159].

Additionally, adiponectin suppresses MMPs, enzymes that degrade the ECM and hinder wound healing. By decreasing the expression of MMP-2 and MMP-9 and increasing the expression of TIMP-1, an inhibitor of MMPs, adiponectin promotes the maintenance of the ECM^[160,161].

Furthermore, myofibroblasts play a crucial role in wound healing and contraction. However, excessive myofibroblast proliferation during the late stage of wound healing can lead to the formation of pathological scars that greatly reduce the quality of wound healing^[162]. Studies have shown that adiponectin may prevent the formation of pathological scars by inhibiting myofibroblast synthesis, proliferation, and migration^[163,164].

Therefore, adiponectin may increase wound contraction by increasing collagen synthesis, inhibiting MMPs, and modulating myofibroblast migration as well as proliferation. More research is needed to understand the molecular mechanisms of these effects and to assess the therapeutic potential of adiponectin in wound healing.

ADIPONECTIN AND OXIDATIVE STRESS

Excessive oxidative stress is a hallmark of diabetic wounds, where high levels of ROS are present. The balance between ROS creation and elimination is crucial for proper wound healing. In diabetes, high glucose levels lead to an increase in energy metabolism substrates, which, in turn, result in elevated levels of superoxide and oxidative stress. This increased oxidative stress enhances the production of advanced glycation end products (AGEs)^[165,166]. Moreover, nitric oxide synthase decoupling in diabetes leads to decreased nitric oxide production^[167], further complicating the healing process. These findings highlight the crucial role that oxidative stress plays in diabetic wound healing and the need to address this issue to improve therapeutic outcomes.

Adiponectin has demonstrated wound healing benefits through its antioxidant properties. Specifically, adiponectin has been shown to increase insulin release^[75], enhance insulin sensitivity^[168], promote glucose uptake^[68,169], and scavenge ROS^[170]. These antioxidant properties of adiponectin provide new avenues for the development of effective therapeutic strategies for diabetic wound healing.

According to a review conducted by Woodward *et al*^[171], adiponectin has additional anti-inflammatory, anti-apoptotic, and antioxidative effects that can reduce cardiovascular oxidative stress. Matsuda and Shimomura^[172] also suggested that adiponectin may protect against oxidative-stress-induced damage in the cardiovascular system, and that circulating adiponectin levels and increased oxidative stress may contribute to the pathogenesis of obesity-associated metabolic diseases. Nguyen^[173] proposed that adiponectin could be explored as a focus of new treatment strategies in various metabolic diseases due to its antioxidative, anti-inflammatory, and anti-fibrotic effects, which help regulate glucose levels, lipid metabolism, and insulin sensitivity. However, further research is needed to investigate the antibacterial effects of adiponectin in DFUs and its potential use as a treatment strategy.

ADIPONECTIN AND NERVE FUNCTION

The development and poor healing of DFUs is influenced by peripheral neuropathy, a complex and multi-factorial condition. Among the identified contributors to DPN are oxidative stress, hypoxia, AGEs, activation of T lymphocytes, and insufficiency of nerve growth factors. Reduced expression of neuropeptides is a hallmark of neuropathy in both autonomic and sensory nerve fibers that arises from diabetes mellitus. These neuropeptides, which act as neuromodulators, play a crucial part in the process of diabetic wound healing^[174]. Adiponectin has been suggested to promote wound healing in diabetics through its neuroprotective role, although further research is required to fully understand the underlying mechanisms involved^[175].

ADIPONECTIN AND INSULIN SENSITIVITY

Persistent hyperglycaemia in diabetic patients is a main factor contributing to delayed wound healing and progression of DFUs^[176]. Several studies reported the beneficial effect of adiponectin on insulin sensitivity through its metabolic effects on various tissues, including skeletal muscle, liver, and adipose tissue. Skeletal muscle, which is a key factor in insulin sensitivity and glucose metabolism, has a high concentration of adiponectin receptors and has been shown to have increased glucose uptake and decreased intramyocellular triacylglycerol content in response to adiponectin binding^[177,178]. The liver, on the other hand, experiences decreased glucose production and increased insulin sensitivity when physiological levels of adiponectin are present^[179]. Adiponectin has also been shown to have anti-inflammatory effects in various tissues, including adipose tissue and liver^[180-183]. In addition, adiponectin has been linked to weight reduction and improved insulin sensitivity, glucose metabolism, and insulin secretion in rodents as evidenced by a recent study^[184]. In addition, adiponectin improves insulin sensitivity through modulating the gut microbiome^[185].

Thus, adiponectin has been proposed to have potential therapeutic and preventive applications in DFUs through various mechanisms, as outlined in Figure 3.

ADIPONECTIN LEVELS IN DFUS: A COMPREHENSIVE REVIEW OF CURRENT EVIDENCE

To review the available evidence on the measurement of adiponectin levels in DFUs in patients with T2DM, a comprehensive search of relevant databases such as PubMed and Google Scholar was conducted to identify relevant studies. The findings of seven selected studies are presented chronologically in Table 1. The results of these studies revealed a consistent pattern, with lower plasma levels of adiponectin found in patients with DFUs compared to those without DFUs. A negative correlation between the duration of diabetes and the development of DFUs was also observed. The findings further indicated a positive association between low plasma levels of adiponectin and DFUs, and that low adiponectin levels could serve as a predictor for DFUs. The results of these investigations imply that reduced levels of adiponectin in the blood of

individuals with T2DM and DFUs may play a role in the emergence of foot ulcers by means of microvascular and inflammatory processes.

ROLE OF ADIPONECTIN IN WOUND HEALING AND METABOLIC CONDITIONS: INSIGHTS FROM *IN VITRO* AND *IN VIVO* STUDIES

Adiponectin has been found to play a crucial role in wound healing, both *in vivo* and *in vitro*. Kumada *et al*^[186] found that the incubation of human monocyte-derived macrophages with human recombinant adiponectin increased tissue inhibitor of metalloproteinases; TIMP-1 mRNA levels in a dose-dependent manner without affecting MMPs mRNA levels. Adiponectin selectively increased TIMP-1 expression in human monocyte-derived macrophages through the induction of IL-10^[186].

Kawai *et al*^[187] investigated the effect of human recombinant adiponectin on an immortalized human keratinocyte cell line (HaCaT) and found that adiponectin suppressed the gene expression of involucrin, a marker of keratinocyte differentiation, in a dose-dependent manner. Adiponectin also upregulated the expression of TGFβ1 in HaCaT cells and promoted apoptosis in keratinocytes, which could inhibit hyperkeratosis during wound healing in diabetic patients^[187,188].

Shibata *et al*^[94] found that adiponectin was a powerful mediator in the regulation of cutaneous wound healing. Adiponectin receptors were found in normal human keratinocytes, and adiponectin increased keratinocyte proliferation and migration via AdipoR1/AdipoR2 and the ERK signaling pathway. Wound closure was significantly delayed in adiponectin-deficient mice compared to wild-type mice, and both systemic and topical adiponectin treatment improved wound healing in adiponectin-deficient and diabetic mice^[94].

In 2013, an orally active adiponectin receptor agonist, AdipoRon, was developed and was found to have similar effects to adiponectin^[76,189], improving insulin sensitivity and glucose tolerance, lipid metabolism^[190], and vascular dysfunction in type 2 diabetic mice^[191].

Salathia *et al*^[192] found that the injection of adiponectin into the skin edges of a wound accelerated healing and enhanced epithelialization at the wound margin. Jin *et al*^[193] found that adiponectin promoted the growth and migration of preadipocytes in an adipose tissue wound healing study. FGF21 has also been shown to stimulate the production of adiponectin, which could contribute to the expansion of subcutaneous fat and improvement of systemic insulin sensitivity^[194].

Kim *et al*^[195] conducted a study on the effects of AdipoRon, a synthetic adiponectin receptor agonist, on diabetic nephropathy in T2DM patients. The study found that AdipoRon treatment reversed kidney abnormalities caused by diabetes in mice. The renoprotective benefits of AdipoRon were achieved through the activation of AdipoR1 and AdipoR2 receptors in the kidneys, which improved pathways related to lipid accumulation and endothelial impairment. AdipoRon also increased intracellular Ca²⁺ levels and activated a CaMKK/phosphorylated Ser431LKB1/phosphorylated Thr172AMPK/PPAR pathway, reducing oxidative stress and apoptosis, and improving endothelial dysfunction. The study also found that AdipoRon had cardioprotective benefits through the same mechanism as shown in the kidney^[195].

Hong *et al*^[95] conducted a study examining the impact of recombinant human full-length adiponectin on human epidermal keratinocyte cell culture. The results showed that adiponectin improved the differentiation and lipid content of keratinocytes through modulation of the expression of multiple enzymes involved in lipid synthesis and regulation within these cells^[95].

Adiponectin replacement therapy has the potential to treat various human diseases, but due to the challenges in using the intact protein, efforts have focused on developing peptide and small molecule agonists of the adiponectin receptor. One such example is ADP355, a peptide that has low nanomolar cellular activity and efficacy in treating fibrotic and inflammation-related diseases. On the other hand, small-molecule therapies like AdipoRon can be taken orally and target multiple metabolic conditions. However, the difficulty in comparing the efficacy of different drug classes due to the use of various *in vivo* models and the limitations of *in vitro* measures makes it challenging to

determine their effectiveness. Adiponectin receptor antagonists can still be useful in target validation studies, but direct receptor agonists have been shown to be more effective in controlling direct signalling than therapies that aim to increase adiponectin production^[196].

Studies have shown the potential benefits of AdipoRon, a small-molecule therapy for multiple metabolic conditions, in improving various aspects of health. A 2020 study by Choi *et al*^[191] found that chronic oral intake of AdipoRon improved vascular function in type 2 diabetic mice through an endothelium-independent mechanism. Lindfors *et al*^[197] discovered that AdipoRon reduced proinflammatory cytokine expression and improved glomerular inflammation and injury in diet-induced obese mice and cultured podocytes. Sun *et al*^[198] showed that AdipoRon reduced inflammation markers and apoptosis, improved mitochondrial function, and accelerated wound healing in aged skin. Zatorski *et al*^[199] found that AdipoRon had a gastroprotective effect and reduced inflammation in stomach ulcers. Tarkhnishvili *et al*^[200] found that AdipoRon changed myocardial substrate preference towards higher glucose consumption in type 2 diabetic mice, although it was insufficient to enhance cardiac output and efficiency. Li *et al*^[201] reported that topical adiponectin was effective in improving clinical signs and reducing inflammation in a mouse model of dry eye or alkali burn, while Baradaran-Rafii *et al*^[137] showed that topical adiponectin decreased recent corneal neovascularization in rabbits.

Hence, adiponectin plays a vital role in wound healing, tissue regeneration, and metabolic regulation. It is typically administered through injections or orally *via* an adiponectin receptor agonist, such as AdipoRon. Studies have shown that adiponectin enhances wound healing, keratinocyte differentiation, and improves insulin sensitivity, glucose tolerance, lipid metabolism, and vascular dysfunction in diabetic patients. AdipoRon, a small-molecule therapy, has demonstrated similar effects to adiponectin and can be taken orally, targeting multiple metabolic conditions. Although challenges exist in comparing the effectiveness of various drug classes due to differing *in vivo* models and *in vitro* limitations, direct receptor agonists have shown promise in controlling signaling better than therapies aiming to increase adiponectin production.

Overall, adiponectin-based therapies appear to be safe and hold potential for treating a range of human diseases.

AREAS FOR FUTURE RESEARCH

¹⁸ Further research is needed to comprehensively understand the effects of adiponectin on DFUs. Although the existing literature has shown favourable outcomes, there is a need for more detailed exploration into the mechanisms underlying adiponectin-mediated promotion of wound healing and tissue regeneration. Research studies should be carried out to establish the safety and efficacy of adiponectin as a therapeutic intervention for DFUs in the clinical setting. Despite positive preclinical outcomes, clinical trials are essential to determine the appropriate dose and treatment schedule, as well as any potential adverse effects. Furthermore, additional research is required to identify subgroups of patients that may benefit the most from adiponectin therapy. This could include examining whether specific genetic or demographic factors influence the effectiveness of adiponectin treatment. Studies should be conducted to identify the optimal delivery method for adiponectin therapy, considering that while topical application has been successful in some studies, other delivery methods such as injection or implantation may be more efficacious in specific cases. Furthermore, ¹⁰ research should be conducted to determine whether adiponectin can be used in combination with other treatments for DFUs, such as antibiotics or growth factors, to enhance wound healing and tissue regeneration. Further investigation is also required into the long-term effects of adiponectin therapy, including its impact on wound recurrence rates and overall wound healing outcomes.

CONCLUSION

In conclusion, the available evidence suggests that adiponectin and its receptors agonist may hold promise as therapeutic targets for the management of DFUs. However, to fully comprehend ⁶ the role of adiponectin in DFU pathogenesis and treatment, additional research is necessary. Future studies should focus on conducting

longitudinal investigations to establish a causal relationship between adiponectin levels and DFU incidence. Moreover, exploring treatment strategies aimed at elevating adiponectin levels in patients with DFUs could provide ⁶ insights into the potential benefits of adiponectin as a therapeutic target. Investigating the specific cellular and molecular mechanisms underlying the relationship between adiponectin and DFUs is also necessary for a comprehensive understanding of the role of adiponectin in the condition. Additionally, clinical trials that evaluate the efficacy and safety of interventions targeting adiponectin levels in DFU prevention and management are needed. Such research could help to identify novel therapeutic targets for DFUs, ultimately leading to more effective management of the condition.

Figure Legends

Figure 1 The common pathways in diabetes mellitus leading to infected foot ulcer.

Figure 2 The role of low adiponectin in contributing to delayed wound healing.

Figure 3 The Potential mechanisms involved in adiponectin-mediated wound healing in diabetic foot ulcers. AdipoRon: Adiponectin receptor agonist; AdipoR1/AdipoR2: adiponectin receptors 1 and 2; MAPK: Mitogen-activated protein kinases; IRS2: Insulin receptor substrate 2; AMAK: Adenosine monophosphate-activated ² kinase; mTOR: Mammalian target of rapamycin; ECM: Extracellular matrix.

1 Table 1 Adiponectin levels in patients with diabetic foot ulcers: A summary of published studies

No	Ref.	Country	Study objective	Study design and sample size	Results	Adiponectin levels (ng/mL)			Conclusion
						Non-Diabetic	Diabetic without FUs	DFU	P value
1	Tuttolo et al ^[202] , 2010	Italy	To investigate the plasma levels of adiponectin, resistin and IL-6 in subjects with diabetic foot in comparison with subjects without foot complication	Case-control; sample size: 34 patients with type 2 DM and 37 type 2 DM with FU and 37 patients with type 2 DM levels of adiponectin; DFU patients have lower median; plasma levels of adiponectin; patients with foot ulcers had a longer duration of DM, higher	The patients with DFUs exhibited higher CRP, HbA1c, lipid profile, IL-6, resistin and lower levels of adiponectin; DFU patients have lower median; plasma levels of adiponectin; patients with foot ulcers had a longer duration of DM, higher	NA	8.48 × 10 ³ (5.15 × 10 ³ -12.87 × 10 ³) ¹	7.145 × 10 ³ (4.470 × 10 ³ -12.170 × 10 ³) ¹	0.022
Adiponectin levels are negatively correlated with the duration of diabetes and the development of DFUs									

and DFU (R2 = - 0.0189)

3	Ahmad <i>et al</i> ^[82] , 2012	India	<p>¹⁴ To evaluate plasma levels of Cathepsin D, adiponectin, TNF-α, IL-6, and hsCRP in subjects with diabetic foot in comparison with subjects without foot complications</p>	Prospective cohort multicentric hospital-based study; sample size: 211 diabetics with FUs, 208 diabetics without FUs	<p>¹⁸ The median of adiponectin were lower in patients with DFUs; adiponectin plasma levels were found to be negatively correlated with various ¹⁴ cardiovascular risk factors, including hypertension, dyslipidemia, and microvascular complications</p>	NA	13.3 (12.1-14.2) ¹	8.5 9.5) ¹	(7.1- < 0.0001	Low adiponectin is a predictor for DFUs; the study ¹¹ suggests that low levels of adiponectin in diabetic patients with foot ulcers could be linked to the development of foot ulcers through microvascular and inflammatory mechanisms. The findings also indicate ⁴ that adiponectin may play a role in inhibiting the	plasma
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11 expression of adhesion molecules on endothelial cells, 11 which are involved in the inflammatory vascular response

such as neuropathy, retinopathy, nephropathy, and PAD; this was found through 14 both multiple linear regression analysis and forward stepwise regression

8 analysis

4	Dhamodharan <i>et al</i> ^[203] , 2015	To investigate the genetic association of IL-6, TNF- α , and SDF-1 polymorphisms with serum cytokine, adiponectin, leptin and	Case-control; sample size: A total of 515 subjects were divided into four groups: Group-I (NGT)/control; <i>n</i> = 106), group-II known T2DM	536.0 (0.1-1787.0) ²	528.6 (6.2-1255.0) ²	524.0 (63.3-1641.0) ² in DFU+ DN; 453.5 (164.9-1078.0) ² in DFU + PVD	< 0.05	Low levels can be a biomarker of DFUs; 18 SNPs in cytokine/chemokine genes are useful biomarkers for DFU and can help predict the risk of developing DFU
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5	Viswana than <i>et al</i> [204], 2018	India	To examine the involvement of IL-6, TNF- α , and SDF-1)	hsCRP levels in diabetic foot ulcers	without (T2DM; $n = 139$); group-III T2DM with neuropathic DFU (DFU-DN; $n = 191$); group-IV T2DM with PVD (DFU-PVD; $n = 79$)	Cross-sectional; Data on adiponectin levels are not reported	NA	NA	NA	9 Screening for SNPs in TNF- α , SDF-1, and IL-6; among DFU subjects would help in identifying high risk individuals and might aid in better patient care
			polymorphisms in determining the susceptibility to foot microbial infection, grade of the ulcer) and treatment-outcome;							

9 (Debridement vs amputation) in DFU subjects and further, the effect of these SNPs on serum cytokine levels and biomarkers such as leptin, adiponectin, CRP and HOMA-IR

6	Anguian o- Hernandez et al ^[205] , 2019	México	To assess the modification in adiponectin, HIF-1α, NF-κB, IGFBP-3, VEGF and adiponectin in diabetic foot ulcers treated with hyperbaric oxygen	Study design: Not specified; sample size: 17 ambulatory patients and one hospitalized; patient with DFUs; 15 were males & 3 females; 17	Adiponectin levels increase	NA	NA	-14943 ± 7915 ² (before therapy); -17281 ± 7962 ² (after therapy)	± 0.035	The study found that while treatment increased adiponectin levels, the increase was not significant; however, all patients showed an increase in angiogenesis and fibrosis and a decrease in ulcer size and
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T2DM and 1
T1DM; grade 3
and 4 on
Wagner scale

infection signs after undergoing HBO₂ therapy. The results suggest that HBO₂ stimulates the expression of IGFBP-3, NF-κB, and HIF-1α and modulates the inflammatory response related to hypoxia.

7 Vangave Aust ti et ralia
al^[206], 2022

7 Vangave Aust
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2022

Prospective, randomized, double-blind, placebo controlled, single-centre study; sample size: 50 participants; 25 were assigned to	Vildagliptin treatment led to significant improvements in key health markers, including reduced HbA1c, total cholesterol, LDL
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NA	NA	11822 ± 1.0
		2584.0 ³ ;
	Placebo;	
	following;	
	treatment	
		13138 ±
		2671 ²

11822 ± 1.0	The	vildagliptin
2584.0 ³ ;	treatment in	DFU
Placebo;	patients	improve
following;	wound healing with	
treatment	an	associated
13138 ±	reduction in	some
2671 ²	inflammatory	
	biomarkers and a non-	
	significant increase in	
	adiponectin	

the placebo and cholesterol, and 25 to the total/HDL treatment group ¹³ compared to the placebo group. Additionally, vildagliptin demonstrated a protective effect on DFU wound healing

¹¹

¹Data presented as median and interquartile (lower and upper quartile);

²Data presented as mean \pm SD;

³Data presented as mean \pm SEM.

DFU: Diabetic foot ulcer; NGT: Normal glucose tolerance; DN: Diabetic neuropathy; PAD: Peripheral artery disease; PVD: Peripheral vascular disease; ¹⁰CRP: C-Reactive protein; hsCRP: High-sensitivity C-reactive protein; HIF-1 α : Hypoxia-inducible factor-1 α ; NF- κ B: Nuclear factor-kappa B; ³⁷IGFBP-3: Insulin-like growth factor-binding protein-3; VEGF: Vascular endothelial growth factor; ⁴⁷HBO₂: Hyperbaric oxygen; ³⁹IL-6: Interleukin-6; TNF- α : Tumour necrosis factor-alpha; SNPs: Single nucleotide polymorphisms; SDF-1: Stromal cell-derived factor; VEGF: Vascular endothelial growth factor; LDL: Low-density lipoprotein; HDL: High-density lipoprotein.

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