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Therapeutic possibilities of gut microbiota modulation in acute decompensation of liver cirrhosis

Garbuzenko DV. Gut microbiota modulation in acute decompensation of LC

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Abstract

The formation of liver cirrhosis (LC) is an unfavorable event in the natural history of chronic liver diseases and with the development of portal hypertension and/or impaired liver function can cause a fatal outcome. Decompensation of LC is considered the most important stratification variable for the risk of death. It is currently postulated that decompensation of LC occurs through acute (including acute-on-chronic liver failure) and non-acute pathway. Acute decompensation of LC is accompanied by the development of life-threatening complications, characterized by an unfavorable prognosis and high mortality. Progress in understanding the underlying molecular mechanisms has led to the search for new interventions, drugs, and biological substances that can affect key links in the pathogenesis of acute decompensation in LC, for example, the impaired gut-liver axis and associated systemic inflammation. Given that particular alterations in the composition and function of gut microbiota play a crucial role here, the study of the therapeutic possibilities of its modulation has emerged as one of the top concerns in modern hepatology. This review summarizes the investigations that describe the theoretical foundations and therapeutic potential of gut microbiota modulation in acute decompensation of LC. Despite the encouraging preliminary data, the majority of the suggested strategies have only been tested in animal models or in preliminary

clinical trials; additional multicenter randomized controlled trials must demonstrate their efficacy in larger patient populations.

Key Words: Liver cirrhosis; Acute decompensation; Pathogenesis; Therapy; Gut microbiota; Modulation

Core tip: Given that particular alterations in the composition and function of gut microbiota play a crucial role in the pathogenesis of acute decompensation in LC, this review summarizes the investigations that describe the theoretical foundations and therapeutic potential of gut microbiota modulation in acute decompensation of LC. Despite the encouraging preliminary data, the majority of the suggested strategies have only been tested in animal models or in preliminary clinical trials; additional multicenter randomized controlled trials must demonstrate their efficacy in larger patient populations.

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INTRODUCTION

The formation of liver cirrhosis (LC) is an unfavorable event in the natural history of chronic liver diseases and with the development of portal hypertension and/or impaired liver function can cause a fatal outcome. During LC, there is a compensated stage, which is usually asymptomatic and characterized by preserved quality of life and a median survival exceeding 12 years, and a decompensated stage associated with the occurrence of life-threatening complications, in which a median survival dropped to 2–4 years. Accordingly, the decompensation of LC is considered as the most important stratification variable for the risk of death[1]. Decompensation of LC presents as acute decompensation in a portion of patients with the development of one or more major complications and is accompanied by high mortality, while in many other, its characteristic signs are usually slow-progressing ascites or mild grade 1-2 hepatic encephalopathy, or jaundice, not requiring hospitalization[2].

The EASL-CLIF Consortium CANONIC study, established diagnostic criteria for acute-on-chronic liver failure (ACLF), introduced the concept of acute

decompensation as a distinct clinical presentation of decompensation of LC defined by the acute development of ≥ 1 major complication(s):

- acute (for less than 2 weeks) development of grade 2 or 3 ascites; this may be the first or a new episode of ascites;
- acute hepatic encephalopathy, which is manifested by a sudden change in the mental status of a patient with no previous cognitive impairment and no signs of acute neurological disease; this may be the first or a new episode of hepatic encephalopathy;
- acute gastrointestinal bleeding from the upper and/or lower gastrointestinal tract of any etiology;
- spontaneous bacterial peritonitis (SBP), spontaneous bacteremia, urinary tract infection, pneumonia, cellulite, as well as any other type of acute bacterial infection[3].

The cause of acute decompensation of LC can be both exogenous factors (e.g. bacterial infections, alcohol abuse, etc.) and endogenous factors (e.g. progressive liver disease, translocation of intestinal bacterial immunogenic material to the systemic circulation)[4]. Its most severe form - ACLF, according to the definition of the American College of Gastroenterology (ACG), is a potentially reversible condition in patients with chronic liver disease with or without LC that is associated with the potential for multiple organ failure and mortality within 3 months in the absence of treatment of the underlying liver disease, liver support, or liver transplantation. The severity of organ failure may be assessed by the EASL-CLIF sequential organ failure assessment score or NACSELD organ failure score[5].

The first investigation derived from the PREDICT study group of the EASL-CLIF Consortium uncovered that acute decompensation of LC without ACLF is a heterogeneous condition with three different clinical courses and two major pathophysiological mechanisms: systemic inflammation and portal hypertension. The first clinical course includes patients who develop ACLF and have an extremely high short-term mortality rate - termed pre-ACLF. The second clinical course includes patients with unstable decompensated LC who require frequent hospitalizations unrelated to ACLF and is associated with a lower mortality risk than

pre-ACLF. The third clinical course includes patients with stable decompensated LC who rarely require hospital admission and have a much lower 1-year mortality risk.

Each clinical course of acute decompensation of LC differs significantly regarding the grade of systemic inflammation and the severity of portal hypertension. A high-grade of systemic inflammation at admission with exacerbation during follow-up is observed in pre-ACLF. A low-grade of systemic inflammation at admission with subsequent steady-course is observed in patients with unstable decompensated LC. A low-grade of systemic inflammation at admission with subsequent improvement is observed in patients with stable decompensated LC. A high-grade severity of portal hypertension is observed in patients with unstable decompensated LC. A low-grade severity of portal hypertension is observed in pre-ACLF and stable decompensated LC[6].

The aim of the second investigation derived from the PREDICT study group of the EASL-CLIF Consortium was to analyze and characterize the precipitants leading to acute decompensation of LC without ACLF, or with ACLF. It turned out that of the clinical events explored potential precipitants, only four (proven bacterial infections, severe acute alcoholic hepatitis, gastrointestinal bleeding associated with shock and toxic encephalopathy) fulfilled the diagnostic criteria of precipitants. Proven bacterial infections and severe alcoholic hepatitis were present in the absolute majority (>96%) of the patients. However, no precipitating event could be identified in 2/3 of acute decompensation of LC without ACLF patients and in 1/3 acute decompensation of LC with ACLF patients. These data suggest that acute decompensation of LC without ACLF develops more frequently in the context of endogenous mechanisms (e.g. progressive liver disease, bacterial translocation). The prevalence and number of precipitants increased with the severity of the acute decompensation-sub-phenotype from stable decompensated LC/unstable decompensated LC to pre-ACLF and ACLF, which were also directly related to clinical course severity and short-term mortality in patients with acute decompensation of LC. These data, therefore, strongly suggest that specific preventive and therapeutic approaches for these precipitants must improve outcomes in decompensated LC[7].

Current therapeutic strategies in acute decompensation of LC provide the removal of the precipitants, the treatment of specific complications, as well as intensive

monitoring and support of vital body functions[8]. Liver transplantation may be a successful treatment option for some of the most severe ACLF patients, but its implementation is usually associated with high costs and worse survival compared to "standard" elective surgery[9].

Progress in understanding the underlying molecular mechanisms has led to the search for new interventions, drugs, and biological substances that can affect key links in the pathogenesis of acute decompensation in LC, for example, the impaired gut-liver axis and associated systemic inflammation[10]. Given that particular alterations in the composition and function of gut microbiota play a crucial role here, this review summarizes the investigations that describe the theoretical foundations and therapeutic potential of gut microbiota modulation in acute decompensation of LC.

LITERATURE SEARCH

This review provided an overview of the current knowledge of the therapeutic possibilities of gut microbiota modulation in acute decompensation of LC. The PubMed and Embase databases, the Web of Science platform, the Google Scholar retrieval system, the Cochrane Database of Systematic Reviews, Reference Citation Analysis (https://www.referencecitationanalysis.com/) and the reference lists from related articles were used to search for relevant publications. Articles corresponding to the aim of the review were selected for 2003–2023 using the keywords: "liver cirrhosis", "acute decompensation", "acute-on-chronic liver failure", "pathogenesis", "therapy", "gut microbiota", "modulation". The investigations that describe the theoretical foundations and therapeutic potential of gut microbiota modulation in acute decompensation of LC was the inclusion criterion.

SYSTEMIC INFLAMMATION AS A MAIN DRIVER OF ACUTE DECOMPENSATION IN LIVER CIRRHOSIS

One of the leading hypotheses in recent years suggests that the main driver of acute decompensation and concomitant multiple organ failure in LC is systemic inflammation. Its cause may be the "spill over" of damage-associated molecular patterns (DAMPs), cytokines and immune regulatory cells from the chronically

inflamed liver and potential other inflamed tissue sites to the systemic circulation. Additionally, pathogen-associated molecular patterns (PAMPs), namely bacterial and bacterial products, in particular, the lipopolysaccharides of the cell wall of Gram-negative bacteria, due to pathological translocation from the intestinal lumen through the portal vein, reach the cirrhotic liver where they may be ineffectively cleared by Kupffer cells or shunted through vascularized septae into the systemic circulation, contributing to systemic inflammation (Figure 1). Over time, immune tolerance may develop, which is characterized by accumulation in the systemic circulation of immune cells with immune suppressive- or immune regulatory properties, along with high serum levels of pro- and anti-inflammatory cytokines, DAMPs and PAMPs. Additionally, with further disease progression to ACLF, cells with a reduced capacity to repel microbial challenges appear in the systemic circulation, which increases the risk of infectious complications and sepsis[11]. Once the first episode of acute decompensation of LC develops, systemic inflammation follows a chronic course, with transient episodes of reactivation caused by exogenous pro-inflammatory factors or to bursts of bacterial translocation. Repeated episodes of acute decompensation during the clinical course of decompensated LC develop in the setting of these reactivation of the immune system. The prognosis of patients with acute decompensation of LC associated with moderate, non-progressive systemic inflammation depends on the severity of portal hypertension. Patients with severe portal hypertension frequently develop an unstable clinical course, requiring frequent hospital readmission, and significant short- and long-term mortality. In contrast, if portal hypertension is moderate, systemic inflammation improves after the episode of acute decompensation of LC, patients develop a benign stable course, and long-term mortality is low[12].

THE ROLE OF GUT MICROBIOTA IN THE PATHOGENESIS OF ACUTE DECOMPENSATION IN LIVER CIRRHOSIS

The alteration of gut microbiota composition in acute decompensation of LC create prerequisites for disruption of the gut-liver axis, and bacterial translocation contributing to systemic inflammation is based on small intestinal bacterial

overgrowth (SIBO), gut dysbiosis and increased permeability of the intestinal epithelial barrier[13].

SIBO, which is characterized by an excessive number of bacteria in the small intestine ($\geq 10^3$ CFU/ml) with a predominance of Gram-negative aerobic and anaerobic species, occurs in about half of the patients with LC, but the mechanism of its development has not been definitively established. One of the possible reasons may be the slowing down of orocecal transit[14]. However, the causal relationship between these pathophysiological conditions remains unclear. In some studies, a more pronounced slowing down of orocecal transit was observed in patients with Child-Turcotte-Pugh (CTP) class B and C LC with hepatic encephalopathy, which was explained by the of autonomic neuropathy, metabolic derangements due to portosystemic shunting, and SIBO[15].

Because gastric acid is an important barrier that prevents bacterial colonization of the stomach and small intestine, it is assumed that proton pump inhibitor therapy may promote the SIBO through chronic acid suppression and subsequent hypochlorhydria. However, a meta-analysis of 19 studies demonstrated that proton pump inhibitor therapy was significantly associated with a moderately increased risk of SIBO [odds ratio (OR) 1.71, 95% CI 1.20–2.43][16]. The immune system also plays a role in the genesis of SIBO, as evidenced by the high prevalence of SIBO in patients who have immunodeficiency. Besides, immunoglobulin A content on the duodenum and jejunum mucosa has been shown significantly increased the patients with SIBO[17].

Gut dysbiosis in LC is manifested by an unfavorable change in the balance of autochthonous species of microorganisms with a reduction in symbiont bacteria belonging to *Firmicutes* phylum (e.g. the *Ruminococcaceae* and *Lachnospiraceae* families, etc.) and growth in pathobiont bacteria of the *Bacteroidetes* and *Actinobacteria* phyla[18]. These changes largely depend on the etiology of LC and are aggravated in its decompensated stage. For example, they were the most significant in patients with alcoholic LC, who had the highest content of bacteria of the *Enterobacteriaceae* and *Halomonadaceae* families and the lowest content of bacteria of the *Lachnospiraceae*, *Ruminococcaceae*, and *Clostridiales Incertae Sedis XIV* families, which was accompanied by an exorbitant level of endotoxemia[19]. Note that the bacteria of the

Enterobacteriaceae family express a powerful immunostimulating endotoxin and are the main pathogens involved in the pathogenesis of SBP[20]. In a study by Shu *et al*[21], in patients with hepatitis B virus (HBV)-related LC, the most common phyla of bacteria were *Firmicutes*, *Bacteroidetes*, *Proteobacteria*, *Actinobacteria*, *Fusobacteria*, and *Verrucomicrobia*, together constituting 92.1% of the total number of microorganisms studied. Patients with compensated LC had a high relative abundance of bacteria of the genus *Faecalibacterium spp.* and the *Ruminococcaceae* family, whereas with decompensated LC, bacteria of the genus *Streptococcus spp.* and the *Enterobacteriaceae* family prevailed.

In a prospective study by Solé *et al*[22], patients with LC had a significant decrease in gene and metagenomic species diversity compared to healthy subjects. This was associated with disease stages and was especially noticeable in patients with ACLF and persisted after antibiotic therapy. ACLF was accompanied by a significant increase in *Enterococcus spp.* and *Peptostreptococcus spp.* and a decrease in some autochthonous bacteria. Gut microbiome alterations correlated with the model for end-stage liver disease (MELD) and CTP scores, and multiple organ failure and were associated with some complications, especially hepatic encephalopathy and infections. Additionally, the gut microbiome predicted 3-month survival with good stable predictors. Functional analysis showed that patients with LC had enriched pathways related to ethanol production, γ -aminobutyric acid metabolism, and endotoxin biosynthesis.

According to a study by Philips *et al*[23] pathogenic genera of bacteria in gut microbiota, in particular, *Leptotrichia spp.*, *Neisseria spp.*, and *Erwinia spp.*, were predominant in patients with decompensated LC with infection, and their survival correlated with the presence of microorganisms with high functional propionate metabolism, for example, bacteria of the genus *Megamonas spp.*

Bajaj *et al*[19] developed a quantitative index to describe microbiome alterations accompanying LC based on the ratio of "good vs. bad" taxa abundance (cirrhosis dysbiosis ratio - CDR). It is designed to predict the course of LC and assess the risk of possible complications. CDR is calculated using the formula:

$CDR = [Lachnospiraceae (\%) + Ruminococcaceae (\%) + Veillonellaceae (\%) + Clostridiales Incertae Sedis XIV (\%)] / [Bacteroides spp. (\%) + Enterobacteriaceae (\%)]$.

The authors found that a low CDR was associated with death and organ failures within 30 days.

In a prospective study by the North American Consortium for the Study of End-Stage Liver Disease (NACSELD), including hospitalized patients with LC across North America, the CDR was lower in subjects who developed ACLF, especially among those with renal failure. Taxa belonging to the Proteobacteria phylum (Enterobacteriaceae, Campylobacteriaceae, and Pasteurellaceae) and Firmicutes phylum (Enterococcaceae and Streptococcaceae) were associated with the development of negative outcomes, whereas other Firmicutes members (Lachnospiraceae and Clostridiales) reduced the risk of negative outcomes. Changes in the microbiota associated with extra-hepatic organ failure, transfer to intensive care, ACLF, and death, independently on logistic regression analyses[24].

To study the influence of gut dysbiosis on prognosis in LC, Maslennikov *et al* modified CDR by placing "bad" bacteria in the numerator, and "good" bacteria in the denominator (MDR)[25]:

$MDR = [Bacilli (\%) + Proteobacteria (\%)] / [Clostridia (\%) + Bacteroidetes (\%)]$.

Their case-control study included 48 patients with LC and 21 healthy controls.

Patients with MDR more the median made up the group with severe dysbiosis, others did the group with non-severe dysbiosis. The follow-up period was 4 years.

The mortality rate of patients with severe dysbiosis was significantly higher than that of patients with non-severe dysbiosis. The abundance of Enterobacteriaceae, Proteobacteria, and Lactobacillaceae was increased and the abundance of Firmicutes and Clostridia was reduced in the deceased patients compared with survivors. The abundance of Bacilli, Enterococcaceae and Lactobacillaceae was higher and the abundance of Clostridia was lower in those who died during the first year of follow-up compared with those who survived this year. The abundance of Enterobacteriaceae and Proteobacteria was higher in those who died in the second to fourth year of follow-up compared with survivors.

The cause of gut dysbiosis in LC is not fully understood. One of the key theories explains its presence by depletion of the pool of bile acids due to a decrease in their

synthesis and secretion by hepatocytes. Bile acid synthesis is regulated mainly through the activation of nuclear receptors, in particular, the farnesoid X receptor (FXR), which also induces genes affecting intestinal permeability and inflammation, preventing bacterial translocation in experimental LC[26]. Bile acids have both direct and indirect antimicrobial effects through the modulation of FXR activity, which is important for the homeostasis of the epithelial and gut-vascular barrier. ²⁰ Colonic microbial groups are responsible for the deconjugation and 7 α dehydroxylation of bile acids, and it is hypothesized that the presence of microbe toxic bile acids (particularly deoxycholic acid) in the intestine is a factor that keep undesirable microbial populations under control [27]. The insufficient content of primary bile acids in feces decreased in 7 α -dehydroxylating bacteria belonging to *Firmicutes* phylum, especially the genera *Blautia spp.* and *Ruminococcus spp.* Their deficiency in the small intestine causes overgrowth of pro-inflammatory pathogenic bacteria belonging to *Proteobacteria* phylum, in particular, the *Enterobacteriaceae* family, which induces the release of markers of intestinal inflammation and exacerbates necroinflammatory changes in liver tissues. This triggers a positive feedback mechanism leading to additional inhibition of bile acid synthesis[28]. On the contrary, oral administration of conjugated bile acids to rats with a model of CCl₄-induced LC and ascites significantly reduced the bacteria in the ileum (especially *Escherichia coli* and *Enterococcus spp.*) to levels comparable to those in healthy rats, decreased the SIBO, bacterial translocation, and endotoxemia[29].

Increased permeability of the intestinal epithelial barrier is associated with both gut dysbiosis[30] and microcirculatory disorders in LC that change the barrier properties of the intestinal mucosa, which include mechanical, biological, immune, and chemical protection factors⁶[31]. Intestinal mucosa and intercellular junctions among epithelial cells form a layer that allows selective passage of the toxins and bacterial products⁶. Intestinal epithelial cells produce mucus, which forms a thick layer on the mucosa that prevents the bacterial translocation. Mucous secretions are rich in immunoglobulin A, which neutralizes toxins and microorganisms and prevents their adhesion and colonization. Bile acid secretion also plays a role in intestinal permeability by affecting the intestinal mucosa and by neutralizing endotoxin[32].

In LC, the thickness of the intestinal mucosa is decreased with the loss of mucus-producing goblet cells. The ultrastructural changes of the mucosa, contributing to increased permeability of the intestinal epithelial barrier, are characterized by atrophy, edema and inflammatory infiltration of the lamina propria, fibromuscular proliferation, expansion of the space between neighboring cells, a reduction in the number of short but thicker microvilli, a decrease in the villi/crypt ratio. These disorders are associated with a diminution in the expression of the tight junction proteins, including occludin and claudin-1, in the intestinal mucosa. Additionally, irregularities of the glandular epithelia, loss of normal cylindrical shape, edematous villi, and loosening of the mucous membrane were revealed. High levels of lipid peroxidation markers in enterocytes lead to mitochondrial dysfunction and cellular instability[33].

The increased stimulation of gut-associated lymphoid tissue leads to the persistent activation of monocytes, dendritic cells, and T lymphocytes both in the intestinal mucosa and mesenteric lymph nodes, and to the consequent production of pro- and anti-inflammatory cytokines. Local inflammatory disorders can be a trigger for systemic inflammation because of immune cells entering the systemic circulation. This is facilitated by a violation of the production of intestinal antimicrobial peptides, in particular, α -defensins and Reg3 lectins, a decrease in the ability to bind and neutralize bacterial endotoxin by albumin, lipopolysaccharide-binding protein, high-density lipoproteins, low-density lipoproteins, very low-density lipoproteins, chylomicrons, apolipoproteins, as well as dysfunction of the immune system in patients with LC[34].

The persistence of systemic inflammation leads to a progressive failure of the immune response, similar to a condition of immunodeficiency. The immune dysregulation in patients with LC can be defined as a "dysbiotic immune-inflammatory disorder", characterized by an abnormal local (gut and liver) and systemic inflammation, triggered by an impaired immune response to gut-derived antigens. The main feature of cirrhotic dysbiotic immune-inflammatory disorder is a perpetual immunologic activation, which involves all the immune cells (neutrophils, monocytes, T and B lymphocytes) that exhibit activation and co-stimulatory markers.

At the molecular level, the recognition of PAMPs by Toll-like receptors (TLRs) activates MyD88-dependent and MyD88-independent signaling pathways, leading to the activation of NF- κ B, production of inflammatory cytokines (tumor necrosis factor (TNF)- α , interleukin (IL)-6, IL-1 β and interferon (IFN) β), chemokines (keratinocyte chemoattractant (CXCL1), MIP-2 (CXCL2), MCP-1 (CCL2), RANTES (CCL5), MIP-1 α (CCL3) and MIP-1 β (CCL4)), nitric oxide (NO) and reactive oxygen species (ROS)[35].

The association of TLRs gene polymorphism with a decrease in the inflammatory response was established, which further increases the load of circulating bacterial antigens that modulate the immune response and contribute to the development of complications[36]. Cytosolic NOD-like receptors (NLRs) are also involved in this process. The NLRP3 inflammasome formed after the oligomerization of the NLRP3 protein activates caspase 1, which cleaves pro-IL-1 β and pro-IL-18, followed by the formation of pro-inflammatory cytokines IL-1 β and IL-18[37].

A cascade of molecular events arising from dysbiotic immuno-inflammatory disorders leads to the enhanced phagocytic activity, vascular endothelial injury, synthesis of acute phase proteins by the liver, chemotaxis of leukocytes to the sites of inflammation, mainly the liver, and activation of leukocytes at the systemic level[38]. This in itself worsens bacterial translocation and contributes to the formation of a vicious circle, which can aggravate the pathological process associated with acute decompensation of LC and predispose to the development of its characteristic complications.

GUT MICROBIOTA AS A POTENTIAL TARGET FOR PROPHYLAXIS AND THERAPY OF ACUTE DECOMPENSATION IN LIVER CIRRHOSIS

In accordance with the current recommendations of the European Association for the Study of the Liver (EASL), one of therapeutic strategy that prevents disease progression in patients with decompensated LC should be aimed to improve the microbiome abnormalities and bacterial translocation to ameliorate the impaired gut-liver axis [39]. In this regard, a potential target for therapy may be the gut microbiota, which is the main regulator of bacterial translocation and systemic inflammation[40].

Antibiotics

The use of non-absorbable or poorly absorbable oral antibiotics is an obvious solution aimed at countering bacterial translocation. They affect the gut microbiota with rare side effects and a favorable long-term safety profile, and are recommended as primary and secondary prevention of bacterial infections and treatment of hepatic encephalopathy in patients with decompensated LC[41].

Selective decontamination of the intestine with norfloxacin can contribute to a significant reduction in bacterial translocation. In a study by Albillos *et al*[42], this was manifested by a reduction in the serum levels of lipopolysaccharide-binding protein, soluble CD14, pro-inflammatory cytokines TNF- α , IL-12 and IFN- γ , as well as metabolites NO. In a multicenter, randomized, prospective, double-blind, placebo-controlled trial in parallel groups (NORFLOCIR), including 291 patients with CTP class C LC, the administration of norfloxacin at a dose of 400 mg once daily for 6 months significantly decreased the incidence of any and Gram-negative bacterial infections without growth infections caused by *Clostridium difficile* or multiresistant bacteria and an increase in survival in patients with ascites fluid protein concentrations <15 g/L[43].

At the same time, it turned out that long-term use of norfloxacin increased the gut microbiota resistance to fluoroquinolones[44]. Given this, a semisynthetic broad-spectrum antibiotic rifaximin was proposed as an alternative. Rifaximin belong to the family of naphthalene-ringed ansamycins (rifamycins group) and has a low risk of bacterial resistance. In a randomized controlled trial (RCT) involving 36 patients with decompensated LC with ascites and mean values of the MELD score of 12 \pm 3.9, after 4 weeks of treatment with rifaximin at a dose of 550 mg twice a day, circulating markers of inflammation, including TNF- α , IL-6, IL-10 and IL-18, stromal cell factor-1 α , transforming growth factor β 1, and high sensitivity C-reactive protein, were unaltered. Rifaximin altered abundance of bacterial taxa in blood marginally, only a decrease in *Pseudomonadaceae* family was observed. In faeces, rifaximin decreased the bacterial richness, but no affecting the particular species was observed[45].

In an observational study involving 30 patients with decompensated LC, after 4 weeks of treatment with rifaximin at a dose of 1200 mg/day, there was an improvement in hyperammonemia and cognitive dysfunction, although no

significant changes in the serum pro-inflammatory cytokine levels were observed. Rifaximin reduced the serum levels of ammonia, bacterial endotoxin, soluble CD163 and the D-mannose receptor. At the same time, the serum pro-inflammatory cytokine levels remained the same. ¹⁴ Gut microbial analysis revealed that the richness and complexity of species were unchanged while the abundance of the genus *Streptococcus* spp. after treatment with rifaximin was reduced [46].

The currently literature data do not provide a clear answer which of the antibiotics is more effective in preventing bacterial translocation in patients with decompensated LC. Nevertheless, in a randomized, double-blind, placebo-controlled trial by Kulkarni *et al*[47], primary prophylaxis with oral norfloxacin (400 mg/day for 30 days) effectively prevented bacterial infections in patients with ACLF. Furthermore, a systematic review and meta-analysis of 17 RCTs showed that ¹⁴ rifaximin is useful for both primary and secondary prevention of SBP whereas norfloxacin daily and alternate norfloxacin and rifaximin are useful for primary prophylaxis[48].

Probiotics

A scientific basis for the use of probiotics in the treatment of liver diseases is their ability to correct gut dysbiosis, elevate the production of short-chain fatty acids and reduce the increased permeability of the intestinal epithelial barrier[49]. The therapeutic potential of probiotics in LC has been studied in both experimental and clinical studies. For example, oral administration of a combined probiotic ²⁵ VSL#3 containing 8 different strains (*Bifidobacterium breve*, *Bifidobacterium longum*, *Bifidobacterium infantis*, *Lactobacillus acidophilus*, *Lactobacillus plantarum*, *Lactobacillus casei*, *Lactobacillus bulgaricus*, *Streptococcus thermophilus*) to rats with different models of LC led to stabilization intestinal epithelial barrier, reduction of bacterial translocation, decrease in severity of endotoxemia and systemic inflammation[50; 51]. Oral administration of probiotics *Bifidobacterium pseudocatenulatum* CECT7765 to mice with a model of CCl₄-induced LC was accompanied by an improvement in the integrity of the intestinal epithelial barrier and prevented bacterial translocation[52]. ²³ It also induced a morphologic, phenotypic and functional transitional change towards an anti-inflammatory profile in blood-derived and ascitic fluid macrophages from patients with CTP class C LC, as well as Kupffer cells from rats with a model of

bile duct ligation (BDL)-induced LC[53]. The combined use of probiotics containing *Clostridium butyricum* and *Bifidobacterium infantis* in patients with HBV-related LC and minimal hepatic encephalopathy significantly decreased the pathogenic bacteria of the genus *Enterococcus spp.* and the *Enterobacteriaceae* family in gut microbiota, as well as reduced the circulating levels of bacterial translocation markers, decreased the permeability and damage of the intestinal epithelial barrier[54].

Some RCTs have studied the effect of probiotics on gut microbiota in patients with LC. In one of them, the administration of the probiotic beverage Yakult 400 (Y400), which contains *Lactobacillus casei* strain Shirota, twice a day during the first half of the four-week study contributed to the normalization of gut microbiota and improved liver function in patients with CTP class A alcoholic LC[55]. This probiotic was safe and effective in cirrhotic patients (CTP score ≤ 10) who took it three times daily for six months. It significantly reduced the plasma monocyte chemotactic protein-1, plasma IL-1 β (alcoholic LC), IL-17a and macrophage inflammatory protein-1 β (non-alcoholic LC), compared with placebo group. At the same time, no significant differences in intestinal permeability, bacterial translocation or metabolomic profile were observed[56].

Bajaj *et al*[57] showed that the administration of the probiotic *Lactobacillus rhamnosus* GG (LGG) for eight weeks to patients with LC (mean values of the MELD score of 8.6 ± 2.2) and minimal hepatic encephalopathy was safe and well tolerated, and also reduced the serum levels of bacterial endotoxin and TNF- α , decreased the relative abundance of *Enterobacteriaceae* family and increased the *Clostridiales Incertae Sedis* XIV and *Lachnospiraceae* families, without changes in cognitive dysfunction.

Daily intake for 6 months of a probiotic powder containing eight different bacterial strains (*Bifidobacterium bifidum* W23, *Bifidobacterium lactis* W52, *Lactobacillus acidophilus* W37, *Lactobacillus brevis* W63, *Lactobacillus casei* W56, *Lactobacillus salivarius* W24, *Lactococcus lactis* W19 and *Lactococcus lactis* W58) by cirrhotic patients (CTP score < 12) had a beneficial effect on immune function, but no any affecting the permeability of the intestinal epithelial barrier and bacterial translocation was observed[58]. In addition, it increased the relative abundance of bacteria of the species *Faecalibacterium prausnitzii*, *Syntrophococcus sucromutans*, *Bacteroides vulgatus*, *Alistipes shahii* and the genus *Prevotella spp.*, compared with placebo group. At the same time, the relative

abundance of bacteria of the species *Bifidobacterium bifidum*, *Lactobacillus acidophilus* and *Lactobacillus casei* remained unchanged[59].

Thus, although in most studies the probiotic use in LC is associated with an improvement in gut microbiota profile, data concerning their impact on permeability of the intestinal epithelial barrier, bacterial translocation and systemic inflammation are scarce and contradictory.

Fecal microbiota transplantation

In recent years, numerous studies have demonstrated the therapeutic possibilities of fecal microbiota transplantation (FMT) from healthy donors to patients with chronic liver diseases[60]. It is assumed that the effectiveness of FMT is associated with the creation of a competitive environment in the intestine due to non-pathogenic microorganisms and their production of antimicrobial substances, such as bacteriocins. In addition, the positive effect of donor fecal material on the gut virome and microbiota, the metabolism of short-chain fatty acids and some bile acids, as well as various immune mechanisms is not excluded[61]. The much attention has been paid to the use of FMT to treat the hepatic encephalopathy in LC. At the same time, the issues of its effectiveness, safety and tolerability, as well as methods of administration of donor fecal material (using enemas, colonoscopy or in encapsulated form), the type and number of transplanted microorganisms necessary to obtain a positive result are discussed[62].

In the first an open-label RCT involving 10 patients with LC (MELD score <17) and recurrent hepatic encephalopathy, ²⁸ three frozen-then-thawed FMT units (90 mL total) instilled by enema and retained for 30 minutes eliminated antibiotic-induced dysbiosis. All patients showed an improvement in cognitive dysfunction, which may have been associated with an increase of relative abundance of bacteria of the *Lactobacillaceae* and *Bifidobacteriaceae* families[63]. With further follow-up (12.9±2.9 months), no cases of hepatic encephalopathy were detected and only one patient from this cohort required hospitalization. Microbiological analysis of the gut microflora showed an increase of relative abundance of bacteria of the *Burkholderiaceae* family and a decrease of relative abundance of bacteria of the *Acidaminococcaceae* family, while the relative abundance of bacteria of the

Lactobacillaceae and *Bifidobacteriaceae* families did not differ from the placebo group[64].

In RCT of phase 1, the administration of 15 capsules with donor fecal microbiota to 10 patients with LC (MELD score <17) and recurrent hepatic encephalopathy had a positive effect on cognitive dysfunction, was safe and well tolerated. After 30 days of monitoring, there was an improvement in duodenal mucosal microbial diversity with higher *Ruminococcaceae* and *Bifidobacteriaceae* families and lower *Streptococcaceae* and *Veillonellaceae* families (P = 0.01). Reduction in *Veillonellaceae* family were seen post-FMT in sigmoid (P = 0.04) and stool (P = 0.05). Duodenal E-cadherin (P = 0.03) and defensin A5 (P = 0.03) increased, while the IL-6 (P = 0.02) and serum levels of lipopolysaccharide-binding protein (P = 0.009) reduced post-FMT[65].

An important problem of FMT is the risk of severe infection transmission, what is especially significant in patients with weakened immunity[66]. For this reason, the US Food and Drug Administration (FDA) in 2019 published a list of minimum requirements for screening and testing of fecal microbiota donors for the presence of multidrug-resistant microorganisms[67].

The COVID-19 pandemic has raised concerns about the possible transmission of the SARS-CoV-2 in FMT. Although the genetic material of SARS-CoV-2, including the live virus, has been detected in the feces of patients with a new coronavirus infection even after the elimination of respiratory symptoms [68], no actual cases of infection through donor fecal material were reported. Stool testing for SARS-CoV-2 is not currently widely available. Nevertheless, experts advocate screening donors for the presence of symptoms of a new coronavirus infection with quarantine of their fecal material during further monitoring of the disease[69].

Obeticholic acid

Obeticholic acid is a semisynthetic bile acid that, in addition to bacteriostatic activity, is an agonist of FXR and thus can modulate the gut microbiota. For example, oral administration of obeticholic acid to rats with a model of CCl₄-induced LC reduced the intestinal content of genus *Enterococcus spp.*[70] and decreased the bacterial translocation[71]. Besides, obeticholic acid prevented the increased expression of monocyte chemo-attractant protein-1 following stimulation with TNF- α and

lipopolysaccharides or TNF- α alone in liver sinusoidal endothelial cells and Kupffer cells respectively in rats with a model of thioacetamide-induced LC[72]. In these studies, obeticholic acid had a beneficial effect on the production of antimicrobial peptides by ileum epithelial cells, the expression of the tight junction proteins, intestinal inflammation, and liver fibrosis.

At present, the therapeutic potential of obeticholic acid in LC has been studied mainly on experimental models, and, for safety reasons, its use in patients with decompensated LC is still considered premature.

Carbon nanoparticles

The newly developed carbon-based enterosorbent Carbalive™ (Yaq-001, Yaqrit Limited, UK) has a high absorption capacity for bacterial toxins and, thus, may be a new strategy to counteract changes in gut microbiota and translocation of bacterial products in patients with decompensated LC. It is non-absorbable carbon nanoparticles with a tailored bimodal distribution of porous domains within the macroporous range (>50nm) and microporous range (<2nm) and a vast surface area. The biological significance of this is that, in addition to binding smaller mediators such as indoles, acetaldehyde, etc., carbon granules exhibit rapid adsorption kinetics for larger molecular weight factors, for example, bacterial endotoxin, exotoxins and cytokines. Yaq-001 was found to reduce liver injury, portal pressure and lipopolysaccharide-induced ROS production in an *in vivo* model of LC and ACLF[73]. Yaq-001 significantly increased the relative abundance of symbiont bacteria belonging to *Firmicutes* phylum and was decreasing the relative abundance of pathobiont bacteria belonging to *Bacteroidetes* phylum in fecal samples from rodents with a model of BDL-induced LC, despite the absence of a direct effect on bacterial growth kinetics[74]. In the first multicenter, randomized, double-blind, placebo-controlled trial of phase 2 (CARBALIVE:634579), 14 patients with decompensated LC with diuretic-responsive ascites and mean values of the MELD score of 12.4±0.9 received 4 g of Yaq-001 for 12 weeks. Yaq-001 was safe, well tolerated, contributed to the restoration of intestinal eubiosis and, by affecting the permeability of the intestinal epithelial barrier, weakened the severity of endotoxemia and systemic inflammation[75].

CONCLUSION

Given that particular ⁵alterations in the composition and function of gut microbiota play a crucial role in the pathogenesis of acute decompensation in LC, the study of the therapeutic possibilities of its modulation has emerged as one of the top concerns in modern hepatology. Despite the encouraging preliminary data, the majority of the suggested strategies have only been tested in animal models or in preliminary clinical trials; additional multicenter RCTs must demonstrate their efficacy in larger patient populations.

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