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***Basic Study***

**Thymoquinone enhances the antioxidant and anticancer activity of Lebanese propolis**

Sima AlDreini, Zaynab Fatfat, Najwa Abou Ibrahim, Maamoun Fatfat, Hala Gali-Muhtasib, Hala Khalife

**Abstract**

**BACKGROUND**

Reactive oxygen species (ROS) are produced by multiple cellular processes and are maintained at optimal levels in normal cells by endogenous antioxidants. In recent years, the search for potential exogenous antioxidants from dietary sources has gained considerable attention to eliminate excess ROS that is associated with oxidative stress related diseases including cancer. Propolis, a resinous honeybee product, has been shown to have protective effects against oxidative stress and anticancer effects against several types of neoplasms.

**AIM**

To investigate the antioxidant and anticancer potential of Lebanese propolis when applied alone or in combination with the promising anticancer compound Thymoquinone (TQ) the main constituent of *Nigella sativa* essential oil.

**METHODS**

Crude extracts of Lebanese propolis collected from two locations, Rashaya and Akkar-Danniyeh, were prepared in methanol and the total phenolic content was determined

by Folin–Ciocalteu method. The antioxidant activity was assessed by the ability to scavenge DPPH free radical and to inhibit H<sub>2</sub>O<sub>2</sub>-induced oxidative hemolysis of human erythrocytes. The anticancer activity was evaluated by MTT assay against HCT-116 human colorectal cancer cells and MDA-MB-231 human breast cancer cells.

## RESULTS

The total phenolic content of propolis extract from Rashaya and Akkar-Danniyeh were 56.81 µg and 83.503 µg of gallic acid equivalent /mg of propolis, respectively. Both natural agents exhibited strong antioxidant activities as evidenced by their ability to scavenge DPPH free radical and to protect erythrocytes against H<sub>2</sub>O<sub>2</sub>-induced hemolysis. They also dose-dependently decreased the viability of both cancer cell lines. The IC<sub>50</sub> value of each of propolis extract from Rashaya and Akkar-Danniyeh or TQ was 22.3, 61.7, 40.44 µg/mL for breast cancer cells at 72 h and 33.3, 50.9, 33.5 µg/mL for colorectal cancer cells at the same time point, respectively. Importantly, the inhibitory effects of propolis on DPPH radicals and cancer cell viability were achieved at half its concentration when combined with TQ.

## CONCLUSION

Our results indicate that Lebanese propolis extract has antioxidant and anticancer potential and its combination with TQ could possibly prevent ROS- mediated diseases.

## INTRODUCTION

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Oxidative stress refers to the imbalance between the generation of reactive oxygen species (ROS) and their neutralization by endogenous antioxidant systems resulting in an excess of ROS which has detrimental effects on key cellular components<sup>[1,2]</sup>. There are two types of ROS: free radicals and nonradicals. Free radicals are highly reactive molecules because they have at least one unpaired electron in their structure and react with different biological macromolecules<sup>[3]</sup>. Although nonradical species are less

reactive than free radicals, they can easily cause free radical reactions in living organisms<sup>[3,4]</sup>. The accumulation of ROS causes the peroxidation of cell membrane lipids and cell membrane disintegration, alters the configuration of proteins resulting in loss of biochemical functionality in addition to inducing DNA mutations and replication errors<sup>[2]</sup>. Ample evidence shows that ROS-mediated oxidative stress is associated with the pathogenesis of various diseases including cancer, cardiovascular diseases, neurodegenerative disorders, and diabetes<sup>[5]</sup>.

Removing excessive ROS by exogenous antioxidants supplementation has long been considered a potential strategy to prevent diseases. Over the last decade, there has been considerable interest in the intake of natural antioxidants from food and diets to strengthen cell antioxidant defense in humans. A recent pilot study demonstrated that a healthy mixed diet rich in antioxidant micronutrients reduced the concentration of ROS in the blood of healthy subjects<sup>[6]</sup>. Another study showed that regular consumption of an antioxidant-rich juice increased plasma antioxidant capacity and reduced plasma lipid oxidation in healthy individuals<sup>[7]</sup>. In addition, several clinical trials showed that intake of foods rich in antioxidants can potentiate plasma antioxidant capacity and reduce oxidative stress markers in subjects with diabetes, obesity, and dyslipidemia<sup>[8]</sup>. Interestingly, the combination of several antioxidants has been suggested to be more potent than the application of single antioxidants given the diverse chemistry and biochemistry of ROS, and the interactions that could arise from antioxidants that have different modes of action<sup>[9]</sup>.

Propolis is a glue-like resinous material produced by honeybees from various plant sources and used in the construction and maintenance of their hives<sup>[10,11]</sup>. Propolis possesses numerous health-promoting potentials including anti-inflammatory<sup>[12]</sup>, antioxidant<sup>[13]</sup>, anticancer<sup>[14]</sup> and antidiabetic effects<sup>[15]</sup>. The chemical composition and therefore the biological effects of propolis vary depending on several factors such as the geographical region, botanical source, and the bee species<sup>[16]</sup>. The bioactive compounds

of propolis were reported to effectively scavenge free radicals<sup>[17]</sup>. Different *in vivo* studies reported the protective effects of propolis against the oxidative stress induced by several exogenous oxidants such as cisplatin<sup>[18]</sup>, isoproterenol<sup>[19]</sup>, nicotine<sup>[20]</sup>, UV<sup>[21]</sup>, and carbon tetrachloride<sup>[22]</sup>. In addition, propolis was demonstrated to reduce the blood pressure and suppress oxidative stress in heart, liver, and renal tissues in animal models of hypertension<sup>[23-25]</sup>.

Thymoquinone (TQ), the major bioactive constituent of *Nigella sativa* (black seed) essential oil, was extensively studied for its diverse therapeutic benefits including antioxidant, anti-inflammatory, anticancer, antibacterial, antifungal and anticonvulsant activity<sup>[26]</sup>. TQ was reported as a strong scavenger of different ROS and was found to inhibit non-enzymatic lipid peroxidation<sup>[27]</sup>. TQ was demonstrated to have a protective effect against oxidative stress induced in rats by different agents such as radiation<sup>[28]</sup>, lead<sup>[29]</sup> and acrylamide<sup>[30]</sup>. In addition, it reduced the oxidative stress in rat models of myocardial infarction<sup>[31]</sup>, diabetes mellitus<sup>[32]</sup>, lung injury<sup>[33]</sup> and dopaminergic neurodegeneration<sup>[34]</sup>.

Although the antioxidant potential of propolis and TQ has been well investigated in previous studies, there are no studies that have evaluated the antioxidant and anticancer effects of the combination of both natural agents. Thus, we aimed to test the antioxidant and anticancer activities of combining TQ and propolis that was collected from two locations in Lebanon (Rashaya and Akkar-Danniyeh). We evaluated the total phenolic content of both propolis extract and determined the antihemolytic and antioxidant activity of propolis and TQ in addition to their anticancer effects against HCT-116 human colorectal cancer cells and MDA-MB-231 human breast cancer cells.

## **MATERIALS AND METHODS**

### ***Preparation of Thymoquinone***

Fresh stocks of the purified synthetic compound TQ (Sigma-Aldrich) were prepared in methanol directly before use.

#### ***Preparation of methanol extracts of propolis***

Two samples of raw propolis material were collected, the first from Rashaya district in the Beqaa governorate of Lebanon and the second from Akkar-Danniyeh in the north of the country. A mass of 10 g of raw propolis from each sample was chopped into small pieces and extracted with 100 mL distilled water. The extraction was carried at 80 °C for 3 h and the obtained solution was subsequently filtered through a Buchner funnel. Residues were then extracted with 100 mL methanol. The extraction was carried at room temperature for 4 h then at 50 °C for 15 min. The propolis extracts were subsequently filtered three times by Buchner funnel. The obtained filtrate was evaporated by nitrogen gas to obtain the methanol propolis extract (MPE). MPE-R denotes MPE from Rashaya and MPE-D denotes MPE from Akkar-Danniyeh.

#### ***Total phenolic content***

The relative content in phenols was determined according to the Folin Ciocalteu method. Briefly, 100 µL of MPE-R or MPE-D (1mg/mL of methanol) from each sample were mixed with 500 µL of Folin Ciocalteu's phenol reagent 10%. After 5 min, 1.5 mL of 2% sodium bicarbonate were added to the solution. The mixture was maintained at room temperature in the dark for 30 min after which the absorbance was recorded at 760 nm using a spectrophotometer. The total phenolic content was calculated using the calibration curve generated from standard solutions of gallic acid ranging from 0 to 50 µg/mL ( $y = 0.2811x - 0.3266$ ;  $R^2 = 0.956$ ). Total phenolic content was expressed as the average of 3 independent experiments performed in triplicates and as µg of gallic acid equivalents (GAE)/mg of propolis.

#### ***DPPH assay***

Free radical-scavenger activity was determined by the 2,2-diphenyl-1-picrylhydrazyl (DPPH) assay. Briefly, 1 mL of MPE-R or MPE-D (10-100 µg/mL) were mixed with 1 mL of DPPH (0.052 mg/mL methanol). The reaction mixtures were homogenized and incubated in the dark at room temperature for 30 min and the absorbance (Abs) was

measured at 515 nm by a Gene Quant 1300 UV-Vis spectrophotometer. The ascorbic acid was used as a reference antioxidant and a mixture of 1 mL DPPH with 1 mL methanol was used as a control.

For combination treatments, TQ (12.5-100 µg/mL) was combined with MPE-R or MPE-D (10- 50 µg/mL) and the experiment was carried as described above.

The DPPH scavenging ability of the different agents was calculated using the following equation:

$$\% \text{ DPPH inhibition} = [(\text{Abs control} - \text{Abs sample})] / (\text{Abs control}) \times 100.$$

#### ***H<sub>2</sub>O<sub>2</sub>- induced hemolysis***

Fresh human blood was washed three times with 1X phosphate-buffered saline (PBS). With every wash, the sample was centrifuged for 12 min at 4 °C and 2500 rpm, the supernatant was discarded, and the pellet was resuspended in PBS. Then, the pellet was resuspended in Dulbecco's PBS and 1 mL of the cell suspension was mixed with 100 µL of each of MPE-R, MPE-D or TQ at 10, 50, and 100 µg/mL. After 5 min, 1 mL of 10% H<sub>2</sub>O<sub>2</sub> was added, and the mixture was incubated at 37 °C for 90 min and shaken every 30 min. This was followed by centrifugation at 4 °C and 2500 rpm for 10 min and measurement of the absorbance of the supernatant at 540 nm. The positive control consisted of a mixture of blood with 10% H<sub>2</sub>O<sub>2</sub>. The results were expressed as percentage of inhibition of hemolysis.

$$\% \text{ inhibition of hemolysis} = [(\text{Abs control} - \text{Abs sample})] / (\text{Abs control}) \times 100$$

#### ***Hemolytic activity***

Fresh human blood was washed three times with 1X PBS. With every wash, the sample was centrifuged at 4 °C and 2500 rpm for 12 min, the supernatant was discarded, and the pellet was resuspended in PBS. The washed blood was mixed with each of MPE-R or MPE-D (10, 100, 200 µg/mL), TQ (20, 50, 100 µg/mL) or their combinations. The mixture was kept at 37 °C for 90 min and was shaken every 30 min. The samples were then centrifuged at 4 °C, 2500 rpm, for 10 min after which the absorbance of the supernatant was recorded at 540 nm. The positive control consisted of a mixture of

blood with 1% SDS which is known to cause hemolysis. The results were expressed as the percentage of hemolysis.

$$\% \text{ hemolysis} = [\text{Abs sample} / \text{Abs control}] \times 100$$

#### ***Cell culture conditions***

HCT-116 human colorectal cancer cells and MDA-MB-231 human breast cancer cells were maintained in RPMI 1640 (Lonza; Cat.N: BE12-115F) supplemented with 10% fetal bovine serum (Sigma F9665) and 1% penicillin/ streptomycin (Sigma, P4333) in a humidified atmosphere at 37 °C in 5% CO<sub>2</sub>.

#### ***MTT cell viability assay***

HCT-116 and MDA-MB-231 cells were seeded overnight in 96-well microtiter plates at a density of 10<sup>4</sup> cells/well. After 24 h, cells were treated with MPE-R, MPE-D or TQ at a concentration ranging from 1-15 µg/mL or with the combination of MPE-R or MPE-D (0.5-7.5 µg/mL) with TQ (0.5-7.5 µg/mL). After 24, 48 and 72 h of treatment, cells in each well were incubated with 20 µL of MTT (3-(4, 5-dimethylthiazolyl-2)-2, 5-diphenyltetrazolium bromide) for 3 to 4 h, then with 100 µL of DMSO for about 1 h. The MTT optical density (O.D.) was then measured by a microplate spectrophotometer at 515 nm. The results are expressed as percentage of viable cells with respect to the untreated control using this formula:

$$\% \text{ viability} = [\text{mean O.D. treatment} / \text{mean O.D. control}] \times 100$$

#### ***Statistical analysis***

Data are presented as means ±SD. Two tailed Student's t-test was performed to evaluate the statistical significance of the difference between the groups using GraphPad Prism V.9.5.0 software. Statistical significance was set with a 95% confidence interval at  $P < 0.05$ ,  $P < 0.01$  and  $P < 0.0001$ .

## **RESULTS**

### ***Total phenolic content of propolis varies depending on location***

The total phenolic content of propolis extracts was determined by Folin Ciocalteu method and is reported as gallic acid equivalents by reference to a standard curve ( $y =$



$0.2811x - 0.3266$ ;  $R^2 = 0.956$ ). The phenolic content was variable depending on location such that the total phenolic content of MPE- D in 1 mg of propolis was 47% higher than that of MPE-R (Table1).

#### ***TQ enhanced the antioxidant activity of propolis***

We then evaluated the ability of propolis extracts to scavenge free radicals using DPPH radical scavenging assay. Both propolis extract exhibited a dose-dependent DPPH inhibition efficiency suggesting antioxidant potential. MPE-D had higher antioxidant activity than MPE-R as reflected by the higher percentages of inhibition recorded at all the concentrations ranging from 20-100  $\mu\text{g/mL}$ . MPE-R showed maximum inhibition of DPPH of 56.5% at 100  $\mu\text{g/mL}$ , while inhibition by MPE-D reached 89% at 75 and 100  $\mu\text{g/mL}$  (Figure 1A).

To determine whether the antioxidant effects of propolis extracts could be potentiated by TQ, we combined each of MPE-R or MPE-D (10-50  $\mu\text{g/mL}$ ) with TQ (12.5-100  $\mu\text{g/mL}$ ) and evaluated their antioxidant activity in comparison to single treatments. Results showed that the combination with TQ enhanced the antioxidant activity of propolis extracts. While a dose of 100  $\mu\text{g/mL}$  of MPE-R induced 56.5% inhibition of DPPH, the combination of 50  $\mu\text{g/mL}$  of MPE-R with 100  $\mu\text{g/mL}$  TQ caused 85.7% inhibition. MPE-D alone showed maximal inhibitory effects of 89% at 75-100  $\mu\text{g/mL}$ , while combination with 50-100  $\mu\text{g/mL}$  TQ resulted in 84% inhibition at lower concentrations of 25-50  $\mu\text{g/mL}$  (Figure 1B).

#### ***Propolis extracts and TQ protected human red blood cells against oxidative hemolysis***

We then evaluated the biological relevance of the antioxidant activity of propolis extracts and of TQ by testing the protective effects of single treatments against oxidative hemolysis induced by  $\text{H}_2\text{O}_2$  in human red blood cells. Treatment with MPE-R, MPE-D or TQ exhibited good antihemolytic potential against  $\text{H}_2\text{O}_2$ -induced hemolysis. A dose of 10  $\mu\text{g/mL}$  of each of MPE-R, MPE-D and TQ induced 46, 49 and 51% decrease in hemolysis, respectively (Figure 2).

#### ***The combination of propolis extract with TQ had no hemolytic activity at low concentrations***

To investigate if propolis extracts or TQ are toxic to human red blood cells, we evaluated their hemolytic potential at concentrations ranging from 10- 200  $\mu\text{g/mL}$  and 20- 100  $\mu\text{g/mL}$ , respectively. Both MPE-R and MPE-D produced less than 5% hemolysis at 10  $\mu\text{g/mL}$ , suggesting that these extracts are not toxic to red blood cells at this concentration. Increasing concentrations of MPE-D up to 100 or 200  $\mu\text{g/mL}$  also showed low hemolytic activity of 7.8%. Similarly, hemolysis by TQ was less than 5% at all the tested concentrations. However, MPE-R induced higher hemolytic response that reached 20% at 200  $\mu\text{g/mL}$  (Figure 3A).

Combining 5  $\mu\text{g/mL}$  MPE-R or MPE-D with 10  $\mu\text{g/mL}$  TQ produced less than 5% hemolysis suggesting that combinations at these low doses have low hemolytic effects. However, increasing concentrations to 50  $\mu\text{g/mL}$  MPE-R and 25  $\mu\text{g/mL}$  TQ or 100  $\mu\text{g/mL}$  MPE-R and 50  $\mu\text{g/mL}$  TQ produced 12.7% and 21.9% hemolysis, respectively. Similar concentrations of MPE-D and TQ produced 7.3% and 13.7% hemolysis, respectively (Figure 3B), suggesting that MPE-R had higher hemolytic effects when combined with TQ at higher doses.

#### ***TQ potentiated the inhibitory effects of propolis extracts on cancer cell viability***

Next, we tested the anticancer activity of propolis extracts when applied alone or in combination with TQ. MDA-MB-231 human breast cancer cells and HCT-116 human colorectal cancer cells were treated with different concentrations for 24, 48 and 72 h after which cell viability was assessed by MTT assay. Single treatments with MPE-R or MPE-D (1-15  $\mu\text{g/mL}$ ) reduced the viability of both cell lines in a dose dependent manner to almost similar levels. Treatment of MDA-MB 231 cells with 15  $\mu\text{g/mL}$  of MPE-R, MPE-D or TQ for 72 h caused 34.6%, 18.5% and 24.52% reduction in cell viability, respectively. The  $\text{IC}_{50}$  value of each of MPE-R, MPE-D or TQ at 72 h was 22.3, 61.7, 40.44  $\mu\text{g/mL}$ , respectively. Combining lower doses of 7.5  $\mu\text{g/mL}$  MPE-R or MPE-D with 7.5  $\mu\text{g/mL}$  TQ decreased cell viability by 48.9% and 39.3%, respectively (Figure 4A and 4B), suggesting enhanced efficacy by combination treatment.

Treatment of HCT-116 cells for 72 h with 15 µg/mL of MPE-R, MPE-D or TQ decreased cell viability by 18.6, 14.3 and 26%, respectively. The IC<sub>50</sub> value of each of MPE-R, MPE-D or TQ at 72 h was 33.3, 50.9, 33.5 µg/mL, respectively. Interestingly, the combination of half doses MPE-R or MPE-D (7.5 µg/mL) with 7.5 µg/mL TQ caused a respective decrease in viability of 40.9% and 34.4% at 72 h (Figure 5A and 5B). Thus, combining propolis extracts with TQ enhanced their anticancer activities against breast and human colon cancer cells.

## **DISCUSSION**

The intake of dietary antioxidants is known to support the endogenous antioxidant system and prevent oxidative stress-mediated diseases<sup>[35]</sup>. Studies have shown that combining dietary antioxidants from different sources produces more potent antioxidant effects and possibly more effective therapeutic potential than single agents. Combining *Nigella sativa* oil with honey was shown to augment its antioxidant capacity<sup>[36]</sup>. In addition, the combination of *Nigella sativa* seeds and honey exhibited antioxidant effects and decreased the viability of ovarian cancer cells<sup>[37]</sup>. Interestingly, oral intake of honey potentiated the protective effect of *Nigella sativa* grains against methylnitrosourea-induced oxidative stress and carcinogenesis in Sprague Dawley rats<sup>[38]</sup>. Here, we evaluated the antioxidant and the anticancer potential of combining propolis, the third most important component of bee products<sup>[39]</sup>, with TQ as the major bioactive constituent of *Nigella sativa* essential oil. The key finding of the present study is that combining TQ with Lebanese propolis at half its concentration resulted in an enhanced antioxidant and anticancer effects in comparison to propolis alone as demonstrated by the improved DPPH radical scavenging activity and inhibitory effects against breast and colorectal cancer cell lines.

First, we assessed the total phenolic content of propolis collected from two different Lebanese regions Rashaya and Akkar- Danniye. The phenolic content is the most widely investigated among all the components of propolis because it was reported to be

mainly responsible for its biological activity<sup>[40]</sup>. According to the results reported by El-Ali *et al* (2021), the total phenolic content of ethanol extract of propolis collected from the two Lebanese regions Debaal and Wadi Faara were similar to our study's finding<sup>[41]</sup>. On the other hand, higher phenolic content values were recorded in the ethanol extract of propolis collected from the Lebanese regions Fakeha and Berqayel and the citrus groves of the Lebanese coast<sup>[41,42]</sup>. This variation in total phenolic content of propolis collected from different Lebanese regions could be attributed to several factors including the botanical origin of the raw material, mode of collection, collecting season, or the solvent used in the extraction method<sup>[40]</sup>.

Next, we assessed the antioxidant activity of MPE-R and MPE-D alone or in combination with TQ using DPPH free radical scavenging test. DPPH is a stable nitrogen-centered free radical which color changes from violet to yellow when it receives a hydrogen- or electron- from an antioxidant<sup>[43]</sup>. MPE-R and MPE-D exhibited significant DPPH scavenging efficacy reflecting the presence of antioxidants within their constituents. Numerous studies reported a positive correlation between antioxidant activity of propolis extracts and their contents of phenolic compounds suggesting that they are responsible of the antioxidant activity of the extracts<sup>[41,44,45]</sup>. Phenolics are known to have a hydroxyl group attached to their aromatic ring which can donate electron to free radicals and therefore stabilize them<sup>[46]</sup>. As for TQ which is a non-phenolic compound, a recent computational study reported that it attacks free radical preferentially at its 3CH position and preferably *via* hydrogen atom transfer<sup>[47]</sup>.

After demonstrating the antioxidant efficacy of each of MPE-R, MPE-D and TQ, we assessed their potential to protect red blood cells from oxidative damage and hemolysis induced by H<sub>2</sub>O<sub>2</sub>. Red blood cells are highly prone to oxidative damage due to its high membrane concentration of polyunsaturated fatty acids<sup>[48]</sup>. When the membrane lipids of red blood cells are subjected to ROS attack, they lose a hydrogen atom from an unsaturated fatty acyl chain. This initiates lipid peroxidation that propagates as a chain

reaction and lead to membrane damage and consequently hemolysis<sup>[49,50]</sup>. Our findings are in the same line with previous research that has shown the anti-hemolytic activity of propolis or TQ under oxidative stress conditions<sup>[40,51]</sup>. The antihemolytic activity of MPE-R and MPE-D could be associated with their phenolic content. Phenolic compounds are supposed to donate electrons to hydrogen peroxide, neutralize it to water and prevent it to induce hemolysis<sup>[52]</sup>.

The assessment of hemolytic activity of blood-contacting compounds is of high importance for their future application *in vivo*<sup>[53]</sup>. Our results are in agreement with those reported by Shubharani *et al* who showed that low concentrations of ethanolic extract of Indian propolis did not have hemolytic activity<sup>[54]</sup>. Although high concentrations of Lebanese propolis showed low to moderate hemolysis, same concentrations of Polish or Brazilian propolis extract did not cause hemolysis<sup>[40,55]</sup>.

Cancer cells exhibit elevated levels of ROS which promote cell cycle progression and lead to an increase in cell proliferation<sup>[56]</sup>. By-products of oxidative damage such as 8-hydroxy-2-deoxyguanosine, malondialdehyde, 4-hydroxy-2-nonenal, and carbonylated proteins were speculated to play a mutagenic role<sup>[57]</sup>. In addition, oxidative stress was found to be responsible for inactivation of several key proteins such as caspases, phosphatases, and phosphatase and tensin homologue, and inhibits p53 binding to gene promoters which reduce apoptosis and increase cell survival<sup>[58]</sup>. Dietary antioxidants have been demonstrated to have chemopreventive and anticancer effects *in vitro* and *in vivo*<sup>[59]</sup>. Numerous studies demonstrated the anticancer effect of each of TQ, propolis and its phenolic compounds in different types of cancer<sup>[60,61]</sup>. To our knowledge, this is the first study that demonstrates the promising anticancer effect of the combination of these agents. Only one study demonstrated the anticancer effect of Lebanese propolis collected from the south of the country on leukemic T cells<sup>[10]</sup>. Although MPE-D had higher antioxidant activity than MPE-R, the inhibitory effect of both extracts on the cell

viability of cancer cell lines was almost the same. This result suggests that phenolic compounds may not be responsible for this inhibitory effect of the extracts.

## **CONCLUSION**

In summary, the Lebanese propolis from Rashaya and Akkar-Danniyeh exhibited promising potential as reflected by their potent DPPH radical scavenging activity, protective effects against H<sub>2</sub>O<sub>2</sub> induced hemolysis and inhibitory effects against breast and colorectal cancer cell lines. The combination of TQ with propolis resulted in enhanced antioxidant and anticancer activities in comparison to single treatments. Thus, this combination could have potential health benefits and holds promise for the prevention of oxidative stress related diseases. Further studies should be conducted to analyze the chemical composition of propolis, decipher the antioxidant and anticancer mechanism of its combination with TQ in addition to evaluating the effects of TQ and propolis in animal models of oxidative stress.

## **ARTICLE HIGHLIGHTS**

### ***Research background***

Oxidative stress is implicated in the pathogenesis of numerous diseases including cancer. Propolis, the third most important component of bee products, and Thymoquinone (TQ), the main constituent of *Nigella sativa* essential oil, were extensively reported to have antioxidant and anticancer effects. However, the antioxidant potential of the combination of these natural products as well as their anticancer activity against breast and colorectal cancer cells have not been investigated yet.

### ***Research motivation***

To establish a new therapeutic approach for oxidative stress induced cancers using a combination of natural agents from food and diets.



### ***Research objectives***

To investigate the antioxidant and anticancer potential of Lebanese propolis and TQ alone and in combination.

### ***Research methods***

<sup>8</sup> Folin–Ciocalteu method was used to determine the total phenolic content of the methanolic extract of propolis. The DPPH free radical assay and the H<sub>2</sub>O<sub>2</sub>-induced <sup>1</sup> oxidative hemolysis of human erythrocytes *in vitro* assay were employed to assess the antioxidant activity of TQ and Lebanese propolis. The MTT assay was used to evaluate the anticancer activity of these natural agents in single and dual treatment against HCT-116 human <sup>10</sup> colorectal cancer cells and MDA-MB-231 human breast cancer cells *in vitro*.

### ***Research results***

Combination of TQ with Lebanese propolis at half its concentration improved the antioxidant and anticancer activity of propolis as reflected by the enhanced DPPH radical scavenging activity and inhibitory effects against breast and colorectal cancer cells.

### ***Research conclusions***

Our results suggest the use of a combination of TQ and Lebanese propolis as potential therapy for the management of oxidative stress and treatment of breast and colorectal cancer. This is the first study to report the promising enhancement in Lebanese propolis antioxidant and anticancer activity when combined with TQ.

### ***Research perspectives***

Further research on the antioxidant and anticancer mechanisms of the combination of these natural agents and its therapeutic effects in animal models of oxidative stress should be performed in the future.

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