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**Challenges and exploration for immunotherapies targeting cold colorectal cancer**

Li DD *et al*. Immunotherapies of cold CRC

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**Abstract**

In recent years, immune checkpoint inhibitors (ICIs) have made significant breakthroughs in the treatment of various tumors, greatly improving clinical efficacy. As the fifth most common antitumor treatment strategy for patients with solid tumors after surgery, chemotherapy, radiotherapy and targeted therapy, the therapeutic response to ICIs largely depends on the number and spatial distribution of effector T cells that can effectively identify and kill tumor cells, features that are also important when distinguishing malignant tumors from “cold tumors” or “hot tumors”. At present, only a small proportion of colorectal cancer (CRC) patients with deficient mismatch repair (dMMR) or who are microsatellite instability-high (MSI-H) can benefit from ICI treatments because these patients have the characteristics of a “hot tumor”, with a high tumor mutational burden (TMB) and massive immune cell infiltration, making the tumor more easily recognized by the immune system. In contrast, a majority of CRC patients with proficient MMR (pMMR) or who are microsatellite stable (MSS) have a low TMB, lack immune cell infiltration, and have almost no response to immune monotherapy; thus, these tumors are “cold”. The greatest challenge today is how to improve the immunotherapy response of “cold tumor” patients. With the development of clinical research, immunotherapies combined with other treatment strategies (such as targeted therapy, chemotherapy, and radiotherapy) have now become potentially effective clinical strategies and research hotspots. Therefore, the question of how to promote the transformation of “cold tumors” to “hot tumors” and break through the bottleneck of immunotherapy for cold tumors in CRC patients urgently requires consideration. Only by developing an in-depth understanding of the immunotherapy mechanisms of cold CRCs can we screen out the immunotherapy-dominant groups and explore the most suitable treatment options for individuals to improve therapeutic efficacy.

**Key Words:** Colorectal cancer; Immune checkpoint inhibitors; Cold tumors; Immunotherapy mechanism; Combination therapy; Effector T cells

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**Core Tip:** This review focuses on the potential mechanism of immunotherapy ineffectiveness in colorectal cancer patients with “cold tumors”, how to increase the infiltration of effector T cells at tumor sites by combination therapy strategies, how to transform “cold tumors” into “hot tumors” to expand the number of people benefiting from immunotherapy, and make recommendations on how to screen out immune-dominant populations in the future.

**INTRODUCTION**

As one of the most common gastrointestinal malignancies, colorectal cancer (CRC) ranks third in incidence and second in tumor-related mortality among common malignant tumors in the world[1]. With the in-depth exploration of the pathogenesis of CRC, immune checkpoint inhibitors (ICIs) have gradually entered the field as an antitumor treatment for CRC. Current tumor immunotherapy strategies include oncolytic virus (OV) therapy, monoclonal antibodies, ICIs, cytokine therapy, tumor vaccines, and adoptive cell therapy[2,3], and these strategies are characterized by the enhancement of innate and adaptive immunity to clear tumor cells, with the aim of inducing a durable and effective antitumor immune response. Regardless of whether the tumor type is classified as deficient mismatch repair (dMMR) or microsatellite instability-high (MSI-H) or proficient MMR (pMMR) or microsatellite stable (MSS), the antitumor effects of ICIs ultimately rely on CD8+ T cells and tumor-infiltrating lymphocytes, which have been confirmed in numerous studies to be important features that distinguish malignancies from “cold tumors” or “hot tumors”[4]. In a phase 2 clinical trial comparing the efficacy of dMMR or pMMR CRC with pembrolizumab immunomonotherapy, the median progression-free survival (mPFS) was 5 mo and 2 mo in patients with dMMR and pMMR, respectively. In addition, the objective response rate (ORR) was 40% for dMMR CRC, and there was no response for patients with pMMR CRC[5]. It can be concluded that patients with pMMR/MSS CRC who exhibit traits of “cold tumors” do not benefit from programmed cell death protein 1 (PD-1) inhibitor therapy[6,7]. The results of a meta-analysis by Li *et al*[8] also confirmed this conclusion. However, dMMR patients make up the vast majority of all CRC patients as accounting for 95%[9]. At present, the exploration of the reasons for the poor efficacy of immunotherapy in dMMR CRC patients and whether its efficacy can be improved has become a hot issue worldwide.

**Classification based on immunological characterizations**

In 2017, Chen and Mellman[6] pointed out that tumors can be divided into three immune phenotypes, the immune-inflamed, the immune-excluded and the immune-desert phenotypes, which are based on PD-1 ligand (PD-L1) expression, the total number of immune cells, and their spatial distribution in the tumor microenvironment ([Figure 1](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7206536/figure/coi200014f1/)). High levels of immunocyte infiltration, a high tumor mutational burden (TMB), and high interferon-γ (IFN-γ) signal transduction and PD-L1 expression are the characteristics that define immunoinflammatory tumors and are also key to the success of antitumor immunotherapy[10,11]. The immune-inflamed phenotype is characterized by a large number of immune cells in the tumor parenchyma, also known as a “hot tumor”, which is often sensitive to ICIs[12]. In contrast, although there are a large number of immune cells in the immune-excluded phenotype, the immune cells are mainly distributed in the stroma around the tumor rather than within the tumor. Tumors of this phenotype are less responsive to immunotherapy. The immune-desert phenotype is characterized by a lack of immune cells in both the parenchyma and stroma of the tumor; such tumors rarely respond to immunotherapy[6]. The immune-excluded and the immune-desert phenotypes also have a low TMB and low expression of histocompatibility complex class I (MHC I); these tumors are called “cold tumors”[11,13]. Studies have shown that poor sensitivity to immune checkpoint suppression monotherapy in patients with “cold tumors” is associated with congenital immune deficiency or the suppression of innate antitumor immunity[6,14]. It can be concluded that the most effective way to increase the response of “cold tumors” to immunotherapy is to eliminate barriers to tumor immunosuppression and immune escape mechanisms, thereby establishing or maintaining an adaptive immune response.

**Potential mechanism of cold tumor formation**

For effector T cells to exert antitumor immunity, a complete immune cycle process, in which many factors that can influence T cell priming and homing to the tumor site, including a lack of tumor antigens, a defect in antigen-presenting cells (APCs), a lack of T-cell activation and defects in homing to the tumor bed, is required. All of these processes can lead to a T cell-inflamed phenotype and antitumor immunity failure[14]. Therefore, an in-depth understanding of the various mechanisms of T cell infiltration impairment and how immunocombination therapy strategies affect the immune system’s anti-tumor ability is critical for understanding the transformation of “cold tumors” into “hot tumors” and may help to clarify which step of the antitumor immune response is not working; thus, the mechanism of action can be used to develop individualized treatment regimens.

***The formation mechanism of the immune-excluded phenotype***

This immunophenotype causes a poor immune response because more effector T cells cannot home from the tumor matrix to the inside of the tumor[15]. The underlying premise for the exertion of antitumor immune effects by effector T cells involves their migration to the tumor site, where their arrival involves multiple steps, as follows: immune cells first reach the blood vessels of tumor tissue through peripheral blood circulation and then penetrate tumor blood vessels to reach tumor tissue[16,17]. Chemokines, an important class of cytokines that affect the occurrence, development and angiogenesis of tumor cells, bind to chemokine receptors on the surface of effector T cells and play a key role in coordinating the transport of immune cells to the tumor core. The transport of immune cells into the tumor core occurs through endothelial adhesion and migration under the guidance of chemokines. Previous studies have shown that a variety of chemokines positive for various ligands can be involved in the recruitment of tumor T cells, including CC-chemokine ligand 4 (CCL4), CCL5, CXC-chemokine ligand 9 (CXCL9), CXCL10 and CXCL16, and that the lack or insufficiency of these chemokines can lead to the absence of immune cells in the tumor core[18-20]. For example, CXCL9 and CXCL10 participate in the recruitment of effector T lymphocytes in the human body mainly by binding to their corresponding receptors, and their expression levels determine the number of effector T lymphocytes infiltrating the tumor core. In the absence of CXCL9 and CXCL10 expression, effector T lymphocytes are not recruited into the tumor[20]. In human ovarian cancer cells, methylation of the CXCL9/10 genetic locus can result in immune evasion, and similar results were confirmed in a related study on colon cancer[21]. CCL5 is a typical cytokine secreted by normal T cells with chemotactic activity, and its increased expression can promote the transport of effector T cells into tumor cells[22]. This chemokine acts synergistically with IFN-γ-induced chemokines to establish an immuno-thermal phenotype, and studies have concluded that the cooperation between CCL5 and CXCL9 elucidates an important mechanism of tumor rejection to the T cell response[23]. However, certain chemokines such as CXCL1, CXCL2, and CXCL3 exert their effect by recruiting subsets of immunosuppressive T cells, such as myeloid-derived suppressor cells (MDSCs), to the tumor site, while CXCL12 mediates effector T lymphocytes into the tumor stroma, thus excluding them from recruitment into tumor cells[24].

The recruitment of effector T lymphocytes to tumors depends not only on the role of chemokines positive for various ligands but also on function of the vascular system and tumor matrix as a physical barrier. Structural and functional abnormalities in the tumor vasculature promote tumor growth and metastasis as well as immunosuppression[25]. An insufficient number of pericyclic cells or loose vascular basement membrane connections in tumor tissue that cover endothelial cells increases the permeability of tumor blood vessels, thereby promoting tumor metastasis[26,27]. As tumor cells continue to grow, the increased supply of nutrients and oxygen enhance the expression of vascular endothelial growth factor (VEGF) and basic fibroblast growth factor (bFGF), which promote the production of tumor cell blood vessels and create a hypoxic and acidic environment[28-30]. Hypoxia recruits immunosuppressive cells to the tumor immune microenvironment (TME) and can inhibit the function of immune effector T cells[31]. The neovascular endothelial cells can interfere with the function of T cells by altering the expression of receptors and their ligands, thereby causing immunosuppression[32]. In contrast to hematologic tumors, the matrix barrier is the last barrier affecting immune cells entering solid tumor tissue. When the tumor matrix is activated and fibrosis occurs, the antitumor treatment effect is inversely proportional to the activation and fibrosis of the matrix; that is, the lower the activation and fibrosis of the tumor matrix are, the better the antitumor effect[33]. Stromal cells in the TME can be divided into infiltrating immune cells, angiogenic vascular cells, cancer-associated fibroblasts (CAFs) and cancer-associated adipocytes[34]. CAFs account for a large proportion of the TME. The fibrous structure induced by CAFs can affect the delivery of immune drugs to tumor tissues and reduce antitumor immune efficacy[33]. It can be concluded that the state of chemokines in the immune microenvironment and the presence of specific vascular factors or stromal barriers are key factors affecting the inability of immune cells to nest inside of immune-repelling tumors.

***The formation mechanism of the immune-desert phenotype***

The main feature of the immune-desert tumor phenotype is the absence of a perinucleoplasm or intratumoral immune cell infiltration[35]; therefore, any infiltration of immune cells is mainly an initiation error (*i.e.*, insufficient antigen release, insufficient antigen presentation, and T cell initiation and activation disorders) in the anticancer immune response[15]. For these “cold tumors” to be transformed into “hot tumors”, characterized by a large amount of immune cell infiltration into the tumor tissues, the immune escape mechanism mainly involves immune initiation and the transfer of immune cells to tumor cells.

Newly emerging or overexpressed antigens in tumor tissue are collectively referred to as tumor antigens, which can be divided into tumor-specific antigens (TSAs) and tumor-associated antigens (TAAs) according to tumor specificity[36]. TSAs, produced during tumor mutations as neoantigens, are specific to tumor cells or are present only in certain tumor cells and are not expressed in normal cells[37]. Neoantigens have stronger immunogenicity and higher specificity than TAAs, making them a target for immune responses[38]. TMB is an index used to evaluate the number of gene mutations in tumor cells. The higher the gene mutation frequency of the tumor cells, the more tumor antigens are carried on the cell surface[39]. Some current evidence suggests that tumors with a higher mutation load can improve immunotherapy efficacy by modulating the degree of infiltration of immune cells to influence the immune properties of the TME[40]. Therefore, the lack of neoantigen production and a low TMB are the reasons for the lack of antigen release.

After tumor antigens are recognized, they must be processed by antigen-presenting cells (APCs) to initiate an immune response. As the most powerful APCs, dendritic cells (DCs) function to process antigen information and play a key role in the immune response[41]. DCs can be highly expressive MHC molecules, which bind to their captured tumor antigens and present them to T cells in the form of peptide-MHC molecular complexes, resulting in the activation of effector T cell responses to cancer-specific antigens. Therefore, the absence of DCs plays a key role in tumor immune evasion and immunotherapy failure[42]. However, the response of T cells is regulated by a series of co-suppressive or co-stimulatory signals; for example, the B7 molecule is the most important co-stimulatory molecule of T lymphocytes, superceding CD80 and CD86, which bind to the co-stimulatory protein CD28 to induce T cell activation and proliferation; however when it binds to cytotoxic T lymphocyte-associated antigen 4 (CTLA-4), immune responses are downregulated[43].

**Combination strategies to help fight cold CRC**

***Combination with targeted therapy***

In solid tumors, vascular system abnormalities lead to restriction of the process of immune cell infiltration into the tumor, while vascular system normalization can improve the immunosuppressive microenvironment and facilitate immunotherapy drug delivery[44]. Targeting growth factors such as VEGF to inhibit the pathological angiogenesis of cancer has become a recognized therapeutic strategy[45]. VEGF and epidermal growth factor receptor (EGFR) are common therapeutic targets in CRC. Studies[46,47]have shown that immunotherapy and EGFR antagonists, VEGF antagonists and mitogen-activated protein kinase antagonists are utilized in the treatment of malignant tumors, which can bring new options and hope for patients with pMMR/MSS CRC.

The results of a series of clinical studies on immune-targeted combination therapies have shown that the efficacy of immunotherapy in CRC patients has made a breakthrough[48]. The mechanism by which immune-targeted combination therapy can enable patients with MSS bowel cancer to achieve immune efficacy (*i.e.*, VEGF inhibition can restore tumor blood vessels to normal, increase the efficient initiation and activation of T cell responses, and recombine the typical immunosuppressive tumor microenvironment, thus indicating a synergistic effect between tumor immunotherapy and antivascular therapy)[49]. Regulatory T cells (Tregs) in the TME can inhibit effector T cells by either direct contact pathways or by secreting inhibitory cytokines, where targeted inhibitors can induce the proliferation of CD8 T cells in lymph nodes or suppress this pathway by infiltrating into tumor tissue, thereby improving the body’s anti-immune response[50]. Targeted inhibitors can improve transcriptional activity, increase the expression of anticancer genes and reduce the proliferation of tumor cells by inhibiting the elimination of acetyl groups[51]. It can be concluded that the application of anti-angiogenic targeted therapy combined with immunotherapy in MSS CRC patients is theoretically supported. A Japanese phase Ib REGONIVO study[48] conducted in recent years used nivolumab (a PD-1 inhibitor) plus regorafenib for CRC as the first treatment strategy. The results showed an ORR of 33.3%, a median progression-free survival (PFS) duration of 7.9 mo (95%CI 2.0, NE) and a 1-year overall survival (OS) rate of 68.0% in MSS CRC patients, which highlighted the benefits of immunotherapy for patients with cold CRC. The study further found that the median PFS of patients with PD-L1 combined positive score (CPS) > 1 was significantly better than that of patients with CPS ≤ 1 (6 mo *vs* not reached, *P* < 0.05). However, several studies, such as the North American Phase II study (NCT04126733), the Chinese population REGOTORI study[52] and the French Phase II REGOMUNE trial[53], all adopted a combination of immune and targeted dual therapy strategies, and although the results showed some efficacy, they were not as significant as those seen in the Japanese population.

***Combination with chemotherapy***

Although chemotherapy drugs may kill immune cells that fight the tumor immune response, there is growing evidence that chemotherapy can activate the body’s immune response through multiple mechanisms. In addition to directly killing tumor cells, chemotherapy drugs stimulate the immune system by enhancing immunogenicity and increasing T cell infiltration, which converts “cold tumors” into “hot tumors”[54,55]. Chemotherapy drugs can enhance the recognition and presentation of DCs, activate cytotoxic T lymphocytes to attack tumors, and stimulate the release of interleukin-2 (IL-2), IL-4, and IFN-γ, causing an antitumor immune response[56,57]. Most chemotherapy regimens use a multidrug combination approach; however, the improvement in drug resistance remains nonsignificant[58]. Other studies have found that the use of PD-L1 inhibitors can reduce the incidence of chemotherapy drug resistance, mainly because chemotherapeutic drugs can lead to drug resistance by inducing the expression of PD-L1[55]. However, ICIs (such as CTLA-4 or PD-1/PD-L1) cannot directly kill tumor cells and must rely on the host immune system to enhance antitumor activity. In conclusion, chemotherapy combined with ICIs can improve the immunogenicity of CRC subtypes, help improve immune drug efficacy, and inhibit chemotherapy drug resistance. In recent years, the combination of chemotherapy and immunotherapy has attracted the attention of most clinicians and researchers and has been explored in multiple clinical trials. For example, in a phase 2 clinical study (NCT03374254) using chemotherapy to treat immunocompromised patients with non-MSI-H or pMMR advanced CRC, pembrolizumab combined with chemotherapy was used in both the first and second lines of treatment[59]. A phase 2 clinical study (METIMMOX study) evaluating the efficacy of the FLOX regimen plus nivolumab in the first-line treatment of MSS metastatic CRC (mCRC) validated the possibility of changing tumor immunogenicity and inducing a response to ICIs in this patient group[60]. Based on the publication of the results of this combination therapy strategy, we expect that the ongoing studies will lead to lasting outcomes for patients who have cold CRC. At present, a series of studies have been carried out to explore the effect of chemotherapy combined with antitumor immunotherapy, which has broad application prospects based on the relevant results. The question of how to achieve the optimum combination effect and the optimum timing and drug dose for combination therapies is still a hot topic for further exploration in future research.

***Combination with radiation therapy***

Radiation therapy is an effective antitumor method for the palliative treatment of most patients with local or solitary metastatic tumor lesions, as well as extensive, advanced metastatic lesions. Radiotherapy can induce the body’s antitumor immune response, and its synergistic effect with ICIs can enhance the ability of the body’s immune system to recognize and eradicate tumor cells[61]. Some studies have shown that radiotherapy can transform immunologically “cold tumors” into “hot tumors” to a certain extent and increase the number of immune-beneficial populations[62]. With the advent of the era of immunotherapy, the immunostimulatory and immunosuppressive effects of radiation therapy have also been given increasing attention. Studies have confirmed that the combination of radiation therapy and ICIs can significantly enhance the regressive changes in tumor tissue. A meta-analysis of ICIs with or without radiotherapy in patients with melanoma showed that radiation therapy plus ICIs improved PFS without serious adverse effects[63]. Irreparable damage to DNA is one of the main mechanisms by which radiation therapy kills tumor cells and is also the key to radiation therapy-induced immune regulation. Radiation therapy can induce abscopal effects and reverse resistance to immunotherapy[64]. Radiotherapy also induces immunogenic cell death (ICD) of tumor cells by upregulating the release of damage-associated molecular patterns (DAMPs)[65]. When radiotherapy rays act on tumor cells, they can enhance their expression of MHC I molecules and improve their antigen presentation capacity[66]. In addition, radiotherapy can induce the secretion of chemokines, such as CXCL9 and CXCL10, and the release of pro-inflammatory cytokines, such as IL-2, tumor necrosis factor and type I interferons (IFN-α or IFN-β), recruit T cells and promote the infiltration of DCs and effector T cells into irradiated tumors, thereby regulating the immune state of the TME[66-69]. Radiotherapy also induces ICD and increases the release of TAAs by upregulating the release of DAMPs[70]. It follows that radiotherapy can promote the effect of immunotherapy drugs by altering changes in the TME. In turn, immunotherapy promotes the effects of radiation therapy by targeting and modulating various T cell populations.

***Combination immunotherapy***

Studies have confirmed that dual immunotherapy can obtain significant clinical efficacy, mainly due to the synergistic effect of the combination of anti-PD-1 and anti-CTLA-4 to increase the infiltration of effector T cells, which can play a role in overcoming the congenital tolerance of immune monotherapy in patients with cold CRC[71]. In a systematic review and meta-analysis by He *et al*[72], it was found that the combination of PD-1 and CTLA-4 inhibitors had the highest efficacy. CTLA-4 and PD-1 are both immune checkpoint proteins on T cells but play different roles in the immune response. Anti-CTLA-4 has the effect of expanding T cells in lymphoid organs and tumor tissues, while PD-1 is associated with ligand binding and overcomes the immunosuppressive antitumor effector T cells in the tumor bed[73]. CTLA-4 is highly expressed in Tregs, and many studies have shown that anti-CTLA-4 monoclonal antibody (mAb) can induce Treg depletion at tumor sites and normalize the TME[74]. A large phase II clinical study (CCTG CO.26) reported at the ASCO GI meeting in 2019 used a PD-L1 mAb combined with a CTLA-4 mAb to treat patients with advanced refractory CRC (98% of whom were pMMR/MSS). The study results showed that the mOS of the dual immunotherapy group reached 6.6 mo, and further exploration showed that MSS CRC patients with a TMB of 28 or more had the greatest OS benefit (HR, 0.34)[75]. Other studies have shown that dual immunotherapy does not significantly aggravate toxic side effects compared with immune monotherapy[76]. Some studies have shown that neoadjuvant immunotherapy can activate more effector T cells that kill tumor cells and induce a stronger tumor-specific T cell response[77]. A clinical study using neoadjuvant combination immunotherapy (NICHE study) showed that 27% of patients with pMMR achieved a pathological response, and 3 patients achieved a major pathological response, which brought hope for the application of dual immunotherapy to neoadjuvant therapy for MSS CRC[78]. Dual immunotherapy exhibits the characteristics of the complementary mechanisms, has long-lasting efficacy (in contrast to immune monotherapy), and therapeutic toxicity that is controllable; thus, it is expected to reverse the resistance of MSS CRC patients to ICIs. Although this study achieved the primary endpoint, the improvement in OS was not significant, so the conclusion that dual immunotherapy is efficacious has not been widely accepted, and the combination of dual ICIs in future clinical practice still needs more exploration.

***Combination with OVs***

As a class of viruses that can lyse tumor cells, OVs destroy tumor cells by selective replication and expansion in tumor cells, releasing TAAs and tumor-associated neutrophils. In this way, OVs induce the ICD of tumor cells while activating and promoting T cells to infiltrate the tumor site to improve the tumor microenvironment; thus, “cold tumors” can be effectively immunoactivated and transformed into “hot tumors”[3,79-81]. However, OVs alone have difficulty maintaining a long-term immune response and are prone to immune resistance. If OVs are used in combination with ICIs, they can not only reduce the immune tolerance of tumor cells but also significantly improve the antitumor efficacy, as demonstrated in the early stages of therapeutic safety studies. It has been shown in patients with advanced melanoma treated with the combination of talimogene laherparepvec and pembrolizumab that OVs can induce elevated PD-L1 expression at tumor sites as well as a significant increase T cell infiltration to improve the tumor microenvironment[82]. Therefore, it can be understood from these research results that OVs combined with ICIs are not only dependent on the degree of infiltration of immune cells but can also improve the infiltration of effector T cells in tumor tissues, successfully reversing the nonimmune inflammatory microenvironment in “cold tumors”.

***Combination with bacterial therapy***

There is increasing evidence that the gut microbiome affects the host’s innate and adaptive immune responses, providing a basis for a new class of antitumor agents that combine immunotherapy and gut microbiome therapy[83-86]. Both gut microbes and their metabolites can transform the immune microenvironment by inducing the release of proinflammatory cytokines and increasing the recruitment of T cells to the tumor site. The combination of gut microbes and immunosuppressants can reduce immune resistance. Pathogen-associated molecular patterns of gut microbes are recognized by Toll-like receptors and activate DCs to initiate adaptive immune responses, thereby alleviating immunosuppressive drug resistance and exerting antitumor effects[87]. The gut microbiome, modulated in specific ways, can improve the efficacy of immunotherapy while also alleviating immune-related adverse events caused by immunosuppressive drugs[88]. These findings suggest that Bifidobacterium may alleviate immunotherapy-associated colitis by altering the composition of the gut microbiota[89]. Microbial metabolites, such as the short-chain fatty acids produced by intestinal microorganisms after fermentation, which can induce the production of CD4 T cells and IL-22 to protect intestinal integrity from foreign microorganisms and maintain intestinal homeostasis, can also participate in the body’s immune regulation[90,91]. Gopalakrishnan *et al*[84] confirmed that the antitumor immune response is affected by the number and type of intestinal microorganisms in the gut microbiota. Specifically, the higher the diversity of gut microbes is, the longer the ORR and survival period of immunotherapy[92]. The gut microbiome of patients with CRC is less diverse and less sensitive to antigen stimulation, so it may be difficult to generate an immune response, which explains why the vast majority of CRC patients have “cold tumors”[93]. As a newer class of antitumor agents, gut microbes play an important role in antitumor immunotherapy, but their mechanisms of immune regulation are complex and are affected by factors such as body weight, age, diet, ethnicity, and living environment[94]. These individual differences have brought great challenges to the development of gut microbial antitumor immunotherapy.

***Combination with tumor vaccines***

Tumor vaccines have been used in a variety of tumor treatments and activate the body’s immune response, amplifying the antitumor immune response, and can induce durable antitumor immunity[95]. Tumor vaccines are divided into targeted antigen vaccines, autologous tumor cell vaccines, and DCs vaccines[96]. The first cancer vaccine approved by the United States of America Food and Drug Administration, Sipuleucel-T, was found to significantly improve the prognosis of patients with hormone refractory prostate cancer[97]. MAGE-A and NY-ESO-1, TAAs with strong antigenicity and specificity in the cancer testis antigen family, are highly expressed in a variety of tumor tissues, including CRC, and are expressed at low levels in normal tissues[98,99]. Therefore, they can be specifically recognized by the body’s immune system and exert antitumor effects. MAGE-A and NY-ESO-1 can induce a robust T cell antitumor response, and their related antibodies can be used as alternative biomarkers in vaccine therapy research[100,101]. At present, research on tumor vaccines based on MAGE-A4 and NY-ESO-1 has been carried out successively, and the results show that these vaccines have broad application prospects[101,102]. However, it is difficult to achieve high immune response rates with a single cancer vaccine alone, as has been confirmed in several clinical trials in patients with CRC[103-105]. Given that vaccination stimulates the body’s immune system to exert an effective antitumor response, the combination of ICIs and vaccines may lead to better outcomes for cold CRC patients.

***Combination with multiple therapies***

Studies have demonstrated that either chemotherapy or targeted combination immunization can improve the antitumor immune response in patients with cold CRC. A series of studies have been carried out in the clinic based on the characteristics of chemotherapy drugs that can release tumor antigens, and targeted drugs that can regulate the immune microenvironment to explore whether the addition of immunotherapy will further improve the immune response of patients[51,54,55]. A phase 2 clinical study, AtezoTRIBE, which used FOLFOXIRI plus bevacizumab with or without atezolizumab for the first-line treatment of unresectable advanced CRC, showed that the addition of atezolizumab not only did not significantly increase the incidence of adverse reactions but also prolonged the PFS of patients with mCRC[106]. Another phase 2 clinical study, BACCI, compared the efficacy of capecitabine plus bevacizumab with or without atezolizumab[107]. The results showed that the combined atezolizumab group did not have significantly improved PFS or OS, but the ORR increased from 4.35% to 8.54%. The MODUL study used fluoropyrimidine plus bevacizumab with or without atezolizumab as first-line therapy and as maintenance therapy after stabilization, but the results showed that the addition of immunotherapy to maintenance therapy did not improve the prognosis of patients with advanced MSS CRC[108]. Neoadjuvant chemoradiotherapy (NCRT) is a standard treatment model for locally advanced rectal cancer. The addition of immunotherapy based on NCRT has been shown in a number of studies to promote the treatment response. For example, in a clinical study conducted in Japan on the use of immunoconsolidation therapy after neoadjuvant radiotherapy (NCT02948348), MSS CRC patients achieved a 29.7% pathological complete response (pCR) rate[109]. A phase 2 clinical study (ANAVA) in Italy used sequential immunotherapy after NCRT, and a pCR rate of 21.8% was observed in patients with MSS CRC[110]. Additional trials are ongoing to test the NCRT strategy with checkpoint inhibitors (NCT04083365 and NCT03994835).

Therefore, the benefits of multiple combination therapies are still controversial, and the balance between immune benefits and adverse reactions is controversial and requires further research and verification. However, we expect that additional clinical studies can allow these patients to achieve higher pathological response rates.

**Selection of the immunodominant population**

As a very complex and dynamic ecosystem, the TME is the site for tumor cell survival and proliferation and contains many different kinds of cell populations, including tumor cells, immune cells, and supporting cells (*e.g.*, fibroblasts, stromal cells, and endothelial cells)[111,112]. The TME can be divided into an immune cell-based microenvironment and a non-immune microenvironment dominated by fibroblasts[113]. Similar to many tumors, the TME plays an integral role in immunotherapy in CRC patients, and immunotherapy responses vary due to differences in the TME[114]. One study based on genomics analysis showed that CRC patients can be divided into different immune subtypes, and through the analysis of molecular phenotypes, the immune status of the tumor microenvironment can be determined, and the degree of TMB in CRC patients can be predicted[115]. Several existing studies have shown that biomarkers such as MSI-H, TMB and the gut microbiome can be used as predictors of immunotherapy prognosis[116-118]. The new study found that the PORE/polymerase delta 1 catalytic subunit (POLD1) mutation present in patients with MSS CRC is associated with TMB and MSI status and that CRC containing this mutation can exhibit immunoinflammatory characteristics. CRC is a highly heterogeneous tumor, and accurate immunomolecular typing can reveal the differences between subtypes, which may help to screen out the immunotherapy-dominant populations. This strategy will increase the accuracy of the biomarkers as prognostic indicators and will allow the development of an individualized hierarchical diagnosis and treatment strategy for MSS CRC patients that maximizes patient benefits[119,120]. POLE/POLD1 mutations can lead to activation of the TME, which elicits an immune response to the tumor. He *et al*[121] and Chen *et al*[122] found that POLAR/POLD1 can be used as a molecular marker for predicting the efficacy of ICIs in CRC patients.

**CONCLUSION**

With developments in the understanding of tumor biology and tumor immune tolerance mechanisms, the existing research results suggest that a combination immunotherapy strategy for cold CRC will allow a good immune response in some patients; however there has been no major breakthrough, and maintaining a long-term antitumor immune response is still the main challenge. It is necessary to further explore the best combination of treatments to expand the populations benefiting from immunotherapy and to find new biomarkers to accurately screen sensitive populations, which can promote the development of future precision biomarkers and precision therapies.

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**Figure Legends**



**Figure 1** **Three different types based on immune characteristics.** Immune-excluded phenotype: T cells cannot homie from the tumor stroma to the interior of the tumor; Immune-desert phenotype: Anticancer immune initiation error, including insufficient antigen release, insufficient antigen presentation, T cell initiation and activation disorders. TMB: Tumor mutation burden; IFN-γ: Interferon- γ; PD-L1: Programmed cell death 1 Ligand 1.



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