WJD

# World Journal of **Diabetes**

Submit a Manuscript: https://www.f6publishing.com

World J Diabetes 2023 July 15; 14(7): 1027-1036

DOI: 10.4239/wjd.v14.i7.1027

ISSN 1948-9358 (online)

MINIREVIEWS

## Klotho: A new therapeutic target in diabetic retinopathy?

Alessandra Puddu, Davide Carlo Maggi

Specialty type: Endocrinology and metabolism

Provenance and peer review: Invited article; Externally peer reviewed.

Peer-review model: Single blind

#### Peer-review report's scientific quality classification

Grade A (Excellent): A Grade B (Very good): B, B Grade C (Good): C Grade D (Fair): 0 Grade E (Poor): 0

P-Reviewer: Cen LS, China; Liu Y, China; Long P, China; Mansour AM, Lebanon

Received: February 16, 2023 Peer-review started: February 16, 2023 First decision: April 11, 2023 Revised: May 12, 2023 Accepted: May 22, 2023 Article in press: May 22, 2023 Published online: July 15, 2023



Alessandra Puddu, Davide Carlo Maggi, Department of Internal Medicine and Medical Specialties, University of Genova, Genova 16132, Italy

Corresponding author: Alessandra Puddu, BSc, Technician, Department of Internal Medicine and Medical Specialties, University of Genova, Viale Benedetto XV 6, Genova 16132, Italy. alep100@hotmail.com

#### Abstract

Klotho (Kl) is considered an antiaging gene, mainly for the inhibition of the insulin-like growth factor-1 signaling. Kl exists as full-length transmembrane, which acts as co-receptor for fibroblast growth factor receptor, and in soluble forms (sKl). The sKl may exert pleiotropic effects on organs and tissues by regulating several pathways involved in the pathogenesis of diseases associated with oxidative and inflammatory state. In diabetic Patients, serum levels of Kl are significantly decreased compared to healthy subjects, and are related to duration of diabetes. In diabetic retinopathy (DR), one of the most common microvascular complications of type 2 diabetes, serum Kl levels are negatively correlated with progression of the disease. A lot of evidences showed that Kl regulates several mechanisms involved in maintaining homeostasis and functions of retinal cells, including phagocytosis, calcium signaling, secretion of vascular endothelial growth factor A (VEGF-A), maintenance of redox status, and melanin biosynthesis. Experimental data have been shown that Kl exerts positive effects on several mechanisms involved in onset and progression of DR. In particular, treatment with Kl: (1) Prevents apoptosis induced by oxidative stress in human retinal endothelial cells and in retinal pigment epithelium (RPE) cells; (2) reduces secretion of VEGF-A by RPE cells; and (3) decreases subretinal fibrosis and preserves autophagic activity. Therefore, Kl may become a novel biomarker and a good candidate for the treatment of DR.

Key Words: Klotho; Diabetic retinopathy; Retinal pigment epithelium; Vascular endothelial growth factor A; Epithelial to mesenchimal transition; Ocular neovascularization

©The Author(s) 2023. Published by Baishideng Publishing Group Inc. All rights reserved.



**Core Tip:** In diabetic Patients, serum levels of Klotho (Kl) are significantly decreased compared to healthy subjects. Moreover, serum Kl levels are negatively correlated with worsening of diabetic retinopathy (DR). Several evidence suggests that retina homeostasis may be affected by altered expression of membrane Kl, as well by reduced levels of soluble Kl. In this review we focused on the role of Kl in DR, highlighting the importance of Kl in maintaining retinal homeostasis and its positive effects on several mechanisms involved in DR onset and progression. Therefore, Kl could be a novel biomarker and a good candidate for the treatment of DR.

**Citation:** Puddu A, Maggi DC. Klotho: A new therapeutic target in diabetic retinopathy? *World J Diabetes* 2023; 14(7): 1027-1036 **URL:** https://www.wjgnet.com/1948-9358/full/v14/i7/1027.htm **DOI:** https://dx.doi.org/10.4239/wjd.v14.i7.1027

#### INTRODUCTION

#### Klotho

The name Klotho (Kl) derives from that of the youngest of the Three Fates who spins the thread of human life[1]. Indeed, it is considered an antiaging gene, since phenotypes of mice with mutation in this gene are similar to those of patients with premature-ageing syndromes. Kl shares sequence similarity with members of the glycosidase family 1 and it has been reported to function as a novel  $\beta$ -glucuronidase[2,3]. It encodes for 3 proteins:  $\alpha$ -Kl,  $\beta$ -Kl and Kl-related protein (Klrp)[4].  $\beta$ -Kl is mainly expressed in liver and adipose tissue and is involved in metabolic processes[4]; whereas Klrp is a cytosolic  $\beta$ -glucocerebrosidase[5].  $\alpha$ -Kl, generally simply referred as Kl, is a type I single-pass trans-membrane glycoprotein mainly expressed in the kidneys, liver, brain, and at lower level in the pituitary, skeletal muscle, urinary bladder, pancreas, testis, ovary, colon, thyroid gland, placenta and vascular tissue[1]. Both the intracellular and the transmembrane domains of α-Kl are very short, whereas the extracellular domain is longer and contains two repeated sequences (KL1 and KL2)[4,6]. After association with fibroblast growth factor receptors (FGFRs), the full-length transmembrane KI (mKI) acts as coreceptor for the bone-derived phosphaturic hormone FGF23, thus taking part to phosphate excretion and calcium homeostasis by regulating the expression and activity of the calcium channel transient receptor potential vanilloid 5 (TRPV5)[7]. Besides mKl, there are 2 isoforms of  $\alpha$ -Kl: A shed soluble form (sKl), which derives from the cleavage of the extracellular domain of Kl from the cell surface by the metalloproteinases ADAM10 and ADAM17, and a secreted form that is produced by alternative splicing of Kl mRNA<sup>[4]</sup>. The shed soluble form of Kl seems to be dominant on both the secreted and the membrane forms in humans<sup>[8]</sup>. It has been proposed that the soluble forms of KI function as a hormone<sup>[9]</sup>. Moreover, since circulating levels of sKI increase following exercise training, it has been also hypothesized that KI may be related to the antiaging effects of physical activity[10]. The sKI has pleiotropic effects on a lot of organs and tissues, thus regulating several pathways<sup>[8]</sup>. Indeed, after the release in blood, urine and cerebrospinal fluid, sKl exerts biological effects involved in preservation of endothelial integrity and permeability, and affect intracellular signaling pathways including those related to insulin, insulin-like growth factor-1 (IGF-1), PI3K, NF-kB, p53/p21, cAMP, protein kinase C and Wnt[8,11-13]. In particular, a lot of evidence demonstrated that the anti-ageing effects of sKl have been associated with the inhibition of IGF-1 signaling and its downstream actions especially by enhancing resistance to oxidative stress[14,15]. Indeed, inhibition of the IGF-1 signaling by sKl results in increased production of antioxidant enzymes[16]. Therefore, activity of sKl may regulate several pathways involved in the pathogenesis of diseases associated with oxidative and inflammatory state.

It is not yet clear whether intracellular signaling of circulating Kl is mediated by a membrane receptor. Recent hypothesis suggests that sKL may act as a circulating co-receptor for membrane-bound FGFRs, thus allowing the interaction with FGF23 and regulating FGFR-mediated signaling also in cells lacking the full length form of Kl[17]. Moreover, it has been demonstrated that sKl is able to bind membrane lipid rafts, alter their organization, and affect caveolae-mediated TRPV5 endocytosis[18], suggesting that the intracellular signaling of sKl may occur at the level of caveoale.

#### **KI AND DIABETES**

In diabetic patients, serum levels of Kl have been found significantly decreased compared with those of healthy subjects [19]. In addition, the amount of sKl is related to duration of diabetes and is negatively correlated to HbA1c. Kidneys are considered the main source of sKl[17], and are also the principal organ involved in the clearance of sKl from the circulation into the urine, thus playing a dual role in the homeostasis of Kl[9]. Therefore, altered kidney function may affect the systemic effects of Kl. Consequently, the anti-aging effects of Kl have been extensively investigated in kidneys, reporting that increased levels of Kl inhibit the progression of various kidney diseases[20,21]. In animal models of diabetes, Kl counteracts podocytic and glomerular albumin permeability induced by hyperglycemia[22], and prevents epithelial-mesenchymal transition (EMT) in diabetic kidneys[21]. Interestingly, expression of Kl has been found decreased in the renal cortices of mice with diabetes[22]. Moreover, Typiak *et al*[23] showed that decreased levels of membrane-bound Kl are associated to increased shedding of Kl, to higher levels in serum of diabetic rats and a to reduced urinary

excretion[23]. In diabetic patients, the amount of soluble Kl is reduced in the early stage of chronic kidney disease (CKD), but increased with disease progression and the decrease of glomerular filtration rate[24]. A recent meta-analysis of data on sKl amount in patients with diabetic nephropathy (DN) confirms that levels of sKl are further lowered in the early stage of DN[25], suggesting that KI might be considered as an early biomarker of DN[23,26]. However, although levels of sKl still remain lower in patients with DN, they seem to increase during the worsening of diabetic CKD probably linked to the decline in glomerular filtration rate that leads to reduced urinary excretion of Kl[23,27].

Expression of KI has been detected also in mouse pancreatic islets and in beta-cell line[28,29]. It has been showed that Kl is involved in regulation of glucose-induced insulin secretion, probably, through regulation of TRPV2 expression [28, 29]. Indeed, overexpression of KI increases both insulin secretion and plasma membrane levels of TRPV2; whereas silencing of KI negatively affects plasma membrane levels of TRPV2, glucose-induced calcium entry and insulin secretion [28]. Moreover, treatment with  $\alpha$ - or  $\beta$ -Kl protects human beta-cells by cytokine-induced apoptosis and improved insulin secretion[30,31].

#### DIABETIC RETINOPATHY

Diabetic retinopathy (DR) is a common microvascular complications of type 2 diabetes and represents the primary cause of blindness in working age adults[32]. Actually, retinal neurodegenerative lesions may occur earlier than microvascular ones, therefore DR has been defined as a highly tissue-specific neurovascular complication of diabetes by the American Diabetes Association[33]. The early manifestations of DR involves damages to both microcirculation and retinal neurons and are associated with oxidative stress[34]. The resulting sustained proinflammatory environment, in turns, increases oxidative stress, due to the reduced levels of antioxidant enzymes in the retina. Photoreceptors and the retinal pigment epithelium (RPE) cells are highly susceptible to oxidative stress in the early stage of DR and their dysfunction lead to progression of retinal degeneration[34]. Furthermore, chronic inflammation causes vasoregression and alters vascular permeability, leading to formation of microaneurysms and exudates. Then, hypoxia and the release of proangiogenic factors, such as vascular endothelial growth factor A (VEGF-A), may promote pathological ocular neovascularization[34]. In the retina, VEGF-A is mainly produced by RPE cells, a monolayer of highly specialized cells located between the choroid and photoreceptors that forms the outer blood-retinal barrier[35]. Due to their localization, RPE cells may affect retinal homeostasis by altering the function and maintenance of both the photoreceptors and capillary endothelium[36]. Indeed, under normal condition, VEGF-A is released at low concentrations from the basal side of the RPE to maintain endothelial cell function [37]. However, under pathological condition, such as chronic hyperglycemia, secretion of VEGF-A increases leading to activation of endothelial cells and altered permeability of the choroidal vessels [37,38]. It is well known that dysfunction of RPE cells contributes to onset and progression of DR. Therefore, maintaining the function of RPE and controlling the levels of VEGF-A are of great importance in preventing worsening of DR to the proliferative state.

#### KI AND RETINAL HOMEOSTASIS

It has been found that Kl is expressed in the human retina, optic nerve, and lens[39,40]. Several evidence showed that Kl regulates a lot of mechanisms involved in maintaining homeostasis and functions of retinal cells[39,41,42]. Firstly, KI knockout mice display several morphological changes as compared to wild type mice: Decreased pigmentation of the RPE layer, large choroidal vessels, thinner and deformed basal membrane, and signs of degeneration in the outer segment of photoreceptors (POS)[41]. Proteomics analysis reveals that proteins involved in eye development, visual perception and mitochondrial function are downregulated in Kl knockout mice[42]. Accordingly, Kl knockout mice have reduced retinal function, with functional deficit comparable to those observed in IGF-1 knockout mice[39]. Considering that KI knockout mice are hypoglycemic, it can be hypothesized that the effects observed in the retina may be attributable to increased sensitive to the insulin and IGF-1 signaling.

Kokkinaki et al[41] demonstrated that KI is expressed in primary cultures of RPE cells, mainly in the cell membrane, and that its depletion compromises several important function of RPE cells[41]. Moreover, they demonstrated that treatment with recombinant KI protein has protective effects on RPE function, including phagocytosis, VEGF-A secretion, oxidative stress response, and melanogenesis.

Phagocytosis of POS is of particular importance in maintaining visual function and the visual cycle. It has been shown that transfection of RPE cells with Kl siRNA significantly reduced phagocytosis[41], suggesting that Kl is involved in the regulation of this important function. Evidences that treatment of RPE cells with KI significantly increased phagocytosis in RPE cells confirm this hypothesis<sup>[41]</sup>. POS phagocytosis is regulated by several factors, among them, the Ca2+ signaling and the expression of Mer Tyrosine Kinase (MerTK) seem to play an important role[43]. Rise in intracellular Calcium is required for maintaining POS phagocytosis rate[44-46]. It has been reported that secreted KI may regulate calcium homeostasis by affecting activity of calcium channels, including TRPVs and the Ca2+ release-activated Ca2+ channel (CRAC)[28,47,48]. Interestingly, human RPE expresses both TRPV5 and CRAC, which regulate calcium entry in this cells[49,50]. However, Kokkinaki et al[41] showed that treatment of RPE cells with Kl did not increase intracellular Calcium concentration<sup>[41]</sup>, suggesting that Kl increases phagocytosis through a mechanism independent to calcium. Internalization of POS requires the engagement of MerTK, a cell surface receptor member of the tyro/Axl/Mer family of receptor tyrosine kinase, therefore MerTK expression is critical for POS phagocytosis[43]. Interestingly, it has been demonstrated that KI regulates phagocytosis by upregulating MerTK expression, indeed treatment of RPE cells with KI



#### Table 1 Main effects of Klotho on retinal cells

Table 1 Wain effects of Kiotho on retinal cells				
Functions	Effects of Klotho depletion	Effects of treatment with Klotho	Type of cell	Ref.
Phagocytosis	Reduced	Improved	RPE cells	[41,43]
		Increased expression of Mertk		
VEGF-A		Decreased secretion	RPE cells	[41]
		Reduced signaling mediated by VEGFR2- and IGF-1R		
Redox balance	Increased oxidative stress	Restored	RPE cells	[41,53]
		Prevention of ROS production		
		Increased NRF2 expression and nuclear translocation		
	Reduced expression of SOD2	Restored expression of SOD2 and CAT		
Pigmentation	Reduced		RPE cells	[41]
	Decreased melanin granules			
Mitochondrial function	Reduced biogenesis of mitochondria	Preserved	RPE cells	[53]
Autophagy		Improved	Retina	[42]
	Decreased activation of AMPK			
	Reduced expression of SIRT1			
EMT		Decreased expression of mesenchymal cell markers	RPE cells	[66]
Apoptosis		Reduced	RPE and retinal endothelial cells	[42,53,54]
		Increased expression of Bcl-2		
		Decreased expression of Bax		
		Decreased activity of Caspase-3		

VEGF-A: Vascular endothelial growth factor A; IGF-1R: Insulin-like growth factor-1; RPE: Retinal pigment epithelium; ROS: Reactive oxygen species; AMPK: 5' adenosine monophosphate-activated protein kinase; SIRT1: Silent information regulator 1; EMT: Epithelial-mesenchymal transition; NRF2: Nuclear factor E2-related factor 2; SOD2: Superoxide dismutase 2; CAT: Catalase.

induces intracellular signaling that leads to increased expression of MerTK and, consequently, improves phagocytosis efficiency<sup>[41]</sup>.

VEGF-A is one of the main important pro-angiogenic factor and its excessive secretion is implicated in promoting the pathological neovascularization of the choroidal vasculature[51,52]. RPE cells are the major responsible of VEGF-A production in the retina. Treatment of the RPE cell line ARPE-19 with Kl significantly decreases VEGF-A secretion from both the apical and the basal sides[41]. Moreover, the presence of Kl inhibits the phosphorylation of VEGFR2 induced by VEGF-A, thus affecting intracellular signaling activated by VEGF-A.

Due to its extremely active metabolism, the retina is one of the organ with major request of oxygen, therefore it may be susceptible to overproduction of reactive oxygen species (ROS). Under normal conditions, ROS take part to the retinal physiological signaling, however, when generation of ROS exceeds the natural antioxidants defenses, oxidative stress may contribute to the pathogenesis of several retinal diseases, including DR. Experimental data demonstrate that Kl contributes to maintain the redox balance in the retina. Indeed, mRNA levels of Kl have been found significantly decreased in ARPE-19 cells treated with hydrogen peroxide ( $H_2O_2$ )[53]. Moreover, Kokkinaki *et al*[41] demonstrated that down-regulation of Kl expression leads to reduced expression of the anti-oxidant Superoxide dismutase 2 (SOD2) in RPE cells[41]. On the contrary, pretreatment with sKl prevented rise in ROS induced by  $H_2O_2$  enhancing the antioxidant activities of ARPE-19[53], and decreased apoptosis induced by oxidative stress in human retinal endothelial cells[54].

Eye pigmentation is essential to maintain visual function. The RPE contribute to absorption of scattered light and to reduce retinal damage from ultraviolet light by forming a dark-brown pigmented wall[35,55]. Studies on models in which Kl expression has been down-regulated revealed that Kl is involved in regulation of genes encoding for melanin biosynthesis[41]. Indeed, pigmentation of eyes from Kl k/o mice was reduced and their RPE cells contained fewer melanin granules than normal RPE cells[41].

All these findings suggest that retina homeostasis may be affected by altered expression of Kl, as well altered levels of soluble Kl (Table 1).

Raishidena® WJD | https://www.wjgnet.com

#### **KI AND DR**

Levels of sKl has been found reduced in ocular pathologies characterized by inflammatory state[56-59], suggesting that the reduced levels of sKI may be a common feature in several ocular diseases. In particular, decreased levels of KI may be associated with increased risk of onset and worsening of DR. Indeed, circulating levels of Kl are lower in diabetic subject with DR than in those without this complication [54,60]. Moreover, serum Kl levels are negatively correlated with progression of DR[54,60]. Following the onset of DR in diabetic patients reveals that patients with progression of retinopathy had lower levels of serum Kl as compared to those without [60]. In addition, Ji et al [54] found that levels of sKl are gradually reduced among patient with diabetes without DR, non-proliferative DR (PDR) and PDR, independently of DN[54]. Corcillo et al[60] hypothesize that a halving of circulating Kl levels may increase the risk of retinopathy progression by 44% [60]. On the other hand, the incidence of the functional "KL-VS" variant of the Kl gene, which is associated with higher longevity in humans, is lower in people with DR and is associated with reduced serum levels of inflammatory markers and pro-angiogenic factors, suggesting that this genotype may be protective against retinopathy incidence<sup>[61]</sup>.

As reported in the previous section, several experimental models demonstrated that depletion of KI negatively affects important function of retinal cells, including oxidative stress response, VEGF-A secretion, and phagocytosis, leading to activation of mechanisms that may contribute to onset and progression of DR. On the other hand, there are also several evidence that treatment with recombinant sKl or overexpression of Kl ameliorate retinal function.

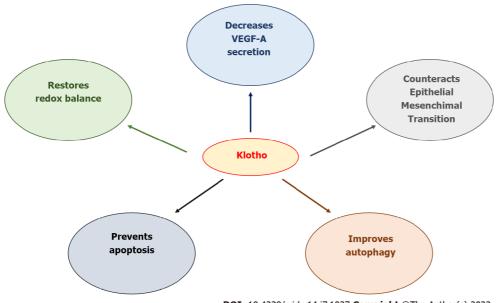
Oxidative stress and inflammation have been causative associated with DR [62,63]. It has been reported that Kl exerts protective effects against oxidative stress in retinal cells[13,41,42,53]. Firstly, it has been observed that pretreatment with sKl prevents increment of ROS production in ARPE-19 cells exposed to  $H_2O_2[41,53]$ . In particular, Wen *et al*[53] demonstrated that sKl improves redox balance in H<sub>2</sub>O<sub>2</sub>-treated ARPE-19 cells by increasing expression and nuclear translocation of nuclear factor E2-related factor 2 (Nrf2), thus restoring glutathione peroxidase, SOD2 and catalase to the levels of untreated cells[53]. In addition, pretreatment with sKl prevents H<sub>2</sub>O<sub>2</sub>-induced apoptosis of ARPE-19 cells[42,53], by increasing expression of Bcl-2 and decreasing the activation of caspase-3[53].

It is well established that VEGF-A plays an important role in driving pathological neovascularization of the retina during DR, and that neovascularization due to severe hypoxia is a hallmark of PDR[34]. The expression of VEGF-A is regulated by hypoxia-inducible factor- $1\alpha$  (HIF- $1\alpha$ ), which is a transcription factor involved in cellular response to hypoxia and hyperglycemia [64,65]. Interestingly, Kl levels have been found decreased in ARPE-19 cells exposed to hypoxia and in laser-induced CNV lesions in mice[66]. Xie et al[66] demonstrated that HIF-1a, besides directly increase VEGF-A transcription, may be responsible of down-regulation of KI expression during hypoxia[66]. Indeed, HIF-1α activates p53, which, in turns, leads to the increased levels of miRNA34, that targets KI thus reducing its expression[66]. Given that KI is expressed in ocular tissues, it is possible that part of the sKl that acts in the eye derives by local shedding of mKl, therefore its contribution may be lost when expression of Kl is down-regulated. It has been reported that treatment with KI reduces VEGF-A secretion from ARPE-19 cells[41]. In particular, KI was able to decrease VEGF-A secretion by reducing phosphorylation of both IGF-1 receptor (IGF-1R) and VEGR2. The pathogenic role of IGF-1 in the development of PDR is still debated, several studies indicate that increased activation of IGF-1 signaling may contribute to retinal neovascularization, however a strong relationship between IGF-1 and the development of proliferative retinopathy has not been still clearly demonstrated[67-69]. Several studies reported that IGF-1R signaling is regulated by lipid raft integrity and interaction with caveolin-1[70-74]. In particular, down-regulation of caveolin-1 expression in RPE cells significantly reduces both basal and IGF-1-stimulated VEGF-A secretion[72]. These data together with the ability of KI to modify the lipid organization within lipid rafts/caveolae[18] suggest that KI may reduce the phosphorylation of IGF-1R by altering these microdomains. Hyperglycemia increases production and secretion of VEGF-A by Muller cells in the retina. In particular, Yu et al[75] demonstrated that hyperglycemia increases the production of VEGF-A in Muller glial cells through the activation of FGFR1[75]. It is well known that sKl acts as a co-receptor for FGFs at non-renal sites and activates protective pathways in several cell types [76,77]. Interestingly, screening the potential pathogenic genes associated with DR revealed that hyperglycemia increases the expression of FGF23[78], and of its membrane receptor FGFR1 on Muller glial cells [75,79]. Considering that absence of Kl may allow Kl-independent activation of FGFRs resulting in pathological cellular changes [17,76,77], and that KI-independent action of FGF23 has been reported to contribute to endothelial dysfunction<sup>[17]</sup>, these findings suggest that lower levels of Kl together with increased production of FGF23 may contribute to the onset of DR and to progression to PDR by increasing VEGF-A production.

Autophagy is a highly conserved lysosomal pathway for the turnover of cytoplasmic organelles and long-lived proteins that acts as an adaptive response to cellular stresses and regulates homeostasis, differentiation, development and survival in several cell types[80]. In retinal cells, autophagy plays an important role by participating to POS degradation, visual pigment recycling, and lipofuscin degradation[81-83]. Altered activation of autophagy has been found in experimental models of DR and in the retina of diabetic patients [84,85]. For instance, RPE cells exposed to high glucose concentration increase formation of autophagosome, suggesting that induction of autophagy is a cytoprotective response against high glucose (HG)[84,85]. However, the excessive activation of this mechanism may lead to its impairment as occur in retinal Muller cells, where the process of degradation cannot be completed due to the lysosomal dysfunction [85]. It has been reported that autophagic activity is reduced in DM mice and human renal proximal tubule cells exposed to HG[86]. Recent studies showed that KI may act as a regulator of autophagy even in diabetic condition[87]. Specific expression of Kl significantly improves autophagy in both pancreatic beta cells and in renal tubule cells exposed to HG[29,86]. Moreover, Zou et al<sup>[21]</sup> showed that activation of 5' adenosine monophosphate-activated protein kinase (AMPK), a positive regulator of autophagy, is significantly decreased in the retina of Kl deficient mice as compared to that of WT mice[42]. Although there is no direct evidence, these finding suggest that KI may affect autophagy also in retinal cells. A decreased activation of AMPK has been observed also in arterial endothelial cells of KI deficient mice[88], confirming that



Puddu A et al. Role of Klotho in DR



DOI: 10.4239/wjd.v14.i7.1027 Copyright ©The Author(s) 2023



AMPK is a crucial mediator of protective effects of Kl. Moreover, Kl deficient mice have also reduced activity of silent information regulator (SIRT) 1[88], another important player in autophagy [89]. Interestingly, the expression of SIRT1 is reduced in DR and intravitreal administration of SIRT1 reverses DR in a mouse model of type 2 diabetes[90]. These results suggest that regulation of SIRT1 may be another mechanism through which KI improve DR.

PDR is also characterized by formation of fibrous proliferative anterior membrane[91]. Subretinal fibrosis is mediated by EMT, a process that leads RPE cells to the acquisition of a mesenchymal phenotype[92]. Several evidence demonstrated that HG induce EMT in RPE[93,94]. It has been shown that KI expression is down-regulated in models of induced fibrosis, suggesting a protective role of KI[22,95,96]. In particular, the protective effects of KI have been related to inhibition of the Wnt/ $\beta$ -catenin and the Egr-mediated signaling pathways. Recently, it has been reported that overexpression of KI decreased the expression of mesenchymal cell markers induced by hypoxia in ARPE-19 cells[66]. Moreover, overexpression of Kl was able to reduce subretinal fibrosis in a mouse laser-induced CNV model[66]. Here, under hypoxic conditions, KI was able to block the axis that through HIF-1 $\alpha$  leads to the activation of p53 and promotes EMT in RPE cells, confirming that Kl may be useful in preventing EMT also in RPE cells.

Besides hyperglycemia, dyslipidemia is another important actor in the progression of DR[97,98]. Palmitic acid (PA) is involved in the onset of DR and may induce endothelial cell damage[98]. It has been demonstrated that KI pretreatment significantly reduces apoptosis induced by PA in human retinal endothelial cells[54]. This effect implies the activation of the PI3K and subsequent phosphorylation of AKT[54]. Moreover, Kl affects expression of proteins involved in apoptosis leading to increased expression of the anti-apoptotic Bcl-2 and down-regulation of the pro-apoptotic Bax[54]. Consistent with these data, pretreatment with Kl reduced the apoptosis rate in ARPE-19 cells exposed to  $H_2O_2$  by up-regulating Bcl-2 expression and decreasing levels of Bax[53]. In addition, KI was able to prevent the decrease of mitochondrial membrane potential and the activation of Caspase-3 induced by  $H_2O_2[53]$ .

#### CONCLUSION

DR is a common complication of diabetes. The International Diabetes Federation estimated the global population with diabetes mellitus to be 463 million in 2019 and 700 million in 2045[99]. These data require the development of strategies able to prevents the onset and the progression of DR. To date, the first line treatment for PDR is intravitreal anti-VEGF therapy. However, it is not so successful for routine treatment of non-PDR[32,100]. Therefore, new molecules in development have been designed to target other pathways involved in pathogenesis of DR[101,102]. It has been demonstrated that Kl has protective effects in DN and that pathological mechanisms between DR and DN share similarities [19,29], suggesting that KI may be a good candidate in counteracting DR. Experimental models targeting KI have been shown to have positive effects on several mechanisms involved in DR onset and progression (Figure 1). Therefore, KI may become a novel biomarker and a good candidate for the treatment of DR[60].

#### FOOTNOTES

Author contributions: Puddu A and Maggi DC contributed equally to this work; Puddu A and Maggi DC contributed to the conception



and design of the article, interpretation of relevant literature, wrote the manuscript, revised the manuscript; All authors approved the final version of the manuscript.

**Conflict-of-interest statement:** All the authors report no relevant conflicts of interest for this article.

**Open-Access:** This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: https://creativecommons.org/Licenses/by-nc/4.0/

#### Country/Territory of origin: Italy

ORCID number: Alessandra Puddu 0000-0002-9084-2636; Davide Carlo Maggi 0000-0003-3928-1295.

S-Editor: Fan JR L-Editor: A P-Editor: Fan JR

#### REFERENCES

- Kuro-o M, Matsumura Y, Aizawa H, Kawaguchi H, Suga T, Utsugi T, Ohyama Y, Kurabayashi M, Kaname T, Kume E, Iwasaki H, Iida A, 1 Shiraki-Iida T, Nishikawa S, Nagai R, Nabeshima YI. Mutation of the mouse klotho gene leads to a syndrome resembling ageing. Nature 1997; **390**: 45-51 [PMID: 9363890 DOI: 10.1038/36285]
- 2 Hayashi Y, Okino N, Kakuta Y, Shikanai T, Tani M, Narimatsu H, Ito M. Klotho-related protein is a novel cytosolic neutral betaglycosylceramidase. J Biol Chem 2007; 282: 30889-30900 [PMID: 17595169 DOI: 10.1074/jbc.M700832200]
- Tohyama O, Imura A, Iwano A, Freund JN, Henrissat B, Fujimori T, Nabeshima Y. Klotho is a novel beta-glucuronidase capable of 3 hydrolyzing steroid beta-glucuronides. J Biol Chem 2004; 279: 9777-9784 [PMID: 14701853 DOI: 10.1074/jbc.M312392200]
- Xu Y, Sun Z. Molecular basis of Klotho: from gene to function in aging. Endocr Rev 2015; 36: 174-193 [PMID: 25695404 DOI: 4 10.1210/er.2013-1079
- 5 Hayashi Y, Ito M. Klotho-Related Protein KLrP: Structure and Functions. Vitam Horm 2016; 101: 1-16 [PMID: 27125736 DOI: 10.1016/bs.vh.2016.02.011]
- Dalton GD, Xie J, An SW, Huang CL. New Insights into the Mechanism of Action of Soluble Klotho. Front Endocrinol (Lausanne) 2017; 8: 6 323 [PMID: 29250031 DOI: 10.3389/fendo.2017.00323]
- 7 Wolf MT, An SW, Nie M, Bal MS, Huang CL. Klotho up-regulates renal calcium channel transient receptor potential vanilloid 5 (TRPV5) by intra- and extracellular N-glycosylation-dependent mechanisms. J Biol Chem 2014; 289: 35849-35857 [PMID: 25378396 DOI: 10.1074/jbc.M114.616649]
- Baranowska B, Kochanowski J. The metabolic, neuroprotective cardioprotective and antitumor effects of the Klotho protein. Neuro 8 Endocrinol Lett 2020; 41: 69-75 [PMID: 33185993]
- Hu MC, Shi M, Zhang J, Addo T, Cho HJ, Barker SL, Ravikumar P, Gillings N, Bian A, Sidhu SS, Kuro-o M, Moe OW. Renal Production, 9 Uptake, and Handling of Circulating αKlotho. J Am Soc Nephrol 2016; 27: 79-90 [PMID: 25977312 DOI: 10.1681/ASN.2014101030]
- Corrêa HL, Raab ATO, Araújo TM, Deus LA, Reis AL, Honorato FS, Rodrigues-Silva PL, Neves RVP, Brunetta HS, Mori MADS, Franco 10 OL, Rosa TDS. A systematic review and meta-analysis demonstrating Klotho as an emerging exerkine. Sci Rep 2022; 12: 17587 [PMID: 36266389 DOI: 10.1038/s41598-022-22123-1]
- Prud'homme GJ, Kurt M, Wang Q. Pathobiology of the Klotho Antiaging Protein and Therapeutic Considerations. Front Aging 2022; 3: 11 931331 [PMID: 35903083 DOI: 10.3389/fragi.2022.931331]
- Wolf I, Levanon-Cohen S, Bose S, Ligumsky H, Sredni B, Kanety H, Kuro-o M, Karlan B, Kaufman B, Koeffler HP, Rubinek T. Klotho: a 12 tumor suppressor and a modulator of the IGF-1 and FGF pathways in human breast cancer. Oncogene 2008; 27: 7094-7105 [PMID: 18762812 DOI: 10.1038/onc.2008.292]
- 13 Wang Y, Kuro-o M, Sun Z. Klotho gene delivery suppresses Nox2 expression and attenuates oxidative stress in rat aortic smooth muscle cells via the cAMP-PKA pathway. Aging Cell 2012; 11: 410-417 [PMID: 22260450 DOI: 10.1111/j.1474-9726.2012.00796.x]
- Kurosu H, Yamamoto M, Clark JD, Pastor JV, Nandi A, Gurnani P, McGuinness OP, Chikuda H, Yamaguchi M, Kawaguchi H, Shimomura I, 14 Takayama Y, Herz J, Kahn CR, Rosenblatt KP, Kuro-o M. Suppression of aging in mice by the hormone Klotho. Science 2005; 309: 1829-1833 [PMID: 16123266 DOI: 10.1126/science.1112766]
- Xie B, Zhou J, Shu G, Liu DC, Chen J, Yuan L. Restoration of klotho gene expression induces apoptosis and autophagy in gastric cancer cells: 15 tumor suppressive role of klotho in gastric cancer. Cancer Cell Int 2013; 13: 18 [PMID: 23432957 DOI: 10.1186/1475-2867-13-18]
- Yamamoto M, Clark JD, Pastor JV, Gurnani P, Nandi A, Kurosu H, Miyoshi M, Ogawa Y, Castrillon DH, Rosenblatt KP, Kuro-o M. 16 Regulation of oxidative stress by the anti-aging hormone klotho. J Biol Chem 2005; 280: 38029-38034 [PMID: 16186101 DOI: 10.1074/jbc.M509039200]
- 17 Richter B, Faul C. FGF23 Actions on Target Tissues-With and Without Klotho. Front Endocrinol (Lausanne) 2018; 9: 189 [PMID: 29770125 DOI: 10.3389/fendo.2018.00189]
- 18 Dalton G, An SW, Al-Juboori SI, Nischan N, Yoon J, Dobrinskikh E, Hilgemann DW, Xie J, Luby-Phelps K, Kohler JJ, Birnbaumer L, Huang CL. Soluble klotho binds monosialoganglioside to regulate membrane microdomains and growth factor signaling. Proc Natl Acad Sci USA 2017; 114: 752-757 [PMID: 28069944 DOI: 10.1073/pnas.1620301114]
- Zhang L, Liu T. Clinical implication of alterations in serum Klotho levels in patients with type 2 diabetes mellitus and its associated 19 complications. J Diabetes Complications 2018; 32: 922-930 [PMID: 30042059 DOI: 10.1016/j.jdiacomp.2018.06.002]
- Xue J, Wang L, Sun Z, Xing C. Basic Research in Diabetic Nephropathy Health Care: A study of the Renoprotective Mechanism of 20 Metformin. J Med Syst 2019; 43: 266 [PMID: 31273547 DOI: 10.1007/s10916-019-1412-4]



- Zou D, Wu W, He Y, Ma S, Gao J. The role of klotho in chronic kidney disease. BMC Nephrol 2018; 19: 285 [PMID: 30348110 DOI: 21 10.1186/s12882-018-1094-z]
- Li Y, Xue M, Hu F, Jia Y, Zheng Z, Yang Y, Liu X, Wang Y. Klotho prevents epithelial-mesenchymal transition through Egr-1 22 downregulation in diabetic kidney disease. BMJ Open Diabetes Res Care 2021; 9 [PMID: 34099438 DOI: 10.1136/bmjdrc-2020-002038]
- Typiak M, Kulesza T, Rachubik P, Rogacka D, Audzeyenka I, Angielski S, Saleem MA, Piwkowska A. Role of Klotho in Hyperglycemia: Its 23 Levels and Effects on Fibroblast Growth Factor Receptors, Glycolysis, and Glomerular Filtration. Int J Mol Sci 2021; 22 [PMID: 34360633 DOI: 10.3390/ijms22157867]
- Kacso IM, Bondor CI, Kacso G. Soluble serum Klotho in diabetic nephropathy: relationship to VEGF-A. Clin Biochem 2012; 45: 1415-1420 24 [PMID: 22836100 DOI: 10.1016/j.clinbiochem.2012.07.098]
- 25 Xin C, Sun X, Li Z, Gao T. Relationship of Soluble Klotho and Early Stage of Diabetic Nephropathy: A Systematic Review and Meta-Analysis. Front Endocrinol (Lausanne) 2022; 13: 902765 [PMID: 35692408 DOI: 10.3389/fendo.2022.902765]
- 26 Piwkowska A, Zdrojewski Ł, Heleniak Z, Dębska-Ślizień A. Novel Markers in Diabetic Kidney Disease-Current State and Perspectives. Diagnostics (Basel) 2022; 12 [PMID: 35626360 DOI: 10.3390/diagnostics12051205]
- Wang K, Mao Y, Lu M, Liu X, Sun Y, Li Z, Li Y, Ding Y, Zhang J, Hong J, Xu D. Association between serum Klotho levels and the 27 prevalence of diabetes among adults in the United States. Front Endocrinol (Lausanne) 2022; 13: 1005553 [PMID: 36440221 DOI: 10.3389/fendo.2022.1005553]
- Lin Y, Sun Z. Antiaging gene Klotho enhances glucose-induced insulin secretion by up-regulating plasma membrane levels of TRPV2 in 28 MIN6 β-cells. Endocrinology 2012; 153: 3029-3039 [PMID: 22597535 DOI: 10.1210/en.2012-1091]
- Lin Y, Sun Z. In vivo pancreatic  $\beta$ -cell-specific expression of antiaging gene Klotho: a novel approach for preserving  $\beta$ -cells in type 2 diabetes. 29 Diabetes 2015; 64: 1444-1458 [PMID: 25377875 DOI: 10.2337/db14-0632]
- Son DO, Liu W, Li X, Prud'homme GJ, Wang Q. Combined effect of GABA and glucagon-like peptide-1 receptor agonist on cytokine-induced 30 apoptosis in pancreatic β-cell line and isolated human islets. J Diabetes 2019; 11: 563-572 [PMID: 30520247 DOI: 10.1111/1753-0407.12881]
- Geng L, Liao B, Jin L, Yu J, Zhao X, Zhao Y, Zhong L, Wang B, Li J, Liu J, Yang JK, Jia W, Lian Q, Xu A. β-Klotho promotes glycolysis 31 and glucose-stimulated insulin secretion via GP130. Nat Metab 2022; 4: 608-626 [PMID: 35551509 DOI: 10.1038/s42255-022-00572-2]
- 32 Tan TE, Wong TY. Diabetic retinopathy: Looking forward to 2030. Front Endocrinol (Lausanne) 2022; 13: 1077669 [PMID: 36699020 DOI: 10.3389/fendo.2022.1077669]
- Solomon SD, Chew E, Duh EJ, Sobrin L, Sun JK, VanderBeek BL, Wykoff CC, Gardner TW. Diabetic Retinopathy: A Position Statement by 33 the American Diabetes Association. Diabetes Care 2017; 40: 412-418 [PMID: 28223445 DOI: 10.2337/dc16-2641]
- Wong TY, Cheung CM, Larsen M, Sharma S, Simó R. Diabetic retinopathy. Nat Rev Dis Primers 2016; 2: 16012 [PMID: 27159554 DOI: 34 10.1038/nrdp.2016.12]
- Strauss O. The retinal pigment epithelium in visual function. Physiol Rev 2005; 85: 845-881 [PMID: 15987797 DOI: 35 10.1152/physrev.00021.2004]
- Ponnalagu M, Subramani M, Jayadev C, Shetty R, Das D. Retinal pigment epithelium-secretome: A diabetic retinopathy perspective. Cytokine 36 2017; 95: 126-135 [PMID: 28282610 DOI: 10.1016/j.cyto.2017.02.013]
- Kannan R, Zhang N, Sreekumar PG, Spee CK, Rodriguez A, Barron E, Hinton DR. Stimulation of apical and basolateral VEGF-A and VEGF-37 C secretion by oxidative stress in polarized retinal pigment epithelial cells. Mol Vis 2006; 12: 1649-1659 [PMID: 17200665]
- Takahashi H, Shibuya M. The vascular endothelial growth factor (VEGF)/VEGF receptor system and its role under physiological and 38 pathological conditions. Clin Sci (Lond) 2005; 109: 227-241 [PMID: 16104843 DOI: 10.1042/CS20040370]
- Reish NJ, Maltare A, McKeown AS, Laszczyk AM, Kraft TW, Gross AK, King GD. The age-regulating protein klotho is vital to sustain 39 retinal function. Invest Ophthalmol Vis Sci 2013; 54: 6675-6685 [PMID: 24045987 DOI: 10.1167/iovs.13-12550]
- 40 Zhang Y, Wang L, Wu Z, Yu X, Du X, Li X. The Expressions of Klotho Family Genes in Human Ocular Tissues and in Anterior Lens Capsules of Age-Related Cataract. Curr Eye Res 2017; 42: 871-875 [PMID: 28095050 DOI: 10.1080/02713683.2016.1259421]
- Kokkinaki M, Abu-Asab M, Gunawardena N, Ahern G, Javidnia M, Young J, Golestaneh N. Klotho regulates retinal pigment epithelial 41 functions and protects against oxidative stress. J Neurosci 2013; 33: 16346-16359 [PMID: 24107965 DOI: 10.1523/JNEUROSCI.0402-13.2013]
- Zhou S, Hum J, Taskintuna K, Olaya S, Steinman J, Ma J, Golestaneh N. The Anti-Aging Hormone Klotho Promotes Retinal Pigment 42 Epithelium Cell Viability and Metabolism by Activating the AMPK/PGC-1α Pathway. Antioxidants (Basel) 2023; 12 [PMID: 36829944 DOI: 10.3390/antiox12020385]
- 43 Kwon W, Freeman SA. Phagocytosis by the Retinal Pigment Epithelium: Recognition, Resolution, Recycling. Front Immunol 2020; 11: 604205 [PMID: 33281830 DOI: 10.3389/fimmu.2020.604205]
- Karl MO, Kroeger W, Wimmers S, Milenkovic VM, Valtink M, Engelmann K, Strauss O. Endogenous Gas6 and Ca2+ -channel activation 44 modulate phagocytosis by retinal pigment epithelium. Cell Signal 2008; 20: 1159-1168 [PMID: 18395422 DOI: 10.1016/j.cellsig.2008.02.005]
- 45 Müller C, Más Gómez N, Ruth P, Strauss O. CaV1.3 L-type channels, maxiK Ca(2+)-dependent K(+) channels and bestrophin-1 regulate rhythmic photoreceptor outer segment phagocytosis by retinal pigment epithelial cells. Cell Signal 2014; 26: 968-978 [PMID: 24407175 DOI: 10.1016/j.cellsig.2013.12.021]
- Strauß O, Reichhart N, Gomez NM, Müller C. Contribution of Ion Channels in Calcium Signaling Regulating Phagocytosis: MaxiK, Cav1.3 46 and Bestrophin-1. Adv Exp Med Biol 2016; 854: 739-744 [PMID: 26427483 DOI: 10.1007/978-3-319-17121-0 98]
- 47 Chang Q, Hoefs S, van der Kemp AW, Topala CN, Bindels RJ, Hoenderop JG. The beta-glucuronidase klotho hydrolyzes and activates the TRPV5 channel. Science 2005; 310: 490-493 [PMID: 16239475 DOI: 10.1126/science.1114245]
- 48 Xuan NT, Hai NV. Changes in expression of klotho affect physiological processes, diseases, and cancer. Iran J Basic Med Sci 2018; 21: 3-8 [PMID: 29372030]
- Cordeiro S, Strauss O. Expression of Orai genes and I(CRAC) activation in the human retinal pigment epithelium. Graefes Arch Clin Exp 49 Ophthalmol 2011; 249: 47-54 [PMID: 20607548 DOI: 10.1007/s00417-010-1445-3]
- Kennedy BG, Torabi AJ, Kurzawa R, Echtenkamp SF, Mangini NJ. Expression of transient receptor potential vanilloid channels TRPV5 and 50 TRPV6 in retinal pigment epithelium. Mol Vis 2010; 16: 665-675 [PMID: 20405023]
- Kwak N, Okamoto N, Wood JM, Campochiaro PA. VEGF is major stimulator in model of choroidal neovascularization. Invest Ophthalmol Vis 51 Sci 2000; 41: 3158-3164 [PMID: 10967078]
- 52 Miller JW, Le Couter J, Strauss EC, Ferrara N. Vascular endothelial growth factor a in intraocular vascular disease. Ophthalmology 2013; 120: 106-114 [PMID: 23031671 DOI: 10.1016/j.ophtha.2012.07.038]



- Wen X, Li S, Zhang Y, Zhu L, Xi X, Zhang S, Li Y. Recombinant human klotho protects against hydrogen peroxide-mediated injury in human 53 retinal pigment epithelial cells via the PI3K/Akt-Nrf2/HO-1 signaling pathway. Bioengineered 2022; 13: 11767-11781 [PMID: 35543385 DOI: 10.1080/21655979.2022.2071023]
- 54 Ji B, Wei H, Ding Y, Liang H, Yao L, Wang H, Qu H, Deng H. Protective potential of klotho protein on diabetic retinopathy: Evidence from clinical and in vitro studies. J Diabetes Investig 2020; 11: 162-169 [PMID: 31197979 DOI: 10.1111/jdi.13100]
- Yang S, Zhou J, Li D. Functions and Diseases of the Retinal Pigment Epithelium. Front Pharmacol 2021; 12: 727870 [PMID: 34393803 DOI: 55 10.3389/fphar.2021.727870]
- Ahoor MH, Ghorbanihaghjo A, Sorkhabi R, Kiavar A. Klotho and Endothelin-1 in Pseudoexfoliation Syndrome and Glaucoma. J Glaucoma 56 2016; **25**: 919-922 [PMID: 27755351 DOI: 10.1097/IJG.00000000000553]
- 57 Ma Z, Liu J, Li J, Jiang H, Kong J. Klotho Levels are Decreased and Associated with Enhanced Oxidative Stress and Inflammation in the Aqueous Humor in Patients with Exudative Age-related Macular Degeneration. Ocul Immunol Inflamm 2022; 30: 630-637 [PMID: 33048602 DOI: 10.1080/09273948.2020.1828488]
- Tokuc EO, Yuksel N, Kır HM, Acar E. Evaluation of serum and aqueous humor klotho levels in pseudoexfoliation syndrome, 58 pseudoexfoliation and primary open-angle glaucoma. Int Ophthalmol 2021; 41: 2369-2375 [PMID: 33738657 DOI: 10.1007/s10792-021-01790-5]
- 59 Yamamoto K, Sato K, Yukita M, Yasuda M, Omodaka K, Ryu M, Fujita K, Nishiguchi KM, Machida S, Nakazawa T. The neuroprotective effect of latanoprost acts via klotho-mediated suppression of calpain activation after optic nerve transection. J Neurochem 2017; 140: 495-508 [PMID: 27859240 DOI: 10.1111/jnc.13902]
- Corcillo A, Fountoulakis N, Sohal A, Farrow F, Ayis S, Karalliedde J. Low levels of circulating anti-ageing hormone Klotho predict the onset 60 and progression of diabetic retinopathy. Diab Vasc Dis Res 2020; 17: 1479164120970901 [PMID: 33225726 DOI: 10.1177/1479164120970901]
- Słomiński B, Ryba-Stanisławowska M, Skrzypkowska M, Myśliwska J, Myśliwiec M. The KL-VS polymorphism of KLOTHO gene is 61 protective against retinopathy incidence in patients with type 1 diabetes. Biochim Biophys Acta Mol Basis Dis 2018; 1864: 758-763 [PMID: 29247834 DOI: 10.1016/j.bbadis.2017.12.015]
- Rübsam A, Parikh S, Fort PE. Role of Inflammation in Diabetic Retinopathy. Int J Mol Sci 2018; 19 [PMID: 29565290 DOI: 62 10.3390/ijms19040942]
- 63 Semeraro F, Cancarini A, dell'Omo R, Rezzola S, Romano MR, Costagliola C. Diabetic Retinopathy: Vascular and Inflammatory Disease. J Diabetes Res 2015; 2015: 582060 [PMID: 26137497 DOI: 10.1155/2015/582060]
- 64 Chang ML, Chiu CJ, Shang F, Taylor A. High glucose activates ChREBP-mediated HIF-1a and VEGF expression in human RPE cells under normoxia. Adv Exp Med Biol 2014; 801: 609-621 [PMID: 24664750 DOI: 10.1007/978-1-4614-3209-8 77]
- Xiao Q, Zeng S, Ling S, Lv M. Up-regulation of HIF-1alpha and VEGF expression by elevated glucose concentration and hypoxia in cultured 65 human retinal pigment epithelial cells. J Huazhong Univ Sci Technolog Med Sci 2006; 26: 463-465 [PMID: 17120749 DOI: 10.1007/s11596-006-0422-x
- 66 Xie L, Wang Y, Li Q, Ji X, Tu Y, Du S, Lou H, Zeng X, Zhu L, Zhang J, Zhu M. The HIF-1α/p53/miRNA-34a/Klotho axis in retinal pigment epithelial cells promotes subretinal fibrosis and exacerbates choroidal neovascularization. J Cell Mol Med 2021; 25: 1700-1711 [PMID: 33438362 DOI: 10.1111/jcmm.16272]
- Arroba AI, Campos-Caro A, Aguilar-Diosdado M, Valverde ÁM. IGF-1, Inflammation and Retinal Degeneration: A Close Network. Front 67 Aging Neurosci 2018; 10: 203 [PMID: 30026694 DOI: 10.3389/fnagi.2018.00203]
- Raman P, Singal AK, Behl A. Effect of Insulin-Like Growth Factor-1 on Diabetic Retinopathy in Pubertal Age Patients With Type 1 Diabetes. 68 Asia Pac J Ophthalmol (Phila) 2019; 8: 319-323 [PMID: 31369407 DOI: 10.1097/APO.00000000000250]
- Wu TE, Chen HS. The role of growth hormone and IGF-1 in retinopathy: a prospective study of retinopathy in patients with acromegaly and 69 impaired fasting glucose. Diabetol Metab Syndr 2022; 14: 38 [PMID: 35248150 DOI: 10.1186/s13098-022-00806-z]
- Hong S, Huo H, Xu J, Liao K. Insulin-like growth factor-1 receptor signaling in 3T3-L1 adipocyte differentiation requires lipid rafts but not 70 caveolae. Cell Death Differ 2004; 11: 714-723 [PMID: 15002041 DOI: 10.1038/sj.cdd.4401405]
- Martins AS, Ordóñez JL, Amaral AT, Prins F, Floris G, Debiec-Rychter M, Hogendoorn PC, de Alava E. IGF1R signaling in Ewing sarcoma 71 is shaped by clathrin-/caveolin-dependent endocytosis. PLoS One 2011; 6: e19846 [PMID: 21611203 DOI: 10.1371/journal.pone.0019846]
- Puddu A, Sanguineti R, Maggi D. Caveolin-1 Down-Regulation Reduces VEGF-A Secretion Induced by IGF-1 in ARPE-19 Cells. Life (Basel) 72 2021; 12 [PMID: 35054437 DOI: 10.3390/life12010044]
- Salani B, Briatore L, Garibaldi S, Cordera R, Maggi D. Caveolin-1 down-regulation inhibits insulin-like growth factor-I receptor signal 73 transduction in H9C2 rat cardiomyoblasts. Endocrinology 2008; 149: 461-465 [PMID: 18039791 DOI: 10.1210/en.2007-0312]
- Salani B, Passalacqua M, Maffioli S, Briatore L, Hamoudane M, Contini P, Cordera R, Maggi D. IGF-IR internalizes with Caveolin-1 and 74 PTRF/Cavin in HaCat cells. PLoS One 2010; 5: e14157 [PMID: 21152401 DOI: 10.1371/journal.pone.0014157]
- 75 Yu Y, Bao Z, Wang X, Gong W, Chen H, Guan H, Le Y, Su S, Chen K, Wang JM. The G-Protein-Coupled Chemoattractant Receptor Fpr2 Exacerbates High Glucose-Mediated Proinflammatory Responses of Müller Glial Cells. Front Immunol 2017; 8: 1852 [PMID: 29312335 DOI: 10.3389/fimmu.2017.01852]
- Han X, Cai C, Xiao Z, Quarles LD. FGF23 induced left ventricular hypertrophy mediated by FGFR4 signaling in the myocardium is attenuated 76 by soluble Klotho in mice. J Mol Cell Cardiol 2020; 138: 66-74 [PMID: 31758962 DOI: 10.1016/j.yjmcc.2019.11.149]
- 77 Yanucil C, Kentrup D, Campos I, Czaya B, Heitman K, Westbrook D, Osis G, Grabner A, Wende AR, Vallejo J, Wacker MJ, Navarro-Garcia JA, Ruiz-Hurtado G, Zhang F, Song Y, Linhardt RJ, White K, Kapiloff MS, Faul C. Soluble α-klotho and heparin modulate the pathologic cardiac actions of fibroblast growth factor 23 in chronic kidney disease. Kidney Int 2022; 102: 261-279 [PMID: 35513125 DOI: 10.1016/j.kint.2022.03.028]
- 78 Gu C, Lhamo T, Zou C, Zhou C, Su T, Draga D, Luo D, Zheng Z, Yin L, Qiu Q. Comprehensive analysis of angiogenesis-related genes and pathways in early diabetic retinopathy. BMC Med Genomics 2020; 13: 142 [PMID: 32993645 DOI: 10.1186/s12920-020-00799-6]
- Hueber A, Wiedemann P, Esser P, Heimann K. Basic fibroblast growth factor mRNA, bFGF peptide and FGF receptor in epiretinal 79 membranes of intraocular proliferative disorders (PVR and PDR). Int Ophthalmol 20: 345-350 [PMID: 9237137 DOI: 10.1007/BF00176889]
- Glick D, Barth S, Macleod KF. Autophagy: cellular and molecular mechanisms. J Pathol 2010; 221: 3-12 [PMID: 20225336 DOI: 80 10.1002/path.2697]
- Lei L, Tzekov R, Li H, McDowell JH, Gao G, Smith WC, Tang S, Kaushal S. Inhibition or Stimulation of Autophagy Affects Early Formation 81 of Lipofuscin-Like Autofluorescence in the Retinal Pigment Epithelium Cell. Int J Mol Sci 2017; 18 [PMID: 28353645 DOI:



10.3390/ijms18040728]

- 82 Mathew B, Chennakesavalu M, Sharma M, Torres LA, Stelman CR, Tran S, Patel R, Burg N, Salkovski M, Kadzielawa K, Seiler F, Aldrich LN, Roth S. Autophagy and post-ischemic conditioning in retinal ischemia. Autophagy 2021; 17: 1479-1499 [PMID: 32452260 DOI: 10.1080/15548627.2020.1767371]
- Villarejo-Zori B, Jiménez-Loygorri JI, Zapata-Muñoz J, Bell K, Boya P. New insights into the role of autophagy in retinal and eye diseases. 83 Mol Aspects Med 2021; 82: 101038 [PMID: 34620506 DOI: 10.1016/j.mam.2021.101038]
- 84 Dehdashtian E, Mehrzadi S, Yousefi B, Hosseinzadeh A, Reiter RJ, Safa M, Ghaznavi H, Naseripour M. Diabetic retinopathy pathogenesis and the ameliorating effects of melatonin; involvement of autophagy, inflammation and oxidative stress. Life Sci 2018; 193: 20-33 [PMID: 29203148 DOI: 10.1016/j.lfs.2017.12.001]
- 85 Lopes de Faria JM, Duarte DA, Montemurro C, Papadimitriou A, Consonni SR, Lopes de Faria JB. Defective Autophagy in Diabetic Retinopathy. Invest Ophthalmol Vis Sci 2016; 57: 4356-4366 [PMID: 27564518 DOI: 10.1167/iovs.16-19197]
- Xue M, Yang F, Le Y, Yang Y, Wang B, Jia Y, Zheng Z, Xue Y. Klotho protects against diabetic kidney disease via AMPK- and ERK-86 mediated autophagy. Acta Diabetol 2021; 58: 1413-1423 [PMID: 34046744 DOI: 10.1007/s00592-021-01736-4]
- 87 Zhou H, Pu S, Zhou H, Guo Y. Klotho as Potential Autophagy Regulator and Therapeutic Target. Front Pharmacol 2021; 12: 755366 [PMID: 34737707 DOI: 10.3389/fphar.2021.755366]
- Gao D, Zuo Z, Tian J, Ali Q, Lin Y, Lei H, Sun Z. Activation of SIRT1 Attenuates Klotho Deficiency-Induced Arterial Stiffness and 88 Hypertension by Enhancing AMP-Activated Protein Kinase Activity. Hypertension 2016; 68: 1191-1199 [PMID: 27620389 DOI: 10.1161/HYPERTENSIONAHA.116.07709
- Kim JY, Mondaca-Ruff D, Singh S, Wang Y. SIRT1 and Autophagy: Implications in Endocrine Disorders. Front Endocrinol (Lausanne) 89 2022; 13: 930919 [PMID: 35909524 DOI: 10.3389/fendo.2022.930919]
- Adu-Agyeiwaah Y, Vieira CP, Asare-Bediako B, Li Calzi S, DuPont M, Floyd J, Boye S, Chiodo V, Busik JV, Grant MB. Intravitreal 90 Administration of AAV2-SIRT1 Reverses Diabetic Retinopathy in a Mouse Model of Type 2 Diabetes. Transl Vis Sci Technol 2023; 12: 20 [PMID: 37070938 DOI: 10.1167/tvst.12.4.20]
- Nawaz IM, Rezzola S, Cancarini A, Russo A, Costagliola C, Semeraro F, Presta M. Human vitreous in proliferative diabetic retinopathy: 91 Characterization and translational implications. Prog Retin Eye Res 2019; 72: 100756 [PMID: 30951889 DOI: 10.1016/j.preteyeres.2019.03.002]
- Lamouille S, Xu J, Derynck R. Molecular mechanisms of epithelial-mesenchymal transition. Nat Rev Mol Cell Biol 2014; 15: 178-196 [PMID: 92 24556840 DOI: 10.1038/nrm3758]
- Che D, Zhou T, Lan Y, Xie J, Gong H, Li C, Feng J, Hong H, Qi W, Ma C, Wu Q, Yang X, Gao G. High glucose-induced epithelial-93 mesenchymal transition contributes to the upregulation of fibrogenic factors in retinal pigment epithelial cells. Int J Mol Med 2016; 38: 1815-1822 [PMID: 27748912 DOI: 10.3892/ijmm.2016.2768]
- You ZP, Chen SS, Yang ZY, Li SR, Xiong F, Liu T, Fu SH. GEP100/ARF6 regulates VEGFR2 signaling to facilitate high-glucose-induced 94 epithelial-mesenchymal transition and cell permeability in retinal pigment epithelial cells. Am J Physiol Cell Physiol 2019; 316: C782-C791 [PMID: 30540496 DOI: 10.1152/ajpcell.00312.2018]
- Li X, Lu P, Shao XF, Jiang T, Liu F, Li G. Klotho Regulates Epithelial-to-Mesenchymal Transition In Vitro via Wnt/β-Catenin Pathway and 95 Attenuates Chronic Allograft Dysfunction in a Rat Renal Transplant Model. Ann Transplant 2021; 26: e930066 [PMID: 33737505 DOI: 10.12659/AOT.930066]
- 96 Yang Z, Zhan YW, Huang YY, Huang W, Zhan F, Lin SD. Regulation of epithelial mesenchymal transition by the renin-angiotensin system: a role for klotho in renal tubular epithelial cells. J Biol Regul Homeost Agents 2020; 34: 57-67 [PMID: 32466632 DOI: 10.23812/19-410-A-27]
- Kowluru RA, Mishra M, Kowluru A, Kumar B. Hyperlipidemia and the development of diabetic retinopathy: Comparison between type 1 and 97 type 2 animal models. Metabolism 2016; 65: 1570-1581 [PMID: 27621192 DOI: 10.1016/j.metabol.2016.07.012]
- Kumar B, Kowluru A, Kowluru RA. Lipotoxicity augments glucotoxicity-induced mitochondrial damage in the development of diabetic 98 retinopathy. Invest Ophthalmol Vis Sci 2015; 56: 2985-2992 [PMID: 26024084 DOI: 10.1167/iovs.15-16466]
- Saeedi P, Petersohn I, Salpea P, Malanda B, Karuranga S, Unwin N, Colagiuri S, Guariguata L, Motala AA, Ogurtsova K, Shaw JE, Bright D, 99 Williams R; IDF Diabetes Atlas Committee. Global and regional diabetes prevalence estimates for 2019 and projections for 2030 and 2045: Results from the International Diabetes Federation Diabetes Atlas, 9(th) edition. Diabetes Res Clin Pract 2019; 157: 107843 [PMID: 31518657 DOI: 10.1016/j.diabres.2019.107843]
- Gonzalez-Cortes JH, Martinez-Pacheco VA, Gonzalez-Cantu JE, Bilgic A, de Ribot FM, Sudhalkar A, Mohamed-Hamsho J, Kodjikian L, 100 Mathis T. Current Treatments and Innovations in Diabetic Retinopathy and Diabetic Macular Edema. Pharmaceutics 2022; 15 [PMID: 36678750 DOI: 10.3390/pharmaceutics15010122]
- Xia HQ, Yang JR, Zhang KX, Dong RL, Yuan H, Wang YC, Zhou H, Li XM. Molecules related to diabetic retinopathy in the vitreous and 101 involved pathways. Int J Ophthalmol 2022; 15: 1180-1189 [PMID: 35919310 DOI: 10.18240/ijo.2022.07.20]
- Muniyandi A, Hartman GD, Song Y, Mijit M, Kelley MR, Corson TW. Beyond VEGF: targeting inflammation and other pathways for 102 treatment of retinal disease. J Pharmacol Exp Ther 2023; 386: 15-25 [PMID: 3714244] DOI: 10.1124/jpet.122.001563]





### Published by Baishideng Publishing Group Inc 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA Telephone: +1-925-3991568 E-mail: bpgoffice@wjgnet.com Help Desk: https://www.f6publishing.com/helpdesk https://www.wjgnet.com

