W J S C World Journal of Stem Cells

Submit a Manuscript: https://www.f6publishing.com

World J Stem Cells 2024 April 26; 16(4): 375-388

DOI: 10.4252/wjsc.v16.i4.375

ISSN 1948-0210 (online)

REVIEW

Interplay between mesenchymal stem cells and macrophages: Promoting bone tissue repair

Fei-Fan Zhang, Yang Hao, Kuai-Xiang Zhang, Jiang-Jia Yang, Zhi-Qiang Zhao, Hong-Jian Liu, Ji-Tian Li

Specialty type: Cell and tissue engineering

Provenance and peer review: Invited article; Externally peer reviewed.

Peer-review model: Single blind

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): 0 Grade C (Good): C, C Grade D (Fair): 0 Grade E (Poor): 0

P-Reviewer: Gutiérrez-Cuevas J, Mexico

Received: December 30, 2023 Peer-review started: December 30, 2023

First decision: January 17, 2024 Revised: February 14, 2024 Accepted: March 19, 2024 Article in press: March 19, 2024 Published online: April 26, 2024



Fei-Fan Zhang, Yang Hao, Kuai-Xiang Zhang, Jiang-Jia Yang, Zhi-Qiang Zhao, Ji-Tian Li, Molecular Biology Lab, Henan Luoyang Orthopedic Hospital (Henan Provincial Orthopedic Hospital), Zhengzhou 450000, Henan Province, China

Fei-Fan Zhang, Jiang-Jia Yang, Ji-Tian Li, Graduate School, Hunan University of Chinese Medicine, Changsha 410208, Hunan Province, China

Yang Hao, Kuai-Xiang Zhang, Ji-Tian Li, Graduate School, Henan University of Chinese Medicine, Zhengzhou 450046, Henan Province, China

Hong-Jian Liu, Department of Orthopaedics, The First Affiliated Hospital of Zhengzhou University, Zhengzhou 450052, Henan Province, China

Corresponding author: Ji-Tian Li, MD, PhD, Doctor, Professor, Molecular Biology Lab, Henan Luoyang Orthopedic Hospital (Henan Provincial Orthopedic Hospital), No. 100 Yongping Road, Zhengzhou 450000, Henan Province, China. jitianlee@hotmail.com

Abstract

The repair of bone tissue damage is a complex process that is well-orchestrated in time and space, a focus and difficulty in orthopedic treatment. In recent years, the success of mesenchymal stem cells (MSCs)-mediated bone repair in clinical trials of large-area bone defects and bone necrosis has made it a candidate in bone tissue repair engineering and regenerative medicine. MSCs are closely related to macrophages. On one hand, MSCs regulate the immune regulatory function by influencing macrophages proliferation, infiltration, and phenotype polarization, while also affecting the osteoclasts differentiation of macrophages. On the other hand, macrophages activate MSCs and mediate the multilineage differentiation of MSCs by regulating the immune microenvironment. The cross-talk between MSCs and macrophages plays a crucial role in regulating the immune system and in promoting tissue regeneration. Making full use of the relationship between MSCs and macrophages will enhance the efficacy of MSCs therapy in bone tissue repair, and will also provide a reference for further application of MSCs in other diseases.

Key Words: Bone tissue damage; Inflammation; Macrophages; Mesenchymal stem cells; Tissue regeneration

©The Author(s) 2024. Published by Baishideng Publishing Group Inc. All rights reserved.



Core Tip: Bone regeneration has always been a challenge and priority in the treatment of orthopedic diseases. The interaction between mesenchymal stem cells (MSCs) and macrophages mediating the multilineage differentiation potential and immunomodulatory capabilities plays a crucial role in bone tissue repair and remodeling. Therefore, we reviewed the interactions between MSCs and macrophages, summarized the roles and potential of MSCs and macrophages in bone tissue regeneration, and looked forward to how to better utilize their relationship to enhance the efficacy of MSCs therapy in orthopedic diseases such as bone defects and osteoarthritis.

Citation: Zhang FF, Hao Y, Zhang KX, Yang JJ, Zhao ZQ, Liu HJ, Li JT. Interplay between mesenchymal stem cells and macrophages: Promoting bone tissue repair. World J Stem Cells 2024; 16(4): 375-388 URL: https://www.wjgnet.com/1948-0210/full/v16/i4/375.htm DOI: https://dx.doi.org/10.4252/wjsc.v16.i4.375

INTRODUCTION

The destruction of bone tissue caused by violent injuries and chronic inflammation is the core of various orthopedic conditions, such as bone defects, osteoarthritis, and osteoporosis. The repair of bone tissue damage is a complex process that is spatiotemporally well orchestrated. It has long been the focus of but also a difficulty in orthopedic treatment. Inflammation, tissue repair, and tissue remodeling are three important stages of tissue regeneration, involving strictly regulated participation of various types of cells. The immune system plays a core role in tissue repair and regeneration, and the immune response to tissue damage determines the speed and outcome of tissue healing [1]. Mesenchymal stem cells (MSCs) and macrophages are the sources of bone tissue. Moreover, MSCs and macrophages play important collaborative roles in immune regulation and in bone tissue regeneration as regulators and effectors of the inflammatory response^[2]. MSCs mediate the phenotypic polarization of macrophages that are involved in immune regulation of the body. The pro-inflammatory and anti-inflammatory factors secreted by polarized macrophages mediate the biological functions of MSCs such as adhesion, migration, immune activity, and osteogenic activity. On these bases, the osteogenesis mediated by MSCs and the bone resorption mediated by macrophages-derived osteoclasts (OCs) together maintain bone homeostasis and bone remodeling[3].

MSCs, versatile and plastic effector cells, play multiple roles in bone tissue repair and regeneration. On the one hand, as pluripotent stem cells, MSCs have the abilities to self-renew and differentiate into multiple lineages. Under the influence of injury signals and chemokines, MSCs migrate to the site of injury, differentiate directionally, and secrete various growth factors, such as transforming growth factor-beta (TGF- β), vascular endothelial growth factor (VEGF), epidermal growth factor, stromal cell-derived factor-1 (SDF-1), and fibroblast growth factor, to promote cell homing and stimulate tissue regeneration [4,5]. Factors in the injured environment, such as interferon gamma (IFN- γ) and tumor necrosis factor- α (TNF- α), stimulate the multilineage differentiation potential of MSCs, including osteogenic, adipogenic, and angiogenic differentiation, forming the various cells required for tissue repair [5]. On the other hand, as the first line of defense of the innate immune system, MSCs maintain the stability of the tissue microenvironment by controlling the inflammatory cascade [6]. MSCs can be activated by biomarkers in the inflammatory environment and respond to these signals through homing or paracrine effects, secreting soluble factors, such as prostaglandin E2 (PGE2), TGF- β 1, indoleamine 2,3-dioxygenase (IDO), interleukin (IL)-6, and IL-10, as well as proteins and RNAs that control macrophages differentiation and promote the anti-inflammatory phenotypic transformation of macrophages to inhibit uncontrolled immune responses[4,5,7-11].

Macrophages, as important innate immune cells, are closely related to cells involved in bone tissue repair and regeneration[12]. In addition to being responsible for clearing invading organisms, as well as cells and cell fragments after tissue damage, macrophages also demonstrate functional plasticity[13,14]. Macrophages play different roles at various stages of tissue repair, regulating inflammation by secreting pro-inflammatory and anti-inflammatory factors, and simultaneously secreting various biologically active factors such as chemokines, matrix metalloproteinases (MMPs), growth factors, and osteogenesis-related proteins, which affect the recruitment, migration, proliferation, and differentiation of various tissue repair-related cells, including MSCs[15-19]. The reduction of macrophages can lead to the disruption of the tissue microenvironment and the extension of tissue healing time[20]. Due to the critical role of macrophages in tissue repair, they have emerged as a potential target for therapeutic tissue regeneration strategies.

In this review, based on the importance of the immune response in the repair of bone tissue damage, we explored the dual role of MSCs and macrophages and their mutual regulation to maintain different functional phenotypes in immune regulation and bone regeneration. We then discussed the prospects of these cells in bone tissue repair.

BIDIRECTIONAL INTERACTION BETWEEN MSCS AND MACROPHAGES

MSCS can interact with macrophages in mutual regulation of the host microenvironment. MSCs can promote the basic functions of macrophages and mediate the polarization of macrophages phenotypes. Macrophages can secrete different active factors to regulate the immune microenvironment and affect the biological functions of MSCs through their own polarization (Figure 1)[4,21-43].



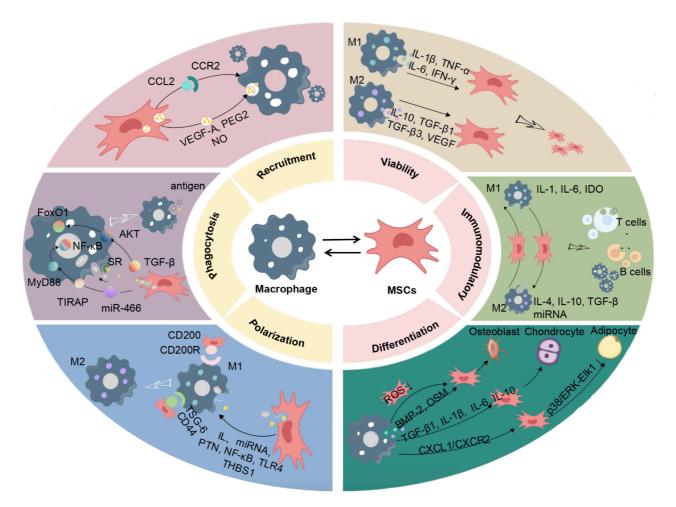


Figure 1 Bidirectional interaction between mesenchymal stem cells and macrophages. Mesenchymal stem cells (MSCs) can facilitate the proliferation and infiltration of macrophages, enhance their phagocytic capabilities, and promote the polarization of macrophages phenotypes. Macrophages are capable of activating MSCs, boosting their proliferation and migration abilities, influencing the immune regulatory microenvironment of MSCs, and promoting the multilineage differentiation of MSCs through the secretion of inflammatory-related factors, chemokines, and osteogenic-related factors. MSC: Mesenchymal stem cell; CCL2: C-C motif chemokine ligand 2; CCR2: C-C motif chemokine receptor 2; VEGF-A: Vascular endothelial growth factor-A; PEG2: Prostaglandin E2; IL: Interleukin; IFN: Interferon; TGF-B: Transforming growth factor beta; FoxO1: Forkhead box transcription factor O1; NF-kB: Nuclear factor kappa-B; MyD88: Myeloid differentiation primary response gene 88; TIRAP: Toll-interleukin-1 receptor domain-containing adaptor protein; TIR: Toll-interleukin-1 receptor; SR: Scavenger receptor; AKT: Protein kinase B; PTN: Pleiotropic hormones; TLR4: Toll-like receptor 4; THBS-1: Thrombospondin-1; BMP-2: Bone morphogenetic protein-2; OSM: Oncostatin-M; IDO: Indoleamine 2,3-dioxygenase; HGF: Hepatocyte growth factor; miRNA: MicroRNA; ROS: Reactive oxygen species; NO: Nitric oxide.

THE ROLES OF MSCS AS RELATED TO MACROPHAGES

Macrophages are an important part of the human immune system. Their polarization states and biological function changes are closely related to the occurrence and development of various diseases[12]. Proteins, RNA, chemokines, and other regulatory factors produced by MSCs enter macrophages through endocytosis, regulating macrophages genes expression, functional phenotypes, and also physiological and pathological states[44,45]. By mediating macrophages phenotype and function, MSCs participate in regulating inflammation, tissue healing, immune diseases, tumor growth, and other biological processes, playing an important role in the treatment of orthopedic diseases, respiratory diseases, urological diseases, etc.[17].

MSCs promote the recruitment of macrophages

MSCs promote macrophages recruitment by interacting with macrophages. Chemokines are considered to be chemotactic agents for immune cells, and macrophages play a key role in inflammatory responses by migrating along an increased chemokine gradient signal. C-C motif chemokine ligand 2 (CCL2) is an inflammatory chemokine that regulates the recruitment of immune cells during inflammation[46]. The release of CCL2 by MSCs and its binding to C-C motif chemokine receptor 2 (CCR2) may increase intracellular calcium ion influx through the Janus kinase (JAK)/signaling and transcription activator (STAT)1/STAT3 signaling pathway, activate important inflammatory signaling cascades, and promote the recruitment and infiltration of macrophages[47,48]. MSCs express various bioactive factors, such as VEGF, PGE2, and nitric oxide (NO) through exosomes, which may regulate macrophages recruitment via endocytosis[22,23]. Additionally, apoptosis of MSCs can inhibit macrophages recruitment. Research conducted by Zheng et al[49] revealed that apoptotic MSCs release apoptotic vesicles, which are engulfed by macrophages and induce macrophages reprogramming at the transcriptional level, inhibiting the production of pro-inflammatory cytokines, such as TNF- α , IL-1 β , IL-

Baishidena® WJSC | https://www.wjgnet.com

6, and, IL-8, and, thus inhibiting the accumulation of macrophages. This mechanism involves the autocrine or paracrine action of MSC-derived TGF- β [49].

MSCs enhance the phagocytic ability of macrophages

MSCs can secrete a variety of active ingredients to regulate the phagocytic ability of macrophages. TGF-β secreted by MSCs can stimulate the polarization of macrophages toward the M2 type and enhance their phagocytic ability through AKT/forkhead box transcription factor O1 signaling pathway[24]. MiR-466 secreted by MSCs, increases the phagocytic ability of macrophages by downregulating the Toll-interleukin-1 receptor domain-containing adaptor protein (TIRAP)-Myeloid differentiation primary response gene 88 (MyD88)-nuclear factor kappa-B (NF-κB) signaling pathway[25]. Scavenger receptor (SR), a receptor with extensive ligand binding ability, is related to the phagocytic function of macrophages. After interacting with macrophages, MSCs can upregulate the expression of SR in macrophages, enhance their phagocytic ability for microbes, and reduce further stimulation of immune cells by invading microbes[26]. In addition, MSCs mitochondria also play an important role in regulating macrophages function. Studies have found that MSCs can transfer mitochondria to macrophages through tunneling nanotube like structures, which not only promotes the oxidative phosphorylation of macrophages but also enhances their phagocytic ability[50].

MSCs mediate the phenotypic transformation of macrophages

MSCs primarily regulate macrophages through paracrine effects, direct cell contact, mitochondrial transfer, and cell burial. Mechanisms mediated by paracrine factors, such as cytokines and hormones are the main pathways by which MSCs participate in tissue damage repair. Macrophages, derived from monocytes, are involved in innate immunity and cell-mediated immunity in the body and can be polarized into M1 and M2 subtypes. Increasing evidence suggests that extracellular vesicles secreted by MSCs play a key role in regulating the balance of M1/M2 macrophages, but the exact mechanism remains unclear[51]. Pleiotropic hormones (PTNs) are critical regulators of macrophages differentiation, and MSCs maintain the M2-like phenotype by secreting PTNs to regulate the syndecan-3 receptor-mediated cell adhesion molecule pathway [30]. Liu *et al* [31] found that thrombos pondin-1 secreted by MSCs could induce the activation of TGF- β and stimulate the polarization of macrophages toward the M2 phenotype. In addition, other microRNAs (miRNAs), through extracellular vesicles, affect the levels of transcription, post-transcriptional modifications, and metabolism in macrophages, regulating their function and phenotype, and participating in physiological and pathological processes. MSC-derived small extracellular vesicles can transfer miR-223-3p, miRNA-124-3p, miRNA-125a, etc., to macrophages, change gene expression and biological activity, and promote the polarization of macrophages from M1 to M2 anti-inflammatory phenotype[28,52,53]. Abe et al[54] found that MSCs could release paracrine factors through cell-cell contact, further inducing the polarization of macrophages from M1 to M2, and significantly reducing the expression of proinflammatory factors such as IL-6 and TNF- α [29,54]. Lu *et al*[55] found that extracellular vesicles derived from MSCs could promote the polarization of M2 macrophages after transferring mitochondria to macrophages. Ghahremani Piraghaj et al[56] found that the apoptotic vesicles of MSCs were engulfed by macrophages, promoting M2 polarization of macrophages, reducing the production of $TNF-\alpha$ and NO, and increasing the level of IL-10.

THE EFFECTS OF MACROPHAGES ON MSCS

The functional phenotype and secretory profile of MSCs, as well as their availability, are regulated by the microenvironment, which determines their tissue repair capabilities. Multiple studies have shown that while MSCs regulate macrophages, the change in the polarization phenotype of macrophages also affects the biological activity and function of MSCs.

Macrophages affect the migration and viability of MSCs

Homing is one of the three major mechanisms of MSCs. In the damaged microenvironment, various chemokines, adhesion factors, and growth factors bind to receptors on the surface of MSCs, driving MSCs to migrate directionally to inflamed or damaged areas and participate in tissue repair. Multiple chemokines are indispensable for MSCs homing. Various chemokine receptors are present on MSCs, such as CCR4, CCR7, CCR9, C chemokine receptor type 6 (CXCR6), and SDF1 α [57-59]. The most studied is the SDF-1/CXCR4 axis, whose main functions include regulating the transplantation, chemotaxis, and homing of precursor cells to damaged sites. It is a key signal mediating the migration and distribution of MSCs[60]. Numerous studies have shown that IL-1β has an important impact on the migration activity of MSCs. IL-1β can enhance the expression of CXCR4 and MMP1 on MSCs, thereby promoting the endogenous recruitment of MSCs[33,61]. TNF- α is considered to be a promoter of MSCs. The anti-inflammatory IL-10 secreted by macrophages can enhance the promoting effect of TNF-a. Leucine-rich alpha-2-glycoprotein 1 (LRG1) is a proangiogenic protein that can promote the adhesion and migration of MSCs. TNF-α secreted by pro-inflammatory M1 macrophages can induce the secretion of LRG1, recruit MSCs, and promote angiogenesis and new bone formation[62]. Under hypoxic conditions, exosomes produced by M1 polarized macrophages can significantly reduce the viability and migration of MSCs, and induce MSC apoptosis by transporting miR-222 into MSCs[63]. High mobility group box 1 (HMGB1), NF-κB, and other inflammatory factors secreted by macrophages in the inflammatory microenvironment of bone injury have been shown to enhance the MSCs attachment and migration via Rap1 activation[64-66]. In addition, macrophages also play a certain regulatory role in the proliferation ability of MSCs. IL-1β, IL-6, TNF-α, and IFN-γ secreted by M1 macrophages can inhibit the proliferation of MSCs, while IL-10, TGF-β1, TGF-β3, and VEGF secreted by M2 macrophages can promote the proliferation of MSCs[4,5].



Macrophages mediate the immunomodulatory activity of MSCs

During the organizational repair process, the interaction between MSCs and immune cells is necessary to activate the immune regulatory activity of MSCs. On the one hand, the immunomodulatory ability of MSCs is triggered by the inflammatory environment. The changes in macrophages functional phenotypes play an essential regulatory role in the inflammatory environment. Inflammatory factors released by M1 macrophages, such as IFN-y, can activate the immune regulatory properties of MSCs, promoting the regulatory effect of MSCs on immune cells, such as secreting IDO to inhibit T cell proliferation and induce T cell apoptosis[35]. Anti-inflammatory factors produced by M2 macrophages, such as IL-10, can enhance the promoter activity of TNF- α , a pro-inflammatory cytokine, in MSCs, reducing the expression of inflammatory factors such as IL-6 and IFN- γ [27]. On the other hand, MSCs demonstrate poor survival in inflammatory areas, mainly through paracrine chemokines, growth factors, and cytokines, or by direct contact, to regulate macrophages polarization, resulting in anti-inflammatory or immunosuppressive effects, and playing an immune regulatory role. Exosomes derived from MSCs can significantly enrich macrophages for activating the HMGB1/toll-like receptor 4 (TLR4)/NF-κB signaling pathway, and promoting the conversion of macrophages to M2 macrophages[67,68]. M2 macrophages secrete anti-inflammatory factors, such as IL-4, IL-10, and TGF- β , to prevent excessive activation of inflammatory responses[34,36].

Macrophages regulate the differentiation of MSCs

Pittenger and colleagues successfully induced the differentiation of MSCs into osteoblasts (OBs), chondrocytes, and adipocytes in vitro[38]. Subsequently, the potential of MSCs to differentiate into muscle, ligament, endotheliocytes, and hepatocytes, among other cell types, was discovered[38,51,69-71]. Multiple studies have shown that macrophages play a crucial role in the MSC differentiation process. Firstly, macrophages provide a certain protective effect on the multilineage differentiation potential of MSCs. Mitochondrial metabolic damage in MSCs can lead to differentiation disorders. Teissier et al^[72] found that co-culture of M0 macrophages with mitochondrially impaired MSCs could correct the redox imbalance in MSCs, restore their metabolic homeostasis, and promote their differentiation. Additionally, macrophages have an inducing effect on MSC differentiation. Current research mainly focuses on the impact of macrophages on the osteogenic, chondrogenic, and adipogenic differentiation of MSCs. Macrophages secrete certain antiinflammatory factors, pro-inflammatory factors, and osteogenic factors such as bone morphogenic protein-2 (BMP-2) and oncostatin-M (OSM), which can mediate the osteogenic differentiation of MSCs[39,73]. Recent studies have also shown that oxidative stress plays a key role in the regulation of MSCs by macrophages. Macrophages can reduce the level of reactive oxygen species in MSCs similarly to the antioxidant N-acetyl cysteine, regulating their osteogenic differentiation [74]. Cao et al[40] discovered that co-culture of macrophages with MSCs could inhibit the expression of C-X-C motif chemokine ligand 1 (CXCL1) and its receptor CXCR2, reduce the phosphorylation of p38/ERK-ELK1 during the early stage of adipogenesis, and regulate the adipogenic differentiation of MSCs. Physical stimuli and changes in chemical composition can enhance the chondrogenic differentiation of MSCs in vitro and in vivo by controlling macrophages activation. Wang et al[41] found that mechanical stimulation could promote the polarization of macrophages toward the M2 phenotype, which then secreted TGF- β 1 to promote MSC-based chondrogenesis. Hu *et al*[42] found that magnesium deficiency can increase the secretion of inflammatory factors, such as IL-1β and IL-6 by M1 macrophages, inhibiting MSCs chondrogenesis. Due to the roles of macrophages in bone, cartilage, and fat formation, they are considered potential targets for the treatment of various related diseases, such as bone defects, osteoarthritis, and vascular diseases.

THE INTERACTION BETWEEN MSCS AND MACROPHAGE IS AN IMPORTANT PART OF BONE TISSUE REPAIR

The process of bone tissue damage includes an inflammatory response, which creates the basis for subsequent bone formation, mineralization, resorption, and bone replacement. In the process of damage repair, the synergistic effect of MSCs and macrophages mainly involves intervention in the occurrence and regression of inflammation after damage and regulation of bone formation and remodeling. The polarization of M1/M2 macrophages is the basis of the inflammatory response, and MSCs can regulate the balance of M1/M2 macrophages to participate in the regulation of inflammation progression and regression [75]. During bone formation and remodeling, the mutual regulation between MSCs and macrophages plays an important role in balancing OBs and OCs and maintaining bone homeostasis. At the same time, the dynamic balance between MSC-drives osteogenic differentiation-mediated bone regeneration and OCs-mediated bone resorption also participates in the bone repair and remodeling process. The interaction between MSCs and macrophages creates a negative feedback loop, which plays an important role in bone tissue repair.

Maintaining tissue microenvironment homeostasis

The occurrence of inflammation is a necessary link in the repair of bone tissue damage. Activation of the innate and adaptive immune system is beneficial for the body to make a sufficient response to bone tissue damage [76,77]. The temporary inflammatory response after damage helps the body to mobilize the immune system and to activate the damage repair mechanism, which is conducive to wound healing. After tissue damage, cells autonomously conduct inflammatory signal transduction. Macrophages are large secretory cells that act as inflammation effectors and can shape the post-damage tissue environment by secreting a variety of bioactive factors. They play pro-anabolic roles throughout the entire stage of bone tissue damage repair and are key cells in the repair of bone tissue damage [78]. MSCs are key regulators of inflammation. Under the action of damage-associated molecular patterns, MSCs exhibit a pro-inflammatory



phenotype[79]. After receiving stimulation signals through pattern recognition receptors, MSCs activate transcriptional effectors such as NF-KB and TLR4 for immune regulation[80]. For example, NF-KB can bind to the receptor activator of NF-κB (RANK) ligand (RANKL) receptor, stimulate the activation of classical M1 macrophages, and release a variety of inflammatory factors, while recruiting various immune cells to enhance the immune response[81,82].

Prolonged inflammatory responses are not conducive to the repair of injuries, as they can damage the osteogenic ability of MSCs and lead to secondary osteopathies such as osteoporosis, bone necrosis, and hyperostosis[83]. Therefore, improving the inflammatory state of the organizational microenvironment is the precursor and basis for the recovery of tissue structure and function. Inflammatory signal transduction stimulation can cause changes in the biological activities and epigenetics of various cells, which is a key adaptation of cells to tissue damage. Once inflammatory factors in the microenvironment reach a certain level, MSCs are activated to inhibit the secretion of pro-inflammatory factors by M1 macrophages in various ways, promoting the secretion of multiple anti-inflammatory factors by M2 macrophages, effectively preventing excessive activation of inflammation, and ensuring the smooth progress of tissue repair and remodeling. On the one hand, MSCs and macrophages interact with each other through direct receptor binding. Li et al [27] found that pro-inflammatory M1 macrophages can interact directly with MSCs, promoting the production of TNF- α stimulated gene 6 protein (TSG-6) and CD200 expression in MSCs. Simultaneously, CD200 on MSCs can interact with CD200 receptor on macrophages, promoting the polarization of M2 macrophages and enhancing the inhibitory effect on inflammation[27]. On the other hand, MSCs and macrophages also regulate each other through paracrine effects. The anti-inflammatory protein TGS-6 is a multifunctional protein associated with inflammation that has anti-inflammatory and tissue-protective properties and plays a critical role in inflammatory cell migration, cell proliferation, and development. Choi et al[32] found that M1 macrophages secrete various pro-inflammatory cytokines, including TNF- α and IL-1β, at sites of inflammation, which can activate MSCs to induce TSG-6 expression. TSG-6 can bind to CD44 receptors on macrophages and decrease zymosan/TLR2-mediated nuclear translocation of NF-KB, inhibiting the activity of M1 macrophages, inducing anti-inflammatory M2 macrophage polarization, and promoting resolution of inflammation. IL-6 is a signaling molecule dependent on polarization of M1 macrophages to M2 macrophages during inflammation. Philipp et al^[29] found that MSCs-derived IL-6 triggers M2 macrophages polarization. IL-12 is an effective inducer of cellmediated immune responses, mainly derived from monocytes and macrophages. Chahal et al[84] found that the expression of pro-inflammatory M1 macrophages and IL-12 in synovial fluid significantly decreased after injecting MSCs into the joints of mice with osteoarthritis. The heterodimer formed by the chemokines CCL2 and CXCL12 secreted by MSCs can upregulate IL-10 expression in CCR2(+) macrophages[85], promoting the polarization of M2 macrophages and driving an anti-inflammatory response. MSCs-secreted miR-466 promotes the polarization of macrophages toward the M2 type by downregulating the TIRAP-MyD88-NF-*k*B signaling pathway^[25].

Formation of the basic multicellular unit for bone tissue

Bone repair and remodeling are processes involving "activation-resorption-formation" of the basic multicellular units [86]. OBs derived from MSCs and OCs derived from macrophages are two major cell types involved in bone tissue repair, playing essential roles in maintaining bone homeostasis[87]. OBs are the key cells for bone formation. MSCs and skeletal stem cells differentiate into OB precursor cells (pre-OBs), while macrophages and endosteal OBs participate in bone mineralization side by side. Mature OBs embed in the gaps of the mineralized matrix and become bone cells. OCs are the key cells for bone resorption. Inflammatory factors and chemokines can attract circulating macrophages to migrate to the inflammatory site and become resident macrophages in bone (also known as bone macrophages)[88]. OBs-derived macrophages secrete colony-stimulating factor (M-CSF), which binds to CSF1 receptor (CSF1R, also known as c-FMS) on the macrophages surface, activating the induction signal for OC differentiation, RANKL/RANK[89]. Key factors, such as M-CSF and RANKL, can activate the expression of the primary switch regulatory factor for OC generation, *i.e.*, the transcription factor nuclear factor of activated T cells (NFATc1), promoting the migration of monocytes/macrophages, which are OC precursor cells (pre-OCs), to the bone surface for activation and differentiation into mature OCs[90-92].

Regulate a balance between bone resorption and bone formation

MSCs and macrophages not only serve as the sources of OBs and OCs for tissue repair but also interact closely with each other through common cellular precursors and molecular mediators of the immune system and bone tissue. This shapes the various bioactive factors required for the bone tissue repair microenvironment, including osteogenic-related factors, TNF family, IL family, IFN family, chemokines, etc., and participating in metabolism related to bone resorption and bone formation (Figure 2)[73,93-107].

In the acute phase of inflammation, the interaction between MSCs and M1 macrophages releases various pro-inflammatory factors and chemokines, mainly involved in the differentiation and maturation of pre-OCs. RANKL serves as a bridge between the inflammatory response and bone resorption[93]. In the presence of RANKL, multiple pro-inflammatory cytokines synergistically coordinate to enhance the genesis of OCs. MSCs-secreted NF-KB binds to the OCs differentiation-inducing RANKL to control the expression of key factors in pre-OCs differentiation. Inflammatory factors, such as TNF-a, IL-1, IL-1β, IL-6, IL-7, IL-8, IL-11, IL-17, IL-18, IL-23, etc., secreted by M1, can directly or indirectly affect OBs to promote RANKL expression, participating in OBs differentiation of MSCs and OCs differentiation of bone macrophages [93-99]. Among them, TNF- α and IL-6 are key regulatory factors. TNF- α acts on signaling pathways such as that of RANKL/RANK, downstream NF-KB, JNK, p38, AKT, etc., to reduce the expression of OCs-specific genes and proteins, promoting the activation of bone macrophages and the differentiation of OCs[81]. After binding to IL-6R and gp130, IL-6 activates the phosphorylation of JAK/STAT[108]. The phosphorylated STAT3 translocate to the nucleus, where it activates the expression of RANKL, directly participating in the OC differentiation of bone macrophages, or indirectly influencing OC differentiation through its action on TNF- α [109,110]. At the same time, these factors also provide key aspects of the feedback regulation mechanism of RANKL induction in bone macrophages. IL-4 inhibits the transcription



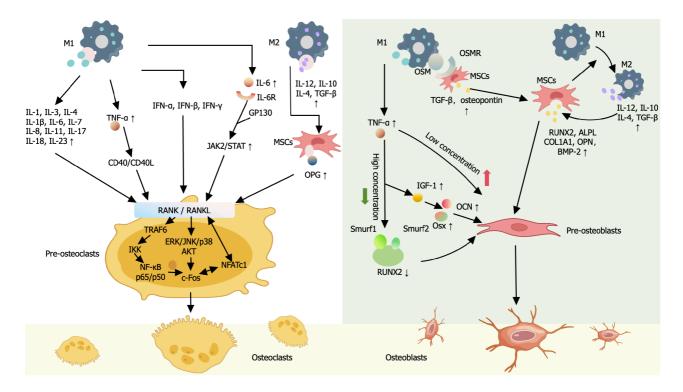


Figure 2 The dialogue between mesenchymal stem cells and macrophages in mediating the immune environment and bone tissue. Multiple inflammatory factors secreted by M1 macrophages can promote the differentiation of osteoclast precursors into osteoclasts. When the inflammatory factors accumulate to a certain extent, the differentiation of osteoclasts is inhibited by the action of osteogenic factors secreted by M2 macrophages and mesenchymal stem cells (MSCs). Macrophages can promote the secretion of various osteogenic induction factors by MSCs through direct contact or by secreting various bioactive factors indirectly, inducing further differentiation of osteogenic precursor cells into osteoblasts. MSC: Mesenchymal stem cell; RANK: Receptor activator of nuclear factor kappa-B; RANKL: Receptor activator of nuclear factor kappa-B ligand; NF-kB: Nuclear factor kappa-B; TRAF6: Tumor necrosis factor receptor-associated factor 6; NFATc1: Transcription factor nuclear factor of activated T cells; OPG: Osteoprotegerin; OSM: Oncostatin-M; OSMR: Oncostatin-M receptor; RUNX2: Runtrelated transcription factor 2; OCN: Osteocalcin; Osx: Osterix; OPN: Osteopontin; ALPL: Alkaline phosphatase; BMP-2: Bone morphogenetic protein-2; TNF: Tumor necrosis factor; IL: Interleukin; IFN: Interferon; TGF-β: Transforming growth factor beta; IGF-1: Insulin-like growth factor-1.

factor NFATc1, weakening RANKL-induced OC formation [104]. IFNs, such as IFN- α , IFN- β , and IFN- γ induce rapid degradation of RANK-associated protein TNF receptor-associated factor 6 or mediate FAS/FASL signaling, strongly inhibiting the activation of RANKL-induced transcription factors NF-KB and *c-fos*, thus inhibiting excessive OC formation [101]. TNF- α activates RANKL-related CD40/CD40L signaling to promote the production of the OC genesis inhibitory factor osteoprotegerin (OPG) and to reduce bone resorption [100]. IL-3 can also down-regulates TNF- α -induced *c-fos* nuclear translocation to inhibit OCs formation and bone resorption[94].

In addition, the pro-inflammatory factors secreted by M1 macrophages are also related to the early and mid-stage osteogenic effects of MSCs. The absence of TNF- α and IL-6 has been proven to delay the differentiation of MSCs. TNF- α can selectively mediate the osteogenic differentiation of OBs. Low concentrations of $TNF-\alpha$ stimulate the differentiation of pre-OBs into OBs, while high concentrations of TNF-α regulate the degradation of RUNT-related transcription factor 2 (RUNX2) protein by signal molecules drosophila mothers against decapentaplegic homologue ubiquitination regulatory factor 1 (SMURF1) and SMURF2 to block the expression of RUNX2, and simultaneously inhibit the expression of insulinlike growth factor-1 and osterix and osteocalcin, inhibiting OB differentiation[106]. The effect of IL-6 on bone marrowderived OCs stimulates the release of "bone messengers", which act through the cortical bone cell network to stimulate the formation of OBs on the bone membrane. Cyclooxygenase-2 and PGE2 regulate the production of the IL-6 family cytokine OSM in classically activated inflammatory M1 macrophages[73]. OSM binds to the OSM receptor on MSCs, then activates the STAT3-CCAAT-enhancer-binding protein delta-C/core binding factor alpha 1 differentiation pathway, mediates the recruitment and proliferation of MSCs, stimulates the maturation of OBs from MSCs, and drives the mineralization of OBs. TGF- β and OPN enhance the activity of alkaline phosphatase (ALP) in MSCs, promoting the maturation and mineralization of the matrix[107].

However, recent studies have shown that the microenvironment that forms during the resolution stage of inflammation has a more important impact on bone formation and remodeling, which can significantly increase the ratio of OBs/OCs and can promote bone formation. On the one hand, MSCs promote M2 macrophages polarization, and release inflammatory factors, chemokines, and other active factors to regulate bone homeostasis indirectly. Anti-inflammatory factors secreted by M2 macrophages, such as IL-4, IL-10, TGF-β, and IL-12, can promote the expression of OBs and inhibit the activity of OCs by regulating the expression of osteogenic markers such as RUNX2, ALP ligand, and collagen1A1, and adjusting the OPG/RANKL signaling pathway [103,106]. Chemokines, such as CXCL3, CXCL6, and CXCL14, are important mediators promoting osteogenic differentiation and bone integration of MSCs. These chemokines regulate pathways downstream of cytoskeletal actin in MSCs and promote the expression of distal-less homeobox 5 and RUNX2 by activating CXCR2 and CCR1[30]. On the other hand, macrophages induce MSCs to release osteogenic factors, such as

Zaishideng® WJSC | https://www.wjgnet.com

BMP-2, activating signal molecules drosophila mothers against decapentaplegic homologue in OBs to promote the osteogenic function of MSCs[111]. These studies suggest that MSCs and macrophages play an important role in bone tissue repair (Figure 3)[30,31,43,81,88,93,100].

FUTURE THERAPIES FOR BONE REGENERATION BASED ON THE CROSSTALK BETWEEN MSCS AND MACROPHAGES

The multilineage differentiation potential of MSCs and their susceptibility to microenvironmental regulation make their roles in bone tissue less clear. Modulating immune cells to enhance the osteogenic effects of MSCs is emerging as a new method to address current challenges in this field. Co-transplantation of macrophages and MSCs may be a potential effective treatment for bone repair. Current research attempts to exploit the characteristics of mutual regulation and microenvironment sensitivity between MSCs and macrophages, exploring their potential applications in acute or chronic bone tissue injuries such as fractures and osteoarthritis. Co-culture of MSCs with macrophages can enhance the immunomodulatory and osteogenic ability of MSCs. Some studies have found that preconditioning can further enhance the regulatory ability of MSCs on macrophages. For example, Philipp *et al*^[29] discovered that the immunosuppressive effect of MSCs mediated by IFN- γ and IL-1 β pretreatment mainly involved the active component IL-6, which stimulates the polarization of macrophages toward the M2 phenotype, promoting the expression of CD86, inducible nitric oxide synthase protein, and IL-10 secretion by macrophages, thus significantly enhancing the immune regulatory capabilities of MSCs. Furthermore, conditions such as hypoxia, lipopolysaccharide treatment, and heat shock pretreatment can enhance the impacts of macrophages on the immune regulatory capacity and osteogenic activity of MSCs, although these treatments may have some inhibitory effects on MSC survival[112]. The depletion of macrophages can lead to a decrease in MSCs, and the use of IL-13 and IL-4 to induce an increase in the number of M2 macrophages can promote MSC osteogenic differentiation and accelerate bone repair [113,114].

Methods of effectively regulating the crosstalk between MSCs and macrophages through biomaterial scaffolds to promote bone healing are becoming a new research focus. The implantation of living cells wrapped in 3D-bioprinted scaffolds can effectively improve the inflammatory microenvironment within the body, further accelerating bone repair. Yu et al[115] discovered that implanting a dual-channel bioprinting scaffold, formed by wrapping macrophages with 8% methacrylamidated gelatin/1% methacrylamidated hyaluronic acid and wrapping MSCs with 3% alginate/0.5 mg/mL graphene oxide, into model rat calvarial defects, can effectively promote the polarization of M2 macrophages in the early bone defect environment, avoid excessive inflammatory responses, and further promote bone repair. The results indicated that this method is more effective than using single-cell MSC scaffolds[115]. Furthermore, some studies have chosen to use special coatings to modify implanted scaffolds or hydrogels, etc., to regulate the biomaterial scaffolds to activate macrophages in the body and promote the differentiation of osteogenic phenotypes. For example, Majrashi et al [116] implanted dexamethasone, which has both anti-inflammatory and osteogenic differentiation promoting effects, into 3D scaffolds. According to the principle that early inflammation benefits MSC-based osteogenesis and that excessive inflammatory responses harm this process, they used excipient-sucrose acetate isobutyrate to achieve sustained release of dexamethasone, regulate the activation of macrophage phenotypes, and control the inflammatory microenvironment within the tissue to promote tissue regeneration. In the early stage of bone tissue injury repair, M1 macrophages can secrete IL-6 to promote osteogenesis. As inflammation progresses, dexamethasone can promote the polarization of M2 macrophages and enhance the osteogenic effect of MSCs by jointly secreting BMP-2 with M2 macrophages[116]. Niu et al [117] wrapped hydrogels containing MSCs with a unique macrophage-affinitive glucomannan polysaccharide and found that, after implantation into mice, it effectively promoted the adherence and activation of macrophages, and induced the release of osteogenic genes in MSCs, thereby effectively promoting bone tissue regeneration[117]. In addition, numerous studies are currently innovating biomaterial compositions for bone tissue engineering, which will provide a foundation for enhancing the application of MSCs and macrophages in bone tissue repair and remodeling[118-121].

CONCLUSION

A comprehensive understanding of the roles of MSCs and macrophages in immunomodulation and bone regeneration is beneficial for the optimal application of cellular therapy in tissue repair and regeneration. We here discussed the important roles of both MSCs and macrophages in the immune microenvironment of bone injury and bone tissue regeneration by summarizing their mutual regulatory interactions. However, the regulatory mechanisms at play between MSCs and macrophages are complex, and the repair and regeneration of bone tissue is a complicated, dynamic process. Our research and discussion are not comprehensive, and with further study, new breakthroughs in their interactive mechanisms may emerge. Furthermore, through further exploration of potential therapeutic models based on the crosstalk between MSCs and macrophages, we have described that special pretreatment of MSCs and macrophages, or the combined use of new biological material carriers (such as hydrogels and 3D scaffolds) for the MSC and macrophage co-transplantation treatment model, may represent effective therapies that can promote the immunomodulatory capacity of MSCs or endogenous tissue regeneration. This approach could provide new methods and strategies for the repair and regeneration of clinical bone defects or other soft tissue injuries. However, this field is currently mostly in the basic research stage, and the efficacy and safety of these approaches will require verification in future.

Zaisbidene® WJSC | https://www.wjgnet.com

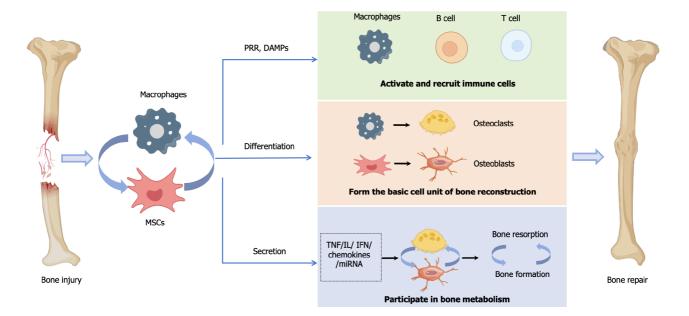


Figure 3 Mesenchymal stem cells and macrophages promote bone tissue repair. The inflammatory microenvironment after injury promotes the activation of mesenchymal stem cells (MSCs) and macrophages. Through immune response patterns such as pattern recognition receptors and damage associated molecular patterns, the interaction between MSCs and macrophages further activates immune cells such as B cells and T cells, enhancing the immune response in the damaged microenvironment. MSCs and macrophages are precursor cells for osteoblasts (OBs) and osteoclasts (OCs), respectively, and play an important role in bone tissue reconstruction. At the same time, MSCs and macrophages also affect the formation of OBs and OCs by secreting chemokines, tumor necrosis factor, interferon, interferon and other bioactive factors, regulating the balance between bone resorption and bone formation, and promoting the repair of bone tissue. MSC: Mesenchymal stem cell; PRR: Pattern recognition receptor; DAMPs: Damage associated molecular patterns; TNF: Tumor necrosis factor; IL: Interleukin; IFN: Interferon; miRNA: MicroRNA.

ACKNOWLEDGEMENTS

I would like to thank all people who helped me in the process of writing this article.

FOOTNOTES

Author contributions: Zhang FF wrote the paper; Hao Y, Zhang KX, Yang JJ, and Zhao ZQ performed the collected the data; Liu HJ and Li JT responsible for review and revision; and all authors read and approved the final manuscript.

Supported by the National Key Research and Development Program of China, No. 2023YFC2508806; Key Research and Development Project in Henan Province, No. 231111310500; Young Elite Scientists Sponsorship Program by CAST, No. 2021-QNRC2-A06; Scientific Research Project of Henan Zhongyuan Medical Science and Technology Innovation and Development Foundation, No. ZYYC2023ZD; Youth Science Award Project of the Provincial-Level Joint Fund for Science and Technology Research and Development Project in Henan Province, No. 225200810084; Special Project on Training Top Talents in Traditional Chinese Medicine in Henan Province, No. 2022ZYBJ24; and 2023 Hunan University of Chinese Medicine Postgraduate Innovation Project, No. 2023CX64.

Conflict-of-interest statement: All the authors report no relevant conflicts of interest for this article.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: https://creativecommons.org/Licenses/by-nc/4.0/

Country/Territory of origin: China

ORCID number: Ji-Tian Li 0000-0001-5449-8084.

S-Editor: Wang JJ L-Editor: A P-Editor: Zhang XD



REFERENCES

- Wynn TA, Vannella KM. Macrophages in Tissue Repair, Regeneration, and Fibrosis. Immunity 2016; 44: 450-462 [PMID: 26982353 DOI: 1 10.1016/j.immuni.2016.02.015]
- 2 Bernardo ME, Fibbe WE. Mesenchymal stromal cells: sensors and switchers of inflammation. Cell Stem Cell 2013; 13: 392-402 [PMID: 24094322 DOI: 10.1016/j.stem.2013.09.006]
- Loi F, Córdova LA, Pajarinen J, Lin TH, Yao Z, Goodman SB. Inflammation, fracture and bone repair. Bone 2016; 86: 119-130 [PMID: 3 26946132 DOI: 10.1016/j.bone.2016.02.020]
- 4 Liu H, Yang R, Zhao S, Zhou F, Liu Y, Zhou Z, Chen L, Xie J. Collagen scaffolds derived from bovine skin loaded with MSC optimized M1 macrophages remodeling and chronic diabetic wounds healing. Bioeng Transl Med 2023; 8: e10467 [PMID: 37206210 DOI: 10.1002/btm2.10467]
- 5 Freytes DO, Kang JW, Marcos-Campos I, Vunjak-Novakovic G. Macrophages modulate the viability and growth of human mesenchymal stem cells. J Cell Biochem 2013; 114: 220-229 [PMID: 22903635 DOI: 10.1002/jcb.24357]
- Shi Y, Su J, Roberts AI, Shou P, Rabson AB, Ren G. How mesenchymal stem cells interact with tissue immune responses. Trends Immunol 6 2012; **33**: 136-143 [PMID: 22227317 DOI: 10.1016/j.it.2011.11.004]
- 7 Yuan Y, Ni S, Zhuge A, Li L, Li B. Adipose-Derived Mesenchymal Stem Cells Reprogram M1 Macrophage Metabolism via PHD2/HIF-1a Pathway in Colitis Mice. Front Immunol 2022; 13: 859806 [PMID: 35757749 DOI: 10.3389/fimmu.2022.859806]
- 8 Cao X, Duan L, Hou H, Liu Y, Chen S, Zhang S, Wang C, Qi X, Liu N, Han Z, Zhang D, Han ZC, Guo Z, Zhao Q, Li Z. IGF-1C hydrogel improves the therapeutic effects of MSCs on colitis in mice through PGE(2)-mediated M2 macrophage polarization. Theranostics 2020; 10: 7697-7709 [PMID: 32685014 DOI: 10.7150/thno.45434]
- 9 Saldaña L, Bensiamar F, Vallés G, Mancebo FJ, García-Rey E, Vilaboa N. Immunoregulatory potential of mesenchymal stem cells following activation by macrophage-derived soluble factors. Stem Cell Res Ther 2019; 10: 58 [PMID: 30760316 DOI: 10.1186/s13287-019-1156-6]
- 10 Ylöstalo JH, Bartosh TJ, Coble K, Prockop DJ. Human mesenchymal stem/stromal cells cultured as spheroids are self-activated to produce prostaglandin E2 that directs stimulated macrophages into an anti-inflammatory phenotype. Stem Cells 2012; 30: 2283-2296 [PMID: 22865689 DOI: 10.1002/stem.1191]
- Vasandan AB, Jahnavi S, Shashank C, Prasad P, Kumar A, Prasanna SJ. Human Mesenchymal stem cells program macrophage plasticity by 11 altering their metabolic status via a PGE(2)-dependent mechanism. Sci Rep 2016; 6: 38308 [PMID: 27910911 DOI: 10.1038/srep38308]
- 12 Wynn TA, Chawla A, Pollard JW. Macrophage biology in development, homeostasis and disease. Nature 2013; 496: 445-455 [PMID: 23619691 DOI: 10.1038/nature12034]
- Peiser L, Mukhopadhyay S, Gordon S. Scavenger receptors in innate immunity. Curr Opin Immunol 2002; 14: 123-128 [PMID: 11790542 13 DOI: 10.1016/s0952-7915(01)00307-7]
- 14 Murray PJ. Macrophage Polarization. Annu Rev Physiol 2017; 79: 541-566 [PMID: 27813830 DOI: 10.1146/annurev-physiol-022516-034339]
- Wynn TA, Barron L. Macrophages: master regulators of inflammation and fibrosis. Semin Liver Dis 2010; 30: 245-257 [PMID: 20665377 15 DOI: 10.1055/s-0030-1255354]
- Rodríguez-Morales P, Franklin RA. Macrophage phenotypes and functions: resolving inflammation and restoring homeostasis. Trends 16 Immunol 2023; 44: 986-998 [PMID: 37940394 DOI: 10.1016/j.it.2023.10.004]
- 17 Arabpour M, Saghazadeh A, Rezaei N. Anti-inflammatory and M2 macrophage polarization-promoting effect of mesenchymal stem cellderived exosomes. Int Immunopharmacol 2021; 97: 107823 [PMID: 34102486 DOI: 10.1016/j.intimp.2021.107823]
- 18 Pirraco RP, Reis RL, Marques AP. Effect of monocytes/macrophages on the early osteogenic differentiation of hBMSCs. J Tissue Eng Regen Med 2013; 7: 392-400 [PMID: 22392849 DOI: 10.1002/term.535]
- 19 Koenders MI, Kolls JK, Oppers-Walgreen B, van den Bersselaar L, Joosten LA, Schurr JR, Schwarzenberger P, van den Berg WB, Lubberts E. Interleukin-17 receptor deficiency results in impaired synovial expression of interleukin-1 and matrix metalloproteinases 3, 9, and 13 and prevents cartilage destruction during chronic reactivated streptococcal cell wall-induced arthritis. Arthritis Rheum 2005; 52: 3239-3247 [PMID: 16200598 DOI: 10.1002/art.213421
- Zhang MZ, Yao B, Yang S, Jiang L, Wang S, Fan X, Yin H, Wong K, Miyazawa T, Chen J, Chang I, Singh A, Harris RC. CSF-1 signaling 20 mediates recovery from acute kidney injury. J Clin Invest 2012; 122: 4519-4532 [PMID: 23143303 DOI: 10.1172/JCI60363]
- 21 Zhu S, Liu M, Bennett S, Wang Z, Pfleger KDG, Xu J. The molecular structure and role of CCL2 (MCP-1) and C-C chemokine receptor CCR2 in skeletal biology and diseases. J Cell Physiol 2021; 236: 7211-7222 [PMID: 33782965 DOI: 10.1002/jcp.30375]
- Ren G, Zhang L, Zhao X, Xu G, Zhang Y, Roberts AI, Zhao RC, Shi Y. Mesenchymal stem cell-mediated immunosuppression occurs via 22 concerted action of chemokines and nitric oxide. Cell Stem Cell 2008; 2: 141-150 [PMID: 18371435 DOI: 10.1016/j.stem.2007.11.014]
- Saldaña L, Vallés G, Bensiamar F, Mancebo FJ, García-Rey E, Vilaboa N. Paracrine interactions between mesenchymal stem cells and 23 macrophages are regulated by 1,25-dihydroxyvitamin D3. Sci Rep 2017; 7: 14618 [PMID: 29097745 DOI: 10.1038/s41598-017-15217-8]
- Liu F, Qiu H, Xue M, Zhang S, Zhang X, Xu J, Chen J, Yang Y, Xie J. MSC-secreted TGF-ß regulates lipopolysaccharide-stimulated 24 macrophage M2-like polarization via the Akt/FoxO1 pathway. Stem Cell Res Ther 2019; 10: 345 [PMID: 31771622 DOI: 10.1186/s13287-019-1447-y
- Shi MM, Zhu YG, Yan JY, Rouby JJ, Summah H, Monsel A, Qu JM. Role of miR-466 in mesenchymal stromal cell derived extracellular 25 vesicles treating inoculation pneumonia caused by multidrug-resistant Pseudomonas aeruginosa. Clin Transl Med 2021; 11: e287 [PMID: 33463070 DOI: 10.1002/ctm2.2871
- Chiossone L, Conte R, Spaggiari GM, Serra M, Romei C, Bellora F, Becchetti F, Andaloro A, Moretta L, Bottino C. Mesenchymal Stromal 26 Cells Induce Peculiar Alternatively Activated Macrophages Capable of Dampening Both Innate and Adaptive Immune Responses. Stem Cells 2016; 34: 1909-1921 [PMID: 27015881 DOI: 10.1002/stem.2369]
- 27 Li Y, Zhang D, Xu L, Dong L, Zheng J, Lin Y, Huang J, Zhang Y, Tao Y, Zang X, Li D, Du M. Cell-cell contact with proinflammatory macrophages enhances the immunotherapeutic effect of mesenchymal stem cells in two abortion models. Cell Mol Immunol 2019; 16: 908-920 [PMID: 30778166 DOI: 10.1038/s41423-019-0204-6]
- Hou L, Zhu Z, Jiang F, Zhao J, Jia Q, Jiang Q, Wang H, Xue W, Wang Y, Tian L. Human umbilical cord mesenchymal stem cell-derived 28 extracellular vesicles alleviated silica induced lung inflammation and fibrosis in mice via circPWWP2A/miR-223-3p/NLRP3 axis. Ecotoxicol Environ Saf 2023; 251: 114537 [PMID: 36646008 DOI: 10.1016/j.ecoenv.2023.114537]



- Philipp D, Suhr L, Wahlers T, Choi YH, Paunel-Görgülü A. Preconditioning of bone marrow-derived mesenchymal stem cells highly 29 strengthens their potential to promote IL-6-dependent M2b polarization. Stem Cell Res Ther 2018; 9: 286 [PMID: 30359316 DOI: 10.1186/s13287-018-1039-2
- 30 Zhang Y, Wei J, Yu X, Chen L, Ren R, Dong Y, Wang S, Zhu M, Ming N, Zhu Z, Gao C, Xiong W. CXCL chemokines-mediated communication between macrophages and BMSCs on titanium surface promotes osteogenesis via the actin cytoskeleton pathway. Mater Today Bio 2023; 23: 100816 [PMID: 37859997 DOI: 10.1016/j.mtbio.2023.100816]
- Liu J, Lai X, Bao Y, Xie W, Li Z, Chen J, Li G, Wang T, Huang W, Ma Y, Shi J, Zhao E, Xiang AP, Liu Q, Chen X. Intraperitoneally 31 Delivered Mesenchymal Stem Cells Alleviate Experimental Colitis Through THBS1-Mediated Induction of IL-10-Competent Regulatory B Cells. Front Immunol 2022; 13: 853894 [PMID: 35371051 DOI: 10.3389/fimmu.2022.853894]
- 32 Choi H, Lee RH, Bazhanov N, Oh JY, Prockop DJ. Anti-inflammatory protein TSG-6 secreted by activated MSCs attenuates zymosan-induced mouse peritonitis by decreasing TLR2/NF-κB signaling in resident macrophages. Blood 2011; 118: 330-338 [PMID: 21551236 DOI: 10.1182/blood-2010-12-327353
- Hengartner NE, Fiedler J, Schrezenmeier H, Huber-Lang M, Brenner RE. Crucial role of IL1beta and C3a in the in vitro-response of 33 multipotent mesenchymal stromal cells to inflammatory mediators of polytrauma. PLoS One 2015; 10: e0116772 [PMID: 25562599 DOI: 10.1371/journal.pone.0116772
- Yu D, Zhao Y, Wang H, Kong D, Jin W, Hu Y, Qin Y, Zhang B, Li X, Hao J, Li G. IL-18 pre-stimulation enhances the therapeutic effects of 34 endometrial regenerative cells on experimental colitis. Stem Cell Res Ther 2021; 12: 324 [PMID: 34090510 DOI: 10.1186/s13287-021-02392-91
- Meisel R, Zibert A, Laryea M, Göbel U, Däubener W, Dilloo D. Human bone marrow stromal cells inhibit allogeneic T-cell responses by 35 indoleamine 2,3-dioxygenase-mediated tryptophan degradation. Blood 2004; 103: 4619-4621 [PMID: 15001472 DOI: 10.1182/blood-2003-11-3909]
- Shapouri-Moghaddam A, Mohammadian S, Vazini H, Taghadosi M, Esmaeili SA, Mardani F, Seifi B, Mohammadi A, Afshari JT, Sahebkar 36 A. Macrophage plasticity, polarization, and function in health and disease. J Cell Physiol 2018; 233: 6425-6440 [PMID: 29319160 DOI: 10.1002/jcp.26429
- Jiang Z, Zhang J. Mesenchymal stem cell-derived exosomes containing miR-145-5p reduce inflammation in spinal cord injury by regulating 37 the TLR4/NF-KB signaling pathway. Cell Cycle 2021; 20: 993-1009 [PMID: 33945431 DOI: 10.1080/15384101.2021.1919825]
- Pittenger MF, Mackay AM, Beck SC, Jaiswal RK, Douglas R, Mosca JD, Moorman MA, Simonetti DW, Craig S, Marshak DR. Multilineage 38 potential of adult human mesenchymal stem cells. Science 1999; 284: 143-147 [PMID: 10102814 DOI: 10.1126/science.284.5411.143]
- Zhang Y, Böse T, Unger RE, Jansen JA, Kirkpatrick CJ, van den Beucken JJJP. Macrophage type modulates osteogenic differentiation of 39 adipose tissue MSCs. Cell Tissue Res 2017; 369: 273-286 [PMID: 28361303 DOI: 10.1007/s00441-017-2598-8]
- Cao D, Ma F, Ouyang S, Liu Z, Li Y, Wu J. Effects of macrophages and CXCR2 on adipogenic differentiation of bone marrow mesenchymal 40 stem cells. J Cell Physiol 2019; 234: 9475-9485 [PMID: 30362570 DOI: 10.1002/jcp.27634]
- 41 Wang L, Li S, Xiao H, Zhang T, Liu Y, Hu J, Xu D, Lu H. TGF-B1 derived from macrophages contributes to load-induced tendon-bone healing in the murine rotator cuff repair model by promoting chondrogenesis. Bone Joint Res 2023; 12: 219-230 [PMID: 37051812 DOI: 10.1302/2046-3758.123.BJR-2022-0368.R1]
- Hu T, Xu H, Wang C, Qin H, An Z. Magnesium enhances the chondrogenic differentiation of mesenchymal stem cells by inhibiting activated 42 macrophage-induced inflammation. Sci Rep 2018; 8: 3406 [PMID: 29467509 DOI: 10.1038/s41598-018-21783-2]
- Franquesa M, Hoogduijn MJ, Bestard O, Grinyó JM. Immunomodulatory effect of mesenchymal stem cells on B cells. Front Immunol 2012; 43 **3**: 212 [PMID: 22833744 DOI: 10.3389/fimmu.2012.00212]
- Biswas S, Mandal G, Roy Chowdhury S, Purohit S, Payne KK, Anadon C, Gupta A, Swanson P, Yu X, Conejo-Garcia JR, Bhattacharyya A. 44 Exosomes Produced by Mesenchymal Stem Cells Drive Differentiation of Myeloid Cells into Immunosuppressive M2-Polarized Macrophages in Breast Cancer. J Immunol 2019; 203: 3447-3460 [PMID: 31704881 DOI: 10.4049/jimmunol.1900692]
- Zhang J, Rong Y, Luo C, Cui W. Bone marrow mesenchymal stem cell-derived exosomes prevent osteoarthritis by regulating synovial 45 macrophage polarization. Aging (Albany NY) 2020; 12: 25138-25152 [PMID: 33350983 DOI: 10.18632/aging.104110]
- Shinohara I, Tsubosaka M, Toya M, Lee ML, Kushioka J, Murayama M, Gao Q, Li X, Zhang N, Chow SK, Matsumoto T, Kuroda R, 46 Goodman SB. C-C Motif Chemokine Ligand 2 Enhances Macrophage Chemotaxis, Osteogenesis, and Angiogenesis during the Inflammatory Phase of Bone Regeneration. *Biomolecules* 2023; 13 [PMID: 38002347 DOI: 10.3390/biom13111665]
- 47 Toya M, Zhang N, Tsubosaka M, Kushioka J, Gao Q, Li X, Chow SK, Goodman SB. CCL2 promotes osteogenesis by facilitating macrophage migration during acute inflammation. Front Cell Dev Biol 2023; 11: 1213641 [PMID: 37457301 DOI: 10.3389/fcell.2023.1213641]
- Rahimi P, Wang CY, Stashenko P, Lee SK, Lorenzo JA, Graves DT. Monocyte chemoattractant protein-1 expression and monocyte 48 recruitment in osseous inflammation in the mouse. Endocrinology 1995; 136: 2752-2759 [PMID: 7750500 DOI: 10.1210/endo.136.6.7750500]
- 49 Zheng C, Sui B, Zhang X, Hu J, Chen J, Liu J, Wu D, Ye Q, Xiang L, Qiu X, Liu S, Deng Z, Zhou J, Shi S, Jin Y. Apoptotic vesicles restore liver macrophage homeostasis to counteract type 2 diabetes. J Extracell Vesicles 2021; 10: e12109 [PMID: 34084287 DOI: 10.1002/jev2.12109
- Yuan Y, Yuan L, Li L, Liu F, Liu J, Chen Y, Cheng J, Lu Y. Mitochondrial transfer from mesenchymal stem cells to macrophages restricts 50 inflammation and alleviates kidney injury in diabetic nephropathy mice via PGC-1α activation. Stem Cells 2021; 39: 913-928 [PMID: 33739541 DOI: 10.1002/stem.3375]
- Jiang X, Yang J, Lin Y, Liu F, Tao J, Zhang W, Xu J, Zhang M. Extracellular vesicles derived from human ESC-MSCs target macrophage and 51 promote anti-inflammation process, angiogenesis, and functional recovery in ACS-induced severe skeletal muscle injury. Stem Cell Res Ther 2023; 14: 331 [PMID: 37964317 DOI: 10.1186/s13287-023-03530-1]
- Li R, Zhao K, Ruan Q, Meng C, Yin F. Bone marrow mesenchymal stem cell-derived exosomal microRNA-124-3p attenuates neurological 52 damage in spinal cord ischemia-reperfusion injury by downregulating Ern1 and promoting M2 macrophage polarization. Arthritis Res Ther 2020; **22**: 75 [PMID: 32272965 DOI: 10.1186/s13075-020-2146-x]
- Chang Q, Hao Y, Wang Y, Zhuo H, Zhao G. Bone marrow mesenchymal stem cell-derived exosomal microRNA-125a promotes M2 53 macrophage polarization in spinal cord injury by downregulating IRF5. Brain Res Bull 2021; 170: 199-210 [PMID: 33609602 DOI: 10.1016/j.brainresbull.2021.02.015
- Abe Y, Ochiai D, Sato Y, Kanzaki S, Ikenoue S, Kasuga Y, Tanaka M. Prophylactic Therapy with Human Amniotic Fluid Stem Cells 54 Improves Long-Term Cognitive Impairment in Rat Neonatal Sepsis Survivors. Int J Mol Sci 2020; 21 [PMID: 33339379 DOI: 10.3390/iims21249590]



- Lu D, Jiao X, Jiang W, Yang L, Gong Q, Wang X, Wei M, Gong S. Mesenchymal stem cells influence monocyte/macrophage phenotype: 55 Regulatory mode and potential clinical applications. Biomed Pharmacother 2023; 165: 115042 [PMID: 37379639 DOI: 10.1016/j.biopha.2023.115042]
- Ghahremani Piraghaj M, Soudi S, Ghanbarian H, Bolandi Z, Namaki S, Hashemi SM. Effect of efferocytosis of apoptotic mesenchymal stem 56 cells (MSCs) on C57BL/6 peritoneal macrophages function. Life Sci 2018; 212: 203-212 [PMID: 30287233 DOI: 10.1016/j.Ifs.2018.09.052]
- Altemus J, Dadgar N, Li Y, Lightner AL. Adipose tissue-derived mesenchymal stem cells' acellular product extracellular vesicles as a potential 57 therapy for Crohn's disease. J Cell Physiol 2022; 237: 3001-3011 [PMID: 35522572 DOI: 10.1002/jcp.30756]
- Sohni A, Verfaillie CM. Mesenchymal stem cells migration homing and tracking. Stem Cells Int 2013; 2013: 130763 [PMID: 24194766 DOI: 58 10.1155/2013/130763]
- 59 Wang M, Chen F, Wang J, Chen X, Liang J, Yang X, Zhu X, Fan Y, Zhang X. Calcium phosphate altered the cytokine secretion of macrophages and influenced the homing of mesenchymal stem cells. J Mater Chem B 2018; 6: 4765-4774 [PMID: 32254303 DOI: 10.1039/c8tb01201f
- Yu Y, Wu RX, Gao LN, Xia Y, Tang HN, Chen FM. Stromal cell-derived factor-1-directed bone marrow mesenchymal stem cell migration in 60 response to inflammatory and/or hypoxic stimuli. Cell Adh Migr 2016; 10: 342-359 [PMID: 26745021 DOI: 10.1080/19336918.2016.1139287]
- Fan H, Zhao G, Liu L, Liu F, Gong W, Liu X, Yang L, Wang J, Hou Y. Pre-treatment with IL-1β enhances the efficacy of MSC 61 transplantation in DSS-induced colitis. Cell Mol Immunol 2012; 9: 473-481 [PMID: 23085948 DOI: 10.1038/cmi.2012.40]
- Wang Y, Xu J, Zhang X, Wang C, Huang Y, Dai K. TNF-a-induced LRG1 promotes angiogenesis and mesenchymal stem cell migration in the 62 subchondral bone during osteoarthritis. Cell Death Dis 2017; 8: e2715 [PMID: 28358372 DOI: 10.1038/cddis.2017.129]
- Qi Y, Zhu T, Zhang T, Wang X, Li W, Chen D, Meng H, An S. M1 macrophage-derived exosomes transfer miR-222 to induce bone marrow 63 mesenchymal stem cell apoptosis. Lab Invest 2021; 101: 1318-1326 [PMID: 34230646 DOI: 10.1038/s41374-021-00622-5]
- Chen YC, Lai YS, Hsuuw YD, Chang KT. Withholding of M-CSF Supplement Reprograms Macrophages to M2-Like via Endogenous CSF-1 64 Activation. Int J Mol Sci 2021; 22 [PMID: 33805444 DOI: 10.3390/ijms22073532]
- Lin F, Xue D, Xie T, Pan Z. HMGB1 promotes cellular chemokine synthesis and potentiates mesenchymal stromal cell migration via Rap1 65 activation. Mol Med Rep 2016; 14: 1283-1289 [PMID: 27314424 DOI: 10.3892/mmr.2016.5398]
- Lin F, Zhang W, Xue D, Zhu T, Li J, Chen E, Yao X, Pan Z. Signaling pathways involved in the effects of HMGB1 on mesenchymal stem cell 66 migration and osteoblastic differentiation. Int J Mol Med 2016; 37: 789-797 [PMID: 26846297 DOI: 10.3892/ijmm.2016.2479]
- 67 Correction to: Hypoxic ASCs-derived Exosomes Attenuate Colitis by Regulating Macrophage Polarization via miR-216a-5p/HMGB1 Axis. Inflamm Bowel Dis 2023; 29: 1515 [PMID: 37196143 DOI: 10.1093/ibd/izad101]
- Biguetti CC, Cavalla F, Silveira EV, Tabanez AP, Francisconi CF, Taga R, Campanelli AP, Trombone APF, Rodrigues DC, Garlet GP. 68 HGMB1 and RAGE as Essential Components of Ti Osseointegration Process in Mice. Front Immunol 2019; 10: 709 [PMID: 31024546 DOI: 10.3389/fimmu.2019.00709
- 69 Chong JJ, Chandrakanthan V, Xaymardan M, Asli NS, Li J, Ahmed I, Heffernan C, Menon MK, Scarlett CJ, Rashidianfar A, Biben C, Zoellner H, Colvin EK, Pimanda JE, Biankin AV, Zhou B, Pu WT, Prall OW, Harvey RP. Adult cardiac-resident MSC-like stem cells with a proepicardial origin. Cell Stem Cell 2011; 9: 527-540 [PMID: 22136928 DOI: 10.1016/j.stem.2011.10.002]
- Vismara I, Papa S, Rossi F, Forloni G, Veglianese P. Current Options for Cell Therapy in Spinal Cord Injury. Trends Mol Med 2017; 23: 831-70 849 [PMID: 28811172 DOI: 10.1016/j.molmed.2017.07.005]
- Kang JY, Oh MK, Joo H, Park HS, Chae DH, Kim J, Lee HR, Oh IH, Yu KR. Xeno-Free Condition Enhances Therapeutic Functions of 71 Human Wharton's Jelly-Derived Mesenchymal Stem Cells against Experimental Colitis by Upregulated Indoleamine 2,3-Dioxygenase Activity. *J Clin Med* 2020; **9** [PMID: 32927587 DOI: 10.3390/jcm9092913]
- Teissier V, Gao Q, Shen H, Li J, Li X, Huang EE, Kushioka J, Toya M, Tsubosaka M, Hirata H, Alizadeh HV, Maduka CV, Contag CH, Yang 72 YP, Zhang N, Goodman SB. Metabolic profile of mesenchymal stromal cells and macrophages in the presence of polyethylene particles in a 3D model. Stem Cell Res Ther 2023; 14: 99 [PMID: 37085909 DOI: 10.1186/s13287-023-03260-4]
- Guihard P, Danger Y, Brounais B, David E, Brion R, Delecrin J, Richards CD, Chevalier S, Rédini F, Heymann D, Gascan H, Blanchard F. 73 Induction of osteogenesis in mesenchymal stem cells by activated monocytes/macrophages depends on oncostatin M signaling. Stem Cells 2012; 30: 762-772 [PMID: 22267310 DOI: 10.1002/stem.1040]
- Schulert GS, Fall N, Harley JB, Shen N, Lovell DJ, Thornton S, Grom AA. Monocyte MicroRNA Expression in Active Systemic Juvenile 74 Idiopathic Arthritis Implicates MicroRNA-125a-5p in Polarized Monocyte Phenotypes. Arthritis Rheumatol 2016; 68: 2300-2313 [PMID: 27014994 DOI: 10.1002/art.39694]
- Xu J, Kong L, Oliver BA, Li B, Creasey EA, Guzman G, Schenone M, Carey KL, Carr SA, Graham DB, Deguine J, Xavier RJ. Constitutively 75 active autophagy in macrophages dampens inflammation through metabolic and post-transcriptional regulation of cytokine production. Cell Rep 2023; 42: 112708 [PMID: 37392388 DOI: 10.1016/j.celrep.2023.112708]
- Julier Z, Park AJ, Briquez PS, Martino MM. Promoting tissue regeneration by modulating the immune system. Acta Biomater 2017; 53: 13-28 76 [PMID: 28119112 DOI: 10.1016/j.actbio.2017.01.056]
- Cooke JP. Inflammation and Its Role in Regeneration and Repair. Circ Res 2019; 124: 1166-1168 [PMID: 30973815 DOI: 77 10.1161/CIRCRESAHA.118.314669]
- Granulocyte-macrophage colony-stimulation factor and infection. Infection 1992; 20 Suppl 2: S79-134 [PMID: 1493938] 78
- Deng Z, Ren Y, Park MS, Kim HKW. Damage associated molecular patterns in necrotic femoral head inhibit osteogenesis and promote 79 fibrogenesis of mesenchymal stem cells. Bone 2022; 154: 116215 [PMID: 34571205 DOI: 10.1016/j.bone.2021.116215]
- Li Y, Dong Y, Ran Y, Zhang Y, Wu B, Xie J, Cao Y, Mo M, Li S, Deng H, Hao W, Yu S, Wu Y. Three-dimensional cultured mesenchymal 80 stem cells enhance repair of ischemic stroke through inhibition of microglia. Stem Cell Res Ther 2021; 12: 358 [PMID: 34154653 DOI: 10.1186/s13287-021-02416-4]
- Nakao Y, Fukuda T, Zhang Q, Sanui T, Shinjo T, Kou X, Chen C, Liu D, Watanabe Y, Hayashi C, Yamato H, Yotsumoto K, Tanaka U, 81 Taketomi T, Uchiumi T, Le AD, Shi S, Nishimura F. Exosomes from TNF-α-treated human gingiva-derived MSCs enhance M2 macrophage polarization and inhibit periodontal bone loss. Acta Biomater 2021; 122: 306-324 [PMID: 33359765 DOI: 10.1016/j.actbio.2020.12.046]
- 82 Hotokezaka H, Sakai E, Kanaoka K, Saito K, Matsuo K, Kitaura H, Yoshida N, Nakayama K. U0126 and PD98059, specific inhibitors of MEK, accelerate differentiation of RAW264.7 cells into osteoclast-like cells. J Biol Chem 2002; 277: 47366-47372 [PMID: 12237315 DOI: 10.1074/jbc.M208284200]
- Vi L, Baht GS, Whetstone H, Ng A, Wei Q, Poon R, Mylvaganam S, Grynpas M, Alman BA. Macrophages promote osteoblastic 83 differentiation in-vivo: implications in fracture repair and bone homeostasis. J Bone Miner Res 2015; 30: 1090-1102 [PMID: 25487241 DOI:



10.1002/jbmr.2422]

- 84 Chahal J, Gómez-Aristizábal A, Shestopaloff K, Bhatt S, Chaboureau A, Fazio A, Chisholm J, Weston A, Chiovitti J, Keating A, Kapoor M, Ogilvie-Harris DJ, Syed KA, Gandhi R, Mahomed NN, Marshall KW, Sussman MS, Naraghi AM, Viswanathan S. Bone Marrow Mesenchymal Stromal Cell Treatment in Patients with Osteoarthritis Results in Overall Improvement in Pain and Symptoms and Reduces Synovial Inflammation. Stem Cells Transl Med 2019; 8: 746-757 [PMID: 30964245 DOI: 10.1002/sctm.18-0183]
- Giri J, Das R, Nylen E, Chinnadurai R, Galipeau J. CCL2 and CXCL12 Derived from Mesenchymal Stromal Cells Cooperatively Polarize IL-85 10+ Tissue Macrophages to Mitigate Gut Injury. Cell Rep 2020; 30: 1923-1934.e4 [PMID: 32049021 DOI: 10.1016/j.celrep.2020.01.047]
- Siddiqui JA, Partridge NC. Physiological Bone Remodeling: Systemic Regulation and Growth Factor Involvement. Physiology (Bethesda) 86 2016; 31: 233-245 [PMID: 27053737 DOI: 10.1152/physiol.00061.2014]
- Hu K, Shang Z, Yang X, Zhang Y, Cao L. Macrophage Polarization and the Regulation of Bone Immunity in Bone Homeostasis. J Inflamm 87 Res 2023; 16: 3563-3580 [PMID: 37636272 DOI: 10.2147/JIR.S423819]
- 88 Gentek R, Molawi K, Sieweke MH. Tissue macrophage identity and self-renewal. Immunol Rev 2014; 262: 56-73 [PMID: 25319327 DOI: 10.1111/imr.12224]
- Mun SH, Park PSU, Park-Min KH. The M-CSF receptor in osteoclasts and beyond. Exp Mol Med 2020; 52: 1239-1254 [PMID: 32801364 89 DOI: 10.1038/s12276-020-0484-z]
- Zhao Q, Wang X, Liu Y, He A, Jia R. NFATc1: functions in osteoclasts. Int J Biochem Cell Biol 2010; 42: 576-579 [PMID: 20035895 DOI: 90 10.1016/j.biocel.2009.12.018
- Asagiri M, Takayanagi H. The molecular understanding of osteoclast differentiation. Bone 2007; 40: 251-264 [PMID: 17098490 DOI: 91 10.1016/j.bone.2006.09.023
- Takayanagi H, Kim S, Koga T, Nishina H, Isshiki M, Yoshida H, Saiura A, Isobe M, Yokochi T, Inoue J, Wagner EF, Mak TW, Kodama T, 92 Taniguchi T. Induction and activation of the transcription factor NFATc1 (NFAT2) integrate RANKL signaling in terminal differentiation of osteoclasts. Dev Cell 2002; 3: 889-901 [PMID: 12479813 DOI: 10.1016/s1534-5807(02)00369-6]
- 93 Huang R, Wang X, Zhou Y, Xiao Y. RANKL-induced M1 macrophages are involved in bone formation. Bone Res 2017; 5: 17019 [PMID: 29263936 DOI: 10.1038/boneres.2017.19]
- Piprode V, Singh K, Kumar A, Josh SR, Wani MR. IL-3 inhibits rat osteoclast differentiation induced by TNF-a and other pro-94 osteoclastogenic cytokines. J Biosci 2021; 46 [PMID: 34183475]
- Fukawa Y, Kayamori K, Tsuchiya M, Ikeda T. IL-1 Generated by Oral Squamous Cell Carcinoma Stimulates Tumor-Induced and RANKL-95 Induced Osteoclastogenesis: A Possible Mechanism of Bone Resorption Induced by the Infiltration of Oral Squamous Cell Carcinoma. Int J Mol Sci 2022; 24 [PMID: 36614130 DOI: 10.3390/ijms24010688]
- Zhao JJ, Wu ZF, Yu YH, Wang L, Cheng L. Effects of interleukin-7/interleukin-7 receptor on RANKL-mediated osteoclast differentiation and 96 ovariectomy-induced bone loss by regulating c-Fos/c-Jun pathway. J Cell Physiol 2018; 233: 7182-7194 [PMID: 29663382 DOI: 10.1002/jcp.26548]
- Ganesan R, Rasool M. Interleukin 17 regulates SHP-2 and IL-17RA/STAT-3 dependent Cyr61, IL-23 and GM-CSF expression and RANKL 97 mediated osteoclastogenesis by fibroblast-like synoviocytes in rheumatoid arthritis. Mol Immunol 2017; 91: 134-144 [PMID: 28898718 DOI: 10.1016/j.molimm.2017.09.003
- 98 Mae M, Alam MI, Yamashita Y, Ozaki Y, Higuchi K, Ziauddin SM, Montenegro Raudales JL, Sakai E, Tsukuba T, Yoshimura A. The Role of Cytokines Produced via the NLRP3 Inflammasome in Mouse Macrophages Stimulated with Dental Calculus in Osteoclastogenesis. Int J Mol Sci 2021; 22 [PMID: 34830316 DOI: 10.3390/ijms222212434]
- Liang M, Ma Q, Ding N, Luo F, Bai Y, Kang F, Gong X, Dong R, Dai J, Dai Q, Dou C, Dong S. IL-11 is essential in promoting osteolysis in 99 breast cancer bone metastasis via RANKL-independent activation of osteoclastogenesis. Cell Death Dis 2019; 10: 353 [PMID: 31040267 DOI: 10.1038/s41419-019-1594-1
- Glass GE, Chan JK, Freidin A, Feldmann M, Horwood NJ, Nanchahal J. TNF-alpha promotes fracture repair by augmenting the recruitment 100 and differentiation of muscle-derived stromal cells. Proc Natl Acad Sci USA 2011; 108: 1585-1590 [PMID: 21209334 DOI: 10.1073/pnas.1018501108]
- Takayanagi H, Ogasawara K, Hida S, Chiba T, Murata S, Sato K, Takaoka A, Yokochi T, Oda H, Tanaka K, Nakamura K, Taniguchi T. T-101 cell-mediated regulation of osteoclastogenesis by signalling cross-talk between RANKL and IFN-gamma. Nature 2000; 408: 600-605 [PMID: 11117749 DOI: 10.1038/350461021
- 102 Johnson RW, McGregor NE, Brennan HJ, Crimeen-Irwin B, Poulton IJ, Martin TJ, Sims NA. Glycoprotein130 (Gp130)/interleukin-6 (IL-6) signalling in osteoclasts promotes bone formation in periosteal and trabecular bone. Bone 2015; 81: 343-351 [PMID: 26255596 DOI: 10.1016/j.bone.2015.08.005
- Chen S, Liang H, Ji Y, Kou H, Zhang C, Shang G, Shang C, Song Z, Yang L, Liu L, Wang Y, Liu H. Curcumin Modulates the Crosstalk 103 Between Macrophages and Bone Mesenchymal Stem Cells to Ameliorate Osteogenesis. Front Cell Dev Biol 2021; 9: 634650 [PMID: 33634135 DOI: 10.3389/fcell.2021.634650]
- Kamel Mohamed SG, Sugiyama E, Shinoda K, Hounoki H, Taki H, Maruyama M, Miyahara T, Kobayashi M. Interleukin-4 inhibits RANKL-104 induced expression of NFATc1 and c-Fos: a possible mechanism for downregulation of osteoclastogenesis. Biochem Biophys Res Commun 2005; 329: 839-845 [PMID: 15752732 DOI: 10.1016/j.bbrc.2005.02.049]
- Chen H, Fang C, Zhi X, Song S, Gu Y, Chen X, Cui J, Hu Y, Weng W, Zhou Q, Wang Y, Jiang H, Li X, Cao L, Su J. Neobavaisoflavone 105 inhibits osteoclastogenesis through blocking RANKL signalling-mediated TRAF6 and c-Src recruitment and NF-KB, MAPK and Akt pathways. J Cell Mol Med 2020; 24: 9067-9084 [PMID: 32604472 DOI: 10.1111/jcmm.15543]
- Vallés G, Bensiamar F, Maestro-Paramio L, García-Rey E, Vilaboa N, Saldaña L. Influence of inflammatory conditions provided by 106 macrophages on osteogenic ability of mesenchymal stem cells. Stem Cell Res Ther 2020; 11: 57 [PMID: 32054534 DOI: 10.1186/s13287-020-1578-1]
- Guihard P, Boutet MA, Brounais-Le Royer B, Gamblin AL, Amiaud J, Renaud A, Berreur M, Rédini F, Heymann D, Layrolle P, Blanchard F. 107 Oncostatin m, an inflammatory cytokine produced by macrophages, supports intramembranous bone healing in a mouse model of tibia injury. Am J Pathol 2015; 185: 765-775 [PMID: 25559270 DOI: 10.1016/j.ajpath.2014.11.008]
- Choe JY, Park KY, Park SH, Lee SI, Kim SK. Regulatory effect of calcineurin inhibitor, tacrolimus, on IL-6/sIL-6R-mediated RANKL 108 expression through JAK2-STAT3-SOCS3 signaling pathway in fibroblast-like synoviocytes. Arthritis Res Ther 2013; 15: R26 [PMID: 23406906 DOI: 10.1186/ar4162]
- 109 Li CH, Xu LL, Jian LL, Yu RH, Zhao JX, Sun L, Du GH, Liu XY. Stattic inhibits RANKL-mediated osteoclastogenesis by suppressing



activation of STAT3 and NF-kB pathways. Int Immunopharmacol 2018; 58: 136-144 [PMID: 29587202 DOI: 10.1016/j.intimp.2018.03.021]

- Hu L, Liu R, Zhang L. Advance in bone destruction participated by JAK/STAT in rheumatoid arthritis and therapeutic effect of JAK/STAT 110 inhibitors. Int Immunopharmacol 2022; 111: 109095 [PMID: 35926270 DOI: 10.1016/j.intimp.2022.109095]
- Wu S, Ma J, Liu J, Liu C, Ni S, Dai T, Wang Y, Weng Y, Zhao H, Zhou D, Zhao X. Immunomodulation of Telmisartan-Loaded PCL/PVP 111 Scaffolds on Macrophages Promotes Endogenous Bone Regeneration. ACS Appl Mater Interfaces 2022; 14: 15942-15955 [PMID: 35353482 DOI: 10.1021/acsami.1c24748]
- Antebi B, Rodriguez LA 2nd, Walker KP 3rd, Asher AM, Kamucheka RM, Alvarado L, Mohammadipoor A, Cancio LC. Short-term 112 physiological hypoxia potentiates the therapeutic function of mesenchymal stem cells. Stem Cell Res Ther 2018; 9: 265 [PMID: 30305185 DOI: 10.1186/s13287-018-1007-x]
- Bosurgi L, Cao YG, Cabeza-Cabrerizo M, Tucci A, Hughes LD, Kong Y, Weinstein JS, Licona-Limon P, Schmid ET, Pelorosso F, Gagliani 113 N, Craft JE, Flavell RA, Ghosh S, Rothlin CV. Macrophage function in tissue repair and remodeling requires IL-4 or IL-13 with apoptotic cells. Science 2017; 356: 1072-1076 [PMID: 28495875 DOI: 10.1126/science.aai8132]
- 114 Kozicky LK, Sly LM. Depletion and Reconstitution of Macrophages in Mice. Methods Mol Biol 2019; 1960: 101-112 [PMID: 30798525 DOI: 10.1007/978-1-4939-9167-9_9]
- Yu K, Huangfu H, Qin Q, Zhang Y, Gu X, Liu X, Zhou Y. Application of Bone Marrow-Derived Macrophages Combined with Bone 115 Mesenchymal Stem Cells in Dual-Channel Three-Dimensional Bioprinting Scaffolds for Early Immune Regulation and Osteogenic Induction in Rat Calvarial Defects. ACS Appl Mater Interfaces 2022; 14: 47052-47065 [PMID: 36194837 DOI: 10.1021/acsami.2c13557]
- 116 Majrashi M, Kotowska A, Scurr D, Hicks JM, Ghaemmaghami A, Yang J. Sustained Release of Dexamethasone from 3D-Printed Scaffolds Modulates Macrophage Activation and Enhances Osteogenic Differentiation. ACS Appl Mater Interfaces 2023; 15: 56623-56638 [PMID: 38016086 DOI: 10.1021/acsami.3c09774]
- 117 Niu Y, Li Q, Xie R, Liu S, Wang R, Xing P, Shi Y, Wang Y, Dong L, Wang C. Modulating the phenotype of host macrophages to enhance osteogenesis in MSC-laden hydrogels: Design of a glucomannan coating material. Biomaterials 2017; 139: 39-55 [PMID: 28582717 DOI: 10.1016/j.biomaterials.2017.05.042]
- Demarquay C, Moussa L, Réthoré G, Milliat F, Weiss P, Mathieu N. Embedding MSCs in Si-HPMC hydrogel decreased MSC-directed host 118 immune response and increased the regenerative potential of macrophages. Regen Biomater 2022; 9: rbac022 [PMID: 35784096 DOI: 10.1093/rb/rbac022]
- 119 Nativel F, Smith A, Boulestreau J, Lépine C, Baron J, Marquis M, Vignes C, Le Guennec Y, Veziers J, Lesoeur J, Loll F, Halgand B, Renard D, Abadie J, Legoff B, Blanchard F, Gauthier O, Vinatier C, Rieux AD, Guicheux J, Le Visage C. Micromolding-based encapsulation of mesenchymal stromal cells in alginate for intraarticular injection in osteoarthritis. Mater Today Bio 2023; 19: 100581 [PMID: 36896417 DOI: 10.1016/j.mtbio.2023.100581]
- Doron G, Wood LB, Guldberg RE, Temenoff JS. Poly(ethylene glycol)-Based Hydrogel Microcarriers Alter Secretory Activity of Genetically 120 Modified Mesenchymal Stromal Cells. ACS Biomater Sci Eng 2023; 9: 6282-6292 [PMID: 37906515 DOI: 10.1021/acsbiomaterials.3c00954]
- Chen Z, Wu C, Gu W, Klein T, Crawford R, Xiao Y. Osteogenic differentiation of bone marrow MSCs by β-tricalcium phosphate stimulating 121 macrophages via BMP2 signalling pathway. Biomaterials 2014; 35: 1507-1518 [PMID: 24268199 DOI: 10.1016/j.biomaterials.2013.11.014]





Published by Baishideng Publishing Group Inc 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA Telephone: +1-925-3991568 E-mail: office@baishideng.com Help Desk: https://www.f6publishing.com/helpdesk https://www.wjgnet.com

