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REVIEW

Challenges and dilemmas in pediatric hepatic Wilson's disease

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Abstract

Wilson disease is an autosomal recessive disorder affecting the ATP7B gene located on chromosome 13q. This leads to copper deposition in various organs, most importantly in the liver and brain. The genetic mutations are vast, well reported in the West but poorly documented in developing countries. Hence the diagnosis is made with a constellation of clinico-laboratory parameters which have significant overlap with other liver diseases and often pose a significant dilemma for clinicians. Diagnostic scoring systems are not fool-proof. The availability and affordability of chelators in developing countries impact the drug compliance of patients. While D-penicillamine is a potent drug, its side effects lead to drug discontinuation. Trientine is cost-prohibitive in developing countries. There is no single test to assess the adequacy of chelation. Exchangeable urinary copper is an essential upcoming diagnostic and prognostic tool. In the presence of cirrhosis, hypersplenism clouds the assessment of myelosuppression of drugs. Similarly, it may be difficult to distinguish disease tubulopathy from druginduced glomerulonephritis. Neurological worsening due to chelators may appear similar to disease progression. Presentation as fulminant hepatic failure requires rapid workup. There is a limited window of opportunity to salvage these patients with the help of plasmapheresis and other liver-assisted devices. This review addresses the challenges and clinical dilemmas faced at beside in developing countries.

Key Words: Wilson's disease; Children; Hepatic Wilson disease; D-penicillamine; Trientine; Exchangeable copper

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Core Tip: Wilson's disease is an important and common cause of chronic liver disease among children with multi-organ affection. There are various biochemical parameters that aid in making the diagnosis but are nonetheless fraught with limitations. Treatment is lifelong and it's important to maintain adequate chelation while monitoring for its adverse effects. It is important to distinguish the adverse effects of chelators from the disease manifestations per se, as the management is diagonally opposite, for example, differentiating D-penicillamine-induced paradoxical neurological worsening from neurological manifestations of Wilson disease. This review discusses these challenges in making a diagnosis and treating pediatric hepatic Wilson's disease.

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INTRODUCTION

Wilson disease (WD) is a genetic disorder of copper metabolism, with an autosomal recessive mode of inheritance. The estimated prevalence of WD is approximately 30 per million population[1]. Among all causes of hepatobiliary disorders in children, in a 3.5-year study period, in a tertiary care centre in India, WD accounted for 7.6% of total cases[2]. It is important to make an early diagnosis for the prevention of disease progression and the identification of pre-symptomatic cases. This paper addresses the management difficulties in Wilson disease in terms of the identification of varied atypical clinical presentations, diagnostic dilemmas, and pharmacotherapy-related problems.

Pathogenesis

Figure 1 depicts the etiopathogenesis of WD. WD is caused by a mutation in the ATP7B gene, located on chromosome no. 13 Long arm, (13q14.3). This gene codes for an ATPase (a metal-transporting P-type adenosine triphosphatase), which is membrane-bound and mediates trans-Golgi migration of copper within the hepatocyte and its subsequent excretion into the biliary canaliculi [3]. Copper (Cu^+) from the diet enters the hepatocyte via the copper transport receptor 1 (CTR1) located in the apical membrane. Then it binds to Atox1 (antioxidant protein1) and is then handed over to ATPase (gene product of ATP7B) for its transport into the trans-Golgi network (TGN)[4]. Ceruloplasmin has two functions- i. it transports copper to the TGN where apoceruloplasmin binds to 6 atoms of copper to form holo-ceruloplasmin ii. in copper loaded condition, ATP7B goes to the endocytic vesicles to mediate copper excretion into the bile canaliculi[5]. Thus, in normal conditions, ATP7B is present in the trans-Golgi network and helps in the synthesis of holo-ceruloplasmin. But, in the copper-excess state, the protein moves towards the canalicular membrane to promote copper excretion[5]. Apo-ceruloplasmin is the inactive form of ceruloplasmin and is less stable. In the absence of this transporter, apoceruloplasmin degrades, and non-TGN-transported copper accumulates in the liver and gets bound to metallothioneins (endogenous chelators). When the copper binding capacity of metallothioneins is overwhelmed, the free copper spills into the lysosomes and causes free radical-mediated cell damage. Ultimately, this free copper comes to the peripheral circulation, where it is weakly bound to albumin and gets deposited in other tissues (brain, kidneys, cornea) (Figure 1). Apoceruloplasmin is a plasma protein and carrier of copper, mainly produced by the liver. In normal physiology, after copper reaches the liver via the portal circulation, 20% of copper is re-excreted into the GI tract via bile and the remaining 80% is transported to the periphery, bound to ceruloplasmin (Apoceruloplasmin + copper = holoceruloplasmin). However, in WD, holoceruloplasmin is reduced, and free copper (non-ceruloplasmin bound copper) is elevated, which leads to high urinary excretion of copper[6,7]. Hepatic involvement is the most common presentation in children with WD, as liver is the major organ responsible for copper homeostasis. Initially, copper binds with metallothionein and accumulates in lysosomes and progressively causes mitochondrial damage leading to hepatic steatosis, fibrosis, and cirrhosis. Subsequently, non-ceruloplasmin-bound excess and toxic copper leaks into the blood and accumulates in the other tissues like brain, leading to neurological manifestations, which is the commonest presentation in older children and adults.

DIAGNOSTIC DIFFICULTIES

Hepatic Wilson disease may present as acute hepatitis, persistent or intermittent elevations of transaminases, organomegaly, fluctuating or worsening jaundice, resembling autoimmune hepatitis, fatty liver, acute liver failure or as decompensated CLD[6,8,9]. Other major presentations of WD include neurological presentation with extrapyramidal symptoms and psychiatric manifestations in the form of tremors, dystonia, dysarthria, gait disturbances, and psychiatric manifestations, like behavioral abnormalities and psychiatric manifestations, like bipolar affective disorder and psychosis with cognitive decline. It is hypothesized that it is because of copper which gets deposited in the basal ganglia by penetrating through the choroid plexus (fenestrated endothelium)[4]. Minor presentations include renal tubular acidosis presenting with microscopic hematuria, osseo-muscular involvement in the form of pathological fractures, arthralgia,

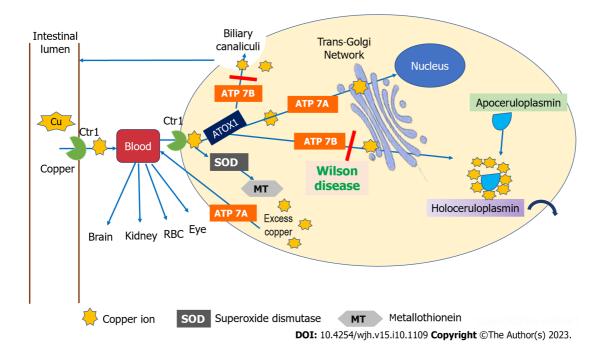


Figure 1 Dietary copper (Cu) is taken up in the small intestine by intestinal enterocytes by copper transport receptor 1 (Ctr1) on the apical site. Most of the newly absorbed copper is taken up by the liver, mediated primarily by Ctr1. Accumulated copper is sequestered by superoxide dismutase and stored in metallothioneins. Specialized chaperons shuttle copper to its specific cellular targets: Antioxidant protein 1 (Atox1) to the copper-transporting ATPases ATP7A and ATP7B. Both ATP7A and ATP7B transport copper into the trans-Golgi network for subsequent incorporation into copper-dependent enzymes. ATP7B provides copper for incorporation into ceruloplasmin as well as for biliary excretion of copper, whereas ATP7A has role in copper mobilization from hepatic stores into the bloodstream during peripheral copper deficiency. Mutation in ATP7B gene leads to impaired biliary excretion of copper, causing copper overload in liver and other tissues as well as failure of loading of ceruloplasmin with copper. MT: Metallothioneins.

proximal muscle weakness.), and importantly haematological presentation in the form of acute fulminant liver failure with Coomb's negative haemolytic anemia.

To make the diagnosis of Wilson disease, we use the presence of a Kayser-Fleischer (K-F) ring on slit lamp exam, 24-h urine copper of > 100 micrograms (mcg) per day, and ceruloplasmin < 20 milligrams/decilitre (mg/dL) commonly. These biochemical parameters are not to be interpreted in isolation; it is prudent to remember various factors that affect them. Table 1 summarizes all the diagnostic tests and their interpretation. Table 2 enlists various diagnostic tests used in suspected WD patients and for family screening.

Ceruloplasmin

Apo-ceruloplasmin combines with copper and is secreted from the liver in the form of holo-ceruloplasmin. Serum ceruloplasmin is measured by two methods; either enzymatically by measuring its copper-dependent ferroxidase activity (holoceruloplasmin) or by antibody-based tests such as radioimmunoassay, radial immunodiffusion, or nephelometry [10]. The enzymatic method measures holo-ceruloplasmin, whereas immunologic assays measure both apo ceruloplasmin and holo-ceruloplasmin and thus can overestimate ceruloplasmin levels[10,11]. It has been suggested that the enzymatic method is better than the immunologic method to diagnose Wilson's disease[12]. In those patients with impaired synthetic functions due to causes other than WD, the specificity of both these tests dropped, but the enzymatic test had better specificity as compared to immunologic assays (84.5% vs 68.9%)[12]. However, in routine practice, immunological assays are more readily available, rapid and commonly used. The normal value of ceruloplasmin ranges between 20-40 mg/dL. Ceruloplasmin level < 20 mg/dL with the presence of Kayser Fleischer (KF) ring is considered consistent with WD[8]. Ceruloplasmin < 5 mg/dL is strongly suggestive of WD[6]. Also, ceruloplasmin level < 14 mg/dL has a 100% positive predictive value for the diagnosis of WD and a value < 5 mg/dL is strongly suggestive of WD[8,13]. Serum ceruloplasmin is also an acute phase reactant and may be falsely elevated in the presence of sepsis and it may be falsely low in conditions like fulminant hepatic failure of any etiology, advanced liver disease, celiac disease and aceruloplasminemia, nephrotic syndrome, and 20% of heterozygotes[10,11,14].

24-h urine copper excretion

It is another indirect marker of non-ceruloplasmin-bound copper, which is elevated in WD. 24-h urine copper excretion of more than 100 mcg per day is diagnostic of Wilson's disease[15]. However, certain other conditions like autoimmune hepatitis, chronic cholestatic liver disease and acute fulminant liver failure of any aetiology may give rise to high 24-h urine copper excretion[10]. Basal copper excretion may be low in children and asymptomatic siblings, thus a normal value doesn't rule out WD. The test may give rise to false results in case of improper urine collection and copper contamination if kept in certain types of containers. 24-h urine copper of more than 40 mcg per day may indicate WD in asymptomatic individuals. In addition to making a diagnosis, this is also a valuable test in monitoring the adequacy of

Table 1 Various laborato	ry investigations us	sed in the diagnosis and	monitoring of Wilson disease
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Diagnostic tests	Cut-off values in diagnosis	Disease monitoring	Problems in interpretation
Serum cerulo- plasmin	$<\!10$ mg/dL (strong evidence), 10-20 mg/dL (needs further evaluation), 20-40 mg/dL (normal value, but does not exclude diagnosis)	Not helpful	Can be normal in fulminant presentation and acute inflammation as it is an acute phase reactant
24-h urine copper	> 100 mcg/d (virtually diagnostic in symptomatic patients), $>$ 40 mcg/d (may indicate disease, needs further evaluation)	> 500 mcg/d during initial phase, 200-500 mcg/d in the maintenance phase	Difficult to perform, to be done in reliable laboratories
Hepatic copper	> 250 mcg/g dry weight (diagnostic), 50-250 mcg/g dry weight (needs further evaluation), $<$ 50 mcg/g dry weight (normal)	Not recommended	Inhomogenous distribution of copper (sampling error), elevated in long-standing cholestasis
Serum total copper	> 25 micromol/L (needs further evaluation), 14-24 micromol/L (90-150 mg/dL) normal	N/Aª	
Non-ceruloplasmin copper	10-15 mcg/dL (normal person), > 25 μ g/dL (untreated patients), Not recommended for diagnosis	> 15 mg/dL (poor compliance) ^a < 5 mg/dL (over-chelation)	
Exchangeable copper	\geq 2.08 micromol/L (more likelihood of extrahepatic organ involvement), 0.62 and 1.15 micromol/L (normal)	Experimental	Requires equipped laboratories and expertise
Relative exchangeable copper	> 15% (100% sensitivity and specificity for diagnosis)	Experimental	Requires equipped laboratories and expertise

^aThe serum free copper or NCC has a role in monitoring: Initial phase: > 25 mcg/dL, maintenance phase: 10-15 mcg/dL, under-chelation: > 15 mcg/dL, over-chelation: < 5 mcg/dL.

Table 2 Diagnostic tests for suspected Wilson disease patients and asymptomatic siblings			
Parameters	Tests for suspected patients	Tests for asymptomatic siblings	
Clinical examination	Yes	Yes	
Liver function test	Yes	Yes	
Slit lamp examination for Kayser-Fleischer ring	Yes	Yes	
Serum ceruloplasmin	Yes	Yes	
24 h urine copper	Yes	Yes	
Genetic analysis for ATP7B gene mutation analysis	Yes, if feasible	Yes (if proband sample is available)	
Liver biopsy	Yes, ancillary test	No	
Hepatic copper	Yes, to be done in cases of ambiguity	No	

chelation, a 24-h urine copper of 200-500 mcg per day is considered the target[8].

Liver biopsy and hepatic copper

On liver biopsy, early changes include steatosis, and fibrosis and late changes include changes of chronic active hepatitis with bridging fibrosis, occasionally resembling autoimmune hepatitis and, Mallory-Denk bodies[16]. Copper staining is demonstrated by orcein, which stains the copper-associated proteins, and rhodanine which stains the elemental copper. Copper-associated proteins may be elevated in various causes of advanced cholestatic liver disease (e.g., Primary biliary cirrhosis or Chronic hepatitis B) other than Wilson's disease, thus histochemical staining for copper is not specific for Wilson's disease[17]. Also, negative histochemical staining for copper does not rule out the possibility of WD[18]. Copper deposition also depends on the stage of disease, early in the disease, copper staining is diffuse in the cytoplasm in the periportal region, and later in the disease, it is mainly seen within the lysosomes of hepatocytes in the periphery of the regenerating nodules. This pattern of distribution suggests that the cytoplasmic distribution leads to hepatocyte damage while copper in the lysosomes is likely to be less toxic. Initial cytoplasmic deposition of copper is due to the high affinity of the sulfhydryl-rich proteins (metallothionines) present in the cytoplasm as compared to lysosomes[19]. Hepatic copper of more than 250 mcg/gram (gm) dry weight of the liver is strongly suggestive of copper deposition in the liver but it may be also found in any advanced liver disease. Liver copper < 50 mcg/gm dry weight almost excludes the diagnosis of WD[8,20]. Also, the regional variation due to regenerative nodules and tissue fibrosis is to be kept in mind while interpreting hepatic copper[10].

Serum copper

It is important to measure the serum copper simultaneously while measuring ceruloplasmin, as it helps in the calculation of the "free copper" also known as "non-ceruloplasmin bound copper (NCC)". Total serum copper comprises of holoceruloplasmin (70%), albumin-bound (20%), peptide-bound, and free copper. Holo-ceruloplasmin level is low in WD. So, in most patients of WD, serum copper is low, unless there is a massive release of free copper from the liver due to liver necrosis[21]. This free copper, which is not bound to ceruloplasmin is known as "non-ceruloplasmin bound copper", is elevated in WD and is toxic. Serum copper is estimated by either atomic absorption or emission spectrometry or by inductively coupled plasma mass spectrometry (ICP-MS) methods [22]. Total serum copper (in mcg/L) = serum copper (in micromol/L) × 63.5[10]. The normal range of serum copper is 14-24 micromol/L (90-150 mg/dL)[11]. Holo-ceruloplasmin contains 3.15 mcg of copper per mg of ceruloplasmin, hence, ceruloplasmin-bound copper may be calculated as serum ceruloplasmin (mg/dL) x 3.15 (mcg copper per mg ceruloplasmin)[11]. Free copper or NCC = (Serum copperceruloplasmin bound copper). Non-ceruloplasmin-bound copper has been proposed as a diagnostic test for WD[23]. In normal individuals, the NCC (free copper) is 10-15 mcg/dL[3]. In untreated patients, it is > 25 mcg/dL[3]. The estimation of NCC depends on the methods used to estimate serum ceruloplasmin and copper levels. Falsely high ceruloplasmin calculated by immunological tests may lead to the calculation of a falsely low or negative value of free copper. In normal conditions in WD, serum copper is low. In acute fulminant WD, due to a massive release of copper from the liver, free copper may rise up to ten times and can cause intravascular hemolysis[23]. NCC is not validated for diagnosis of WD but is helpful in monitoring treatment response. With adequate chelation, the NCC decreases to < 5 mcg/dL. Among patients who are on chelation therapy, the target 24-hour urine copper excretion is 200-500 mcg/d. However, patients with poor compliance to chelators can also have low 24-h urine copper excretion as there is adequate chelator in the circulation to leach out copper. Such patients can be differentiated by estimating the NCC. Those with high NCC > 15 mg/dL have poor compliance while those with NCC $\leq 5 \text{ mg/dL}$ are over-chelated[10].

Exchangeable copper and relative exchangeable copper

Exchangeable copper is measured by adding ethylenediaminetetraacetic acid (EDTA) to the serum sample. As EDTA is a copper chelator, it binds with copper, and EDTA-bound copper is measured as exchangeable copper. It measures copper bound to albumin, and other amino acids, which easily gets exchanged with EDTA (copper-chelating agents)[21]. Unlike NCC, it doesn't depend on serum ceruloplasmin levels. Normal values for exchangeable copper (CuEx) are between 0.62 and 1.15 micromol/L[24]. Poujois et al[25] demonstrated that CuEx is normal to moderately increased in WD, but elevated in those with acute fulminant WD and WD with extrahepatic involvement. In patients with extra-hepatic manifestations, CuEx was the only biological marker to be positively correlated with the neurologic disease burden (assessed by the Unified Wilson Disease Rating Score). CuEx determination is, in consequence, useful when diagnosing WD with a value > 2.08 micromol/L, and is indicative of the severity of the extra-hepatic involvement. However, CuEx did not indicate the severity of liver damage. CuEx is an interesting experimental biomarker but needs to be interpreted with caution, especially in WD patients with hepatic manifestation. In this study, it was postulated that there may be a threshold concentration of CuEx above which organs like the brain and eyes (and possibly others such as kidneys and heart) may be affected by copper overload [25]. Relative exchangeable copper (REC) is the ratio of exchangeable copper to total serum copper. El Balkhi et al[26] showed that REC > 18.5% can be used to diagnose WD with a sensitivity and specificity of 100%. Trocello et al[27] showed that REC can also be used to diagnose WD in asymptomatic siblings, by taking REC cut-off of 15%, it can distinguish asymptomatic WD from heterozygotes accurately.

KF ring

It represents copper deposition of copper in Descemet's membrane of the cornea, visible on slit lamp examination, it is pathognomonic of WD. It is not specific to Wilson's disease and may be found in various cholestatic conditions. It is present in 44%-62% of hepatic WD and 95% of neurological WD[3]. KF-like rings have been described in primary biliary cirrhosis, autoimmune hepatitis and severe cholestasis where the total bilirubin is > 10 g/dL. They are called bilirubin rings and disappear with the resolution of jaundice[28]. It is important to note that the appearance of the KF ring is not related to disease severity and its resolution may take variable time, months to years [29,30]. Anterior segment-optical coherence tomography (AS-OCT) can also be used to diagnose the KF ring, where the KF ring is visualized as an intense reflectivity in Descemet's membrane and it may have better accuracy in diagnosing KF ring as compared to slit-lamp examination[31]. Broniek-Kowalik et al[32] showed that AS-OCT detected copper deposition in 15 additional patients of WD, in whom the KF ring was not seen in slit lamp examination.

Coomb's negative hemolytic anemia with a disproportionate elevation of aspartate transaminase (AST) as compared to alanine transaminase (ALT) is suggestive of acute fulminant WD. The ratio of AST/ALT > 2.2 and ALP/total bilirubin ratio < 4, when combined is suggestive of acute fulminant Wilson disease with a sensitivity and specificity of 100% [33]. Alkaline phosphatases are metalloenzymes and zinc is a co-factor. In fulminant WD, during the period of massive release of copper into circulation, copper may compete with zinc for incorporation into alkaline phosphatase apoenzymes. Copper-containing enzymes released into the circulation would have little or no enzymatic activity, and serum alkaline phosphatase values would be low[34].

Neurological imaging

The face of the giant panda sign, which is caused by the normal intensity of red nuclei, preservation of signal intensity in the lateral portion of pars reticulata of substantia nigra and hypointensity in the superior colliculus with hyperintense surrounding tegmentum, is considered pathognomonic of neurological WD[35]. This finding has been shown to reverse with chelation[36,37]. Other findings include lesions in the putamen, globus pallidus, caudate, thalamus, midbrain, pons, and cerebellum as well as cortical atrophy and white matter changes. Usually, the lesions are hyperintense in T2 and hypointense in T1 weighted images. Magnetic resonance imaging (MRI) changes correlate with disease severity. Diffuse atrophy of the brain is one of the most common features [35]. MRI findings are universal in symptomatic patients and occasionally reported in pre-symptomatic patients[38].

Genetic analysis

Lastly, genetic analysis can be done in patients having ambiguity in diagnosis or for sibling screening. More than 600 pathogenic variants have been identified[39]. Most common mutations are single-nucleotide missense and nonsense mutations[39]. There is no genotype-phenotype correlation. Siblings may have varied presentations[40]. On screening the proband, the chances of a sibling being affected is 25% while either parent being affected is 0.5% [6].

Leipzig score

A diagnostic score named Leipzig score was proposed in 2001, which took into account various clinical, and biochemical, parameters to diagnose WD. A Leipzig score of more than 4 is highly suggestive of WD, while a score of 2-3 would merit further investigations and a score of less than 1 makes the diagnosis of WD unlikely[41]. There are practical difficulties in implementing the Leipzig score in developing countries. Hepatic copper is not universally available and inconsistent in reporting. Genetic testing is cumbersome and region-specific. Most regions do not report the mutations. The d-penicillamine challenge is not reliable. A revised Leipzig score was devised for the Asian setting, which gave importance to family history, greater points for the KF ring and an additional score for a ceruloplasmin level < 5 mg/dL[6].

Family screening

First-degree relatives of newly diagnosed patients should be screened by clinical examination, LFT, slit-lamp examination, serum ceruloplasmin, 24-h urine copper estimation and genetic testing, especially if the mutation is known in the proband. Screening by laboratory tests is usually deferred till 2 years of age though genetic screening is possible [6]. Preservation of the DNA sample of the proband is essential.

MANAGEMENT DIFFICULTIES

Differentiating manifestations of WD from chelation-related complications/Atypical symptoms of WD masquerading as chelation-related complications:

Kidney-related issues in WD

The renal manifestation of WD can be categorized into (1) renal involvement of underlying disease, or (2) treatmentrelated nephrotoxicity. The differentiating features in pathogenesis, laboratory investigations and management of these two conditions are summarized in Table 3. The renal involvement of WD can be in the form of renal calculi, hypercalciuria or tubulopathy. Renal manifestations of WD are due to copper deposition in the renal tubular cells. It has been shown that glomerular and tubular functions improve and normalize after starting D-penicillamine[42]. It is vital to differentiate the renal manifestations from the drug-induced glomerulonephritis as the management differs. Renal involvement in WD was first reported by Litin et al[43] in 1959 in the form of hypercalciuria. Acute renal failure, which is the most severe form, can be precipitated by massive intravascular hemolysis seen in WD[44]. Renal involvement can manifest with glomerular or non-glomerular (tubular) injury. Aminoaciduria was first detected in patients with Wilson disease by Uzman and Denny-Brown in 1948[45]. Tubular causes are tubulopathies [proximal or distal renal tubular acidosis (RTA)] presenting as renal rickets, polyuria, polydipsia, or macroscopic hematuria (nephrocalcinosis or renal stone), aminoaciduria, glucosuria, proteinuria, hyperphosphaturia, hypercalcemia and defective urinary acidification [46]. Sözeri et al[42] in their study of 10 patients of Wilson's disease, for whom excretion of tubular markers could be $repeated \ between \ 3 \ mo \ to \ 12 \ years, \ N-acetyl-\beta-d-glucosaminidase \ (NAG), \ beta \ 2 \ microglobulin \ and \ low \ molecular \ weight$ proteins were higher in the first year, as compared to the subsequent period. This is in contrast to high molecular weight proteinuria, which increases after 1 year of treatment with D-penicillamine. Renal impairment may be mild to severe. It may present as sub-nephrotic or nephrotic range proteinuria, microscopic hematuria, hypercalciuria, renal stones, renal failure, and lastly D-penicillamine-associated glomerulonephritis[47].

Tubular damage/ RTA

The disease per se can cause glomerular as well as tubular dysfunction, however, tubulopathy is more common. Excess copper deposits in the epithelium of the proximal and distal convoluted tubules, and the thickening of the basement membrane interfere with reabsorptive function of the renal tubules, thereby leading to RTA[47]. Distal, as well as proximal RTA, has been reported in WD. In a cross-sectional study by Kapoor et al[48], done over 1 year duration, 14 out of 25 patients (56%) of WD, had renal tubular acidosis of which 24% (6/25) had distal RTA, 16% (4/25) had mixed RTA, and another 16% (4/25) had proximal RTA. Wolff et al [49] described post-mortem kidney biopsies of 5 patients, wherein, on histopathology, glomeruli were normal in all 5 patients, focal areas of degeneration and necrosis of tubular epithelial cells along with copper staining (rubeanic-acid staining) were seen in all patients. Elsas et al [50] showed progressive renal impairment in an adolescent with Wilson's disease for whom D-penicillamine was stopped for almost 18 mo (owing to Dpenicillamine induced lupus nephritis), with simultaneous renal biopsy showing an increased number of conspicuous electron-dense bodies in the subapical areas of cytoplasm suggestive of metalloprotein complexes. Azizi et al[51]

Table 3 Differentiating features between Wilson disease-related renal tubular acidosis and D-penicillamine induced glomerulonephritis

	Wilson disease-related renal tubular acidosis	D-penicillamine induced glomerulonephritis
Mechanism	Copper induced tubular damage	Immune complex deposition
Presentation	Prior to starting chelation/during chelation	After starting chelation
Tests to differentiate	Normal anion gap metabolic acidosis, Urine pH, Urine for glucosuria, aminoaciduria, acidification test of urine	Urine for proteinuria, autoantibodies for glomer- ulonephritis, renal biopsy
Chelation	To be continued	To be stopped

described a 9-year-old boy, who first presented with renal colic due to hypercalciuria, only to be diagnosed as WD 1 year later. There are few case reports of children presenting with renal rickets who were finally diagnosed as WD[52]. In a retrospective study of 85 children with WD by Zhuang et al [47], renal impairment was found in 25 (29.4%) of the treatment-naïve patients. Seven of the twenty-five WD patients (28%) had symptoms of renal impairment (one each had acute nephritis, persistent glomerulonephritis, hemolytic uremic syndrome, and 2 had nephrotic syndrome). Five children had evidence of the glomerular cause of hematuria while the remaining two had non-glomerular hematuria. Twelve of the 25 (48%) had proteinuria, 14/25 (56%) had hematuria, and 5/25 (20%) had both proteinuria and hematuria, 4/25 (16%) had glucosuria[47]. Thus, by characterizing the type of hematuria (glomerular or non-glomerular) and proteinuria (low molecular weight or high molecular weight), the site of renal involvement can be ascertained. In the treatment -naïve patients, albuminuria with elevated serum creatinine, and low creatinine clearance suggest glomerular involvement. On the other hand, glycosuria, LMW proteinuria, non-glomerular hematuria, an increase in NAG and beta2 microglobulin, nephrocalcinosis, hypercalciuria, and non-anion gap metabolic acidosis indicate tubular involvement.

Glomerular involvement can be due to (1) copper deposition in the mesangium, leading to membranoproliferative glomerulonephritis, (2) IgA nephropathy caused by IgA deposits in the glomerulus, due to the loss of scavenging capacity of the liver, and (3) chelation (D-penicillamine) induced. IgA nephropathy: Gündüz et al[53] have reported a 13year-old boy who presented with nephritic syndrome after 4 mo of diagnosis of Wilson disease, and was diagnosed as membranoproliferative glomerulonephritis on renal biopsy with positive immunofluorescence for IgA, suggestive of IgA nephropathy. Similar case of IgA vasculitis is described by Acharya et al[54] in an 11-year-old boy with Wilson disease with F2 fibrosis on liver biopsy (METAVIR staging), who presented with palpable purpura without any arthritis or gastrointestinal involvement. He was diagnosed with IgA nephropathy and mild tubular epithelial degeneration and atrophy on renal biopsy and had complete resolution of the rash and achievement of normal renal function within 3 to 6 mo of chelation. This is similar to primary IgA nephropathy, abnormally glycosylated IgA1 form large soluble IgA1 immune complexes by combining with IgG and IgA, which deposit in the mesangium and lead to mesangial injury. Clinical and histopathological changes may reverse after liver transplantation.

Treatment-related nephrotoxicity

D-penicillamine-induced nephrotic syndrome was first reported by Fellers and Shahidi in 1959[55]. D-penicillamine can cause inhibition of enzymes required for collagen synthesis, thus can damage the glomerular basement membrane and reduce GFR or it can act as a hapten and induce the formation of immune complexes leading to membranous glomerulonephritis[56]. D-penicillamine-induced proteinuria occurs in < 10% of patients of Wilson disease and usually begins after 1 year of treatment [57,58]. The spectrum of D-penicillamine-induced nephrotoxicity ranges from membranous glomerulonephritis, tubule-interstitial disease, crescentic glomerulonephritis, Goodpasture's syndrome and renal-limited vasculitis[59,60].

Proteinuria is the most common manifestation and is the first abnormality to be detected followed by progressive renal disease if D-penicillamine is not stopped[60]. In a case series reported by Sternlieb, all the patients were started on racemic penicillamine, the clinical features resembled nephrotic syndrome, 3 of whom improved following discontinuation of drugs, and another 5 required steroids and there was a recurrence of nephrotic syndrome in all 3 patients in whom penicillamine was re-started while 2 of them didn't have any recurrence after they were started on D isomer of penicillamine[61]. Initially, the racemic mixture of penicillamine was incriminated in the causation of nephrotic syndrome, however, later on, it was shown that even the D- isomer can cause proteinuria. Since 1960, the D-isomer of penicillamine has been available and was approved by the FDA in 1963[61]. Histological examination shows membranous nephropathy (most commonly) minimal change disease or membranoproliferative or rarely crescentic nephropathy[62-64]. D-penicillamine causes nephrotoxicity by immunological mechanism, ultrastructural changes include immune-complex deposits, subepithelial deposits and IgG deposits on immunofluorescence.

The incidence of nephrotoxicity due to D-penicillamine was more in conditions unrelated to copper metabolism, e.g. rheumatoid arthritis, cystinurias, and scleroderma[65,66]. In a case series of 33 patients, studied by Hall et al[59], who were followed up serially for a mean duration of 74 mo, it was found that the onset of proteinuria peaked after the first 6 mo, with 27/33 developing proteinuria within 1 year of starting D-penicillamine. There are reports to show that the time period from exposure of D-penicillamine to proteinuria may range from a few weeks to years[60]. Siafakas et al[67] report a 12-year-old boy who presented with nephrotic syndrome (nephrotic range proteinuria without hematuria, with renal biopsy showing minimal change disease and a negative immunofluorescence study) after 2 wk of D-penicillamine and showed resolution of proteinuria in 3 wk, after being started on steroids. There are studies to show that proteinuria increases even after stoppage of D-penicillamine (within 1-5 mo), and then gradually improves over a few months, which can be explained by Wilson disease-related tubulopathy, as it improves over time with chelation [42,60,63]. In the study by Hall et al [59] the median first and last creatinine clearance almost remained the same, and none went into renal failure or required corticosteroids. The spontaneous resolution was seen within 2-32 mo of the stoppage of the drug [63]. According to Hall et al[59], among 33 patients of patients on D-penicillamine who had proteinuria, the number of patients in whom proteinuria resolved by 6, 12 and 18 mo were 12 (36%), 21(63%), 29 (88%) respectively. Renal biopsies done in all patients, 29/33 showed ultrastructural changes characteristic of membranous glomerulonephritis with IgG deposits and complement deposits on immunofluorescence, the remaining 4 patients' biopsies showed mesangial proliferation without any deposition of immunoglobulins or complement on immunofluorescence [59]. The severity of proteinuria is not related to the duration or dose of d-penicillamine or HLA typing[59]. In a retrospective study of 63 patients who developed nephrotic syndrome after being started on D-penicillamine, (75% had rheumatoid arthritis, 10% had Wilson disease), the mean duration of D-pen exposure to proteinuria was 7.6 (± 3.90) months and mean duration of drug exposure until the diagnosis of nephrotic syndrome was 11.9 (± 18.8) mo. Fifty-five percent of them had membranous glomerulonephritis and 27% had minimal change disease [63].

In a study of 8 patients of rheumatoid arthritis, who developed proteinuria while on D-penicillamine of variable duration, ranging from 3 to 48 mo and dosage ranging from 250 mg/d to 1.2 gm per day, 5 patients had scanty subepithelial deposits (spikes), 1 had plenty of sub-epithelial deposits, 2 had associated findings of glomerulosclerosis, interstitial scarring and, on immunofluorescence, 3 showed a positive granular pattern of fluorescence along basement membrane for IgG and C3[68]. After stopping D-penicillamine, proteinuria resolved in 3, progressive proteinuria was seen in one patient, mild proteinuria persisted in 4 others, with the persistence of proteinuria being seen up to 27 mo after stoppage of D-penicillamine [68]. Some studies have been done to see whether D-penicillamine can be resumed after the resolution of proteinuria [69]. In one of the studies in rheumatoid arthritis patients, D-penicillamine was re-introduced after 3 mo of resolution of proteinuria at a low dose of 50 mg/d, escalated to a maximum of 250 mg/d, showed that none of them had a relapse of proteinuria. However, this low dose is not enough to treat WD. In another study by Bacon et al [64] in 14 patients of rheumatoid arthritis, 3 developed proteinuria, and 11 developed nephrotic syndrome in a mean duration of 7.5 mo of starting D-penicillamine. Renal biopsies showed a picture similar to membranous nephropathy on light microscopy, granular deposits on immunofluorescence, positive for IgG and C3 and sub-epithelial dense deposits on electron microscopy, even after 3-12 mo of stoppage of treatment. A renal biopsy done soon after stopping D-penicillamine showed effacement of podocytes with discrete electron-dense deposits in the subepithelial region and positive fine granular immunofluorescence with IgG and C3. When the biopsy was repeated after 6 mo of discontinuation, it showed normal foot processes with occasional dense deposits and focal weak immunofluorescence with IgG and C3. Among 3 patients in whom proteinuria persisted at a high level even after stoppage of the drug, biopsy done at 6, 8, 12 mo of discontinuing D-penicillamine, electron microscopy still showed discrete but much smaller electron-dense deposits, and brilliant fluorescence with IgG[64]. D-penicillamine-induced glomerulonephritis and Goodpasture syndrome have also been described[70].

To summarize, diagnosis of D-penicillamine-induced nephropathy can be made with routine urine microscopy, 24-h urine estimation for proteinuria, and type of proteinuria (tubular or glomerular), and if feasible, histopathological and electron microscopic examination (renal biopsy) especially in the first 18 mo. Membranous nephropathy is the most common nephropathy caused by D-penicillamine [59,63,68]. Serum MPLA2R antibodies (an immunofluorescence test) can be done as a supportive test. These antibodies are prevalent in primary membranous glomerulonephritis in up to 68.5% of cases and their absence on the face of renal biopsy picture of membranous nephropathy suggests the secondary cause of membranous glomerulonephritis[71]. Kumar et al[72] reported nephrotic syndrome in a 24-year-old lady, a case WD on D-penicillamine for 18 mo. Renal biopsy showed membranous nephropathy with positive immunofluorescence for IgG and C3, and with negative serum auto-antibodies to M-type phospholipase A2 receptor (MPLA₂R, which is usually seen in primary membranous nephropathy), thus supporting the diagnosis of secondary membranous glomerulonephritis.

Management of D-penicillamine nephrotoxicity includes stoppage of the drug and use of trientine as an alternative. In the study done by Neild et al[63], glucocorticoids had no effect on the natural history of nephropathy. In another case series by De Silva, among 35 patients who developed proteinuria while on D-penicillamine, 60% developed nephrotic syndrome (NS). Among NS patients, D-penicillamine was continued for 62%, stopped at a variable interval, and it was shown that within 4 mo of stopping D-penicillamine, proteinuria decreased to < 2 g/d and 12 mo after stopping Dpenicillamine, proteinuria was 0-0.3 g/d[73]. AASLD recommends prompt withdrawal of D-penicillamine immediately.

Haematological manifestations

These include Coomb's negative hemolytic anaemia and acute renal failure, D-penicillamine-induced myelosuppression and lastly hypersplenism (splenomegaly).

Acute fulminant WD

Acute fulminant Wilson disease usually presents with acute intravascular hemolysis, which is due to free coppermediated damage to the RBC membranes, mortality is 95% [74]. It needs to be tackled with plasmapheresis while awaiting liver transplantation[75].

Drug-induced cytopenia

The rate of myelotoxicity varies from D-penicillamine 0 to 7% and it is one of the most fatal adverse effects [76]. It has been hypothesized that marrow toxicity could be of two types: The first being an idiosyncratic reaction, leading to cytopenias occurring within the first year of treatment, and the other being a dose-dependent gradual fall[77]. Day et al[78] showed that of the 69 patients of rheumatoid arthritis, who were on D-penicillamine for more than 1 year, 15 (21.7%) had

developed dose-dependent hematological adverse effects; while those who were on < 500 mg per day, had no cytopenias. However in another case series of 10 patients of rheumatic arthritis with D-penicillamine induced myelosuppression, 7 had sudden onset myelosuppression, of these, 6 patients died, and the remaining showed gradual recovery of marrow over 1 year [79]. These studies are to be interpreted keeping in mind 2 points; first, both the studies were in rheumatoid arthritis patients, and myelosuppression was documented even with a low dose of D-penicillamine (in contrast to WD, where a higher dose of D-penicillamine is required), secondly, these patients didn't have concomitant hypersplenism (to confound the picture), thereby, cytopenia could easily be attributed to D-penicillamine, leading to faster decision making in terms of drug-discontinuation. Once myelosuppression is there, D-penicillamine is to be stopped. European Society of Pediatric Gastroenterology, Hepatology and Nutrition (ESPGHAN) has suggested monitoring blood counts initially for 1-3 monthly and later 3-6 monthly [80]. Table 4 enlists the risk factors that may complicate/mimic drug toxicity features.

Neurologic symptoms

Neurological symptoms include dystonia, dysarthria, gait abnormalities, and tremors, with dysarthria being the most common[8]. Other manifestations include abnormal gait, musculoskeletal symptoms, seizures, behavioral problems, drooling of saliva, and chorea [81]. Pediatric patients present with hepatic WD whereas neurological manifestations are more common among adults [82]. Neurological manifestations of WD are due to high concentrations of copper in brain tissue as well in CSF and anti-copper therapy leads to improvement in symptoms [83]. It is essential to characterize the disease extent in order to monitor the evolution of symptoms carefully. It is important to identify the subtle neurologic manifestations at the time of diagnosis as it has a bearing on choosing the medical therapy. Multiple studies have shown that neurological worsening on starting chelation is more commonly seen in those having neurological manifestations, within 1-3 mo of starting therapy. In one of the early studies, Brewer et al [84] showed that neurological worsening post-Dpenicillamine occurred in 50% of neurologic WD, with more than half of the patients showing deterioration within 4 wk of chelation. Ranjan et al[85] performed an MRI brain at the time of neurological worsening after D-penicillamine and showed the appearance of new lesions in MRI in white matter, thalamus, pons, and midbrain which showed diffusion restriction with simultaneous blood investigations revealed increased free serum copper, malondialdehyde and reduced glutathione. Litwin et al [86] showed neurological worsening in 11% of neuro-hepatic WD in a mean time of 2.3 ± 1.9 mo from the time of treatment initiation independent of the type of chelation used. One hypothesis proposed is that there is a sudden release of free copper from the liver, which enters the blood-brain barrier and causes brain damage [84]. The second hypothesis is that the free copper is generated by the chelators within the brain tissue itself. This was substantiated by an increase in free copper concentration in serum and brain tissue, within 3 d of starting on D-penicillamine, measured by ultrafiltration in an animal model of WD. It was noted that at the time of the increase in brain tissue-free copper, there was a fall in protein-bound copper in the brain tissue. A simultaneous increase in immunofluorescent staining of ATP7A (copper transporting protein in neurons) and CTR1 (mediates copper uptake in mammalian tissue) in the cortex and basal ganglia, and not in the blood-brain barrier, suggests that the free copper is generated probably by mobilization of copper within the brain parenchymal cells and not via peripherally released free copper entry via the blood-brain barrier[87]. The expression of ATP7A present in the subcellular level to mediate biliary excretion of copper (in this animal model) correlated with the free copper in the cortex and basal ganglia. The third hypothesis proposed by Miki et al[88] is that the penicillamine-copper complexes that are generated are non-toxic, so, shouldn't cause tissue damage. However, these complexes can catalyze oxidation of the ghost membranes, due to changes in the redox potential of copper, Cu²⁺ to Cu¹⁺), thereby causing neurological symptoms.

TREATMENT

Once the diagnosis is established, the patient is to be started on lifelong pharmacotherapy, depending on the stage of the disease. WD patients can be broadly classified into three subtypes; symptomatic WD, asymptomatic WD with active disease (on biochemical, histological or imaging findings), and pre-symptomatic WD (mostly those detected on family screening). Those with symptomatic or asymptomatic active disease need to be started on chelation. Patients who are asymptomatic or without active disease can be treated with lower maintenance doses of chelators or zinc alone. It takes around 6-18 mo of consistent chelation therapy to improve the organ function and then patients can be shifted to a lower dose of chelator and zinc[8].

Table 5 summarizes all the drugs used in the treatment of WD, their mechanism of action, dose, storage and important side effects. There is a fundamental difference in the mechanism of action of chelators and zinc. Chelators decrease the copper load in the body by chelating the copper in the liver, enterocytes and extrahepatic circulation. Whereas zinc induces the synthesis of metallothioneins, which in turn bind to copper and sequester it in the enterocyte (copper is lost as the enterocytes shed off). Thus, the action of zinc is slow. In scenarios, which demand fast and immediate action, for example, advanced liver disease, chelators are a better choice than zinc.

D-penicillamine

D-penicillamine (D-3β,3β-dimethylcysteine; C5H11NO2S) has been the first line of treatment, since its discovery in 1956. It acts by chelating divalent metal ions using its thiol (-SH) group and forming a water-soluble complex which is excreted in urine. It chelates extracellular copper and mediates its excretion in urine. Also, it chelates the intracellular copper from tissue complexes[81]. In WD, the excess free copper in the cytoplasm of hepatocytes, after saturating the metallothioneins, deposits in the lysosomes and causes free radicle-mediated cell damage. D-penicillamine, trientine, as well as zinc, increase the expression of metallothioneins, which bind to copper ions. Metallothionein is a cysteine-rich protein that is

Table 4 Risk factors for side effects of drug therapy in Wilson's disease			
Side effect of drugs	Risk factor	How to differentiate	
Neurological worsening with D-penicillamine or trientine	Pre-existing neurological Wilson disease	Exchangeable copper and relative exchangeable copper	
Cytopenia due to D-penicillamine	Co-existing hypersplenism due to portal hypertension	Bone marrow biopsy	

Table 5 Drugs,	their mechanism o	f action, dosag	ge and side effects
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Name of drug	Mechanism of action	Dose	When to start	Side effects
D-penicil- lamine	Induces cuprieuresis, induces hepatic metallothionine synthesis, reduces fibrosis (by preventing collagen formation)	20 mg/kg/d (maximum induction dose of 1500 mg/d and maintenance dose of 1000 mg/d), to be taken 1 h before or 2 h after meal, storage at room temperature	Chelator of choice in all hepatic phenotype	Early (1-3 wk): Fever, rash, arthralgia, cytopenia, proteinuria; Late: (1) Skin: degenerative dermatoses elastosis perforans serpingosa, cutis laxa, pseudoxanthoma elasticum, bullous dermatoses, psoriasiform dermatoses, lichen planus, seborrheic dermatitis alopecia, aphthous ulcerations, hair loss; (2) Connective tissue disorders: Lupus like syndrome, arthralgia, Rheumatoid arthritis, polymyositis; (3) Renal: proteinuria, hematuria, glomerulonephritis, nephrotic syndrome, renal vasculitis, Goodpasture's syndrome; (4) Nervous system: Paradoxical neurological worsening, neuropathies, myasthenia, hearing abnormalities, serous retinitis; (5) Gastrointestinal: Nausea, vomiting, diarrhea, elevated transaminases, cholestasis, hepatic siderosis; (6) Respiratory: Pneumonitis, pulmonary fibrosis, pleural effusion; (7) Hematological: Cytopenia, agranulocytosis, aplastic anemia, hemolytic anemia; and (8) Others: Immunoglobulin deficiency, breast enlargement, pyridoxine deficiency
Trientine	Induces cuprieuresis, induces hepatic metallothionine synthesis	20 mg/kg/d (maximum induction dose of 1500 mg/d and maintenance dose of 1000 mg/d), to be taken 1 h before or 2 h after meal, storage at 2 ⁰ -8 ⁰ temperature	Indicated if intolerant to D-penicillamine	Paradoxical neurological worsening (10%-50%), sideroblastic anemia, bone marrow suppression, gastritis, skin rash, arthralgia, myalgia, hirsutism
Zinc	Induces intestinal synthesis of metallo- thioneins, prevents copper absorption	25 mg thrice daily (weight < 50 kg), 50 mg thrice daily (weight > 50 kg), taken in empty stomach	Maintenance phase of symptomatic hepatic WD; First-line induction treatment in selected patient subgroups (neurologic WD, intolerant to chelators, presymptomatic patients)	Gastric irritation (30%-40%)
Ammonium Tetra-thiomo- lybdate	Forms complexes with copper in blood, binds the copper present in food		Neurological WD	Neurological dysfunction (rare), hepatotoxicity, bone marrow suppression

WD: Wilson disease.

an endogenous chelator of copper[5,81]. D-penicillamine also solubilizes the copper deposited in the lysosomes, without much affecting the metallothionein-bound copper and reduces cell damage[89]. Thus, D-penicillamine causes the excretion of copper but can also lead to the sequestration of free intracellular copper with metallothioneins[5]. D-penicillamine also has an anti-inflammatory effect, which is why it is a second-line drug in rheumatic diseases[90]. The standard dose of D-penicillamine is 20 mg/kg/d in 2-3 divided doses, it is usually started at a dose of 1000-1500 mg/d, and after liver functions improve it can be reduced to a maintenance dose of 10-15 mg/kg/d or 750-1000 mg/d[8]. Around 40%-70% of D-penicillamine is absorbed in the proximal intestine, and peak plasma concentration is reached within 1-3 h. After long-standing treatment, it is eliminated within 4-6 d (slow pool of drug bound reversibly to the tissue). Eighty percent of the drug circulates in bound form with D-penicillamine and the rest in free form. The drug is eliminated via the kidneys[91]. In neurohepatic WD, the dictum is to start at a low dose and increase slowly. Indian National Association for the Study of the Liver (INASL) recommends starting D-penicillamine at 250 mg on alternate days and increasing by 250 mg every 2-3 wk, until the maximum dose of 1000-1500 mg per day, while keeping a watch on neurological symptoms [6]. As D-penicillamine causes depletion of pyridoxine by direct effect, pyridoxine needs to be supplemented at a dose of 25-40 mg per day. D-penicillamine should be given either 1 hour before or 2 h after meals, as it may get bound to the copper in the diet before its absorption into the intestine [92]. Antacids and iron significantly decrease its absorption [8]. Brewer et al [93] showed that fecal excretion of copper is significantly less with penicillamine and zinc combination than with zinc alone.

Adverse effects of D-penicillamine are acute (early onset) and late-onset

Acute adverse effects include hypersensitivity reaction, in the form of fever, rash, urticaria, arthralgia, proteinuria and leukopenia, usually within the initial 3 wk of starting D-penicillamine, seen in up to 15% of patients [6,94]. Delayed hypersensitivity include pemphigus, lupus-like syndrome[6]. Other delayed adverse effects include various organ systems, such as hematological (both idiosyncratic and dose-dependent marrow suppression), renal (membranous glomerulonephritis, and neurological (worsening of neurological symptoms). Renal and hematological adverse effects have already been discussed in the article earlier. In case of early hypersensitivity reactions, the drug is to be stopped immediately. In case of isolated dermatological involvement, it may be re-started under steroid cover[6]. Other dermatological adverse effects include dermatopathies due to elastic fibre abnormalities e.g. elastosis perforans serpiginosa, and pseudo-pseudoxanthoma elasticum. About 15%-30% of patients develop these adverse effects due to high doses and a long duration of drug intake. These skin changes are caused due to inhibition of the aldol crosslinking of tropocollagen and take months to years to manifest (as it takes a long time for new weakened collagen to be synthesized)[95]. Penicillamine-induced autoimmune dermatoses include pemphigus, epidermolysis bullosa and lupus-erythematosus-like syndrome. Immune-mediated toxicities like Goodpasture syndrome, systemic lupus erythematosus, and nephrotic syndrome warrant immediate drug withdrawal [8,58] (Table 5).

In some cases, paradoxical neurological worsening is seen after starting d-penicillamine, likely due to peripheral mobilization of copper from the liver to the bloodstream and its subsequent deposition in CNS, seen in 11%-50% [84,86,96, 97]. Large-scale comparative studies should be done to study the effect of chelators on neurological WD by correlating the dose of chelators with MRI changes and copper transporting molecules by in vivo studies. Reversal of neurologic deterioration was seen in only 50% of those affected as D-penicillamine was being continued[84]. In another retrospective study, partial/complete reversal was seen in 53% (8/15) and a further partial response in 13% (2/15) of cases during 9.2 ± 5.2 mo[86]. Thus, the solution to this problem is to start at a low dose and increase it gradually.

Trientine

It is also known as triethylenetetramine. It is an alternative to D-penicillamine for chelation, particularly for those who do not tolerate D-penicillamine. One molecule of trientine combines with copper in a 1:1 ratio to form a stable complex, which is excreted in the urine. Trientine dihydrochloride is the oral preparation, which requires cold storage at 20-80 Celsius to maintain its stability, failure to maintain cold chain is a common cause of drug discontinuation [98]. Since 2018, trientine is also available as trientine tetrahydrochloride. Woimant et al[99] showed that there was no difference in efficacy of the two drugs and in terms of adverse effects, there was a case of recurrence of lupus erythematosus-like syndrome with trientine dihydrochloride. Trientine tetrahydrochloride is stable at room temperature, has slightly more rapid absorption and higher bioavailability and greater systemic exposure. Also, it is a cheaper alternative. It is poorly absorbed with a bioavailability of 8-30%. It reaches maximum plasma concentration in 1-3 h[100,101]. Plasma concentration of trientine is significantly reduced when given after food[95]. Trientine has fewer side effects. Pancytopenia is rarely caused. It should not be given with oral iron because trientine-iron complexes are toxic. There are no hypersensitivity reactions reported. Other minor adverse effects include hemorrhagic gastritis, loss of taste and nausea, sideroblastic anemia and allergic rash[6,102,103]. In a multicenter retrospective study of 77 patients with WD, where patients were treated with trientine for an average duration of 8 years (range: 5 mo to 32.5 years), 49.4% had improved hepatic functions, 10.4% of patients remained unchanged, 5.2% showed worsening, and remaining were asymptomatic to begin with. Twenty-two percent of patients had trientine-associated minor adverse effects, with only one patient requiring treatment discontinuation due to anaemia[104]. Neurologic deterioration is reported with trientine, hence, it should be started at a low dose and increased gradually in patients having neurologic manifestations[8]. In a large retrospective cohort of 471 patients (326 receiving D-penicillamine vs. 141 receiving trientine), it was shown that hepatic and neurologic improvement was comparable in either group. Stable neurologic disease as first-line drug was comparable in either group (27.2% for D-penicillamine vs 20% for trientine). A higher rate of neurologic worsening was reported with trientine as the first-line agent (20% vs 5.3%)[105]. In another randomized control trial among neurologic WD patients, for comparison between tetrathiomolybdate, 6/23 (26%) in trientine had neurologic worsening as compared to 1/23 in the tetrathiomolybdate group[106]. The dose is 750-1000 mg per day or 20 mg/kg/d in 3 divided doses. It is to be given 1 h before or 2 h after food[100].

Zinc

It induces synthesis of metallothioneins, and thereby promotes copper-binding to metallothionein in the enterocyte and ultimately hinders its absorption (as copper is lost when the enterocyte is shed). It is a slow de-coppering agent, it decreases the copper absorption but doesn't lead to a sudden massive increase in free copper, hence, it is of choice in neurological WD. However, because of the slow reduction in copper, it is not suitable for treating florid symptoms[8]. Disease may show progression in the initial few months, because of its slow onset of action [106]. In the first pediatric study, in which trientine was used as the initial chelator, it was shown that once adequate chelation was achieved, zinc combination therapy and subsequently zinc monotherapy maintained normal ALT/AST levels[102]. Among those having weight < 50 kg, it is to be given in a dose of 25 mg thrice a day and in those weighing > 50 kg, 50 mg thrice a day[6].

Various zinc preparations are available; zinc acetate, zinc gluconate, and zinc sulfate. There is no statistically significant difference in various zinc preparations in terms of improvement of liver function [107]. There are few adverse effects, gastric irritation is the most common side effect, in 30%-40% of patients. It can also cause an asymptomatic elevation in amylase and lipase[8]. It is recommended to use zinc in pre-symptomatic patients or in the maintenance phase of treatment in symptomatic patients[8]. In a systematic review and meta-analysis, zinc showed better improvement as compared to D-penicillamine in neurologic WD, however, there was no difference in hepatic WD. Also, the incidence of adverse effects and neurologic deterioration was higher with D-penicillamine as compared to zinc (RR: 2.42, 95%CI: 1.20% - 4.88%; P = 0.014) and RR: 1.96, 95% CI: 1.31% - 2.93%; P = 0.001, respectively)[108].

Ammonium tetra-thiomolybdate

It acts by forming a tripartite complex with copper and protein, which is stable. It binds the copper present in food and prevents its absorption. When given without food, it is absorbed into the blood and forms complexes with copper bound to albumin, thus preventing its deposition in various organs[109]. Also, it enters the blood-brain barrier and enters neuronal cells[110]. In an open-label study, 55 neurologic WD were treated with tetrathiomolybdate (120-410 mg for 8 wk), followed by zinc maintenance therapy. Only 3.6% (2/55) showed neurologic deterioration. Among the other adverse effects, 5/22 (23%) of treatment-naïve patients had bone marrow suppression and 3/22 (14%) had an elevation in liver enzymes and both these adverse effects quickly responded to drug dose reduction[111]. Bone marrow suppression is caused by the depletion of copper and is reversed by decreasing the dose of tetrathiomolybdate. The rise in the liver enzymes could be due to the mobilization of hepatic copper (from hepatic pools including metallothioneins) in a heavily copper-loaded liver, which is reversible on dose reduction[111]. In a subsequent double-blind RCT between tetrathiomolybdate (120 mg per day) and trientine (1000 mg/d) among 48 patients of neurologic WD, as mentioned above, tetrathiomolybdate was better than trientine, only 4% worsened neurologically vs 26% in trientine arm[106]. Ammonium tetrathiomolybdate is unstable for routine use, bis-choline tetrathiomolybdate is a more stable complex and has a better availability. Recently, bis-choline tetrathiomolybdate underwent a phase II trial among 28 neurologic WD, where it was given for a span of 24 wk, with the target to achieve a primary end-point of normal value of non-ceruloplasmin bound copper_{corrected} (NCC_{corrected} stands for NCC corrected for copper contained in tetrathiomolybdate-copper-albumin $complexes) \ or \ achievement \ of \ 25\% \ reduction \ from \ baseline \ NCC_{corrected}. \ Twenty-two \ patients \ completed \ the \ study \ up \ to \ 24$ wk and by week 24, 20 (71%) had achieved the primary end-point (treatment success) accompanied by improvement in neurologic status (without any paradoxical neurological worsening, being reported). There were 11 (25%) serious adverse effects which included 6 events of psychiatric disorders in 4 patients, gait disturbance in one patient and two events of raised aminotransferases, one agranulocytosis and a decline in neurologic functioning [112]. Also, Brewer et al [113] analysed free copper levels in patients treated with tetrathiomolybdate vs trientine and found that the mean free copper was significantly less in the tetrathiomolybdate group at week 4 and week 8 of treatment. Thus, tetrathiomolybdate is a fast copper-lowering agent with minimal adverse effects. The dose recommended is initially 120 mg per day for the first 2 wk, as, 20 mg thrice a day with meals and 20 mg daily between meals followed by 60 mg daily as 10 mg thrice a day with meals and 10 mg thrice daily between meals[113]. Currently, a phase 3 trial is going on and results are keenly awaited (NCT03403205).

COMPARISON BETWEEN VARIOUS CHELATORS

D-penicillamine vs trientine

In a large retrospective cohort of median follow-up of 13.3 years, both D-penicillamine and trientine were comparable in terms of improvement in hepatic WD (> 90% cases) as well as in neurologic WD (> 55% cases) while neurologic worsening was more common in trientine group as compared to D-penicillamine group (20% vs 5.3%, P = 0.042). Treatment discontinuation due to adverse effects was more common in the D-penicillamine group as compared to trientine (28.8% vs 7.1%, P = 0.039) [105]. In the most recent randomized open-label non-inferiority trial of D-penicillamine vs trientine, for maintenance therapy after 1 year of chelation with D-penicillamine, trientine was found non-inferior to Dpenicillamine[114].

D-penicillamine vs zinc

There are studies to show that zinc was found to be more effective in ameliorating neurological symptoms in 90% of patients in the zinc arm as compared to 25% alone in the D-penicillamine arm[115]. In a head-to-head comparison between D-penicillamine and zinc, among 67 new patients, of whom the majority were asymptomatic or had neurological disease, 44% discontinued D-penicillamine owing to its adverse effects as compared to 12% in the zinc arm[116]. In Another retrospective study of 288 patients for a period of 17.1 years by Weiss et al[117] showed that zinc monotherapy led to 15.9% (14/88) hepatic treatment failure as compared to 1.2% (4/313) in the D-penicillamine group without any statistically significant difference in the adverse effects in either group. Further, these zinc non-responders responded to chelators (either D-penicillamine or trientine). This supports the use of chelators in asymptomatic patients with active

Dhawan et al[118], Askari et al[119] and Santos et al[120] have shown favourable outcomes with combination therapy but earlier studies by Brewer et al [93] show no added advantage. A systematic review of 17 articles involving 1056 on combination therapy showed that combination therapies are significantly less effective than individual therapies (47.1 vs 78.6%)[121].

Adequacy of treatment

Treatment targets on chelators: Asymptomatic patients should remain asymptomatic and symptomatic patients should show improvement in liver functions in the initial 6-18 mo[8]. Patients with decompensation might take longer to improve. Patients are to be monitored clinically for symptom improvement/new symptom onset. Initially, LFTs are to be done, every 3 mo, and thereafter 3-6 mo depending on the disease severity [8]. Adequacy is monitored by monitoring liver function tests and by quantifying 24-h urine copper excretion or free copper estimation. Free copper estimation, is not a full-proof test if the ceruloplasmin is calculated by the immunologic method as it calculates both apoceruloplasmin and holoceruloplsmin[6]. Also, urinary copper excretion is to be interpreted carefully after taking proper treatment history. Urinary copper excretion may be high if chelation has been re-started after a period of non-adherence or it can be falsely low, in case of poor drug absorption or inadequate dosing itself. Another way to monitor adequacy is to measure REC.

For de-coppering agents, free copper is a marker of the adequacy of chelation, while for zinc, urine copper levels below a certain cut-off is recommended for good copper control (Table 1).

When to decrease chelation?

Transition to maintenance dose or zinc can be used once liver function improves. AASLD recommends a transition to maintenance after clinical and biochemical parameters improve (usually seen after 1 year of therapy). The drug of choice drug could be low-dose chelating agents or full-dose zinc. In a retrospective study of 31 symptomatic hepatic WD who were transitioned from D-penicillamine to zinc (28 due to financial constraints and 3 due to adverse effects), wherein the majority of patients belonged to Child's class C (54%) patients, the average duration of zinc therapy on follow-up was 363 (35-728) weeks. In the Child C cirrhosis group at presentation, who received D-penicillamine for 111 (2-230) weeks followed by zinc for 344 (41-652) weeks, 15 had significant improvement in liver function and disease severity scores [122]. In a prospective study of 44 hepatic WD patients, who received D-penicillamine plus zinc combination therapy for more than 2 years, and were in biochemical remission (defined by AST and ALT > 1.5 times upper limit of normal, serum albumin > 3.5 gm/dL and INR < 500 mcg/d and NCC < 15 mcg/dL, were shifted to zinc monotherapy and biochemical parameters were assessed on follow-up. They showed that 9/44 (20.4%) relapsed till the last follow-up [123]. More prospective studies are needed to establish the correct time of transition and establish guidelines for the same.

CONCLUSION

Wilson's disease is a treatable metabolic liver disease. Early diagnosis is imperative. Wilson's disease has varied manifestations. Certain disease manifestations need to be differentiated from drug-toxicities, e.g., tubulopathies due to WD vs. Dpenicillamine-induced nephrotoxicity and hypersplenism vs D-penicillamine-induced myelosuppression. Though there are difficulties in making the correct diagnosis, with the help of non-ceruloplasmin bound copper, relative exchangeable copper and newer methods to detect early Kayser-Fleischer ring (AS-OCT), diagnosis can be made in resource-limited conditions, where mutational analysis is cost-forbidding. Chelation remains the mainstay of treatment and is to be preferred in active disease whether symptomatic or asymptomatic. Regarding the transition to maintenance therapy, the exact timeline is not yet defined, but depends on the liver function and is to be decided on a case-to-case basis with close follow-up of the copper load in the body. Trientine has been shown to have a good clinical response in a recent clinical trial and its availability as tetrahydrochloride, which is cheaper and doesn't require cold storage gives hope in resourcelimited conditions. Bis-choline tetrathiomolybdate is the new addition in the armamentarium, which rapidly decreases the free copper load and is undergoing phase III trials. In fulminant hepatic failure, plasmapheresis has shown (while awaiting liver transplantation) some hope.

FOOTNOTES

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REFERENCES

- Sandahl TD, Laursen TL, Munk DE, Vilstrup H, Weiss KH, Ott P. The Prevalence of Wilson's Disease: An Update. Hepatology 2020; 71: 722-732 [PMID: 31449670 DOI: 10.1002/hep.30911]
- Yachha SK, Sharma BC, Khanduri A, Srivastava A. Current spectrum of hepatobiliary disorders in northern India. Indian Pediatr 1997; 34: 885-890 [PMID: 9567550]
- 3 Roberts EA, Schilsky ML; Division of Gastroenterology and Nutrition, Hospital for Sick Children, Toronto, Ontario, Canada. A practice guideline on Wilson disease. Hepatology 2003; 37: 1475-1492 [PMID: 12774027 DOI: 10.1053/jhep.2003.50252]
- Stremmel W, Weiskirchen R. Therapeutic strategies in Wilson disease: pathophysiology and mode of action. Ann Transl Med 2021; 9: 732 [PMID: 33987430 DOI: 10.21037/atm-20-3090]
- Ala A, Walker AP, Ashkan K, Dooley JS, Schilsky ML. Wilson's disease. Lancet 2007; 369: 397-408 [PMID: 17276780 DOI: 5 10.1016/S0140-6736(07)60196-2]
- Nagral A, Sarma MS, Matthai J, Kukkle PL, Devarbhavi H, Sinha S, Alam S, Bavdekar A, Dhiman RK, Eapen CE, Goyal V, Mohan N, 6 Kandadai RM, Sathiyasekaran M, Poddar U, Sibal A, Sankaranarayanan S, Srivastava A, Thapa BR, Wadia PM, Yachha SK, Dhawan A. Wilson's Disease: Clinical Practice Guidelines of the Indian National Association for Study of the Liver, the Indian Society of Pediatric Gastroenterology, Hepatology and Nutrition, and the Movement Disorders Society of India. J Clin Exp Hepatol 2019; 9: 74-98 [PMID: 30765941 DOI: 10.1016/j.jceh.2018.08.009]
- Wu F, Wang J, Pu C, Qiao L, Jiang C. Wilson's disease: a comprehensive review of the molecular mechanisms. Int J Mol Sci 2015; 16: 6419-6431 [PMID: 25803104 DOI: 10.3390/ijms16036419]
- 8 Schilsky ML, Roberts EA, Bronstein JM, Dhawan A, Hamilton JP, Rivard AM, Washington MK, Weiss KH, Zimbrean PC. A multidisciplinary approach to the diagnosis and management of Wilson disease: Executive summary of the 2022 Practice Guidance on Wilson disease from the American Association for the Study of Liver Diseases. Hepatology 2023; 77: 1428-1455 [PMID: 36152019 DOI: 10.1002/hep.32805]
- Boga S, Ala A, Schilsky ML. Hepatic features of Wilson disease. Handb Clin Neurol 2017; 142: 91-99 [PMID: 28433114 DOI: 10.1016/B978-0-444-63625-6.00009-4]
- European Association for Study of Liver. EASL Clinical Practice Guidelines: Wilson's disease. J Hepatol 2012; 56: 671-685 [PMID: 10 22340672 DOI: 10.1016/j.jhep.2011.11.007]
- 11 Walshe JM; Clinical Investigations Standing Committee of the Association of Clinical Biochemists. Wilson's disease: the importance of measuring serum caeruloplasmin non-immunologically. Ann Clin Biochem 2003; 40: 115-121 [PMID: 12662398 DOI: 10.1258/0004563037630460211
- Merle U, Eisenbach C, Weiss KH, Tuma S, Stremmel W. Serum ceruloplasmin oxidase activity is a sensitive and highly specific diagnostic 12 marker for Wilson's disease. J Hepatol 2009; 51: 925-930 [PMID: 19720421 DOI: 10.1016/j.jhep.2009.06.022]
- Mak CM, Lam CW, Tam S. Diagnostic accuracy of serum ceruloplasmin in Wilson disease: determination of sensitivity and specificity by 13 ROC curve analysis among ATP7B-genotyped subjects. Clin Chem 2008; 54: 1356-1362 [PMID: 18556333 DOI: 10.1373/clinchem.2008.1034321
- Lu X, Li S, Zhang W, Lin Y, Lu Z, Cai Y, Su X, Shao Y, Liu Z, Sheng H, Huang Y, Liu L, Zeng C. Assessment of the diagnostic value of 14 serum ceruloplasmin for Wilson's disease in children. BMC Gastroenterol 2022; 22: 124 [PMID: 35296237 DOI: 10.1186/s12876-022-02186-0]
- Ferenci P, Członkowska A, Merle U, Ferenc S, Gromadzka G, Yurdaydin C, Vogel W, Bruha R, Schmidt HT, Stremmel W. Late-onset 15 Wilson's disease. Gastroenterology 2007; 132: 1294-1298 [PMID: 17433323 DOI: 10.1053/j.gastro.2007.02.057]
- Lackner C, Denk H. Histopathology of Wilson disease. In: Kerkar N, Roberts E, editors. Clinical and translational perspectives on Wilson 16 disease. New York: Academic Press; 2018; 257-270 [DOI: 10.1016/B978-0-12-810532-0.00023-9]
- Jain S, Scheuer PJ, Archer B, Newman SP, Sherlock S. Histological demonstration of copper and copper-associated protein in chronic liver 17 diseases. J Clin Pathol 1978; 31: 784-790 [PMID: 80410 DOI: 10.1136/jcp.31.8.784]
- Pilloni L, Lecca S, Van Eyken P, Flore C, Demelia L, Pilleri G, Nurchi AM, Farci AM, Ambu R, Callea F, Faa G. Value of histochemical 18 stains for copper in the diagnosis of Wilson's disease. Histopathology 1998; 33: 28-33 [PMID: 9726045 DOI: 10.1046/j.1365-2559.1998.00455.x]
- Goldfischer S, Sternlieb I. Changes in the distribution of hepatic copper in relation to the progression of Wilson's disease (hepatolenticular 19 degeneration). Am J Pathol 1968; 53: 883-901 [PMID: 4177374]
- Mounajjed T, Oxentenko AS, Qureshi H, Smyrk TC. Revisiting the topic of histochemically detectable copper in various liver diseases with 20 special focus on venous outflow impairment. Am J Clin Pathol 2013; 139: 79-86 [PMID: 23270902 DOI: 10.1309/AJCPDZR4OHDQNG3L]
- Woimant F, Djebrani-Oussedik N, Poujois A. New tools for Wilson's disease diagnosis: exchangeable copper fraction. Ann Transl Med 2019; 21 7: S70 [PMID: 31179307 DOI: 10.21037/atm.2019.03.02]
- McMillin GA, Travis JJ, Hunt JW. Direct measurement of free copper in serum or plasma ultrafiltrate. Am J Clin Pathol 2009; 131: 160-165 22 [PMID: 19141375 DOI: 10.1309/AJCP7Z9KBFINVGYF]
- Rodriguez-Castro KI, Hevia-Urrutia FJ, Sturniolo GC. Wilson's disease: A review of what we have learned. World J Hepatol 2015; 7: 2859-23 2870 [PMID: 26692151 DOI: 10.4254/wjh.v7.i29.2859]
- Poujois A, Poupon J, Woimant F. Direct determination of non-ceruloplasmin-bound copper in plasma. In: Kerkar N, Eve A. Clinical and 24 translational perspectives on Wilson disease. Elsevier, 2019; 249-255
- Poujois A, Trocello JM, Djebrani-Oussedik N, Poupon J, Collet C, Girardot-Tinant N, Sobesky R, Habès D, Debray D, Vanlemmens C, 25 Fluchère F, Ory-Magne F, Labreuche J, Preda C, Woimant F. Exchangeable copper: a reflection of the neurological severity in Wilson's disease. Eur J Neurol 2017; 24: 154-160 [PMID: 27739240 DOI: 10.1111/ene.13171]
- El Balkhi S, Trocello JM, Poupon J, Chappuis P, Massicot F, Girardot-Tinant N, Woimant F. Relative exchangeable copper: a new highly sensitive and highly specific biomarker for Wilson's disease diagnosis. Clin Chim Acta 2011; 412: 2254-2260 [PMID: 21878323 DOI: 10.1016/j.cca.2011.08.019]
- Trocello JM, El Balkhi S, Woimant F, Girardot-Tinant N, Chappuis P, Lloyd C, Poupon J. Relative exchangeable copper: a promising tool for 27 family screening in Wilson disease. Mov Disord 2014; 29: 558-562 [PMID: 24375554 DOI: 10.1002/mds.25763]
- Nagral A, Jhaveri A, Nalawade S, Momaya N, Chakkarwar V, Malde P. Kayser-Fleischer rings or bile pigment rings? Indian J Gastroenterol 2015; **34**: 410-412 [PMID: 26541339 DOI: 10.1007/s12664-015-0603-2]



- Sethi M, Madan S, Beri S. Fading Kayser-Fleischer ring revisited. Saudi J Ophthalmol 2021; 35: 146-148 [PMID: 35391817 DOI: 29 10.4103/1319-4534.337854]
- Taly AB, Meenakshi-Sundaram S, Sinha S, Swamy HS, Arunodaya GR. Wilson disease: description of 282 patients evaluated over 3 decades. 30 Medicine (Baltimore) 2007; **86**: 112-121 [PMID: 17435591 DOI: 10.1097/MD.0b013e318045a00e]
- 31 Sridhar MS, Rangaraju A, Anbarasu K, Reddy SP, Daga S, Jayalakshmi S, Shaik B. Evaluation of Kayser-Fleischer ring in Wilson disease by anterior segment optical coherence tomography. Indian J Ophthalmol 2017; 65: 354-357 [PMID: 28573989 DOI: 10.4103/ijo.IJO_400_16]
- Broniek-Kowalik K, Dzieżyc K, Litwin T, Członkowska A, Szaflik JP. Anterior segment optical coherence tomography (AS-OCT) as a new 32 method of detecting copper deposits forming the Kayser-Fleischer ring in patients with Wilson disease. Acta Ophthalmol 2019; 97: e757-e760 [PMID: 30635971 DOI: 10.1111/aos.14009]
- 33 Korman JD, Volenberg I, Balko J, Webster J, Schiodt FV, Squires RH Jr, Fontana RJ, Lee WM, Schilsky ML; Pediatric and Adult Acute Liver Failure Study Groups. Screening for Wilson disease in acute liver failure: a comparison of currently available diagnostic tests. Hepatology 2008; 48: 1167-1174 [PMID: 18798336 DOI: 10.1002/hep.22446]
- Shaver WA, Bhatt H, Combes B. Low serum alkaline phosphatase activity in Wilson's disease. Hepatology 1986; 6: 859-863 [PMID: 3758940 34 DOI: 10.1002/hep.1840060509]
- Sinha S, Taly AB, Ravishankar S, Prashanth LK, Venugopal KS, Arunodaya GR, Vasudev MK, Swamy HS. Wilson's disease: cranial MRI 35 observations and clinical correlation. Neuroradiology 2006; 48: 613-621 [PMID: 16752136 DOI: 10.1007/s00234-006-0101-4]
- Stefano Zagami A, Boers PM. Disappearing "face of the giant panda". Neurology 2001; 56: 665 [PMID: 11245722 DOI: 36 10.1212/WNL.56.5.665]
- Akhan O, Akpinar E, Oto A, Köroglu M, Ozmen MN, Akata D, Bijan B. Unusual imaging findings in Wilson's disease. Eur Radiol 2002; 12 37 Suppl 3: S66-S69 [PMID: 12522607 DOI: 10.1007/s00330-002-1589-6]
- Barbosa ER, Caramelli P, Bacheschi LA, Haddad MS, Magalhães AC, Menezes JR, Scaff M, Canelas HM. Wilson's disease: magnetic 38 resonance imaging (MRI) with clinical correlations in 16 cases. Rev Paul Med 1993; 111: 407-411 [PMID: 8108634]
- Chang IJ, Hahn SH. The genetics of Wilson disease. Handb Clin Neurol 2017; 142: 19-34 [PMID: 28433102 DOI: 39 10.1016/B978-0-444-63625-6.00003-3]
- Merle U, Schaefer M, Ferenci P, Stremmel W. Clinical presentation, diagnosis and long-term outcome of Wilson's disease: a cohort study. Gut 40 2007; **56**: 115-120 [PMID: 16709660 DOI: 10.1136/gut.2005.087262]
- 41 Ferenci P, Caca K, Loudianos G, Mieli-Vergani G, Tanner S, Sternlieb I, Schilsky M, Cox D, Berr F. Diagnosis and phenotypic classification of Wilson disease. Liver Int 2003; 23: 139-142 [PMID: 12955875 DOI: 10.1034/j.1600-0676.2003.00824.x]
- Sözeri E, Feist D, Ruder H, Schärer K. Proteinuria and other renal functions in Wilson's disease. Pediatr Nephrol 1997; 11: 307-311 [PMID: 42 9203178 DOI: 10.1007/s004670050282]
- Litin RB, Randall RV, Goldstein NP, Power MH, Diessner GR. Hypercalciuria in hepatolenticular degeneration (Wilson's disease). Am J Med 43 Sci 1959; 238: 614-620 [PMID: 14417517]
- Reynolds HV, Talekar CR, Bellapart J, Leggett BA, Boots RJ. Copper removal strategies for Wilson's disease crisis in the ICU. Anaesth 44 Intensive Care 2014; 42: 253-257 [PMID: 24580393]
- Mensing B, Nowak A, Zweifel S, Terracciano L, Bernsmeier C, Filipowicz Sinnreich M. [Wilson's disease or hepatolenticular degeneration]. 45 Ther Umsch 2018; **75**: 241-248 [PMID: 30468117 DOI: 10.1024/0040-5930/a000995]
- 46 Leu ML, Strickland GT, Gutman RA. Renal function in Wilson's disease: response to penicillamine therapy. Am J Med Sci 1970; 260: 381-398 [PMID: 5511397 DOI: 10.1097/00000441-197012000-00007]
- Zhuang XH, Mo Y, Jiang XY, Chen SM. Analysis of renal impairment in children with Wilson's disease. World J Pediatr 2008; 4: 102-105 47 [PMID: 18661763 DOI: 10.1007/s12519-008-0019-5]
- Kapoor N, Cherian KE, Sajith KG, Thomas M, Eapen CE, Thomas N, Paul TV. Renal Tubular Function, Bone Health and Body Composition 48 in Wilson's Disease: A Cross-Sectional Study from India. Calcif Tissue Int 2019; 105: 459-465 [PMID: 31317233 DOI: 10.1007/s00223-019-00588-z]
- 49 Wolff SM. Renal Lesions in Wilson's Disease. Lancet 1964; 1: 843-845 [PMID: 14129796 DOI: 10.1016/s0140-6736(64)91573-9]
- Elsas LJ, Hayslett JP, Spargo BH, Durant JL, Rosenberg LE. Wilson's disease with reversible renal tubular dysfunction. Correlation with 50 proximal tubular ultrastructure. Ann Intern Med 1971; 75: 427-433 [PMID: 5315082 DOI: 10.7326/0003-4819-75-3-427]
- 51 Azizi E, Eshel G, Aladjem M. Hypercalciuria and nephrolithiasis as a presenting sign in Wilson disease. Eur J Pediatr 1989; 148: 548-549 [PMID: 2744017 DOI: 10.1007/BF00441555]
- Palkar AV, Shrivastava MS, Padwal NJ, Padhiyar RN, Moulick N. Renal tubular acidosis due to Wilson's disease presenting as metabolic 52 bone disease. BMJ Case Rep 2011; 2011 [PMID: 22688476 DOI: 10.1136/bcr.04.2011.4121]
- Gündüz Z, Düşünsel R, Anarat A. Wilson cirrhosis associated with membranoproliferative glomerulonephritis. Nephron 1996; 74: 497-498 53 [PMID: 8893211 DOI: 10.1159/000189412]
- Acharya R, Zeng X, Clapp WL, Upadhyay K. IgA vasculitis with nephritis in cirrhotic Wilson disease: Is there an association? Clin Nephrol Case Stud 2020; 8: 80-84 [PMID: 33062584 DOI: 10.5414/CNCS110268]
- Begum A, Rahman MA, Akter AM, Akter M, Islam MA, Uddin GM. Nephrotic Syndrome after Treatment with D-Penicillamine in a Patient 55 with Wilson's Disease- A Case Report. J Ped Nephrol [Internet]. 2016 Jul. 4 [cited 2023 Sep. 22]; 4: 70-73
- Davison AM, Day AT, Golding JR, Thomson D. Effect of penicillamine on the kidney. Proc R Soc Med 1977; 70 Suppl 3: 109-113 [PMID: 56
- Mao H, Qin J, Kang T, Ou S. Wilson disease and IgA nephropathy: accidental or related? Int Urol Nephrol 2022; 54: 2441-2443 [PMID: 57 35260977 DOI: 10.1007/s11255-022-03159-8]
- Litwin T, Członkowska A, Socha P. Oral chelator treatment of Wilson Disease: d-penicillamine. InClinical and translational perspectives on 58 Wilson disease 2019; 357-364
- 59 Hall CL, Jawad S, Harrison PR, MacKenzie JC, Bacon PA, Klouda PT, MacIver AG. Natural course of penicillamine nephropathy: a long term study of 33 patients. Br Med J (Clin Res Ed) 1988; 296: 1083-1086 [PMID: 3132218 DOI: 10.1136/bmj.296.6629.1083]
- Habib GS, Saliba W, Nashashibi M, Armali Z. Penicillamine and nephrotic syndrome. Eur J Intern Med 2006; 17: 343-348 [PMID: 16864010 60 DOI: 10.1016/j.ejim.2006.03.001]
- Sternlieb I. Penicillamine and the nephrotic syndrome. JAMA 1966; 198: 1311-1312 [PMID: 5953529 DOI: 10.1001/jama.198.12.1311] 61
- Jaffe IA, Treser G, Suzuki Y, Ehrenreich T. Nephropathy induced by D-penicillamine. Ann Intern Med 1968; 69: 549-556 [PMID: 4175137



- DOI: 10.7326/0003-4819-69-3-549]
- Neild GH, Gärtner HV, Bohle A. Letter: D-penicillamine-induced membranous glomerulonephritis. Lancet 1975; 1: 1201-1202 [PMID: 48830 63 DOI: 10.1016/S0140-6736(75)93197-9]
- Bacon PA, Tribe CR, Mackenzie JC, Verrier-Jones J, Cumming RH, Amer B. Penicillamine nephropathy in rheumatoid arthritis. A clinical, 64 pathological and immunological study. Q J Med 1976; 45: 661-684 [PMID: 1005658]
- 65 Jaffe IA. Induction of auto-immune syndromes by penicillamine therapy in rheumatoid arthritis and other diseases. Springer Semin Immunopathol 1981; 4: 193-207 [PMID: 7034252 DOI: 10.1007/BF01857095]
- Walshe JM. Management of penicillamine nephropathy in Wilson's disease: a new chelating agent. Lancet 1969; 2: 1401-1402 [PMID: 66 4188283 DOI: 10.1016/S0140-6736(69)90940-4]
- Siafakas CG, Jonas MM, Alexander S, Herrin J, Furuta GT. Early onset of nephrotic syndrome after treatment with D-penicillamine in a 67 patient with Wilson's disease. Am J Gastroenterol 1998; 93: 2544-2546 [PMID: 9860423 DOI: 10.1111/j.1572-0241.1998.00715.x]
- Ross JH, McGinty F, Brewer DG. Penicillamine nephropathy. Nephron 1980; 26: 184-186 [PMID: 7432580 DOI: 10.1159/000181984] 68
- Hill H, Hill A, Davison AM. Resumption of treatment with penicillamine after proteinuria. Ann Rheum Dis 1979; 38: 229-231 [PMID: 485579 69 DOI: 10.1136/ard.38.3.2291
- Lee Y, Lee ST, Cho H. D-penicillamine-induced ANA (+) ANCA (+) vasculitis in pediatric patients with Wilson's disease. Clin Nephrol 2016; 70 85: 296-300 [PMID: 26784915 DOI: 10.5414/CN108763]
- Hoxha E, Harendza S, Zahner G, Panzer U, Steinmetz O, Fechner K, Helmchen U, Stahl RA. An immunofluorescence test for phospholipase-71 A2-receptor antibodies and its clinical usefulness in patients with membranous glomerulonephritis. Nephrol Dial Transplant 2011; 26: 2526-2532 [PMID: 21633097 DOI: 10.1093/ndt/gfr247]
- Kumar RP, Prasad ND, Tirumavalavan S, Fernando ME. D-penicillamine-induced membranous nephropathy. Indian J Nephrol 2014; 24: 195-72 196 [PMID: 25120302 DOI: 10.4103/0971-4065.132024]
- DeSilva RN, Eastmond CJ. Management of proteinuria secondary to penicillamine therapy in rheumatoid arthritis. Clin Rheumatol 1992; 11: 73 216-219 [PMID: 1617896 DOI: 10.1007/BF02207960]
- Walshe JM. The liver in Wilson's disease. In: Schiff L, Schiff ER, editors. Diseases of the Liver. 6th ed. Philadelphia: Lippincott; 1987; 74 1037-1050
- Proost R, Cassiman D, Levtchenko E, Morava-Kozicz E, Neirynck J, Witters P. Fulminant Wilson Disease in Children: Recovery After 75 Plasma Exchange Without Transplantation. J Pediatr Gastroenterol Nutr 2020; 71: 720-725 [PMID: 32804913 DOI: 10.1097/MPG.0000000000002894]
- Steen VD, Blair S, Medsger TA Jr. The toxicity of D-penicillamine in systemic sclerosis. Ann Intern Med 1986; 104: 699-705 [PMID: 2938530] 76 DOI: 10.7326/0003-4819-104-5-699]
- Jaffe IA. Adverse effects profile of sulfhydryl compounds in man. Am J Med 1986; 80: 471-476 [PMID: 2937293 DOI: 77 10.1016/0002-9343(86)90722-9]
- 78 Day AT, Golding JR, Lee PN, Butterworth AD. Penicillamine in rheumatoid disease: a long-term study. Br Med J 1974; 1: 180-183 [PMID: 4590670 DOI: 10.1136/bmi.1.5900.1801
- Kay AG. Myelotoxicity of D-penicillamine. Ann Rheum Dis 1979; 38: 232-236 [PMID: 485580 DOI: 10.1136/ard.38.3.232] 79
- 80 Socha P, Janczyk W, Dhawan A, Baumann U, D'Antiga L, Tanner S, Iorio R, Vajro P, Houwen R, Fischler B, Dezsofi A, Hadzic N, Hierro L, Jahnel J, McLin V, Nobili V, Smets F, Verkade HJ, Debray D. Wilson's Disease in Children: A Position Paper by the Hepatology Committee of the European Society for Paediatric Gastroenterology, Hepatology and Nutrition. J Pediatr Gastroenterol Nutr 2018; 66: 334-344 [PMID: 29341979 DOI: 10.1097/MPG.0000000000001787]
- Taly AB, Prashanth LK, Sinha S. Wilson's disease: An Indian perspective. Neurol India 2009; 57: 528-540 [PMID: 19934550 DOI: 81 10.4103/0028-3886.57789]
- 82 Ferenci P, Stremmel W, Członkowska A, Szalay F, Viveiros A, Stättermayer AF, Bruha R, Houwen R, Pop TL, Stauber R, Gschwantler M, Pfeiffenberger J, Yurdaydin C, Aigner E, Steindl-Munda P, Dienes HP, Zoller H, Weiss KH. Age and Sex but Not ATP7B Genotype Effectively Influence the Clinical Phenotype of Wilson Disease. Hepatology 2019; 69: 1464-1476 [PMID: 30232804 DOI: 10.1002/hep.30280]
- Dusek P, Litwin T, Członkowska A. Neurologic impairment in Wilson disease. Ann Transl Med 2019; 7: S64 [PMID: 31179301 DOI: 83 10.21037/atm.2019.02.431
- Brewer GJ, Terry CA, Aisen AM, Hill GM. Worsening of neurologic syndrome in patients with Wilson's disease with initial penicillamine 84 therapy. Arch Neurol 1987; 44: 490-493 [PMID: 3579660 DOI: 10.1001/archneur.1987.00520170020016]
- 85 Ranjan A, Kalita J, Kumar V, Misra UK. MRI and oxidative stress markers in neurological worsening of Wilson disease following penicillamine. Neurotoxicology 2015; 49: 45-49 [PMID: 26004675 DOI: 10.1016/j.neuro.2015.05.004]
- Litwin T, Dzieżyc K, Karliński M, Chabik G, Czepiel W, Członkowska A. Early neurological worsening in patients with Wilson's disease. J 86 Neurol Sci 2015; **355**: 162-167 [PMID: 26071888 DOI: 10.1016/j.jns.2015.06.010]
- Chen DB, Feng L, Lin XP, Zhang W, Li FR, Liang XL, Li XH. Penicillamine increases free copper and enhances oxidative stress in the brain 87 of toxic milk mice. PLoS One 2012; 7: e37709 [PMID: 22629446 DOI: 10.1371/journal.pone.0037709]
- Miki M. Penicillamine as antioxidant. Methods Enzymol 1994; 234: 542-547 [PMID: 7808330 DOI: 10.1016/0076-6879(94)34126-5] 88
- Mohr I, Weiss KH. Current anti-copper therapies in management of Wilson disease. Ann Transl Med 2019; 7: S69 [PMID: 31179306 DOI: 89 10.21037/atm.2019.02.48]
- Klein D, Lichtmannegger J, Heinzmann U, Summer KH. Dissolution of copper-rich granules in hepatic lysosomes by D-penicillamine prevents 90 the development of fulminant hepatitis in Long-Evans cinnamon rats. J Hepatol 2000; 32: 193-201 [PMID: 10707858 DOI: 10.1016/S0168-8278(00)80063-X]
- Netter P, Bannwarth B, Péré P, Nicolas A. Clinical pharmacokinetics of D-penicillamine. Clin Pharmacokinet 1987; 13: 317-333 [PMID: 91 3319347 DOI: 10.2165/00003088-198713050-00003]
- Shimizu N, Yamaguchi Y, Aoki T. Treatment and management of Wilson's disease. Pediatr Int 1999; 41: 419-422 [PMID: 10453198 DOI: 92 10.1046/j.1442-200x.1999.01093.x]
- Brewer GJ, Yuzbasiyan-Gurkan V, Johnson V, Dick RD, Wang Y. Treatment of Wilson's disease with zinc: XI. Interaction with other 93 anticopper agents. J Am Coll Nutr 1993; 12: 26-30 [PMID: 8440814 DOI: 10.1080/07315724.1993.10718278]
- Levy RS, Fisher M, Alter JN. Penicillamine: review and cutaneous manifestations. J Am Acad Dermatol 1983; 8: 548-558 [PMID: 6222087 94 DOI: 10.1016/S0190-9622(83)70062-9]



- Ishak R, Abbas O. Penicillamine revisited: historic overview and review of the clinical uses and cutaneous adverse effects. Am J Clin Dermatol 2013; 14: 223-233 [PMID: 23605177 DOI: 10.1007/s40257-013-0022-z]
- Kalita J, Kumar V, Chandra S, Kumar B, Misra UK. Worsening of Wilson disease following penicillamine therapy. Eur Neurol 2014; 71: 126-96 131 [PMID: 24356057 DOI: 10.1159/000355276]
- Kumar M, Murugan TP, Lionel AP, Thomas MM, Mannam P, Yoganathan S. Management of Children and Adolescents with Wilson Disease 97 and Neurological Worsening Following D-Penicillamine Therapy: A Single Centre Experience. Ann Indian Acad Neurol 2022; 25: 698-702 [PMID: 36211139 DOI: 10.4103/aian.aian_519_21]
- Lu J. Triethylenetetramine pharmacology and its clinical applications. Mol Cancer Ther 2010; 9: 2458-2467 [PMID: 20660601 DOI: 98 10.1158/1535-7163.MCT-10-0523]
- 99 Woimant F, Debray D, Morvan E, Obadia MA, Poujois A. Efficacy and Safety of Two Salts of Trientine in the Treatment of Wilson's Disease. J Clin Med 2022; 11 [PMID: 35887738 DOI: 10.3390/jcm11143975]
- Seetharaman J, Sarma MS. Chelation therapy in liver diseases of childhood: Current status and response. World J Hepatol 2021; 13: 1552-1567 [PMID: 34904029 DOI: 10.4254/wjh.v13.i11.1552]
- Weiss KH, Thompson C, Dogterom P, Chiou YJ, Morley T, Jackson B, Amin N, Kamlin COF. Comparison of the Pharmacokinetic Profiles of 101 Trientine Tetrahydrochloride and Trientine Dihydrochloride in Healthy Subjects. Eur J Drug Metab Pharmacokinet 2021; 46: 665-675 [PMID: 34357516 DOI: 10.1007/s13318-021-00704-1]
- Arnon R, Calderon JF, Schilsky M, Emre S, Shneider BL. Wilson disease in children: serum aminotransferases and urinary copper on triethylene tetramine dihydrochloride (trientine) treatment. J Pediatr Gastroenterol Nutr 2007; 44: 596-602 [PMID: 17460493 DOI: 10.1097/MPG.0b013e3180467715]
- Taylor RM, Chen Y, Dhawan A; EuroWilson Consortium. Triethylene tetramine dihydrochloride (trientine) in children with Wilson disease: experience at King's College Hospital and review of the literature. Eur J Pediatr 2009; 168: 1061-1068 [PMID: 19066958 DOI: $10.1007/s00431\hbox{-}008\hbox{-}0886\hbox{-}8]$
- Weiss KH, Kruse C, Manolaki N, Zuin M, Ferenci P, van Scheppingen D, Wijnberg L, de Koning CE, Dhawan A. Multicentre, retrospective study to assess long-term outcomes of chelator based treatment with trientine in Wilson disease patients withdrawn from therapy with dpenicillamine. Eur J Gastroenterol Hepatol 2022; 34: 940-947 [PMID: 35482910 DOI: 10.1097/MEG.00000000000002387]
- Weiss KH, Thurik F, Gotthardt DN, Schäfer M, Teufel U, Wiegand F, Merle U, Ferenci-Foerster D, Maieron A, Stauber R, Zoller H, Schmidt HH, Reuner U, Hefter H, Trocello JM, Houwen RH, Ferenci P, Stremmel W; EUROWILSON Consortium. Efficacy and safety of oral chelators in treatment of patients with Wilson disease. Clin Gastroenterol Hepatol 2013; 11: 1028-35.e1 [PMID: 23542331 DOI: 10.1016/j.cgh.2013.03.012]
- Brewer GJ, Askari F, Lorincz MT, Carlson M, Schilsky M, Kluin KJ, Hedera P, Moretti P, Fink JK, Tankanow R, Dick RB, Sitterly J. Treatment of Wilson disease with ammonium tetrathiomolybdate: IV. Comparison of tetrathiomolybdate and trientine in a double-blind study of treatment of the neurologic presentation of Wilson disease. Arch Neurol 2006; 63: 521-527 [PMID: 16606763 DOI: 10.1001/archneur.63.4.521]
- Camarata MA, Ala A, Schilsky ML. Zinc Maintenance Therapy for Wilson Disease: A Comparison Between Zinc Acetate and Alternative Zinc Preparations. Hepatol Commun 2019; 3: 1151-1158 [PMID: 31388634 DOI: 10.1002/hep4.1384]
- Tang S, Bai L, Hou W, Hu Z, Chen X, Zhao J, Liang C, Zhang W, Duan Z, Zheng S. Comparison of the Effectiveness and Safety of d-Penicillamine and Zinc Salt Treatment for Symptomatic Wilson Disease: A Systematic Review and Meta-Analysis. Front Pharmacol 2022; 13: 847436 [PMID: 35370752 DOI: 10.3389/fphar.2022.847436]
- Brewer GJ, Dick RD, Yuzbasiyan-Gurkin V, Tankanow R, Young AB, Kluin KJ. Initial therapy of patients with Wilson's disease with tetrathiomolybdate. Arch Neurol 1991; 48: 42-47 [PMID: 1986725 DOI: 10.1001/archneur.1991.00530130050019]
- Stremmel W. Bis-choline Tetrathiomolybdate as Old Drug in a New Design for Wilson's Disease: Good for Brain and Liver? Hepatology 2019; **69**: 901-903 [PMID: 30070374 DOI: 10.1002/hep.30130]
- Brewer GJ, Hedera P, Kluin KJ, Carlson M, Askari F, Dick RB, Sitterly J, Fink JK. Treatment of Wilson disease with ammonium tetrathiomolybdate: III. Initial therapy in a total of 55 neurologically affected patients and follow-up with zinc therapy. Arch Neurol 2003; 60: 379-385 [PMID: 12633149 DOI: 10.1001/archneur.60.3.379]
- Weiss KH, Askari FK, Czlonkowska A, Ferenci P, Bronstein JM, Bega D, Ala A, Nicholl D, Flint S, Olsson L, Plitz T, Bjartmar C, Schilsky ML. Bis-choline tetrathiomolybdate in patients with Wilson's disease: an open-label, multicentre, phase 2 study. Lancet Gastroenterol Hepatol 2017; 2: 869-876 [PMID: 28988934 DOI: 10.1016/S2468-1253(17)30293-5]
- Brewer GJ, Askari F, Dick RB, Sitterly J, Fink JK, Carlson M, Kluin KJ, Lorincz MT. Treatment of Wilson's disease with tetrathiomolybdate: V. Control of free copper by tetrathiomolybdate and a comparison with trientine. Transl Res 2009; 154: 70-77 [PMID: 19595438 DOI: 10.1016/j.trsl.2009.05.002]
- Schilsky ML, Czlonkowska A, Zuin M, Cassiman D, Twardowschy C, Poujois A, Gondim FAA, Denk G, Cury RG, Ott P, Moore J, Ala A, D'Inca R, Couchonnal-Bedoya E, D'Hollander K, Dubois N, Kamlin COF, Weiss KH; CHELATE trial investigators. Trientine tetrahydrochloride versus penicillamine for maintenance therapy in Wilson disease (CHELATE): a randomised, open-label, non-inferiority, phase 3 trial. Lancet Gastroenterol Hepatol 2022; 7: 1092-1102 [PMID: 36183738 DOI: 10.1016/S2468-1253(22)00270-9]
- Medici V, Trevisan CP, D'Incà R, Barollo M, Zancan L, Fagiuoli S, Martines D, Irato P, Sturniolo GC. Diagnosis and management of Wilson's disease: results of a single center experience. J Clin Gastroenterol 2006; 40: 936-941 [PMID: 17063115 DOI: 10.1097/01.mcg.0000225670.91722.59]
- Czlonkowska A, Gajda J, Rodo M. Effects of long-term treatment in Wilson's disease with D-penicillamine and zinc sulphate. J Neurol 1996; **243**: 269-273 [PMID: 8936358 DOI: 10.1007/BF00868525]
- Weiss KH, Gotthardt DN, Klemm D, Merle U, Ferenci-Foerster D, Schaefer M, Ferenci P, Stremmel W. Zinc monotherapy is not as effective as chelating agents in treatment of Wilson disease. Gastroenterology 2011; 140: 1189-1198.e1 [PMID: 21185835 DOI: 10.1053/j.gastro.2010.12.034]
- Dhawan A, Taylor RM, Cheeseman P, De Silva P, Katsiyiannakis L, Mieli-Vergani G. Wilson's disease in children: 37-year experience and revised King's score for liver transplantation. Liver Transpl 2005; 11: 441-448 [PMID: 15776453 DOI: 10.1002/lt.20352]
- Askari FK, Greenson J, Dick RD, Johnson VD, Brewer GJ. Treatment of Wilson's disease with zinc. XVIII. Initial treatment of the hepatic decompensation presentation with trientine and zinc. J Lab Clin Med 2003; 142: 385-390 [PMID: 14713890 DOI: 10.1016/S0022-2143(03)00157-4
- 120 Santos Silva EE, Sarles J, Buts JP, Sokal EM. Successful medical treatment of severely decompensated Wilson disease. J Pediatr 1996; 128:



- 285-287 [PMID: 8636833 DOI: 10.1016/S0022-3476(96)70412-2]
- Chen JC, Chuang CH, Wang JD, Wang CW. Combination Therapy Using Chelating Agent and Zinc for Wilson's Disease. J Med Biol Eng 2015; **35**: 697-708 [PMID: 26692828 DOI: 10.1007/s40846-015-0087-7]
- Gupta P, Choksi M, Goel A, Zachariah U, Sajith KG, Ramachandran J, Chandy G, Kurian G, Rebekah G, Eapen CE. Maintenance zinc therapy after initial penicillamine chelation to treat symptomatic hepatic Wilson's disease in resource-constrained setting. Indian J Gastroenterol 2018; **37**: 31-38 [PMID: 29457214 DOI: 10.1007/s12664-018-0829-x]
- Panda K, Lal BB, Sood V, Khanna R, Alam S. Incidence of Relapse Following Withdrawal of D-Penicillamine from Combination (D-Penicillamine+ Zinc) Therapy in Previously Symptomatic Hepatic Wilson Disease. Journal of Clinical and Experimental Hepatology 2022; 12: S80 [DOI: 10.1016/j.jceh.2023.07.222]





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