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Please find enclosed our revised submission of the manuscript 28883 entitled "Overview of the progress on haploidentical hematopoietic transplantation". We declare that this manuscript is original and has not been published before and is not currently being considered for publication elsewhere. The following cover letter is in response to the reviewer's comments.

Reviewer(s) comments to Author:

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First author: Noshad Farhadfar

Corresponding author: William J Hogan

Comment 2: All the authors' work should be given in this section. Thank you

All authors contributed to drafting, revision and final approval of the article.

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Comment 6: Please add PubMed citation numbers and DOI citation to the reference list and list all authors.

Pubmed citation numbers were added to the reference list. Some of the older references (1980s and early 1990s) do not have DOI citation number. All authors are listed.

We hope you find the responses above and the revised manuscript to be acceptable for publication.

Thank you,

Noshad Farhadfar, M.D.

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Overview of the progress on haploidentical hematopoietic transplantation

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Abstract

Allogeneic hematopoietic stem cell transplant (HSCT) remains the only potentially curative option for variety of hematologic disorders. Lack of a suitable fully HLA-matched donor limits this option for many patients. Without a suitable related or unrelated HLA-matched donor, umbilical cord blood and haploidentical family members provide a potential source of stem cells. Timely donor availability make haploidentical donors an attractive alternative donor source. Initial attempts at haploidentical HSCT was associated with significantly increased mortality owing to high rates of graft rejection and severe graft-versus-host disease caused by major donor-recipient HLA-disparity. However, over the past decade, outcomes of haploidentical HSCT have improved significantly. Here, we review the advantages and challenges of haploidentical transplantation. We also discuss new developments to attempt to overcome the challenges to a successful haploidentical transplantation.

Keywords: Haploidentical donor, Hematopoietic stem cell transplantation, Hematological malignancies, Transplant related mortality

Core tip: Over the past decade, haploidentical donors have emerged as a viable alternate graft source for patients without a HLA-matched donor. Several strategies including graft manipulation, conditioning regimen optimization and better graft-versus-host disease control have significantly improved the outcomes of haploidentical HSCT. Here, we summarize some of the recent advances in the field of haploidentical HSCT in adults.

Audio Core Tip

Introduction

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Hematopoietic stem-cell transplantation (HSCT) is a potentially curative treatment for several malignant and non-malignant hematologic diseases. Most institutions currently consider a HLA-matched sibling as a preferred donor source, typically followed by either HLA-matched unrelated donor or an alternative donor source depending on the clinical scenario. The probability of having an HLA-matched sibling donor is approximately thirty percent after consideration of factors such as donor consent and health status. The likelihood of finding a suitable matched unrelated donor is strongly influenced by the patient's ethnicity and can range from more than 75% for Caucasians to less than 20% for certain ethnic groups such as African Americans^[1]. For patients without a suitable related or unrelated HLA-matched donor, umbilical cord blood and haploidentical family members provide a potential source of stem cells. The use of haploidentical hematopoietic stem cell transplantation as an alternative graft source has been substantially increasing in the past decade.

The use of haploidentical related donors has a number of advantages including immediate donor availability for many patients facilitating a shorter interval to transplant. In addition, having a related donor makes post-transplant donor-derived cellular therapy more easily accessible. Challenges include major donor-recipient HLA-disparity and T cell depletion/modification which can lead to delayed immune reconstitution, graft failure and severe graft versus host disease (GVHD) due to T-cell alloreactivity^[2, 3]. This review highlights the major advances over the past decade to overcome the obstacles to successful haploidentical transplantation.

Donor selection

In contrast to unrelated donor transplant HSCT where finding the best HLA matched donor is the most important factor in determining transplant outcome, increasing HLA disparity in—haploidentical matching does not have the same detrimental impact with dedicated techniques such as modification of post-transplant T cell reconstitution with cyclophosphamide. In 2010, *Kasamon et al.* evaluated the impact of HLA disparity between donor and recipient in 185 patients with hematologic malignancies who underwent un-manipulated bone marrow haploidentical transplant^[4].

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GVHD prophylaxis consisted of post-transplant cyclophosphamide and the number of HLA-mismatches did not influence the rate of acute GVHD or disease free survival.

Donor characteristics that influence the outcome of haploidentical transplant were also investigated in a large study by *Wang et al.* involving 1210 patients with hematologic diseases[5]. Grafts consisted of G-CSF mobilized T-cell replete bone marrow and peripheral stem cells. Similar to the prior studies, the degree of HLA disparity had no effect on acute GVHD and treatment related mortality (TRM). Younger donor age (< 30 years) correlated with lower incidence of acute GVHD compared to older donor age (> 30 years). Younger donor age and male gender were also associated with less TRM and better overall survival (OS). The benefit of male recipient gender was lost when maternal donors were excluded. There was a higher risk of grade II-IV acute GVHD with maternal donors compared to paternal donors. In a male recipient, a maternal donor also correlated with a higher TRM rate and decreased OS. The impact of non-inherited maternal antigen (NIMA) disparities was evaluated in 264 patients. NIMA mismatched donors conferred a lower incidence of acute GVHD compared to non-inherited paternal antigen (NIPA) mismatched donors. Based on these results, authors concluded younger, male, NIMA-mismatched donor is a preferred donor in setting of T-cell replete haploidentical transplant. This study did not evaluate the impact of natural killer (NK) cell alloreactivity and CMV status of the donor. In contrast to *Wang et al.* study, several trials demonstrated decreased risk of relapse and survival advantage with using maternal donors [6]. A more potent anti-leukemic effect of maternal donor grafts has been attributed to the maternal immune system exposure to fetal antigens during pregnancy [7].

Another factor influencing haploidentical transplant outcome is donor versus recipient NK cell alloreactivity. Tumor cells are able to escape T-cell adoptive immune response by down regulating cell surface MHC class I. NK cells play a crucial role in innate immunity and have MHC-unrestricted ability to target malignant cells. Cytotoxic activity of NK cells are mainly under the negative feedback control from inhibitory killer immunoglobulin-like receptors (KIRs) through binding to self HLA class I antigen.

This phenomenon is known as “missing self” [8-10]. KIR-KIR ligand mismatched in the donor-recipient direction lead to loss of the inhibitory feedback and activation of donor NK cells targeting recipient hematopoietic cells and leukemic cells. In contrast to allo-reactive T-lymphocytes, NK cells are thought to be capable of inducing Graft versus leukemia (GVL) effect without promoting GVHD. In 2002, a study by the Perugia group demonstrated therapeutic efficacy of allo-reactive NK cells in 57 patients with acute myeloid leukemia (AML) following haploidentical transplant [11]. Twenty out of 57 patients had KIR-ligand incompatibility in the graft versus host direction. The probability of OS at 5 years was markedly improved in patients with AML who had NK allo-reactive donors (60% versus 5%, p= 0.0005). A beneficial effect of NK cell mismatching was not seen in patients with acute lymphoblastic leukemia (ALL). Similar results were observed in the updated analysis of 112 patients with high risk AML who received T-cell depleted haploidentical transplants[12]. Fifty-one of 112 patients had NK cell allo-reactive donors. The conditioning regimen included TBI (8 Gy), fludarabine (40 mg/m²/d for 4 days), thioguanine (5mg/kg/d for 2 days) and rabbit ATG. A significantly lower relapse rate (3% versus 47%, p<0.003) and better EFS (67% versus 18%, p=0.02) was observed in patients transplanted in any CR with NK allo-reactive donors compared to recipients of non-allo-reactive grafts. Although transplantation from NK allo-reactive donors improved survival in the entire cohort, subset analysis suggested that transplantation from NK allo-reactive donors did not decrease the incidence of relapse in patients transplanted at chemo-resistant relapse. Incidence of acute GVHD was similar between the two cohorts (10% versus 11%). These findings reinforced the theory that GVL activity by allo-reactive NK cells translated into prolonged overall survival. Subsequently, several studies revealed a favorable impact of allo-reactive NK cells on transplant outcome in patients undergoing HLA-haploidentical transplant [11, 13-15]. An important role of donor-recipient KIR mismatch was also demonstrated after non-myeloablative T cell-replete haploidentical transplantation using post-transplant cyclophosphamide in a retrospective study involving 86 patients with high risk hematologic malignancies [16]. On the contrary, a deleterious effect of KIR mismatches

was seen in the earlier studies [17, 18]. Due to ongoing controversy, currently the KIR testing is not considered mandatory for donor selection in haploidentical transplant setting.

Haploidentical stem cell transplant strategies

T-cell depletion

The first successful haploidentical transplants were performed in the 1980s in children with severe combined immunodeficiency syndrome (SCIDS) using T-cell depleted bone marrow grafts. T-lymphocyte depletion in this setting mitigated GVHD associated with crossing a major HLA-barrier without compromising engraftment [19]. Subsequently, this approach was implemented successfully in several studies of patients with SCIDS. In contrast to SCIDS, haploidentical transplantation was less successful in the setting of acute leukemia owing to a high rate of graft failure. Increased risk of graft failure was attributed to host derived T-lymphocytes that survived the conditioning regimen [20-22]. A decade later, it was shown in preclinical studies (murine models) that infusion of a large number of donor hematopoietic stem cells can overcome the MHC barrier and promote engraftment[23]. In 1993, cell dose escalation approach was tested in 36 patients with acute leukemia following myeloablative conditioning using a total body irradiation (TBI) based regimen (-TBI 8 Gy in single fraction, thiotepa 10mg/kg over 2 days, Cyclophosphamide 50 mg/kg for 2 days). Mega doses of stem cells (on average $> 10 \times 10^6$ CD34⁺ cells/kg body weight) were obtained by supplementing T cell-depleted bone marrow transplants with granulocyte colony-stimulating factor (G-CSF) mobilized peripheral blood stem cells. Using this approach, nearly -80% of patients achieved primary engraftment. The sole GVHD prophylaxis consisted of T-cell depletion of the graft by the soybean agglutination and E-rosetting technique. Only 18% of the patients developed grade II-IV acute GVHD [24, 25]. Subsequently, several modifications were introduced to optimize the T-cell depletion of the graft including positive immuno-selection of the CD34⁺ cells using the Ceprate system in 1995 and Clinimacs device in 1999 [13, 26]. In addition, to

reduce the toxicity associated with the myeloablative TBI based conditioning regimen, fludarabine was substituted for cyclophosphamide in 1995 [27]. After optimizing the conditioning regimen and graft processing, Aversa *et al.* investigated haploidentical transplantation in 284 patients with acute leukemia. Ninety five percent of the patients achieved engraftment with minimal GVHD. The relapse rate was 17% in acute myeloid leukemia (AML) and 27 % in acute lymphoblastic leukemia (ALL) patients transplanted in any CR.— Incidence of TRM was 40% mainly due to opportunistic infections. Seventeen year DFS was 30% in ALL and 43% in AML patients transplanted in any CR. Among the long-term survivors, chronic GVHD was not observed in any patients [28].

The major disadvantages of using T-cell depleted grafts are the high rate of relapse and life-threatening infections post-transplant [29]. Due to poor thymic function in adults, T-cell immune recovery after transplant depends of peripheral expansion of donor T-lymphocytes. In a T-cell depleted graft, passive transfer of T-lymphocytes is minimal leading to profound delay in immune recovery. To overcome these obstacles several strategies have evolved over the past decade including selective T-cell depletion, adoptive transfer of donor T-cells post- transplant, and T-regulatory cell (T-reg) add backs.

Selective T cell depletion:

The principle behind adoptive T-cell therapy is to eliminate donor allo-reactive T cells responsible for GVHD while sparing other immune cells, which facilitate immune reconstitution. To selectively deplete allo-reactive donor T-cells, ex vivo T-cells are activated against host antigen presenting cells. Activated T-cells are removed using several methods including immunotoxin, immune-magnetic selection and photodynamic purging [30-32].

Another innovative approach is to selectively remove T-cells responsible for GVHD (TCR alpha-beta) while sparing gamma-delta T-cells ($\gamma\delta$ T-cells). Gamma-delta T-cells account for 1% to 10% of peripheral T-cells. Based on in-vitro studies, human T lymphocytes which express $\gamma\delta$ T-cells receptor have MHC-unrestricted innate cytotoxic

activity against tumor cells [33, 34]. In a recent study, *Lang et al.* retrospectively evaluated the immune recovery after TCR $\alpha\beta$ /CD19-depleted haploidentical HSCT in 41 pediatric patients with acute leukemia, myelodysplasia and nonmalignant disease[35]. Eighty eight percent of the patients achieved primary engraftment. The incidence of grade II and grade III-IV acute GVHD was 10% and 15% respectively. At one year follow up, the event free survival (EFS) of patients with acute leukemia or myelodysplasia transplanted in CR1-CR3 was 100%. One year EFS of patients with subsequent HSCT (CR2-CR6) or with active disease was 29% and 11%, respectively. The use of TCR $\alpha\beta$ /CD19-depleted stem cells substantially accelerated immune recovery. In comparison to CD34+ selected grafts (historic control), patients achieved a higher CD3+ at days +30 and +90 (267 versus 27 and 397 versus 163 cells/ μ L), CD3+4+ at day +30 (58 vs 11 cells/ μ L) and CD56+ at day +14 (622 versus 27 cells/ μ L). The Italian group also reported similar results in 16 adults with high risk acute leukemia after TCR $\alpha\beta$ /CD19-depleted haploidentical HSCT.

A more recent strategy to separate GVHD and the GVL effect involves selectively depleting naïve T cells identified by CD45RA+ expression [36, 37]. Naïve T-cells are shown to be the most allo-reactive amongst the T-cell subsets. Ex vivo depletion of CD45RA+ T-cells and adoptive transfer of CD45RA- memory T cells hasten the immune reconstitution post-transplant, enhances the GVL effect while abrogating GVHD. This strategy was recently evaluated in a study of 17 patients with high risk hematologic malignancies (16 acute myeloid leukemia and 1 myelodysplasia) with KIR receptor-ligand mismatched haploidentical donor [38]. The conditioning regimen included total lymphoid irradiation (8 Gy), fludarabine (150 mg/m²), cyclophosphamide (60 mg/kg), thioguanine (10 mg/kg) and melphalan (140 mg/m²). Patients received a CD34+ selected stem cell graft on day 0 followed by an infusion of CD45RA-depleted stem cells on day +1. NK cell infusion was given on day +6. Post-transplant GVHD prophylaxis included sirolimus and mycophenolate mofetil (MMF). All patients achieved primary engraftment. Neutrophil and platelet engraftment was rapidly achieved at median day +11 and +17 respectively. Acute GVHD was not seen in

any of the patients. There was no infection related mortality. A phase II study of selective depletion of CD45RA+ T Cells from allogeneic peripheral blood stem cell grafts from HLA-matched related and unrelated donors for prevention of GVHD is currently under investigation [39].

Selective T-cell add back

Donor-derived regulatory T- cells (Tregs), co-infused with conventional T-cells (Tcons) is another approach to manipulate the T-cell depleted graft to improve transplant outcome in patients with high risk hematologic malignancy. In pre-clinical studies of bone marrow transplantation, infusion of donor-type CD4⁺CD25⁺Tregs abrogated GVHD without compromising the cytotoxic ability of T-cons against tumor cells[40, 41]. A first in human study by *Di lanni et al.* investigated infusion of Tregs, followed by Tcons in 28 patients with high risk hematologic malignancies who underwent haploidentical transplantation[42]. After TBI containing conditioning regimens, patients received infusion of donor derived T-regs (2×10^6 Tregs) on day -4. CD34⁺ stem cells were infused on Day 0 followed by Tcons. Two out of five patients who received 2×10^6 Tcons/kg developed acute GVHD which led to decreasing the cell dose of Tcons to 1×10^6 cells/kg. Chronic GVHD was not observed in any patients. All patients achieved primary engraftment. Compared to conventional mismatched HSCT, pathogen specific CD4⁺ and CD8⁺ were detected earlier in the study cohort (as early as 2 months versus 9-12 months). CMV- related death, a major cause of mortality in original T-cell depleted HSCT, was not observed. At median 1 year follow up, 46% of the patients were disease free. Subsequently, *Martelli et al.* evaluated the impact of Tregs - Tcons infusion in reducing post-transplant relapse risk in 43 adults with high risk acute leukemia [43]. This approach significantly reduced the risk of relapse and ameliorated GVHD. Grade 2 or more acute GVHD was seen in 15% of patients. At median follow up of 46 months, only two patients relapsed resulting in an incidence of relapse that was significantly lower than historical controls. Despite promising results of T-cell depleted haploidentical transplant, this approach is costly, technically

demanding and labor intensive which limits its application to highly experienced centers.

T cell replete graft

Earlier attempts at using un-manipulated haploidentical transplant were associated with an unacceptably high rate of GVHD related mortality due to donor T-cell alloreactivity. To overcome this obstacle, several strategies have evolved over the past decade including G-CSF primed graft [44, 45] and more recently post-transplant high dose cyclophosphamide.

High-dose post-transplant cyclophosphamide

Cyclophosphamide is an alkylating chemotherapeutic agent which has been used for many years as a component of conditioning regimens. Preclinical trials in the early 1970s revealed short course of cyclophosphamide after bone marrow transplantation can target allo-reactive T-cells and reduce the risk of GVHD [46-48]. In contrast to calcineurin inhibitors, cyclophosphamide is capable of inducing T- lymphocyte apoptosis [49]. Hematopoietic stem cells are resistant to high dose cyclophosphamide due to expression of high levels of aldehyde dehydrogenase [50]. Original clinical trials exploring cyclophosphamide efficacy as the post-transplant GVHD prophylactic agent were performed in the haploidentical transplant setting. In 2002, O'Donnell *et al.* evaluated the transplant outcome of 13 patients with high risk hematologic malignancies who received T-cell replete haploidentical transplant after a non-myeloablative conditioning regimen with TBI and fludarabine^[51]. GVHD prophylaxis included post-transplant cyclophosphamide 50 mg/kg on Day +3 in combination with MMF and tacrolimus. Due to high rate of graft failure (2 out of 3 patients) the protocol was amended to add cyclophosphamide 14.5 mg/kg to the conditioning regimen. Subsequently, primary donor cell engraftment was achieved in 8 of 10 patients. At a median of 99 days, 6 patients (46%) developed acute GVHD. At median of 191 days, incidence of DFS was 50%. This study demonstrated the feasibility and possibility of rapid engraftment with a non-myeloablative conditioning regimen in haploidentical transplant setting using post-transplant cyclophosphamide.

Subsequently, *Luznik et al.* compared safety and efficacy of administration of cyclophosphamide on day +3 and +4 rather than only on day +3 among 68 patients with hematologic malignancies after non-myeloablative haploidentical bone marrow transplant [52]. Primary engraftment was achieved in 87% of the patients. Notably, a very low incidence of grade III acute GVHD (6%) with no grade IV acute GVHD was observed at one year follow up. The only difference between the two cohorts was a trend toward a lower incidence of chronic GVHD after two doses of post-transplant cyclophosphamide (5% versus 25%, p= 0.05). The 2-year OS and EFS rates were 36% and 26%, respectively. A major contributor to the low OS rate was a high incidence of relapse (58% at 2 years).

A similar outcome was observed in a large phase II study of high dose post-transplantation cyclophosphamide as GVHD prophylaxis after non-myeloablative HLA-haploidentical bone marrow transplantation in 210 patients with hematologic malignancies [53]. Sustained donor cell engraftment was obtained in 87% of the patients. The cumulative incidences of grades II-IV acute GVHD was 27%. At 5 year follow up, OS and EFS were 35% and 27%, respectively. As seen in the prior studies, relapse was a major cause of mortality. Five year cumulative incidence of relapse was 55%.

In parallel multicenter phase 2 trials, BMT CTN 0603 and BMT CTN 0604, patients with acute leukemia or lymphoma underwent reduced intensity bone marrow haploidentical transplantation (0603) or double cord blood transplant (0604)[54]. The conditioning regimens contained 200 Gy TBI in addition to fludarabine and cyclophosphamide. In CTN 0603, the GVHD prophylaxis consisted of post-haploidentical transplant cyclophosphamide 50 mg/kg on day +3 and +4 followed by tacrolimus and MMF. In CTN 0604, GVHD prophylaxis included MMF and cyclosporine after double umbilical cord transplant. Among haploidentical transplant recipients, 100-day incidence of grade II-IV acute GVHD and 1- year incidence of chronic GVHD were 32% and 13%, respectively. After double cord transplant 100-day incidence of grade II-IV acute GVHD and 1- year incidence of chronic GVHD were 40% and 24%, respectively. One year cumulative incidence of relapse after haploidentical

and double umbilical cord transplant were 45% and 31%, respectively. The OS and EFS rates were 62% and 48% respectively after the haploidentical transplants. Similar OS (54%) and EFS (46%) were seen after double cord transplant. The authors concluded that both RIC haploidentical and double umbilical cord HSCT are valid options in patients with hematologic malignancy. Currently a multicenter randomized phase III trial (BMT CTN 1101) is investigating the effectiveness of haploidentical and double umbilical transplant in patients with leukemia or lymphoma [55].

Despite relatively low rates of GVHD with non-myeloablative haploidentical transplant, a high incidence of relapse has remained the main challenge in patients with high risk hematologic malignancies. To address this obstacle, use of more intense (myeloablative) preparative regimens and peripheral blood stem cell graft was explored. In a prospective study by *Solomon et al*, 20 patients with high risk (relapsed/refractory) hematologic malignancies were treated with myeloablative conditioning followed by peripheral blood derived haploidentical transplant [56]. The conditioning regimen consisted of fludarabine 30 mg/m² for 4 days, intravenous busulfan 130 mg/m²/d for 4 days, and Cy 14.5 mg/kg/d for 2 days. GVHD prophylaxis included high dose cyclophosphamide on day +3 and +4 followed by tacrolimus and MMF. All patients achieved primary engraftment. One year cumulative incidence of grade II-IV acute GVHD and chronic GVHD were 10% and 5%, respectively. At median follow up of 20 months, DFS and OS were 69% and 50%, respectively. The cumulative incidence of relapse was approximately 40%. The major drawback of this trial was non-fatal BK virus associated hemorrhagic cystitis which was seen in 75% of patients. This was attributed to the combination of high dose busulfan and cyclophosphamide. Association of BK induced hemorrhagic cystitis and high dose busulfan in setting of mismatched HSCT was reported previously in several studies [57]. To alleviate this problem, the conditioning regimen was changed to TBI-based myeloablative regimen in the subsequent study[58]. In this phase II prospective trial, 30 patients underwent peripheral stem cell haploidentical transplant using fludarabine 25 mg/m²/d for three days and 1200 cGy TBI as the preparative regimen. All patients achieved primary

engraftment. Median time to neutrophil and platelet engraftment was 16 days and 25 days, respectively. Incidence of grade II-IV acute GVHD was 23%, whereas moderate to severe chronic GVHD occurred in 22% of patients. In the entire cohort, 2-year NRM and OS were 3% and 78%, respectively. Among patients with low or intermediate risk disease NRM and OS were 0% and 100%, respectively. Relapse rate was significantly lower in comparison to patients treated at the same center with matched related transplant. Incidence of post-transplant BK virus associated hemorrhagic cystitis was significantly reduced after TBI-based regimen compared to the busulfan- based conditioning regimen (30% versus 75%, p= 0.005).

Similar results were observed in several other trials of myeloablative haploidentical transplant [59, 60]. Raiola *et al.* confirmed the low rate of GVHD and encouraging rate of DFS and OS in 50 patients with high risk hematologic disease- (23 patients in CR and 27 patients with active disease) after un-manipulated myeloablative haploidentical transplant [59]. Myeloablative conditioning consisted of thioguanine, busulfan, fludarabine or TBI based regimen. GVHD prophylaxis contained post-transplant cyclophosphamide on day +3 and +5 followed by cyclosporine and MMF. In the entire cohort, the cumulative incidence of grade II-III acute GVHD was 12%, and of moderate chronic GVHD was 10%. The actuarial 22-month DFS for patients transplanted in CR and patients with active disease was 68% and 37%, respectively[61]. The overall risk of relapse after myeloablative haploidentical HSCT was approximately 40% which compares favorably with that reported for non-myeloablative haploidentical HSCT. Therefore, despite the lack of randomized studies, myeloablative haploidentical transplant may be a reasonable option in younger patients with high risk hematologic malignancy in absence of timely access to a conventional donor.

Haploidentical related donor versus matched related sibling or matched unrelated donor (Table 1)

Encouraging results of haploidentical transplant compared to matched related or matched unrelated transplant has been suggested by several non-randomized studies.

In 2015, a large retrospective study compared the transplant outcome of 868 patients with acute leukemia after haploidentical transplant and 9[–]815 patients with HLA-matched sibling donor (MRD)^[62]. However, leukemia free survival was significantly longer after matched sibling donor transplant compared to haploidentical transplant (T-cell depleted or T-cell replete grafts). Haploidentical transplant was associated with higher TRM. The probability of relapse was not significantly different between the two cohorts. Therefore, the authors concluded haploidentical GVL effect is similar to MRD.

Ciurea et al. also retrospectively compared the transplant outcome of patients with AML after haploidentical transplant (n=192) using post-transplant cyclophosphamide and MUD (n=1982)^[63]. In the haploidentical cohort, 104 patients received MA and 88 had reduced intensity conditioning. In MUD cohort, 1245 patients (63%) received MA and 737 (37%) received RIC regimens. Compared to MUD, thirty day neutrophil engraftment was lower after haploidentical transplant in MA setting (97% versus 90%, P = 0.02). In RIC setting, day 30 neutrophil engraftment rate was similar between the two cohorts (96% and 93%, P = 0.25). Acute and chronic GVHD was notably lower after haploidentical transplant. In the MA setting, three month incidence of acute GVHD (16% versus 33%, P< .0001) and 3-year incidence of chronic GVHD (30% versus 53%, P< 0.0001) were significantly lower with haploidentical in comparison to MUD transplant. Similar results were obtained in RIC setting. A lower rate of GVHD with haploidentical transplant was attributed to the use of bone marrow as a graft source and the use of post-transplant cyclophosphamide. Among patients receiving myeloablative and RIC regimens, three-year DFS and OS were comparable in haploidentical and MUD transplant.

Transplant results of matched sibling donor (MSD) transplant and T-cell replete haploidentical transplant was also evaluated by *Wang et al* [64]. In this prospective, multicenter, nonrandomized trial, 450 patients with acute leukemia in CR1 underwent MSD (n=219) or haploidentical (n=231) transplant. The GVHD prophylaxis regimen in both groups consisted of cyclosporine, MMF, and mini-methotrexate. All patients in both cohorts achieved donor-cell engraftment. The median time to achieve neutrophil

engraftment was 2 days longer after MSD transplant. The 100-day cumulative incidence of grade II-IV acute GVHD after haploidentical and MSD transplant was 36% and 13% ($p=0.001$), respectively. The incidence of chronic GVHD was significantly higher after haploidentical transplant compared to MSD (42% versus 15%, $p < 0.001$). However, the rate of GVHD related death was similar in both groups. Among haploidentical and MSD recipients, the 3 year probability of DFS (74% versus 78%, $p= 0.34$) and OS (79% versus 82%, $P = 0.36$) were comparable. There was no difference in 3-year cumulative incidence of relapse between the two cohorts (15% versus 15%, $p =0.98$). Lower incidence of GVHD after MSD was attributed to combination of cyclosporine, methotrexate and MMF for GVHD prophylaxis. Prior studies also reported significantly lower rate of GVHD using this combination in recipients of MSD transplant [65, 66].

More recently *Ghosh et al.* performed a registry analysis comparing outcomes of 987 patients with lymphoma following reduced intensity haploidentical HSCT ($n=180$) with MSD HSCT ($n=807$) [67]. GVHD prophylaxis for the haploidentical group consisted of post-transplant cyclophosphamide with or without calcineurin inhibitor and MMF. GVHD prophylaxis for the MSD group contained calcineurin inhibitor based approaches. The cumulative incidence of grade II-IV acute GVHD was similar between the two cohorts (27% in haploidentical cohort versus 25% in MSD cohort, $p= 0.84$). Cumulative incidence of chronic GVHD was significantly lower with haploidentical HSCT (12% versus 45%, $p <0.001$). Chronic GVHD was the main cause of death in 5 patients in MSD group. Only one patient died of GVHD in haploidentical cohort. There was no significant difference in the three-year cumulative incidence of relapse (37% in haploidentical versus 40% in MSD, $p=0.51$), DFS (48% versus 48%, $p=0.96$) and OS (61% versus 62%, $p=0.82$). Therefore, based on this retrospective registry study in patients with lymphoma, RIC haploidentical HSCT using post-transplant cyclophosphamide provides comparable survival outcome to MSD HSCT with significantly lower risk of chronic GVHD.

Conclusion

HSCT is the only curative option for a large number of hematologic diseases. A minority of patients (30%) have a suitable HLA-identical sibling donor. For patients who lack MSD, MUD HSCT is frequently the preferred graft source. However, the presence of a suitably matched unrelated donor depends on factors such as the ethnicity of the patient, with a likelihood of finding an acceptably matched unrelated donor less than 20% in certain minorities compared to approximately 80% in Caucasians. A major disadvantage of MUD transplant is the prolonged time from patient referral to donor identification and collection of stem cells. Delay in the process of unrelated donor search due to logistical issues may increase the risk of disease progression or relapse [68]. Immediate availability of a haploidentical donor makes this approach an attractive treatment option for patients who lack an HLA-identical MSD or those for whom a MUD cannot be found in a timely manner. The field of haploidentical HSCT has matured significantly over the past two decades. In earlier studies of haploidentical HSCT, HLA-incompatibility barrier resulted in unacceptably high rate of GVHD and graft rejection leading to inferior overall survival. While effective T-cell depletion followed by infusion of mega doses of highly purified stem cells permitted high engraftment rates and reduced incidence of GVHD, higher risk of relapse and delay in immune reconstitution remained a significant obstacle. Newer methods of graft manipulation including adoptive T-cell immunotherapy and selective T-cell depletion have been shown to hasten immune recovery and reduce the risk of relapse. Despite the promising results, these approaches are costly and labor intensive, hence may not be globally available. In recent years, use of post-transplantation cyclophosphamide for GVHD prophylaxis after T-cell replete haploidentical HSCT has yielded encouraging results in adults. In several non-randomized studies, survival outcomes following haploidentical HSCT with post-transplant cyclophosphamide have been comparable to MSD or MUD transplant. Ultimately, a prospective randomized controlled trial such as [BMT](#) CTN 1101 is needed to determine the optimal approach to haploidentical transplant.

Table 1: Unmanipulated haploidentical HSCT versus matched related and matched unrelated HSCT

Disease		Conditioning regimen (N)	Graft type (N)	GVHD prophylaxis	Neutrophil engraftment	Grade II-IV acute GVHD	Chronic GVHD	Relapse rate	DFS	OS
Bashey et al [69] 2013 N= 271	Acute leukemia / CML/myeloma /lymphoma/MDS	RIC (102) MUD (101) MA(169)	MRD (117) MUD (101) Haplo (53)	CNI based CNI based CNI+MMF+PT-Cy	NR	6 months 27% 39% 30% (p= NS)	2 yrs 54% 54% 38% (p< 0.05)	2 yrs 34% 34% 33% (p= NS)	2 yrs 53% 52% 60% (p= NS)	2 yrs 76% 67% 64% (p= NS)
Di Stasi et al [70] 2014 N= 227	AML/MDS	RIC (227)	MRD (81) MUD (108) Haplo (32)	CNI+MTX CNI+MTX+ATG CNI + MMF + PT-Cy	30 days 99% 96% 97% (p= 0.44)	100 days 24% 19% 26% (p= 0.68)	3 yrs 46% 42% 24% (p= 0.52)	1 yr 28% 23% 33% (p= 0.75)	3 yrs 36% 27% 30% (p= 0.12)	NR
Luo et al [71] 2014 N= 305	Acute leukemia /lymphoma/ MDS	MA +ATG (305)	MRD (90) MUD (116) Haplo (99)	CNI+MMF+MTX CNI+MMF+MTX CNI+MMF+MTX	15 days 97% 97% 78% (p < 0.001)	3 months 15.6% 39% 42% (p< 0.0001)	2 yrs 24% 41% 41% (P= NS)	5 yrs 34% 21% 14% Haplo vs MRD p=0.008 Haplo vs MUD p= 0.17	5 yrs 63% 58% 58% (p= 0.57)	5 yrs 77% 63% 60.8% Haplo vs MRD p=0.026 Haplovs MUD p= 0.38
Ciurea et al [63] 2015 N= 2, 174	AML	RIC (825)	MUD (737) Haplo (88)	CNI + MMF or MTX CNI + MMF + PT-Cy	30 days 93% 96% (p=0.25)	3 months 19% 28% (P= 0.05)	3 yrs 34% 52% (p=0.002)	3 yrs 58% 42% (p= 0.006)	3 yrs 9% 23% (p=0.0001)	3 yrs 46% 44% (p= 0.71)
Wang et al[64] 2015 N= 450	AML in CR1	MA (ATG in haplo cohort)	MRD (219) Haplo (231)	CNI+MMF+MTX CNI+MMF+MTX	NE engraftment 2 days longer after MRD p=0.004	100 days 36% 13% (p< 0.001)	1yr 42% 15% (p< 0.001)	3 yrs 15% 15% (p= 0.98)	3 yrs 74% 78% (p= 0.34)	3 yrs 79% 82% (p=0.36)
Ghosh et al[67] 2016 N= 987	Lymphoma	RIC (987)	MRD (807) Haplo (180)	CNI based PT-Cy +/- CNI	28 days 95% 97% (p=0.31)	100 days 25% 27% (0.84)	1yr 45% 12% (P<0.001)	3 yrs 37% 40% (p=0.51)	3 yrs 48% 48% (p=0.98)	3 yrs 62% 61% (p=0.82)
Kanate et al[72] 2016 N= 917	Lymphoma	RIC (917)	MUD + ATG (241) MUD (491) Haplo (185)	CNI based CNI based PT-Cy based	28 days 97% 97% (p=0.32)	100 days 17% 12% (p= 0.44)	1yr 33% 51% (p< 0.001)	3 yrs 36% 28% 36% (p= 0.07)	3 yrs 38% 49% 47% (p=0.02)	3 yrs 50% 62% 60% (p= 0.2)

Abbreviations: AML, acute myeloid leukemia; ATG, anti-thymocyte globulin; CR, complete remission; CNI, calcineurin inhibitor; DFS, disease free survival; GVHD, graft versus host disease; Haplo, haploidentical; MMF, mycophenolate mofetil; MTX, methotrexate; RIC, reduced intensity conditioning; MA, myeloablative; MDS, myelodysplasia; MUD, matched unrelated donor; MRD, matched related donor; NE, neutrophil; NR, not reported; NS, not significant; OS, overall survival.

REFERENCES

1. Gragert L, Eapen M, Williams E, Freeman J, Spellman S, Baity R, Hartzman R, Rizzo JD, Horowitz M, Confer D, Maiers M. *HLA match likelihoods for hematopoietic stem-cell grafts in the U.S. registry*. N Engl J Med, 2014. **371**(4): p. 339-48 [PMID:25054717 DOI: 10.1056/NEJMsa1311707].
2. Beatty PG, Clift RA, Mickelson EM, Nisperos BB, Flounoy N, Martin PJ, Sanders JE, Stewart P, Buckner CD, Storb R, et al., *Marrow transplantation from related donors other than HLA-identical siblings*. N Engl J Med, 1985. **313**(13): p. 765-71 [PMID:3897863 DOI:10.1056/NEJM198509263131301]
3. Ash RC, Horowitz MM, Gale RP, van Bekkum DW, Casper JT, Gordon-Smith EC, Henslee PJ, Kolb HJ, Lowenberg B, Masaoka T. *Bone marrow transplantation from related donors other than HLA-identical siblings: effect of T cell depletion*. Bone Marrow Transplant, 1991. **7**(6): p. 443-52. [PMID:1873591]
4. Kasamon YL, Luznik L, Leffell MS, Kowalski J, Tsai HL, Bolaños-Meade J, Morris LE, Crilley PA, O'Donnell PV, Rossiter N, Huff CA, Brodsky RA, Matsui WH, Swinnen LJ, Borrello I, Powell JD, Ambinder RF, Jones RJ, Fuchs EJ. *Nonmyeloablative HLA-haploidentical bone marrow transplantation with high-dose posttransplantation cyclophosphamide: effect of HLA disparity on outcome*. Biol Blood Marrow Transplant, 2010. **16**(4): p. 482-9. [PMID:19925877 DOI:10.1016/j.bbmt.2009.11.011]
5. Wang Y, Chang YJ, Xu LP, Liu KY, Liu DH, Zhang XH, Chen H, Han W, Chen YH, Wang FR, Wang JZ, Chen Y, Yan CH, Huo MR, Li D, Huang XJ. *Who is the best donor for a related HLA haplotype-mismatched transplant?* Blood, 2014. **124**(6): p. 843-50. [PMID:24916508 DOI:10.1182/blood-2014-03-563130]
6. Stern M, Ruggeri L, Mancusi A, Bernardo ME, de Angelis C, Bucher C, Locatelli F, Aversa F, Velardi A. *Survival after T cell-depleted haploidentical stem cell transplantation is improved using the mother as donor*. Blood, 2008. **112**(7): p. 2990-5. [PMID:18492955 DOI:10.1182/blood-2008-01-135285]
7. Van Rood JJ, Eernisse JG, Van Leeuwen A. *Leucocyte antibodies in sera from pregnant women*. Nature, 1958. **181**(4625): p. 1735-6. [PMID:13566127]
8. Moretta A, Bottino C, Pende D, Tripodi G, Tambussi G, Viale O, Orengo A, Barbaresi M, Merli A, Ciccone E, Moretta L. *Identification of four subsets of human CD3-CD16+ natural killer (NK) cells by the expression of clonally distributed functional surface*

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PMID number was added to the references. Some of the older references (1980s and early 1990s) do not have DOI numbers.

All authors are listed.

molecules: correlation between subset assignment of NK clones and ability to mediate specific alloantigen recognition. *J Exp Med*, 1990. **172**(6): p. 1589-98. [PMID:2147946]

9. **Moretta A**, Bottino C, Mingari MC, Biassoni R, Moretta L. *What is a natural killer cell?* *Nat Immunol*, 2002. **3**(1): p. 6-8. [PMID:11753399 DOI:10.1038/ni0102-6]
10. **Vivier E**, Raulet DH, Moretta A, Caligiuri MA, Zitvogel L, Lanier LL, Yokoyama WM, Ugolini S. *Innate or adaptive immunity? The example of natural killer cells.* *Science*, 2011. **331**(6013): p. 44-9. [PMID:21212348 DOI:10.1126/science.1198687]
11. **Ruggeri L**, Capanni M, Urbani E, Perruccio K, Shlomchik WD, Tosti A, Posati S, Roggia D, Frassoni F, Aversa F, Martelli MF, Velardi A. *Effectiveness of donor natural killer cell alloreactivity in mismatched hematopoietic transplants.* *Science*, 2002. **295**(5562): p. 2097-100. [PMID:11896281 DOI:10.1126/science.1068440]
12. **Ruggeri L**, Mancusi A, Capanni M, Urbani E, Carotti A, Aloisi T, Stern M, Pende D, Perruccio K, Burchielli E, Topini F, Bianchi E, Aversa F, Martelli MF, Velardi A. *Donor natural killer cell allorecognition of missing self in haploidentical hematopoietic transplantation for acute myeloid leukemia: challenging its predictive value.* *Blood*, 2007. **110**(1): p. 433-40. [PMID:17371948 DOI:10.1182/blood-2006-07-038687]
13. **Aversa F**, Terenzi A, Tabilio A, Falzetti F, Carotti A, Ballanti S, Felicini R, Falcinelli F, Velardi A, Ruggeri L, Aloisi T, Saab JP, Santucci A, Perruccio K, Martelli MP, Mecucci C, Reisner Y, Martelli MF. *Full haplotype-mismatched hematopoietic stem-cell transplantation: a phase II study in patients with acute leukemia at high risk of relapse.* *J Clin Oncol*, 2005. **23**(15): p. 3447-54. [PMID:15753458 DOI:10.1200/JCO.2005.09.117]
14. **Pende D**, Marcenaro S, Falco M, Martini S, Bernardo ME, Montagna D, Romeo E, Cognet C, Martinetti M, Maccario R, Mingari MC, Vivier E, Moretta L, Locatelli F, Moretta A. *Anti-leukemia activity of alloreactive NK cells in KIR ligand-mismatched haploidentical HSCT for pediatric patients: evaluation of the functional role of activating KIR and redefinition of inhibitory KIR specificity.* *Blood*, 2009. **113**(13): p. 3119-29. [PMID:18945967 DOI:10.1182/blood-2008-06-164103]
15. **Pfeiffer MM**, Feuchtinger T, Teltschik HM, Schumm M, Müller I, Handgretinger R, Lang P. *Reconstitution of natural killer cell receptors influences natural killer activity and*

relapse rate after haploidentical transplantation of T- and B-cell depleted grafts in children.
Haematologica, 2010. 95(8): p. 1381-8.[PMID:20145268
DOI:10.3324/haematol.2009.021121]

16. **Symons HJ**, Leffell MS, Rossiter ND, Zahurak M, Jones RJ, Fuchs EJ. *Improved survival with inhibitory killer immunoglobulin receptor (KIR) gene mismatches and KIR haplotype B donors after nonmyeloablative, HLA-haploidentical bone marrow transplantation.* Biol Blood Marrow Transplant, 2010. 16(4): p. 533-42. [PMID:19961944
DOI:10.1016/j.bbmt.2009.11.022]
17. **Sun JY**, Dagis A, Gaidulis L, Miller MM, Rodriguez R, Parker P, Nademanee A, Falk P, Rosenthal J, Forman SJ, Senitzer D. *Detrimental effect of natural killer cell alloreactivity in T-replete hematopoietic cell transplantation (HCT) for leukemia patients.* Biol Blood Marrow Transplant, 2007. 13(2): p. 197-205. [PMID:17241925
DOI:10.1016/j.bbmt.2006.09.009]
18. **Cooley S**, McCullar V, Wangen R, Bergemann TL, Spellman S, Weisdorf DJ, Miller JS. *KIR reconstitution is altered by T cells in the graft and correlates with clinical outcomes after unrelated donor transplantation.* Blood, 2005. 106(13): p. 4370-6. [PMID:16131567 DOI:10.1182/blood-2005-04-1644]
19. **Reisner Y**, Kapoor N, Kirkpatrick D, Pollack MS, Cunningham-Rundles S, Dupont B, Hodes MZ, Good RA, O'Reilly RJ. *Transplantation for severe combined immunodeficiency with HLA-A,B,D,DR incompatible parental marrow cells fractionated by soybean agglutinin and sheep red blood cells.* Blood, 1983. 61(2): p. 341-8. [PMID:6217853]
20. **Kernan NA**, Flomenberg N, Dupont B, O'Reilly RJ. *Graft rejection in recipients of T-cell-depleted HLA-nonidentical marrow transplants for leukemia. Identification of host-derived antidonor allocytotoxic T lymphocytes.* Transplantation, 1987. 43(6): p. 842-7. [PMID:3296349]
21. **Martin P.J.** *The role of donor lymphoid cells in allogeneic marrow engraftment.* Bone Marrow Transplant, 1990. 6(5): p. 283-9.[PMID:2291990]

22. **Reisner Y**, Ben-Bassat I, Douer D, Kaploon A, Schwartz E, Ramot B. *Demonstration of clonable alloreactive host T cells in a primate model for bone marrow transplantation*. Proc Natl Acad Sci U S A, 1986. **83**(11): p. 4012-5. [PMID:3520563]
23. **Bachar-Lustig E**, Rachamim N, Li HW, Lan F, Reisner Y. *Megadose of T cell-depleted bone marrow overcomes MHC barriers in sublethally irradiated mice*. Nat Med, 1995. **1**(12): p. 1268-73. [PMID:7489407]
24. **Aversa F**, Tabilio A, Terenzi A, Velardi A, Falzetti F, Giannoni C, Iacucci R, Zei T, Martelli MP, Gambelunghe C, et al. *Successful engraftment of T-cell-depleted haploidentical "three-loci" incompatible transplants in leukemia patients by addition of recombinant human granulocyte colony-stimulating factor-mobilized peripheral blood progenitor cells to bone marrow inoculum*. Blood, 1994. **84**(11): p. 3948-55. [PMID:7524753]
25. **Aversa F**, Velardi A, Tabilio A, Reisner Y, Martelli MF. *Haploidentical stem cell transplantation in leukemia*. Blood Rev, 2001. **15**(3): p. 111-9. [PMID:11735159 DOI:10.1054/blre.2001.0157]
26. **Aversa F**, Tabilio A, Velardi A, Cunningham I, Terenzi A, Falzetti F, Ruggeri L, Barbabietola G, Aristei C, Latini P, Reisner Y, Martelli MF. *Treatment of high-risk acute leukemia with T-cell-depleted stem cells from related donors with one fully mismatched HLA haplotype*. N Engl J Med, 1998. **339**(17): p. 1186-93. [PMID:9780338 DOI:10.1056/NEJM199810223391702]
27. **Terenzi A**, Aristei C, Aversa F, Perruccio K, Chionne F, Raymond C, Latini P, Martelli MF. *Efficacy of fludarabine as an immunosuppressor for bone marrow transplantation conditioning: preliminary results*. Transplant Proc, 1996. **28**(6): p. 3101. [PMID:8962201]
28. **Aversa, F.**, *Setting the standard in T-cell-depleted haploidentical transplantation and beyond*. Best Pract Res Clin Haematol, 2011. **24**(3): p. 325-9. [PMID:21925085 DOI:10.1016/j.beha.2011.04.008]
29. **Ciceri F**, Labopin M, Aversa F, Rowe JM, Bunjes D, Lewalle P, Nagler A, Di Bartolomeo P, Lacerda JF, Lupo Stanghellini MT, Polge E, Frassoni F, Martelli MF, Rocha V. *A survey of fully haploidentical hematopoietic stem cell transplantation in adults with high-risk acute leukemia: a risk factor analysis of outcomes for patients in remission at*

transplantation. Blood, 2008. **112**(9): p. 3574-81. [PMID:18606875 DOI:10.1182/blood-2008-02-140095]

30. **André-Schmutz I**, Le Deist F, Hacein-Bey-Abina S, Vitetta E, Schindler J, Chedeville G, Vilmer E, Fischer A, Cavazzana-Calvo M. *Immune reconstitution without graft-versus-host disease after haemopoietic stem-cell transplantation: a phase 1/2 study.* Lancet, 2002. **360**(9327): p. 130-7. [PMID:12126823 DOI:10.1016/S0140-6736(02)09413-8]
31. **Amrolia PJ**, Muccioli-Casadei G, Huls H, Adams S, Durett A, Gee A, Yvon E, Weiss H, Cobbold M, Gaspar HB, Rooney C, Kuehnle I, Ghetie V, Schindler J, Krance R, Heslop HE, Veys P, Vitetta E, Brenner MK. *Adoptive immunotherapy with allogeneic donor T-cells improves immune reconstitution after haploidentical stem cell transplantation.* Blood, 2006. **108**(6): p. 1797-808. [PMID:16741253 DOI:10.1182/blood-2006-02-001909]
32. **Perruccio K**, Topini F, Tosti A, Carotti A, Burchielli E, Ruggeri L, Mancusi A, Urbani E, Aversa F, Martelli MF, Velardi A. *Optimizing a photoallodepletion protocol for adoptive immunotherapy after haploidentical SCT.* Bone Marrow Transplant, 2012. **47**(9): p. 1196-200. [PMID:22139067 DOI:10.1038/bmt.2011.237]
33. **Kunzmann V**. and Wilhelm M. *Anti-lymphoma effect of gammadelta T cells.* Leuk Lymphoma, 2005. **46**(5): p. 671-80. [PMID:16019504 DOI:10.1080/10428190500051893]
34. **Minculescu L**. and Sengelov H. *The role of gamma delta T cells in haematopoietic stem cell transplantation.* Scand J Immunol, 2015. **81**(6): p. 459-68. [PMID:25753378 DOI:10.1111/sji.12289]
35. **Lang P**, Feuchtinger T, Teltschik HM, Schwinger W, Schlegel P, Pfeiffer M, Schumm M, Lang AM, Lang B, Schwarze CP, Ebinger M, Urban C, Handgretinger R. *Improved immune recovery after transplantation of TCRalphabeta/CD19-depleted allografts from haploidentical donors in pediatric patients.* Bone Marrow Transplant, 2015. **50 Suppl 2:** p. S6-10. [PMID:26039210 DOI:10.1038/bmt.2015.87]
36. **Bleakley M**, Heimfeld S, Jones LA, Turtle C, Krause D, Riddell SR, Shlomchik W. *Engineering human peripheral blood stem cell grafts that are depleted of naive T cells and retain functional pathogen-specific memory T cells.* Biol Blood Marrow Transplant, 2014. **20**(5): p. 705-16. [PMID:24525279 DOI:10.1016/j.bbmt.2014.01.032]

37. **Teschner D**, Distler E, Wehler D, Frey M, Marandiuc D, Langeveld K, Theobald M, Thomas S, Herr W. *Depletion of naive T cells using clinical grade magnetic CD45RA beads: a new approach for GVHD prophylaxis.* Bone Marrow Transplant, 2014. **49**(1): p. 138-44. [PMID:23933765 DOI:10.1038/bmt.2013.114]
38. **Triplett BM**, Shook DR, Eldridge P, Li Y, Kang G, Dallas M, Hartford C, Srinivasan A, Chan WK, Suwannasaen D, Inaba H, Merchant TE, Pui CH, Leung W. *Rapid memory T-cell reconstitution recapitulating CD45RA-depleted haploidentical transplant graft content in patients with hematologic malignancies.* Bone Marrow Transplant, 2015. **50**(7): p. 1012. [PMID:25665048 DOI:10.1038/bmt.2014.324]
39. **Clinicaltrials.gov**. *Selective Depletion of CD45RA+ Naive T Cells From Allogeneic Peripheral Blood Stem Cell Grafts From HLA-Matched Related and Unrelated Donors for Prevention of GVHD.* Available at:www.clinicaltrials.gov (NCT02220985), 2016.
40. **Hoffmann P**, Ermann J, Edinger M, Fathman CG, Strober S. *Donor-type CD4(+)CD25(+) regulatory T cells suppress lethal acute graft-versus-host disease after allogeneic bone marrow transplantation.* J Exp Med, 2002. **196**(3): p. 389-99. [PMID:12163567]
41. **Nguyen VH**, Shashidhar S, Chang DS, Ho L, Kambham N, Bachmann M, Brown JM, Negrin RS. *The impact of regulatory T cells on T-cell immunity following hematopoietic cell transplantation.* Blood, 2008. **111**(2): p. 945-53. [PMID:17916743 DOI:10.1182/blood-2007-07-103895]
42. **Di Ianni M**, Falzetti F, Carotti A, Terenzi A, Castellino F, Bonifacio E, Del Papa B, Zei T, Ostini RI, Cecchini D, Aloisi T, Perruccio K, Ruggeri L, Balucani C, Pierini A, Sportoletti P, Aristei C, Falini B, Reisner Y, Velardi A, Aversa F, Martelli MF. *Tregs prevent GVHD and promote immune reconstitution in HLA-haploidentical transplantation.* Blood, 2011. **117**(14): p. 3921-8. [PMID:21292771 DOI:10.1182/blood-2010-10-311894]
43. **Martelli MF**, Di Ianni M, Ruggeri L, Falzetti F, Carotti A, Terenzi A, Pierini A, Massei MS, Amico L, Urbani E, Del Papa B, Zei T, Iacucci Ostini R, Cecchini D, Tognellini R, Reisner Y, Aversa F, Falini B, Velardi A. *HLA-haploidentical transplantation with regulatory and conventional T-cell adoptive immunotherapy prevents acute leukemia*

relapse. Blood, 2014. **124**(4): p. 638-44. [PMID:24923299 DOI:10.1182/blood-2014-03-564401]

44. **Huang XJ**, Han W, Xu LP, Chen YH, Liu DH, Lu J, Chen H, Zhang YC, Jiang Q, Liu KY, Lu DP. *A novel approach to human leukocyte antigen-mismatched transplantation in patients with malignant hematological disease.* Chin Med J (Engl), 2004. **117**(12): p. 1778-85. [PMID:15603704]
45. **Huang XJ**, Liu DH, Liu KY, Xu LP, Chen H, Han W, Chen YH, Zhang XH, Lu DP. *Treatment of acute leukemia with unmanipulated HLA-mismatched/haploidentical blood and bone marrow transplantation.* Biol Blood Marrow Transplant, 2009. **15**(2): p. 257-65. [PMID:19167686 DOI:10.1016/j.bbmt.2008.11.025]
46. **Mayumi H**, Himeno K, Shin T, Nomoto K. *Drug-induced tolerance to allografts in mice. VI. Tolerance induction in H-2-haplotype-identical strain combinations in mice.* Transplantation, 1985. **40**(2): p. 188-94. [PMID:3895627]
47. **Berenbaum MC**. and Brown IN. *Prolongation of Homograft Survival in Mice with Single Doses of Cyclophosphamide.* Nature, 1963. **200**: p. 84. [PMID:14074645]
48. **Berenbaum MC**. and Brown IN. *Dose-Response Relationships for Agents Inhibiting the Immune Response.* Immunology, 1964. **7**: p. 65-71. [PMID:14113077]
49. **Strauss G**, Osen W, Debatin KM. *Induction of apoptosis and modulation of activation and effector function in T cells by immunosuppressive drugs.* Clin Exp Immunol, 2002. **128**(2): p. 255-66. [PMID:11985515]
50. **Kanakry CG**, Ganguly S, Zahurak M, Bolaños-Meade J, Thoburn C, Perkins B, Fuchs EJ, Jones RJ, Hess AD, Luznik L. *Aldehyde dehydrogenase expression drives human regulatory T cell resistance to posttransplantation cyclophosphamide.* Sci Transl Med, 2013. **5**(211): p. 211ra157. [PMID:24225944 DOI:10.1126/scitranslmed.3006960]
51. **O'Donnell PV**, Luznik L, Jones RJ, Vogelsang GB, Leffell MS, Phelps M, Rhubarb P, Cowan K, Piantadosi S, Fuchs EJ. *Nonmyeloablative bone marrow transplantation from partially HLA-mismatched related donors using posttransplantation cyclophosphamide.* Biol Blood Marrow Transplant, 2002. **8**(7): p. 377-86. [PMID:12171484]

52. **Luznik L**, O'Donnell PV, Symons HJ, Chen AR, Leffell MS, Zahurak M, Gooley TA, Piantadosi S, Kaup M, Ambinder RF, Huff CA, Matsui W, Bolaños-Meade J, Borrello I, Powell JD, Harrington E, Warnock S, Flowers M, Brodsky RA, Sandmaier BM, Storb RF, Jones RJ, Fuchs EJ. *HLA-haploidentical bone marrow transplantation for hematologic malignancies using nonmyeloablative conditioning and high-dose, posttransplantation cyclophosphamide*. Biol Blood Marrow Transplant, 2008. **14**(6): p. 641-50. [PMID:18489989 DOI:10.1016/j.bbmt.2008.03.005]
53. **Munchel A**, Kesserwan C, Symons HJ, Luznik L, Kasamon YL, Jones RJ, Fuchs EJ. *Nonmyeloablative, HLA-haploidentical bone marrow transplantation with high dose, post-transplantation cyclophosphamide*. Pediatr Rep, 2011. **3 Suppl 2**: p. e15. [PMID:22053277 DOI:10.4081/pr.2011.s2.e15]
54. **Brunstein CG**, Fuchs EJ, Carter SL, Karanes C, Costa LJ, Wu J, Devine SM, Wingard JR, Aljitawi OS, Cutler CS, Jagasia MH, Ballen KK, Eapen M, O'Donnell PV. *Alternative donor transplantation after reduced intensity conditioning: results of parallel phase 2 trials using partially HLA-mismatched related bone marrow or unrelated double umbilical cord blood grafts*. Blood, 2011. **118**(2): p. 282-8. [PMID:21527516 DOI:10.1182/blood-2011-03-344853]
55. **Clinicaltrials.gov**. Double Cord Versus Haploidentical www.clinicaltrials.gov (NCT01597778), 2016.
56. **Solomon SR**, Sizemore CA, Sanacore M, Zhang X, Brown S, Holland HK, Morris LE, Bashey A. *Haploidentical transplantation using T cell replete peripheral blood stem cells and myeloablative conditioning in patients with high-risk hematologic malignancies who lack conventional donors is well tolerated and produces excellent relapse-free survival: results of a prospective phase II trial*. Biol Blood Marrow Transplant, 2012. **18**(12): p. 1859-66. [PMID:22863841 DOI:10.1016/j.bbmt.2012.06.019]
57. **Silva Lde P**, Patah PA, Saliba RM, Szewczyk NA, Gilman L, Neumann J, Han XY, Tarrand J, Ribeiro R, Gulbis A, Shpall EJ, Jones R, Popat U, Walker JA, Petropoulos D, Chiattone A, Stewart J, El-Zimaity M, Anderlini P, Giralt S, Champlin RE, de Lima M. *Hemorrhagic cystitis after allogeneic hematopoietic stem cell transplants is the complex result of*

BK virus infection, preparative regimen intensity and donor type. Haematologica, 2010. **95**(7): p. 1183-90. [PMID:20410183 DOI:10.3324/haematol.2009.016758]

58. **Solomon SR**, Sizemore CA, Sanacore M, Zhang X, Brown S, Holland HK, Morris LE, Bashey A. *Total Body Irradiation-Based Myeloablative Haploidentical Stem Cell Transplantation Is a Safe and Effective Alternative to Unrelated Donor Transplantation in Patients Without Matched Sibling Donors.* Biol Blood Marrow Transplant, 2015. **21**(7): p. 1299-307. [PMID:25797174 DOI:10.1016/j.bbmt.2015.03.003]
59. **Raiola AM**, Dominietto A, Ghiso A, Di Grazia C, Lamparelli T, Gualandi F, Bregante S, Van Lint MT, Geroldi S, Luchetti S, Ballerini F, Miglino M, Varaldo R, Bacigalupo A. *Unmanipulated haploidentical bone marrow transplantation and posttransplantation cyclophosphamide for hematologic malignancies after myeloablative conditioning.* Biol Blood Marrow Transplant, 2013. **19**(1): p. 117-22. [PMID:22940057 DOI:10.1016/j.bbmt.2012.08.014]
60. **Grosso D**, Carabasi M, Filicko-O'Hara J, Kasner M, Wagner JL, Colombe B, Cornett Farley P, O'Hara W, Flomenberg P, Werner-Wasik M, Brunner J, Mookerjee B, Hyslop T, Weiss M, Flomenberg N. *A 2-step approach to myeloablative haploidentical stem cell transplantation: a phase 1/2 trial performed with optimized T-cell dosing.* Blood, 2011. **118**(17): p. 4732-9. [PMID:21868572 DOI:10.1182/blood-2011-07-365338]
61. **Symons HJ**, Chen A, Gamper C, Cooke Kr, Showel M, Bolanos-Meade J, Luznik L, Jones RJ, Fuchs EJ. *Biology of Blood and Marrow Transplant.* **21**(2): p. S29, 2015. [DOI: <http://dx.doi.org/10.1016/j.bbmt.2014.11.019>]
62. **Ringdén O**, Labopin M, Ciceri F, Velardi A, Bacigalupo A, Arcese W, Ghavamzadeh A, Hamladji RM, Schmid C, Nagler A, Mohty M. *Is there a stronger graft-versus-leukemia effect using HLA-haploidentical donors compared with HLA-identical siblings?* Leukemia, 2016. **30**(2): p. 447-55. [PMID:26293645 DOI:10.1038/leu.2015.232]
63. **Ciurea SO**, Zhang MJ, Bacigalupo AA, Bashey A, Appelbaum FR, Aljitawi OS, Armand P, Antin JH, Chen J, Devine SM, Fowler DH, Luznik L, Nakamura R, O'Donnell PV, Perales MA, Pingali SR¹, Porter DL, Riches MR, Ringdén OT, Rocha V, Vij R, Weisdorf DJ, Champlin RE, Horowitz MM, Fuchs EJ, Eapen M. *Haploidentical transplant with posttransplant cyclophosphamide vs matched unrelated donor transplant for*

acute myeloid leukemia. Blood, 2015. **126**(8): p. 1033-40. [PMID:26130705 DOI:10.1182/blood-2015-04-639831]

64. **Wang Y**, Liu QF, Xu LP, Liu KY, Zhang XH, Ma X, Fan ZP, Wu DP, Huang XJ. *Haploidentical vs identical-sibling transplant for AML in remission: a multicenter, prospective study*. Blood, 2015. **125**(25): p. 3956-62. [PMID:25940714 DOI:10.1182/blood-2015-02-627786]
65. **Lai YR**, Chen YH, Hu DM, Jiang M, Liu QF, Liu L, Hou J, Schwarzenberger P, Li QC, Zhang ZM, Liu KY, Huang XJ. *Multicenter phase II study of a combination of cyclosporine a, methotrexate and mycophenolate mofetil for GVHD prophylaxis: results of the Chinese Bone Marrow Transplant Cooperative Group (CBMTG)*. J Hematol Oncol, 2014. **7**: p. 59. [PMID: 25139202 DOI:10.1186/s13045-014-0059-3]
66. **Lai Y**, Ma J, Schwarzenberger P, Li W, Cai Z, Zhou J, Peng Z, Yang J, Luo L, Luo J, Deng D, Li Q, Zhou Y, Liang J. *Combination of CsA, MTX and low-dose, short-course mycophenolate mofetil for GVHD prophylaxis*. Bone Marrow Transplant, 2009. **43**(1): p. 61-7. [PMID:18724395 DOI:10.1038/bmt.2008.265]
67. **Ghosh N**, Karmali R, Rocha V, Ahn KW, DiGilio A, Hari PN, Bachanova V, Bacher U, Dahi P, de Lima M, D'Souza A, Fenske TS, Ganguly S, Kharfan-Dabaja MA, Prestidge TD, Savani BN, Smith SM, Sureda AM, Waller EK, Jaglowski S, Herrera AF, Armand P, Salit RB, Wagner-Johnston ND, Fuchs E, Bolaños-Meade J, Hamadani M. *Reduced-Intensity Transplantation for Lymphomas Using Haploidentical Related Donors Versus HLA-Matched Sibling Donors: A Center for International Blood and Marrow Transplant Research Analysis*. J Clin Oncol, 2016. 10.1200/jco.2015.66.3476. [PMID:27269951 DOI:10.1200/JCO.2015.66.3476]
68. **Davies SM**, Ramsay NK, Weisdorf DJ. *Feasibility and timing of unrelated donor identification for patients with ALL*. Bone Marrow Transplant, 1996. **17**(5): p. 737-40. [PMID:8733690]
69. **Bashey A**, Zhang X, Sizemore CA, Manion K, Brown S, Holland HK, Morris LE, Solomon SR. *T-cell-replete HLA-haploidentical hematopoietic transplantation for hematologic malignancies using post-transplantation cyclophosphamide results in outcomes equivalent to*

those of contemporaneous HLA-matched related and unrelated donor transplantation. J Clin Oncol, 2013. **31**(10): p. 1310-6. [PMID:23423745 DOI:10.1200/JCO.2012.44.3523]

70. **Di Stasi A**, Milton DR, Poon LM, Hamdi A, Rondon G, Chen J, Pingali SR, Konopleva M, Kongtim P, Alousi A, Qazilbash MH, Ahmed S, Bashir Q, Al-atrash G, Oran B, Hosing CM, Kebriaei P, Popat U¹, Shpall EJ, Lee DA, de Lima M, Rezvani K, Khouri IF, Champlin RE, Ciurea SO. *Similar transplantation outcomes for acute myeloid leukemia and myelodysplastic syndrome patients with haploidentical versus 10/10 human leukocyte antigen-matched unrelated and related donors.* Biol Blood Marrow Transplant, 2014. **20**(12): p. 1975-81. [PMID:25263628 DOI:10.1016/j.bbmt.2014.08.013]
71. **Luo Y**, Xiao H, Lai X, Shi J, Tan Y, He J, Xie W, Zheng W, Zhu Y, Ye X, Yu X, Cai Z, Lin M, Huang H. *T-cell-replete haploidentical HSCT with low-dose anti-T-lymphocyte globulin compared with matched sibling HSCT and unrelated HSCT.* Blood, 2014. **124**(17): p. 2735-43. [PMID:25214441 DOI:10.1182/blood-2014-04-571570]
72. **Kanate AS**, Mussetti A, Kharfan-Dabaja MA, Ahn KW, DiGilio A, Beitinjaneh A, Chhabra S, Fenske TS, Freytes C, Gale RP, Ganguly S, Hertzberg M, Klyuchnikov E, Lazarus HM, Olsson R, Perales MA, Rezvani A, Riches M, Saad A, Slavin S, Smith SM, Sureda A, Yared J, Ciurea S, Armand P, Salit R, Bolaños-Meade J, Hamadani M. *Reduced-intensity transplantation for lymphomas using haploidentical related donors vs HLA-matched unrelated donors.* Blood, 2016. **127**(7): p. 938-47. [PMID:26670632 DOI:10.1182/blood-2015-09-671834]